## <u>Original Article</u> The Prevalence of Antibodies against Sandfly Fever Viruses and West Nile Virus in Cyprus

### Gaetan Billioud; Christina Tryfonos; \*Jan Richter

Department of Molecular Virology, Cyprus Institute of Neurology and Genetics, Nicosia, Cyprus

#### (Received 14 Oct 2017; accepted 2 Jan 2019)

#### Abstract

**Background:** Sandfly fever is an incapacitating disease caused by sandfly-borne Phleboviruses that can lead to meningitis, encephalitis or meningoencephalitis. West Nile virus (WNV), a mosquito-borne Flavivirus, can induce neuroinvasive disease manifested by meningitis, encephalitis or acute flaccid paralysis. Both vectors are endemic in Cyprus and very active during summer. The aims of this study were to determine first the prevalence of sandfly fever viruses (SFV) and WNV infections in Cyprus and second, to investigate their role in central nervous system (CNS) infections.

**Methods:** For the prevalence study, 327 sera collected in 2013 and 2014 were tested for anti-SFV and anti-WNV IgG using indirect immunofluorescence assay and ELISA, respectively. In order to investigate a possible role of SFV and WNV in CNS infections, 127 sera of patients presenting symptoms of SFV or WNV infections were screened for IgM specific to SFV and WNV.

**Results**: The overall anti-SFV IgG seroprevalence was 28% and was increasing with age (P < 0.01). The seroprevalence rate for anti-WNV IgG in Cyprus was 5%. Concerning the role of SFVs in CNS infections, anti-SFV IgM was detected in 8 out of 127 sera from selected patients presenting relevant symptoms of infections during vector's active period. In addition, anti-WNV IgM were detected in 17 out of the 127 patients with compatible symptoms.

**Conclusion:** The findings confirm the presence of sandfly fever and WNV in Cyprus and should, therefore, be considered in the differential diagnosis of patients with febrile illness/meningitis.

Keywords: Phleboviruses; West-Nile virus; Cyprus; Seroprevalence

## Introduction

Sandfly (Diptera: Psychodidae)-transmitted phleboviruses may cause a transient and moderate febrile illness (named pappatasi fever, 3-day fever or sandfly fever) but can also affect the central nervous system (CNS) leading to serious infections. The most common vectors are *Phlebotomus papatasi*, *P. perfiliewi or P. perniciosus*. The genus *Phlebovirus* has been recently assigned to the newly created family *Phenuiviridae* and the order *Bunyavirales* (1).

Sandfly-borne phleboviruses are enveloped negative-sense and single-stranded tripartite RNA viruses classified around two viral species (Sandfly fever Naples (SFNV) and Salehabad) and two tentative species (Sicilian and Corfu). Infections with SFNV and Sandfly fever Sicilian (SFSV) viruses are clinically similar and characterized by high fever, headache, retro-orbital pain, malaise, diarrhoea, myalgia, photophobia and anorexia along with thrombocytopenia, leukopenia and elevated liver enzymes (2, 3). Toscana virus, a member of the Naples serogroup, generally induces a mild febrile illness without CNS involvement.

However, when the CNS is affected, high fever, headache, nausea, vomiting, Kernig signs, neck rigidity, myalgia and sometimes unconsciousness, tremors, paresis and nystagmus as well as encephalitis, severe meningoencephalitis and long-lasting sequelae have been reported (4, 5). The Mediterranean basin is endemic for sandfly-borne phleboviruses infections (4, 6-10) and particularly Cyprus where phlebotomines (11, 12) and outbreaks have been reported (13-16).

Regarding West Nile fever, the majority of clinical cases are mild and present with flu-like symptoms characterized by fever, malaise, and myalgia frequently accompanied by rash. Severe cases with signs of encephalitis, meningoencephalitis or meningitis are rare (<1%), most often observed among elderly and associated with a 10% fatality rate (17). The disease results from an infection by West Nile Virus (WNV) (genus Flavivirus, family Flaviviridae, order unassigned) transmitted through a mosquito (Diptera: Culicidae) bite, mostly from the genus Culex and usually from the Culex pipiens complex (18), abundant in Cyprus (19, 20). Human infection is solely incidental of the enzootic transmission cycle between avian natural hosts and mosquito vectors (21). The virus is considered to be one of the most important emerging arboviruses in recent years because of the multiple outbreaks reported in Europe and particularly in the Mediterranean area (Greece, Turkey, Spain) (22, 23). To date, no epidemiological data for WNV have been reported for Cyprus but the first neuroinvasive case was reported in 2016 (24, 25) and a second confirmed case in 2018 (ECDC update), confirming the presence of the WNV in the island.

Because of outdated or absent epidemiological data concerning Cyprus, a cross-sectional serologic survey was conducted to estimate SFV and WNV IgG respective seroprevalences in order to assess the exposure of the Cypriot population to these pathogens. In addition, the possible role of these pathogens in CNS infections in Cyprus was investigated by testing for markers of recent infection (IgM) and viral RNA in samples of patients with febrile illness and/or CNS infection.

## **Materials and Methods**

### Sample collection

Serum and cerebrospinal fluid (CSF) samples from patients were received at the Cyprus Institute of Neurology and Genetics, stored appropriately at -20 °C and used subsequently. For IgG seroprevalence, 327 samples between Jan 2013 and Dec 2014 were retrospectively selected following a leftover sampling methodology matching the 2013 census Cypriot demography according to patient's sexes (male or female), age and origin of the sample (Famagusta, Larnaca, Limassol, Nicosia or Paphos district) (Fig. 1). Informed consent was taken from the patients before participation.

The patients with known disease, medical procedure or treatments affected the antibodies detection were excluded (i.e. autoimmune disease, immunomodulatory treatments, transplanted, neoplasm, etc.). Determination of specific IgM was performed on samples received during 2013 and 2014 vector active season (warm months, from May to Nov in Cyprus) from 127 selected patients with infectious pathologic features relating to Sandfly or West Nile fevers. Based on similar selection criteria, 72 CSF samples from patients collected between 2015 and 2016 during vector active season were tested for circulating viral RNA.

### SFV IgG and IgM detection

Overall, 327 and 127 sera were analyzed for the presence of anti-SFV IgG and IgM, respectively, by indirect immunofluorescence test (IIFT) BIOCHIP Mosaic<sup>TM</sup>: Sandfly Fever Virus IgG or IgM (EUROIMMUN, Lübeck, Germany). This test is the only commercially available test that offers the possibility to assess the presence of antibody reacting against two species members of the Sandfly Sicilian serogroup (SFSV and Sandfly fever Cyprus viruses, SFCV) and two members of the Sandfly Naples serogroup (SFNV and Toscana virus, TOSV), without excluding the possibilities of cross-reactivity.

Sera were diluted 100 and 50 times in sample buffer for IgG and IgM, respectively, and tested according to the manufacturer's instructions. Before determining specific anti-SFV IgM, IgG antibodies and rheumatoid factors of class M were removed by immune-absorption using the Siemens RF-Absorbent (Marburg, Germany) [Ref: OUCG194] for 15min at room temperature. Samples were considered positive if in some cells, but not all, cytoplasmic finely granular structures and inclusion bodies fluoresced with the same pattern obtained as for the positive control serum.

### WNV IgG and IgM detection

Overall, 327 sera were collected for specific anti-WNV IgG detection and 127 sera for IgM detection, respectively, using an FDA-approved enzyme-linked immunosorbent assay (ELISA) (EUROIMMUN, Lübeck, Germany). Sera were diluted 100 times for both IgG and IgM in sample buffer (containing IgG/RF-Absorbent for IgM detection) and tested according to the manufacturer's instructions. The samples were considered positive when the ratio between the extinction values of the samples over the calibrator (20 RU/ml for IgG) was  $\geq 1.1$ .

# Viral RNA extraction and RT–PCR amplification

Viral RNA was extracted from 72 CSF samples using 400µl of clinical specimen by the iPrep<sup>TM</sup> PureLink® Virus kit according to manufacturer's instructions and resuspended in 100µl DEPC water. Ten microliters of the extracted viral RNA was used as a template using specific primers with the SuperScript® III Platinium One-Step qRT-PCR kit (Invitrogen, USA), according to the manufacturer's instructions. For Phlebovirus detection and quantification, the specific primers used amplified the 3' end of the small segment N gene and were based on published protocols (26, 27). For *Flavivirus* detection and quantification, the specific primers targeted the NS5 gene and were based on a previously published protocol (28).

### Statistics

Results were statistically analyzed with the Prism<sup>TM</sup> version 5.01 software from GraphPad (La Jolla, CA, USA). Along with descriptive statistics, univariate analysis was conducted on the results obtained from the seroprevalence study by Fisher's exact tests. Comparison of groups' average age was analyzed by nonparametric two-tailed Mann-Whitney tests. The logistic regression model was generated from the GNU package R environment. P< 0.05 indicated statistical significance.

## Results

### Prevalence study of anti-sandfly fever viruses and West Nile virus IgG antibodies

To determine the exposure of Cypriots to sandfly fever viruses, 327 human serum samples from 2013–2014 were screened for anti-SFVs and anti-WNV IgG. The sampling was distributed by sex and age proportional to Cyprus' population (Table 1). By sex, 166 were females (~51%) and 161 males (~49%). By age groups, 81 were <20yr old (~25%), 184 aged 20–60yr (~56%), and 62 were aged >60yr (~19%) with an overall mean age of 37yr old. By districts, 9 specimens were from Famagusta (~3%), 32 from Larnaca (~10%), 107 from Limassol (~33%), 170 from Nicosia (~52%) and 9 were from Paphos District (~3%) (Fig. 1).

The overall seroprevalence of anti-SFVs IgG was 28% (CI95%: 23.5–33.4%) and increased with age (Table 1). Difference of mean age in anti-SFV seropositive group (47yr old) compared to the seronegative patient's group (33yr old) was statistically significant (P< 0.0001). Logistic regression model confirmed the trend (P< 0.001) and computed an increased risk of ~3% per year (SE 0.6%). Differences observed in seroprevalence for anti-SFV IgG between females (~25%) and males (~31%)

or between districts (Famagusta: ~0%, Larnaca: ~19%, Limassol: ~31%, Nicosia: ~30% and Paphos: ~33%) were not statistically significant (P> 0.05) (Table 1). Difference in prevalence between sera reacting only against SFSV and/or SFCV (~12%) and sera reacting only against SFNV and/or TOSV (~11%) was not statistically significant (P> 0.05) (data not shown).

The same 327 human sera from 2013–2014 were also screened for anti-WNVs IgG. The overall seroprevalence of anti-WNV IgG was ~5% (CI95%: 2.5–7.2%) (Table 1). The difference of mean age in anti-WNV seropositive group (33yr old) compared to the seronegative patients group (37yr old) was not statistically significant. Differences in frequencies observed between each age group, sex and district were also not statistically significant.

Five patients (~1.5%) were found to have IgG antibodies reacting against both SFVs and WNV (data not shown), giving seroprevalence rates of ~31% (5/16) and ~5% (5/93) of anti-SFVs and anti-WNV IgG in anti-WNV and anti-SFVs IgG positive groups, respectively. The differences of rates in positive groups compared to the general population (~28% and ~5% for anti-SFV and anti-WNV IgG, respectively) were not statistically significant.

# Role of SFV and WNV infections in CNS infections in Cyprus

To assess the morbidity of SFV infections in Cyprus, human sera from patients displaying infectious pathologic features in 2013 and 2014 during sandflies' active period were tested for markers of recent SFV infection, IgM. Eight sera (~6%) were found positive for anti-SFV IgM out of the 127 selected patients (Table 2). Difference of SFV IgM seroprevalence rates between each age group, sex and district were not statistically significant. However, a higher frequency of samples reacting only against SFV serotypes belonging to the Naples serogroup (SFNV and TOSV, 7/127) than to the Sicilian serogroup (SFSV and SFCV, 0/ 127) (data not shown) was statistically significant (P< 0.05). Three of the 8 anti-SFV IgM positive patients (~38%) showed neuroinvasive features (meningitis or meningoencephalitis) but no statistical significance was found compared to the anti-SFV IgM negative group (34/127, ~26%, P> 0.05) (data not shown). Of the 8 seropositive patients for anti-SFV IgM, four patient's CSF were available and three of them were also found positive for anti-SFV IgM (data not shown). All CSF positive for SFV IgM reacted only against the Naples serocomplex, further confirming their neuroinvasion capabilities.

Detection of *Phlebovirus* nucleic acids by specific quantitative RT-PCR in 72 human cerebrospinal fluid samples from patients displaying infectious pathologic features in 2015 and 2016 during sandflies' active period was performed, but no specimen was found positive (data not shown).

Seventeen (~13%) out of the 127 tested human sera from patients displaying infectious pathologic features in 2013 and 2014 during mosquito's active period were found positive for anti-WNV IgM (Table 2). The difference of WNV IgM seroprevalence rates between sexes, age groups or districts were not found statistically significant. Two of the 17 anti-WNV IgM positive patients (~12%) showed neuroinvasive features (meningitis or meningoencephalitis) but no statistical significance was found compared to the anti-WNV IgM negative group (29/110, ~26%) (data not shown).

With regard to nucleic acid testing, no WNV RNA was detected in the 72 human CSF samples from selected patients displaying infectious pathologic features in 2015 and 2016 during mosquitos' active period (data not shown).

		SFV IgG		WNV IgG	
	Tested	Positive	%	Positive	%
Total	327	93	~28	16	~5
Males	161	52	~32	6	~4
Females	166	41	~25	10	~6
< 20 y	81	6 (*)	~7	4	~5
20–60 y	184	60 (*)	~33	11	~6
> 60c y	62	27 (*)	~44	1	~2
Famagusta	9	0	0	0	0
Larnaca	32	6	~19	1	~3
Limassol	107	33	~31	5	~5
Nicosia	170	51	~30	10	~6
Paphos	9	3	~33	0	0

Table 1. Sandfly fever virus and West Nile IgG seropositivity

SFV, sandfly fever viruse	; WNV; (*), Statistically	significant ( $P < 0.05$ )
---------------------------	---------------------------	----------------------------

		SFV IgM		WNV IgM	
	Tested	Positive	%	Positive	%
Total	127	8	~6	17	~13
Males	58	6	~10	5	~9
Females	69	2	~3	12	~17
< 20 y	52	2	~4	8	~15
20–60 y	54	4	~7	7	~13
> 60c y	21	2	~10	2	~10
Famagusta	5	0	0	2	~40
Larnaca	10	0	0	0	0
Limassol	41	6	~15	9	~22
Nicosia	66	2	~3	6	~9
Paphos	5	0	0	0	0

Table 2. Sandfly fever virus and West Nile IgM seropositivity

SFV, sandfly fever viruses; WNV

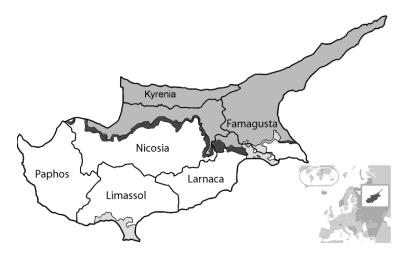


Fig. 1. Illustrative map of Cyprus' districts. White, Area studied; Grey, Non-government controlled area; Dark grey, UN buffer zone; Light grey, UK sovereign bases

## Discussion

Because the presence of arboviruses in Cyprus had been reported previously (15, 24, 29-32), we evaluated the residents' exposure to SFV and WNV following Cyprus' demography in terms of age, sex and location.

The overall seroprevalence rate for anti-SFVs IgG estimated in this study was ~28%. Higher seroprevalence of antibodies against SFSV virus (~62%) and neutralizing antibodies against SFNV (~57%), SFSV (~32%) and TOSV (~20%) viruses were reported in previous studies on Cypriot residents (33, 34). A recent seroprevalence study of phleboviruses in Cypriot dogs indicated also high neutralizing rates for SFSV (~60%) and TOSV (8.4%) (35). Lower rates found here may reflect the different specificity and sensitivity of the assay used (IIFT or plaque neutralization reduction assay (PRNT) (36), sampling method skewed towards specific geographical areas (Famagusta, Larnaca or Paphos districts), test subjects (healthy donors or symptomatic patients), species tested (humans or dogs) or simply a lower exposition of the modern Cypriot population, possibly due to arthropods abatement programs to eradicate malaria in the island (37).

Direct comparison with studies depicting SFV seroprevalences from neighbouring countries is limited, mostly because of the wide use of PRNT and the focus on TOSV. However, our study reports comparable anti-SFV IgG seroprevalence rates described in Turkish Central/Northern Anatolia (~33%) (38) but lower compared to the Turkish Mersin Province (~67%) (39), both tested by IIF.

The anti-SFV IgG seroprevalence rate increasing with age is in agreement with the conclusions of previous studies (40, 41) and demonstrates that the Cypriot population is exposed to SFVs throughout life. Unfortunately, other risk factors previously identified such as living in rural areas or high levels of outdoor activity could not be assessed because of the lack of patient's information.

Concerning WNV, the overall IgG seroprevalence found in Cyprus was 5% and the difference of rates between sexes, age groups or districts were not statistically significant. It is the first time that anti-WNV IgG seroprevalence has been estimated in Cyprus. WNV-specific IgG rates were lower in neighbouring countries such as Greece (2.1%) (42), Northern Italy (0.3–2.1%) (43) or South-Eastern France (1.4%) and higher in Libya (13.1%) or Tunisia (12.5%) (44) but more similar to Turkey (2.4– 12.1%) (20, 45), Algeria (6.7%) or Serbia (~4%) (46).

In addition, the SFV IgG seropositivity rate in the WNV IgG seropositive group (~31%) was comparable to the rate found in the entire population (29.4%). Similarly, the WNV IgG seropositivity proportion in the SFV IgG seropositive group (~5%) was analogous to the sample population tested in this study (~5%). This supports the hypothesis of random chance to contract each viral infection.

Regarding the role of SFV in CNS infections in Cyprus, we identified samples (8/ 127) with markers of recent SFV infection (IgM) in patients presenting symptoms akin to sandfly fever, supporting a role of SFVs in the aetiology of febrile illnesses/meningitis in Cyprus.

Distinction between antigenically related viruses amongst the sandfly fever Naples species (i.e. SFNV and TOSV) and virus closely related to Sandfly fever Sicilian virus (i.e. SFSV and SFCV) based on IIFT can be difficult due to cross-reactivity. However, there was a significantly higher rate of sera from symptomatic patients tested for anti-SFV IgM reacting only against viruses from the Naples serogroup than sera reacting only against viruses belonging to the Sicilian serogroup. Because of similar rates observed for IgG between serogroups and that patients were selected based on relevant symptoms, it confirms that infections from viruses belonging to the Naples serogroup are more symptomatic, particularly TOSV (47). Anti-SFV IgM specific to members of the Naples serogroup are at higher concentrations or last longer than the ones specific to Sicilian-like viruses (34). However, the detection of anti-SFV IgM specific to the Naples serocomplex in the CSF of affected patients strongly supports their neuroinvasive pathogenesis.

Possible cross-reactivity with anti-Hantavirus and anti-Crimean-Congo hemorrhagic fever virus (CCHFV) immunoglobulins cannot be ruled out based on the IIFT assay used. However, no hantavirus or CCHFV infection nor hemorrhagic fever have ever been reported in Cyprus so far. Even though their vectors are present on the island (48), it is unlikely that the anti-SFV antibodies seroprevalence rates depicted in this report are markedly influenced due to such cross-reactivity.

Markers of recent WNV infection (IgM) were also detected in patients displaying relevant symptoms, highlighting its circulation and morbidity in the Cypriot population. Few seropositive patients had neurological symptoms such as meningitis or meningoencephalitis. Together with the first Cypriot symptomatic case of a 75yr-old man with confirmed WNV infection during summer 2016 (24), these results demonstrate the circulation of WNV in Cyprus. All the anti-WNV IgM seropositive cases were from 2013, following the trend of marked decreased West Nile fever autochthonous cases reported in Europe during these yr (49).

Attempts to detect viral genetic material in more recent (2015 and 2016) patients' CSF by RT-qPCR following published protocols (26-28) were unsuccessful. The most likely causes for this shortcoming are probably low SFV and WNV viremia detectable only in a small period of time, usually before patients present to the healthcare personnel, the limited number of samples tested and the use of direct RTqPCR.

## Conclusion

This report provides evidence of significant SFV and WNV circulation in Cyprus as well as evidence of their pathogenicity among Cypriot patients. For these reasons, the surveillance and study of arboviral infections in Cyprus should be strengthened as these pathogens are potential causes of incapacitating and severe neurological diseases.

## Acknowledgements

The authors are grateful to Astero Constantinou and Sylvie Pavlides for technical assistance and Dr Christiana Demetriou for statistical assistance. This study was supported by the Cyprus Institute of Neurology and Genetics.

The authors declare that there is no conflict of interests.

## References

- Adams MJ, Lefkowitz EJ, King AMQ, Harrach B, Harrison RL, Knowles NJ, Kropinski AM, Krupovic M, Kuhn JH, Mushegian AR, Nibert M, Sabanadzovic S, Sanfaçon H, Siddell SG, Simmonds P, Varsani A, Zerbini FM, Gorbalenya AE, Davison AJ (2017) Changes to taxonomy and the International Code of Virus Classification and Nomenclature ratified by the International Committee on Taxonomy of Viruses (2017). Arch Virol. 162 (8): 2505–2538.
- Guler S, Guler E, Caglayik DY, Kokoglu OF, Ucmak H, Bayrakdar F, Uyar Y (2012) A sandfly fever virus outbreak in the East Mediterranean region of Turkey. Int J Infect Dis. 16(4): e244–246.
- Schultze D, Korte W, Rafeiner P, Niedrig M (2012) First report of sandfly fever virus infection imported from Malta into Switzerland, October 2011. Euro Surveill. 17(27): 20209.

- Charrel RN, Bichaud L, de Lamballerie X (2012) Emergence of Toscana virus in the mediterranean area. World J Virol. 1 (5): 135–141.
- Ayhan N, Charrel RN (2017) Of phlebotomines (sandflies) and viruses: a comprehensive perspective on a complex situation. Curr Opin Insect Sci. 22: 117–124.
- Kocak Tufan Z, Weidmann M, Bulut C, Kinikli S, Hufert FT, Dobler G, Demiroz AP (2011) Clinical and laboratory findings of a sandfly fever Turkey Virus outbreak in Ankara. J Infect. 63(5): 375–381.
- 7. Alkan C, Moin Vaziri V, Ayhan N, Badakhshan M, Bichaud L, Rahbarian N, Javadian EA, Alten B, de Lamballerie X, Charrel RN (2017) Isolation and sequencing of Dashli virus, a novel Sicilian-like virus in sandflies from Iran; genetic and phylogenetic evidence for the creation of one novel species within the Phlebovirus genus in the Phenuiviridae family. PLoS Negl Trop Dis. 11(12): e0005978.
- Ergunay K, Ayhan N, Charrel RN (2017) Novel and emergent sandfly-borne phleboviruses in Asia Minor: a systematic review. Rev Med Virol. 27(2): 10.1002/rmv.1898.
- Calzolari M, Ferrarini G, Bonilauri P, Lelli D, Chiapponi C, Bellini R, Dottori M (2018) Co-circulation of eight different phleboviruses in sand flies collected in the Northern Apennine Mountains (Italy). Infect Genet Evol. 64: 131–134.
- Marchi S, Trombetta CM, Kistner O, Montomoli E (2017) Seroprevalence study of Toscana virus and viruses belonging to the Sandfly fever Naples antigenic complex in central and southern Italy. J Infect Public Health. 10(6): 866–869.
- Dokianakis E, Tsirigotakis N, Christodoulou V, Poulakakis N, Antoniou M (2018) Identification of wild-caught phlebotomine sand flies from Crete and Cyprus using DNA barcoding. Parasit Vectors.

11(1): 94.

- 12. Leger N, Depaquit J, Ferte H (2000) Phlebotomine sandflies (Diptera-Psychodidae) of the isle of Cyprus. I--Description of *Phlebotomus* (*Transphlebotomus*) *economidesi* n. sp. Parasite. 7(2): 135– 141.
- 13. Niklasson B, Eitrem R (1985) Sandfly fever among Swedish UN troops in Cyprus. Lancet. 1(8439): 1212.
- Eitrem R, Vene S, Niklasson B (1990) Incidence of sand fly fever among Swedish United Nations soldiers on Cyprus during 1985. Am J Trop Med Hyg. 43 (2): 207–211.
- Papa A, Konstantinou G, Pavlidou V, Antoniadis A (2006) Sandfly fever virus outbreak in Cyprus. Clin Microbiol Infect. 12(2): 192–194.
- 16. Konstantinou GN, Papa A, Antoniadis A (2007) Sandfly-fever outbreak in Cyprus: are phleboviruses still a health problem? Travel Med Infect Dis. 5(4): 239–242.
- 17. Petersen LR, Brault AC, Nasci RS (2013) West Nile virus: review of the literature. JAMA. 310(3): 308–315.
- Hayes EB, Sejvar JJ, Zaki SR, Lanciotti RS, Bode AV, Campbell GL (2005) Virology, pathology, and clinical manifestations of West Nile virus disease. Emerg Infect Dis. 11(8): 1174–1179.
- 19. Violaris M, Vasquez MI, Samanidou A, Wirth MC, Hadjivassilis A (2009) The mosquito fauna of the Republic of Cyprus: a revised list. J Am Mosq Control Assoc. 25(2): 199–202.
- 20. Ergunay K, Gunay F, Erisoz Kasap O, Oter K, Gargari S, Karaoglu T, Tezcan S, Cabalar M, Yildirim Y, Emekdas G, Alten B, Ozkul A (2014) Serological, molecular and entomological surveillance demonstrates widespread circulation of West Nile virus in Turkey. PLoS Negl Trop Dis. 8(7): e3028.
- 21. Kilpatrick AM (2011) Globalization, land use, and the invasion of West Nile virus.

Science. 334(6054): 323-327.

- Papa A (2017) Emerging arboviral human diseases in Southern Europe. J Med Virol. 89(8): 1315–1322.
- 23. Burki T (2018) Increase of West Nile virus cases in Europe for 2018. Lancet. 392 (10152): 1000.
- 24. Paphitou NI, Tourvas A, Floridou D, Richter J, Tryfonos C, Christodoulou C (2017) The first human case of neuroinvasive West Nile virus infection identified in Cyprus. J Infect Public Health. 10(6): 891–893.
- 25. Richter J, Tryfonos C, Tourvas A, Floridou D, Paphitou NI, Christodoulou C (2017) Complete Genome Sequence of West Nile Virus (WNV) from the First Human Case of Neuroinvasive WNV Infection in Cyprus. Genome Announc. 5 (43): e01110–01117.
- 26. Perez-Ruiz M, Collao X, Navarro-Mari JM, Tenorio A (2007) Reverse transcription, real-time PCR assay for detection of Toscana virus. J Clin Virol. 39(4): 276–281.
- 27. Cordey S, Bel M, Petty TJ, Docquier M, Sacco L, Turin L, Cherpillod P, Emonet S, Louis-Simonet M, Zdobnov EM, Ambrosioni J, Kaiser L (2015) Toscana virus meningitis case in Switzerland: an example of the ezVIR bioinformatics pipeline utility for the identification of emerging viruses. Clin Microbiol Infect. 21 (4): e1–4.
- 28. Patel P, Landt O, Kaiser M, Faye O, Koppe T, Lass U, Sall AA, Niedrig M (2013) Development of one-step quantitative reverse transcription PCR for the rapid detection of flaviviruses. Virol J. 10: 58.
- 29. Ergunay K, Kasap OE, Orsten S, Oter K, Gunay F, Yoldar AZ, Dincer E, Alten B, Ozkul A (2014) Phlebovirus and Leishmania detection in sandflies from eastern Thrace and northern Cyprus. Parasit Vectors. 7: 575.
- 30. Sellers RF, Herniman KA (1981) Neutral-

ising antibodies to Akabane virus in ruminants in Cyprus. Trop Anim Health Prod. 13(1): 57–60.

- Polydorou K (1978) The 1977 outbreak of bluetongue in Cyprus. Trop Anim Health Prod. 10(4): 229–232.
- 32. Stahn B, Sudeck H, Frickmann H, Kruger A, Burchard HG, Wiemer D (2018) [Sandfly fever-a "neglected" disease]. Hautarzt. 69(11): 928–937.
- 33. Eitrem R, Vene S, Niklasson B (1991) ELI-SA for detection of IgM and IgG antibodies to sandfly fever Sicilian virus. Res Virol. 142(5): 387–394.
- 34. Eitrem R, Stylianou M, Niklasson B (1991) High prevalence rates of antibody to three sandfly fever viruses (Sicilian, Naples, and Toscana) among Cypriots. Epidemiol Infect. 107(3): 685–691.
- 35. Alwassouf S, Christodoulou V, Bichaud L, Ntais P, Mazeris A, Antoniou M, Charrel RN (2016) Seroprevalence of Sandfly-Borne Phleboviruses Belonging to Three Serocomplexes (Sandfly fever Naples, Sandfly fever Sicilian and Salehabad) in Dogs from Greece and Cyprus Using Neutralization Test. PLoS Negl Trop Dis. 10(10): e0005063.
- 36. Ergunay K, Litzba N, Lo MM, Aydogan S, Saygan MB, Us D, Weidmann M, Niedrig M (2011) Performance of various commercial assays for the detection of Toscana virus antibodies. Vector Borne Zoonotic Dis. 11(6): 781–787.
- 37. Vasquez MI, Violaris M, Hadjivassilis A, Wirth MC (2009) Susceptibility of *Culex pipiens* (Diptera: Culicidae) field populations in Cyprus to conventional organic insecticides, *Bacillus thuringiensis* subsp. *israelensis*, and methoprene. J Med Entomol. 46(4): 881–887.
- 38. Ergunay K, Saygan MB, Aydogan S, Lo MM, Weidmann M, Dilcher M, Sener B, Hascelik G, Pinar A, Us D (2011) Sandfly fever virus activity in central/northern Anatolia, Turkey: first report of Tos-

cana virus infections. Clin Microbiol Infect. 17(4): 575–581.

- Tezcan S, Dincer E, Ulger M, Ozgur D, Erdogan S, Ozkul A, Emekdas G, Ergunay K (2015) Serological investigation of phlebovirus exposure in blood donors from the Mediterranean Province of Mersin, Turkey. Mikrobiyol Bul. 49(3): 403– 413.
- 40. de Ory-Manchon F, Sanz-Moreno JC, Aranguez-Ruiz E, Ramirez-Fernandez R (2007) Age-dependent seroprevalence of Toscana virus in the Community of Madrid: 1993–1994 and 1999–2000. Enferm Infecc Microbiol Clin. 25(3): 187–189.
- 41. Anagnostou V, Papa A (2013) Prevalence of antibodies to phleboviruses within the sand fly fever Naples virus species in humans, northern Greece. Clin Microbiol Infect. 19(6): 566–570.
- 42. Hadjichristodoulou C, Pournaras S, Mavrouli M, Marka A, Tserkezou P, Baka A, Billinis C, Katsioulis A, Psaroulaki A, Papa A, Papadopoulos N, Mamuris Z, Tsakris A, Kremastinou J, Project M (2015) West Nile Virus Seroprevalence in the Greek Population in 2013: A Nationwide Cross-Sectional Survey. PLoS One. 10(11): e0143803.
- 43. Barzon L, Pacenti M, Franchin E, Squarzon L, Lavezzo E, Cattai M, Cusinato R, Palu G (2013) The complex epidemiological scenario of West Nile virus in Italy. Int J Environ Res Public Health. 10(10): 4669–4689.
- 44. Bahri O, Dhifallah I, Ben Alaya-Bouafif N, Fekih H, Gargouri J, Triki H (2011) Sero-epidemiological study of West Nile virus circulation in human in Tunisia. Bull Soc Pathol Exot. 104(4): 272–276.
- 45. Ergünay K, Tufan ZK (2014) Overview of West Nile virus and sandfly-borne Phlebovirus infections in Anatolia. J Microbiol Infect Dis. Special Issue 1: S22– S31.

- 46. Failloux AB, Bouattour A, Faraj C, Gunay F, Haddad N, Harrat Z, Jancheska E, Kanani K, Kenawy MA, Kota M, Pajovic I, Paronyan L, Petric D, Sarih M, Sawalha S, Shaibi T, Sherifi K, Sulesco T, Velo E, Gaayeb L, Victoir K, Robert V (2017) Surveillance of arthropod-borne viruses and their vectors in the Mediterranean and Black Sea Regions within the MediLabSecure Network. Curr Trop Med Rep. 4(1): 27–39.
- 47. Alkan C, Bichaud L, de Lamballerie X, Alten B, Gould EA, Charrel RN (2013) Sandfly-borne phleboviruses of Eurasia and Africa: epidemiology, genetic diversity, geographic range, control measures. Antiviral Res. 100(1): 54–74.
- 48. Chochlakis D, Ioannou I, Sandalakis V, Dimitriou T, Kassinis N, Papadopoulos B, Tselentis Y, Psaroulaki A (2012) Spotted fever group Rickettsiae in ticks in Cyprus. Microb Ecol. 63(2): 314–323.
- 49. ECDC (2017) West Nile fever maps\_ Historical data. European Centre for Disease Prevention and Control. Available at: http://ecdc.europa.eu/en/healthtopics/w est\_nile\_fever/West-Nile-fevermaps/Pages/historical-data.aspx.