Lin et al. BMC Genomics 2014, 15(Suppl 6):S6
http://www.biomedcentral.com/1471-2164/15/S6/S6

BMC Genomics

RESEARCH Open Access

What is the difference between the breakpoint graph and the de Bruijn graph?

Yu Lin[1], Sergey Nurk[2,3], Pavel A Pevzner[1*]

From Twelfth Annual Research in Computational Molecular Biology (RECOMB) Satellite Workshop on Comparative Genomics
Cold Spring Harbor, NY, USA. 19-22 October 2014

Abstract

The breakpoint graph and the de Bruijn graph are two key data structures in the studies of genome rearrangements and genome assembly. However, the classical breakpoint graphs are defined on two genomes (represented as sequences of synteny blocks), while the classical de Bruijn graphs are defined on a single genome (represented as DNA strings). Thus, the connection between these two graph models is not explicit. We generalize the notions of both the breakpoint graph and the de Bruijn graph, and make it transparent that the breakpoint graph and the de Bruijn graph are mathematically equivalent. The explicit description of the connection between these important data structures provides a bridge between two previously separated bioinformatics communities studying genome rearrangements and genome assembly.

Introduction

The de Bruijn graph is a data structure first brought to bioinformatics as a method to assemble genomes from the experimental data generated by sequencing by hybridization [1]. It later became the key algorithmic technique in genome assembly [2,3] that resulted in dozens of software tools [4-12]. In addition, the de Bruijn graphs have been used for repeat classification [13], de novo protein sequencing [14], synteny block construction [15,16], multiple sequence alignment [17], and other applications in genomics and proteomics.

The breakpoint graph is a data structure introduced to study the reversal distance [18], which has formed the basis for much algorithmic research on rearrangements over the last two decades [19].

Since the connections between the breakpoint graphs and the de Bruijn graphs was never explicitly described, researchers studying genome rearrangements often do not realize that breakpoint graphs are merely de Bruijn graphs in disguise. As a result, they often do not know how to move from the traditional breakpoint graphs on *synteny blocks* to the breakpoint graphs on genomes (with "single nucleotide" resolution), particularly in the case of double-stranded *genomes* with inverted repeats. Likewise, researchers working in genome assembly are often unaware about the connections between the de Bruijn graphs and the breakpoint graphs. As a result, the exchange of ideas between these two communities has been limited. For example, Iqbal et al. [20] recently introduced the notion of the *colored de Bruijn graphs* that resulted in a popular Cortex assembler. While the notion of the colored de Bruijn graphs is essentially identical to the notion of the breakpoint graph, authors of [20] are probably unaware about this connection since they provided no references to previous genome rearrangement studies. This is unfortunate since various results about the breakpoint graphs (e.g., the connection between rearrangements and alternating cycles) remained beyond the scope of this very useful study.

Recently, genome rearrangement studies moved from the level of synteny blocks to the level of single nucleotides [21]. Likewise, genome assembly experts recently moved towards the analysis of structural variations and comparative assembly of related species based on the analysis of the de Bruijn graphs [20]. We thus argue that the time has come to explain that the breakpoint graphs and the de

* Correspondence: ppevzner@ucsd.edu
[1]Department of Computer Science and Engineering, University of California, San Diego, 9500 Gilman Dr, CA 92093 La Jolla, USA
Full list of author information is available at the end of the article

Bruijn graphs are two identical data structures (if one ignores a cosmetic difference between them) as they both represent specific instances of a general notion of the *A-Bruijn graph* introduced in [13]. The A-Bruijn graphs are based on representing genomes as sets of labeled paths and further gluing identically labeled edges (breakpoint graphs) or vertices (de Bruijn graphs) in the resulting paths.

We argue that a unified framework covering both breakpoint and de Bruijn graphs is important to bridge the divide between researchers working with breakpoint graphs (that usually focus in rearrangements and ignore repeats) and researchers working with de Bruijn graphs (that usually focus on repeats and ignore rearrangements). In reality, there exists a complex interplay between rearrangements and repeats, e.g., LINE repeats and segmental duplications often trigger rearrangements [22-24]. However, this interplay is not explicitly revealed by the breakpoint graphs since they do not even encode repeats (repeats are intentionally masked at the synteny block construction step). For example, the interplay between LINEs and rearrangements cannot be derived from the breakpoint graph alone forcing Zhao and Bourque [23] to perform additional analysis. Our goal is to introduce the graphs that encode both rearrangements and repeats and immediately reveal this interplay. For example, encoding repeats present in the breakpoint regions (that may potentially trigger rearrangements) leads to gluing alternating cycles in the breakpoint graphs and requires development of new algorithms that integrate rearrangements and repeats. In such graphs, the classical *non-self-intersecting alternating cycles* formed by edges alternating between two colors (the workhorse of genome rearrangement studies) may turn into *self-intersecting* cycles formed by edges alternating between 3 colors, where the third color corresponds to repeated elements (see Figure 1). Nurk and Pevzner [25] recently used this framework to develop a new comparative genome analysis tool SPArcle and applied it to analyzing multiple bacterial strains resulting from the "controlled evolution" experiments [26]. SPArcle is based on SPAdes assembler and, in difference from Cortex, it uses ideas from the previous genome rearrangement studies (e.g., alternating cycles) to analyze the resulting A-Bruijn graphs.

Genome rearrangement studies usually start from constructing a set of *synteny blocks* shared by two genomes (see Figure 2). Each genome is defined as a sequence of synteny blocks separated by *breakpoint regions* and is represented as a path formed by alternating colored and black edges, where synteny blocks correspond to directed and labeled black edges and breakpoint regions correspond to undirected colored edges. Figure 3(a) presents paths corresponding to 11 synteny blocks shared by Human and Mouse × chromosomes. Each synteny block S_i is represented as an directed black edge (S_i^t, S_i^h), where S_i^t and S_i^h refer to the endpoints of the synteny blocks representing its tail and head, respectively. Two consecutive synteny blocks are separated by a breakpoint region in the Human (Mouse) × chromosome that is modeled by a red (blue) edge connecting the corresponding endpoints of these synteny blocks. The (traditional) breakpoint graph of Human and Mouse × chromosomes is obtained by "gluing" identically labeled black edges in these two paths as shown in Figure 3.

The multiple breakpoint graphs [27] are constructed from a set of $k > 2$ genomes using the same procedure. While every synteny block appears just once in each of the genomes shown above, the definition of the breakpoint graph naturally extends to the case when a synteny block appears multiple times (or does not appear in a particular genome).

The classical de Bruijn graph is defined on a single genome (represented as a DNA string), while the

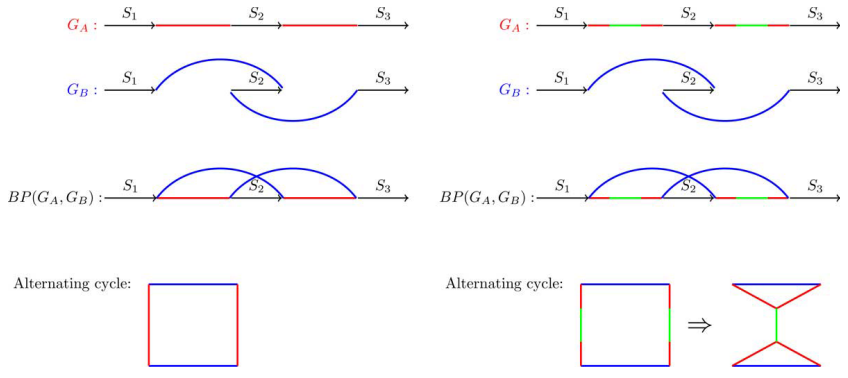


Figure 1 Genomes $G_A = S_1, S_2, S_3$ and $G_B = S_1, -S_2, S_3$ (represented as bicolored paths) differ from each other by a single reversal of segment S_2. The breakpoint graph $BP(G_A, G_B)$ and the alternating cycle constructed from genomes G_A and G_B (left: no repeats at the breakpoint regions; right, a pair of repeats colored in green at the breakpoint regions).

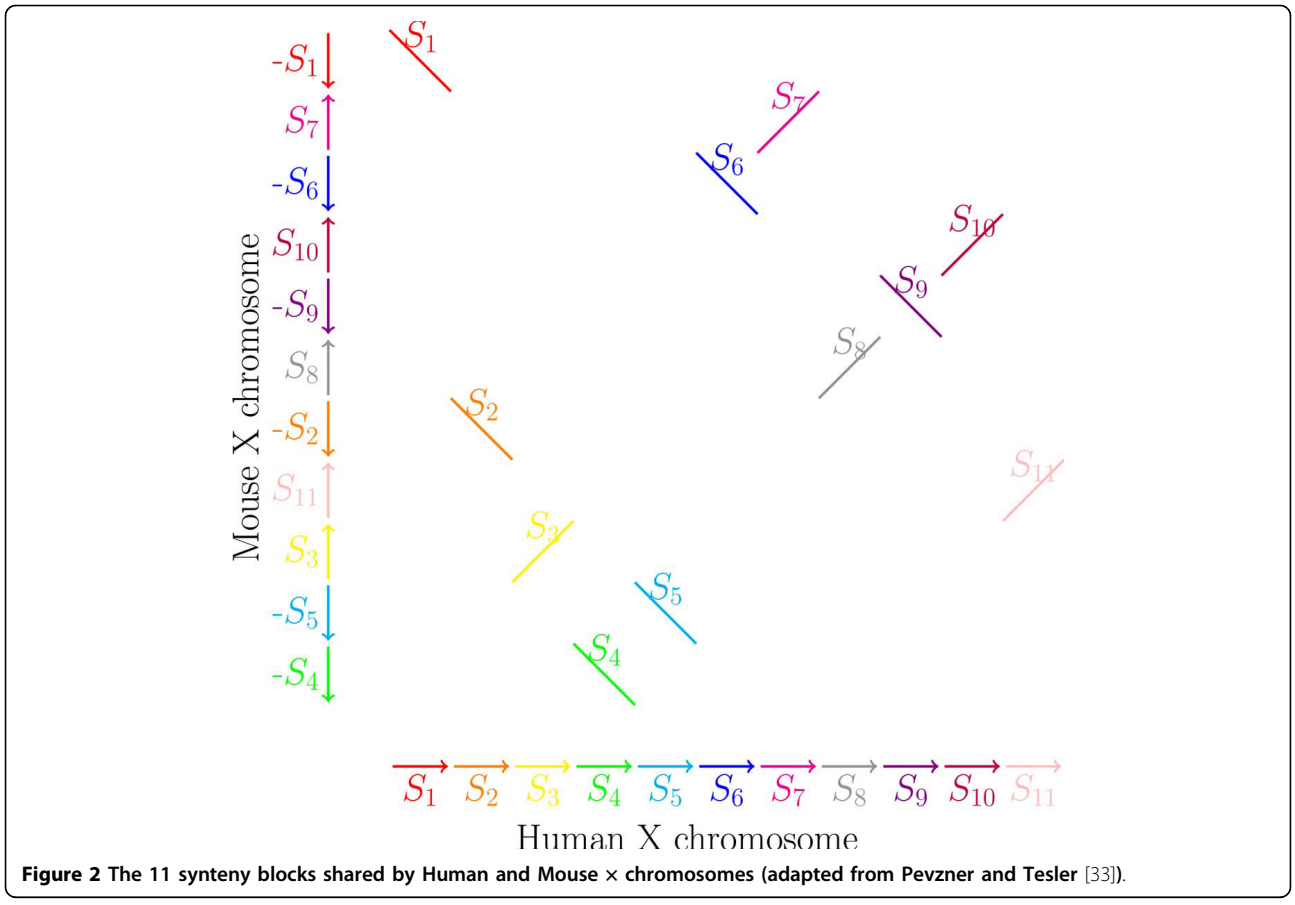


Figure 2 The 11 synteny blocks shared by Human and Mouse × chromosomes (adapted from Pevzner and Tesler [33]).

classical breakpoint graph is defined on two genomes (represented as sequences of synteny blocks). Since synteny blocks are DNA strings, the question arises whether one can study both the de Bruijn graph and the breakpoint graph in the same framework, and what is the difference between the de Bruijn graph and the breakpoint graph.

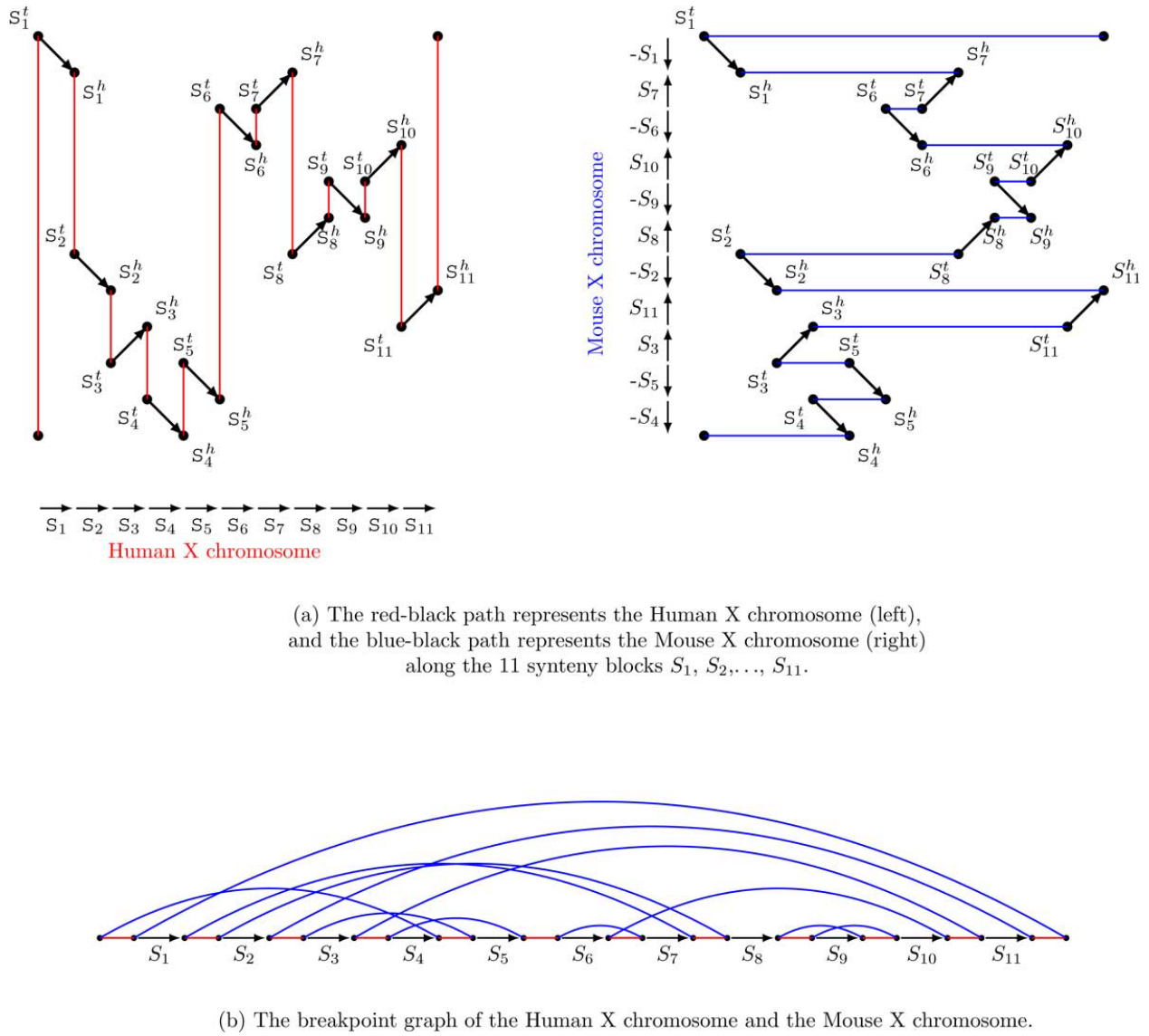
In this paper, we generalize the definitions of both the de Bruijn graph and the breakpoint graph for both single and multiple genomes and for both single-stranded and double-stranded cases. We further show that the breakpoint graph and the de Bruijn graph are mathematically equivalent.

A single genome (single-stranded case)

We now generalize the definition of the breakpoint graph to a single DNA strings. Traditionally, breakpoint graphs were defined on the set of synteny blocks rather than directly on DNA strings. Given a string *String*, we define its "*synteny blocks*" as its (k-1)-mers, represent each (k-1)-mer by a directed black edge, and construct an alternating path, denoted $Path_{BP}$ (*String, k*), by inserting directed colored edges between consecutive (k-1)-mers. The breakpoint graph is obtained by gluing identical (k-1)-mers (directed black edges) in this path. It is easy to see that each breakpoint region connects 2 consecutive (k-1)-mers and thus corresponds to a kmer. The breakpoint graph of a string *String*, denoted *BP* (*String, k*), is shown in Figure 4.

Given a string $String = s_1 s_2 \ldots s_n$, the de Bruijn graph *DB*(*String, k*) is defined as follows. The string *String* is represented as a colored path $Path_{DB}$ (*String, k*), whose $(n - k + 1)$ edges are labeled by k-mers, $s_1 s_2 \ldots s_k$, $s_2 s_3 \ldots s_{k+1}$, . . ., and $s_{n-k+1} s_{n-k+2} \ldots s_n$. Implicitly, each vertex in the path is labeled by a (k-1)-mer. The de Bruijn graph *DB*(*String, k*) results from gluing identically labeled vertices in the path [28]. Figure 5 shows the de Bruijn graph on the same string as in Figure 4.

Comparison of Figure 4 and 5 reveals that *BP* (*String, k*) is equivalent to *DB*(*String, k*). *BP* (*String, k*) can be converted into *DB*(*String, k*) by collapsing all black edges, while *DB*(*String, k*) can be converted into *BP* (*String, k*) by expanding each vertex v into a directed edge (v', v'') in such a way that all incoming edges into v (outgoing edges from v) become incoming edges into v' (outgoing edges from v''). Table 1 summarizes the correspondence between *BG*(*String, k*) and *DB*(*String, k*).



(a) The red-black path represents the Human X chromosome (left), and the blue-black path represents the Mouse X chromosome (right) along the 11 synteny blocks $S_1, S_2, \ldots, S_{11}$.

(b) The breakpoint graph of the Human X chromosome and the Mouse X chromosome.

Figure 3 The synteny blocks shared by Human and Mouse × chromosomes and the resulting breakpoint graph. Human × chromosome is represented as a path formed by alternating red and black edges, while Mouse × chromosome is represented as a path formed by alternating blue and black edges.

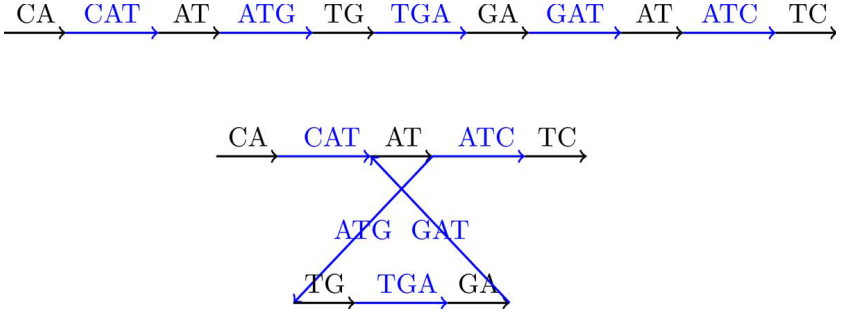


Figure 4 The breakpoint graph *BP* (*CATGATC*, 3). 2-mers represent synteny blocks and 3-mers represent breakpoint regions.

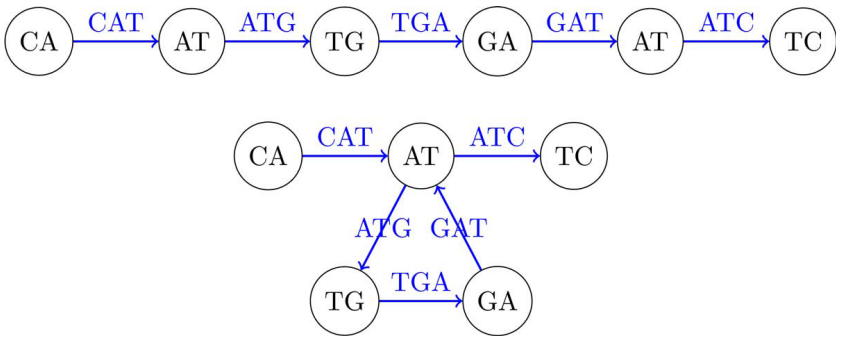


Figure 5 The de Bruijn graph DB(*CATGATC*, 3). 2-mers are represented by vertices and 3-mers are represented by edges.

We note that while the notions of the breakpoint graph and the de Bruijn graph were defined for a single string, the same definition works for a set of strings (see Figure 6).

Multiple genomes (single-stranded case)

Given genomes G_A and G_B (represented as sets of strings), we classify their k-mers into 3 classes: A (occur only in G_A), B (occur only in G_B) and AB (occur in both G_A and G_B).

The breakpoint graph $BP(G_A, G_B, k)$ is simply coloring the k-mer edges of the breakpoint graph $BP(G_A \cup G_B, k)$ into 3 colors: A (blue), B (red) and AB (green) (Figure 7). Similarly, the de Bruijn graph $DB(G_A, G_B, k)$ is simply coloring the edges of the de Bruijn graph $DB(G_A \cup G_B, k)$ into 3 colors: A (blue), B (red) and AB (green) (see Figure 8).

In practice, both $BP(G_A, G_B, k)$ and $DB(G_A, G_B, k)$ are often *condensed* as follows.

A non-branching directed path consisting of 3 edges is called *condensible* in $BP(G_A, G_B, k)$ if its middle edge is black and its starting and ending edges have the same color C. We substitute a condensible path by a single directed edge colored C with the same direction as the direction of the path. The condensed breakpoint graph $CBP(G_A, G_B, k)$ iteratively replaces all condensible paths by single edges in $BP(G_A, G_B, k)$ (Figure 7(c)). A new edge resulting from condensing a nonbranching path formed by edges e_1, e_2, and e_3 is assigned a label whose prefix is a label of e_1 and whose suffix is the label of e_3 (labeling of edges in the condensed graphs is not the focus of this paper and is not discussed in the examples below).

A non-branching directed path consisting of 2 edges is called *condensible* in $DB(G_A, G_B, k)$ if its starting and ending edges have the same color C. We substitute a condensible path by a single edge colored C with the same direction as the direction of the path. The condensed de Bruijn graph $CDB(G_A, G_B, k)$ iteratively replaces all condensible paths by single edges in $DB(G_A, G_B, k)$ (Figure 8(c)).

Note that both the condensed breakpoint graph in Figure 7(c) and the condensed de Bruijn graph in Figure 8(c) reveal an alternating red-blue cycle on 6 edges, a signature of a transposition. While researchers working on genome rearrangements are well aware about alternating cycles in the breakpoint graphs, the researchers working on de Bruijn graphs may not know that these cycles represent fingerprints of rearrangements.

Figure 7 and 8 illustrate that $BP(G_A, G_B, k)$ is mathematically equivalent to $DB(G_A, G_B, k)$. $BP(G_A, G_B, k)$ can be converted into $DB(G_A, G_B, k)$ by collapsing all black edges, while $DB(G_A, G_B, k)$ can be converted into $BP(G_A, G_B, k)$ by expanding all vertices. It is the same for the condensed case. Table 2 summarizes the comparison between $DB(G_A, G_B, k)$ and $BP(G_A, G_B, k)$.

While the above notions of the breakpoint graph and the de Bruijn graph were defined for 2 genomes, they naturally generalize to any number of genomes [27].

Table 1 The correspondence between *BP*(*String, k*) and *DB*(*String, k*)

	DB(*String, k*)	*BP* (*String, k*)
(k-1)-mer	a vertex	a black directed edge
k-mer	a directed blue edge	a directed blue edge
glue	the vertex	the black directed edge
synteny block	the vertex	the black directed edge
breakpoint region	the directed blue edge	the directed blue edge

A single genome (double-stranded case)

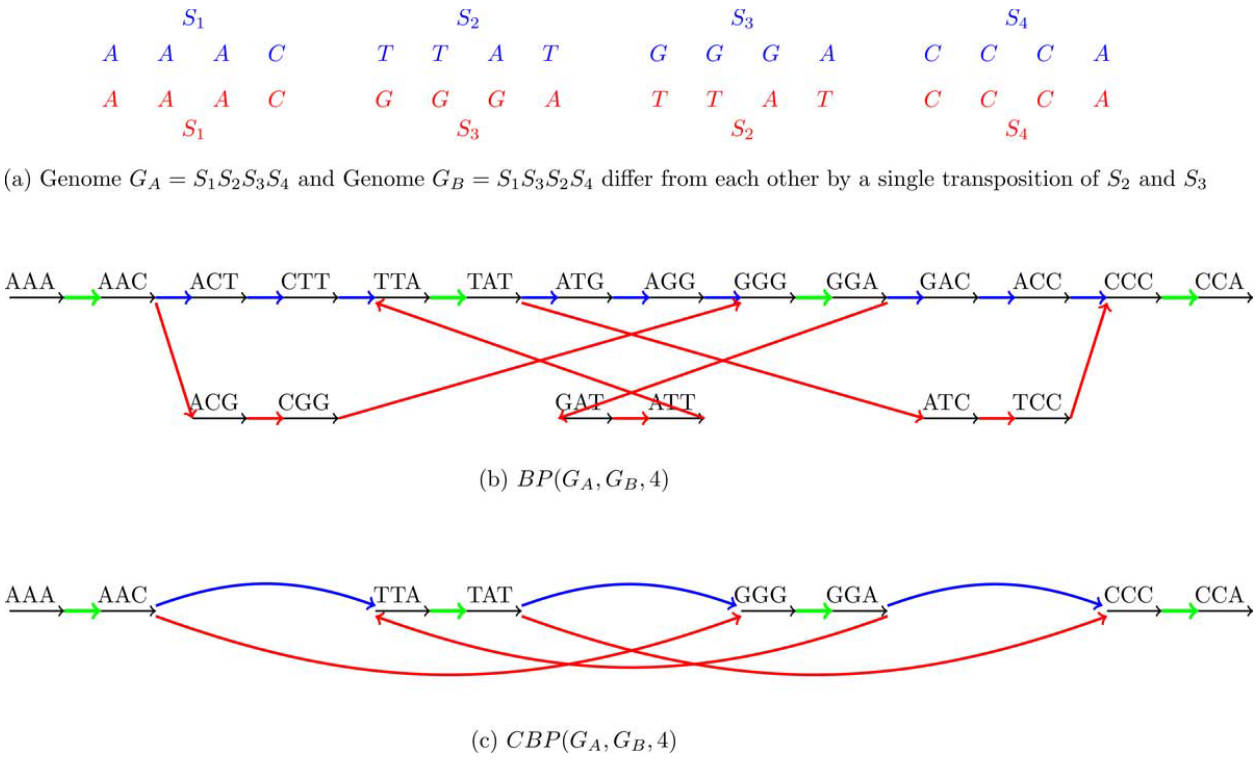
We now generalize the notions of the de Bruijn and breakpoint graphs from single-stranded to double-stranded genomes. Instead of the explicit representation of both strands (like in most existing assemblers), we describe a more efficient representation that encodes both strands in a single *canonical strand* (compare with similar representations of both strands in SPAdes [12] and some other assemblers).

Figure 6 The the breakpoint graph (left) and the de Bruijn graph (right) of two strings *CATGATC* and *CTGAG*.

For a nucleotide x, we denote its complementary nucleotide as $\overline{x}$, e.g., $\overline{A} = T$ T and $\overline{C} = G$. Given a k-mer $s = s_1 s_2 \ldots s_k$, we define its *reverse complement* as the k-mer $\overline{s} = \overline{s_k} \ldots \overline{s_2}\ \overline{s_1}$. We call a k-mer *canonical* if it is smaller or equal (in the lexicographic order) than its reverse complement. For example, AG and AT are canonical 2-mers but CT is not a canonical 2-mer. In this section, we will represent strings as paths labeled only by canonical k-mers and will define the breakpoint and de Bruijn graphs as the results of gluing these paths.

While *DB*(*String, k*) and *BP* (*String, k*) glue *identical* (k-1)-mers in *String*, these graphs were not designed to glue a (k-1)-mer with its reverse complement (see Figure 9). In many applications, gluing (k-1)-mers with their reverse complements would be beneficial; for example, developers of many genome assemblers invest significant efforts in maintaining the "symmetric" de Bruijn graphs at all stages of the assembly so that the subgraph representing one strand is topologically identical to the subgraph representing another strand [12]. Since the breakpoint graph and the de Bruijn graph in Figure 9 do not allow one to analyze the interplay between rearrangements and inverted repeats, we need to come up with a new graph-theoretical model for double-stranded genomes. We note that since A-Bruijn graphs were designed to accommodate gluing of *arbitrary* positions in *String* (as long as they are defined as "aligned" in the A-Bruijn graph framework [13]), gluing (k-1)-mers with their reverse complements perfectly fits in the framework of the A-Bruijn graphs.

To model double-stranded strings as paths, we introduce the concept of a *canonical* path representing a string *String* (that differs from the standard representation of *String* as a path from Section 2). To transform a standard path $Path_{BP}$ (*String, k*) into a canonical path *CPathBP* (*String, k*), we reverse directions of all black edges labeled by non-canonical (k-1)-mers and change their labels into their reverse complements (see Figure 10, left). As a result,

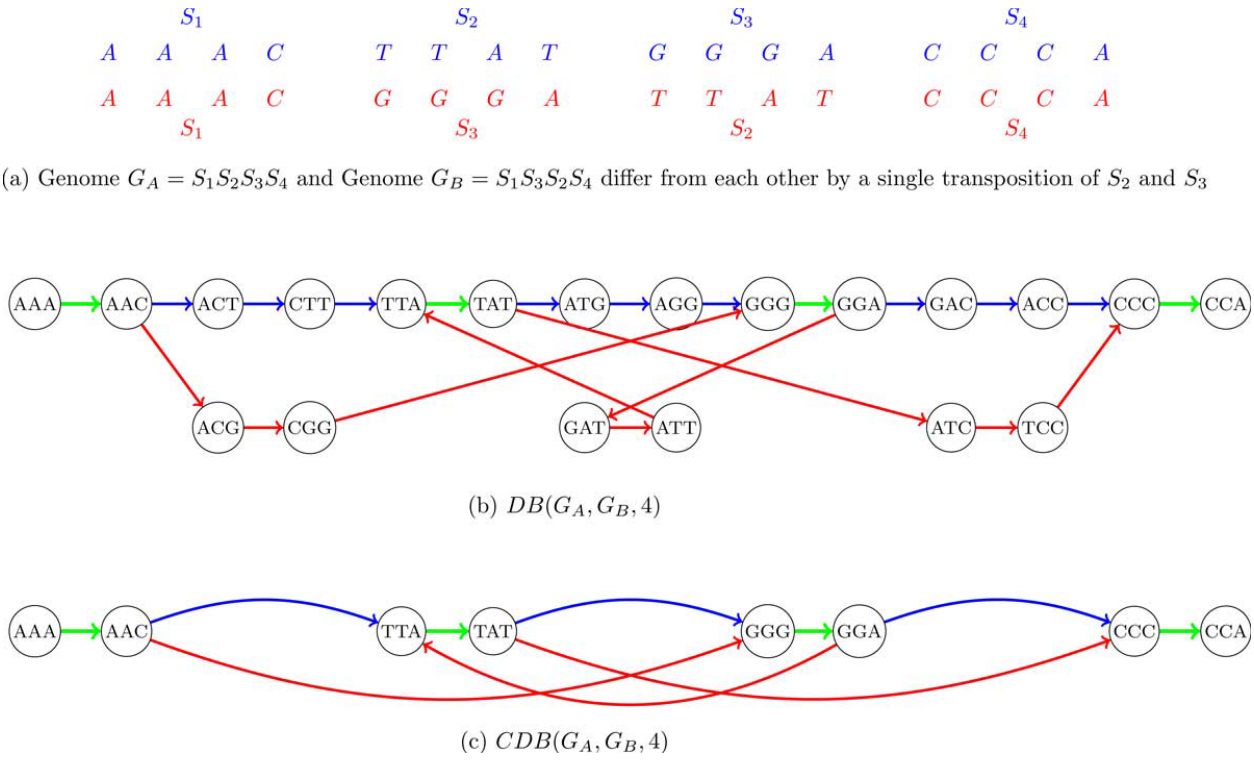


(a) Genome $G_A = S_1S_2S_3S_4$ and Genome $G_B = S_1S_3S_2S_4$ differ from each other by a single transposition of S_2 and S_3

(b) $BP(G_A, G_B, 4)$

(c) $CBP(G_A, G_B, 4)$

Figure 7 The breakpoint graph *BP*(G_A, G_B, 4) and the condensed breakpoint graph *CBP*(G_A, G_B, 4) of two genomes G_A and G_B.



(a) Genome $G_A = S_1S_2S_3S_4$ and Genome $G_B = S_1S_3S_2S_4$ differ from each other by a single transposition of S_2 and S_3

(b) $DB(G_A, G_B, 4)$

(c) $CDB(G_A, G_B, 4)$

Figure 8 The de Bruijn graph *DB*(G_A, G_B, 4) and the condensed de Bruijn graph *CDB*(G_A, G_B, 4) of two genomes G_A and G_B.

all black edges in the canonical path are labeled by canonical (*k*-1)-mers.

To transform a standard path $Path_{DB}$ (*String, k*) into a canonical path $CPath_{DB}$ (*String, k*), we simply change labels of all vertices labeled by non-canonical (*k*-1)-mers into their reverse complements (see Figure 10, right).

After transforming standard paths $Path_{BP}$ (*String, k*) and $Path_{DB}$ (*String, k*) into a canonical paths $CPath_{BP}$ (*String, k*) and $CPath_{DB}$ (*String, k*) respectively, the definition of the breakpoint graph (gluing of identically labeled black edges in $CPath_{BP}$ (*String, k*)) and the de Bruijn graph (gluing of identically labeled vertices in

Table 2 The correspondence between *DB*(G_A, G_B, *k*) and *BP*(G_A, G_B, *k*) (under the single-stranded representation)

	***DB*(G_A, G_B, *k*)**	***BP*(G_A, G_B, *k*)**
(*k*-1)-mer	vertex	black directed edge (E_1)
k-mer	directed edge	directed edge (E_2)
color	red/blue/green directed edge blue in G_A, red in G_B, green in both G_A and G_B	red/blue/green directed edge (E_2) blue in G_A, red in G_B, green in both G_A and G_B
glue	vertex	E_1
synteny block as a path	vertex-green edge-. . .-vertex	E_1 -green E_2-. . .-E_1
breakpoint region as a path	red edge-vertex. . .-red edge blue edge-vertex-. . .-blue edge	red E_2 -E_1-. . .-red E_2 blue E_2 -E_1-. . .-blue E_2
condensing paths into edges	red edge-vertex-red edge → red edge blue edge-vertex-blue edge → blue edge green edge-vertex-green edge → green edge	red E_2 -E_1-red E_2 → red E_2 blue E_2-E_1-blue E_2 → blue E_2 green E_2-E_1-green E_2 → green E_2
after condensation	*CDB*(G_A, G_B, *k*)	*CBP*(G_A, G_B, *k*)
synteny block in condensed graph	vertex-green edge-vertex	E_1-green E_2-E_1
breakpoint region in condensed graph	red edge blue edge	red E_2 blue E_2

We refer to black directed edges in the de Bruijn graph as E_1-edges and to colored directed edges as E_2-edges

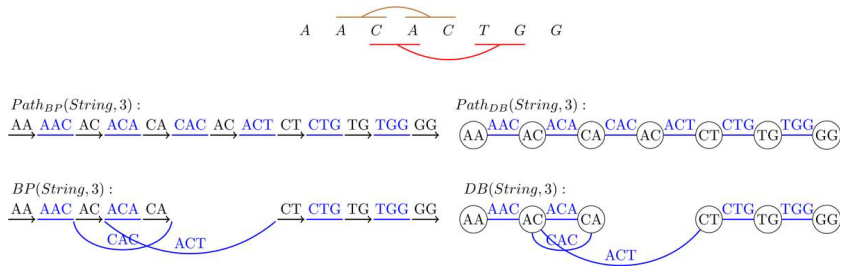


Figure 9 A string *String* = *ACAGTC A* (top), *BP*(*String*, 3) (left) and *DB*(*String*, 3) (right). A pair of repeats in *String* is shown in brown and a pair of inverted repeats is shown in red.

$CPath_{BP}$ (*String, k*)) remain unchanged. Pairs of *k*-mer edges are also glued if they represent the reverse complement *k*-mer of each other and are labeled by both *k*-mers. The graphs obtained after gluing these paths are called the *double-stranded breakpoint graph BP** (*String, k*) and the *double-stranded de Bruijn graph DB** (*String, k*) (Figure 10). While *BP**(*String, k*) makes use of the direction of the black edge to represent whether the canonical string or its reverse complement is used, *DB** (*String, k*) collapses all black edges into vertices and no longer maintains the direction information to distinguish these two possibilities. A pair of vertices in *DB** (*String, k*) labeled by canonical (*k*-1)-mers v and w may potentially correspond to 4 types of edges depending on whether the edge connects (i) v and w, (ii) v and $\overline{w}$, (iii) $\overline{v}$ and w, and (iv) $\overline{v}$ and $\overline{w}$ (compare to the *bi-directed de Bruijn graph* [29]).

As before, *DB**(*String, k*) is obtained from *BP**(*String, k*) by collapsing all black edges, while *BP**(*String, k*) is obtained from *DB**(*String, k*) by expanding all vertices into black edges (and connecting black edges according to the labels on the colored edges).

One may notice that while the double-stranded de Bruijn graph is similar to the bi-directional de Bruijn graph introduced in [29], it does not require the explicit introduction of the bi-directional edges. The notions of the double-stranded breakpoint graphs and de Bruijn graphs also can be naturally extended from a single double-stranded string to multiple double-stranded strings.

Multiple genomes (double-stranded case)

Given a string $String = s_1 s_2 \ldots s_{i-1} s_i s_{i+1} \ldots s_{j-1}\, s_j\, s_{j+1} \ldots s_{n-1} s_n$, a reversal of the segment $s_i s_{i+1} \ldots s_{j-1} s_j$ results in a string $s_1 s_2 \cdots \overline{s_j}\ \overline{s_{j-1}} \cdots \overline{s_{i+1}}\ \overline{s_i}\ s_{j+1} \cdots s_{n-1} s_n$ where this segment is substituted by its reverse complement. Below we illustrate that reversals are represented as usual alternating blue-red cycles in the condensed double-stranded breakpoint and de Bruijn graphs.

Given two genomes G_A and G_B, we classify each *k*-mer *s* into 3 classes: *A* (if either *s* or $\overline{s}$ belongs to G_A but neither *s* or $\overline{s}$ belongs to G_B), *B* (if either *s* or $\overline{s}$ belongs to G_B but neither *s* or $\overline{s}$ belongs to G_A) and *AB* (if both G_A and G_B contain either *S* or $\overline{S}$).

The *double-stranded breakpoint graph BP**(G_A, G_B, *k*) is simply coloring the edges in *BP** ($G_A \cup G_B$, *k*) into 3 colors: *A* (blue), *B* (red) and *AB* (green) (Figure 11). Similarly, the *double-stranded de Bruijn graph DB**(G_A, G_B, *k*) is simply coloring the edges in *DB**($G_A \cup G_B$, *k*) into 3 colors: *A* (blue), *B* (red) and *AB* (green) (Figure 12).

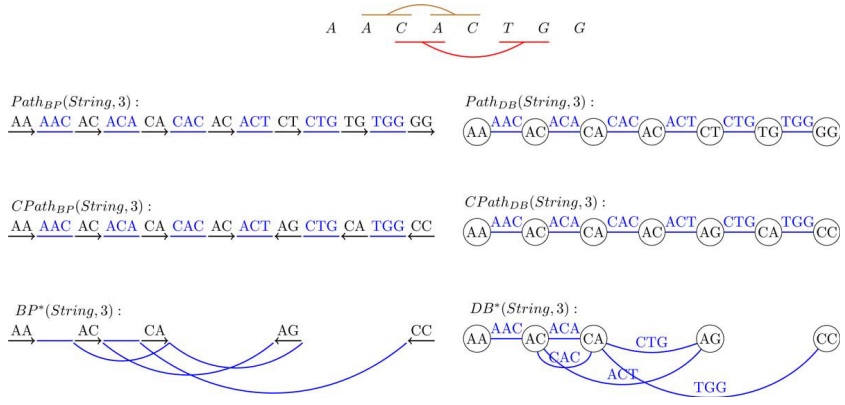


Figure 10 A string *String* = *ACAGTCA* (top) represented by canonical paths $CPath_{BP}$ (*String*, 3) and $CPath_{DB}$ (*String*, 3), the double-stranded breakpoint graph *BP(*String*, 3) (left) and the double-stranded de Bruijn graph *DB**(*String*, 3) (right)**. A pair of repeats in *String* is shown in brown and a pair of inverted repeats is shown in red.

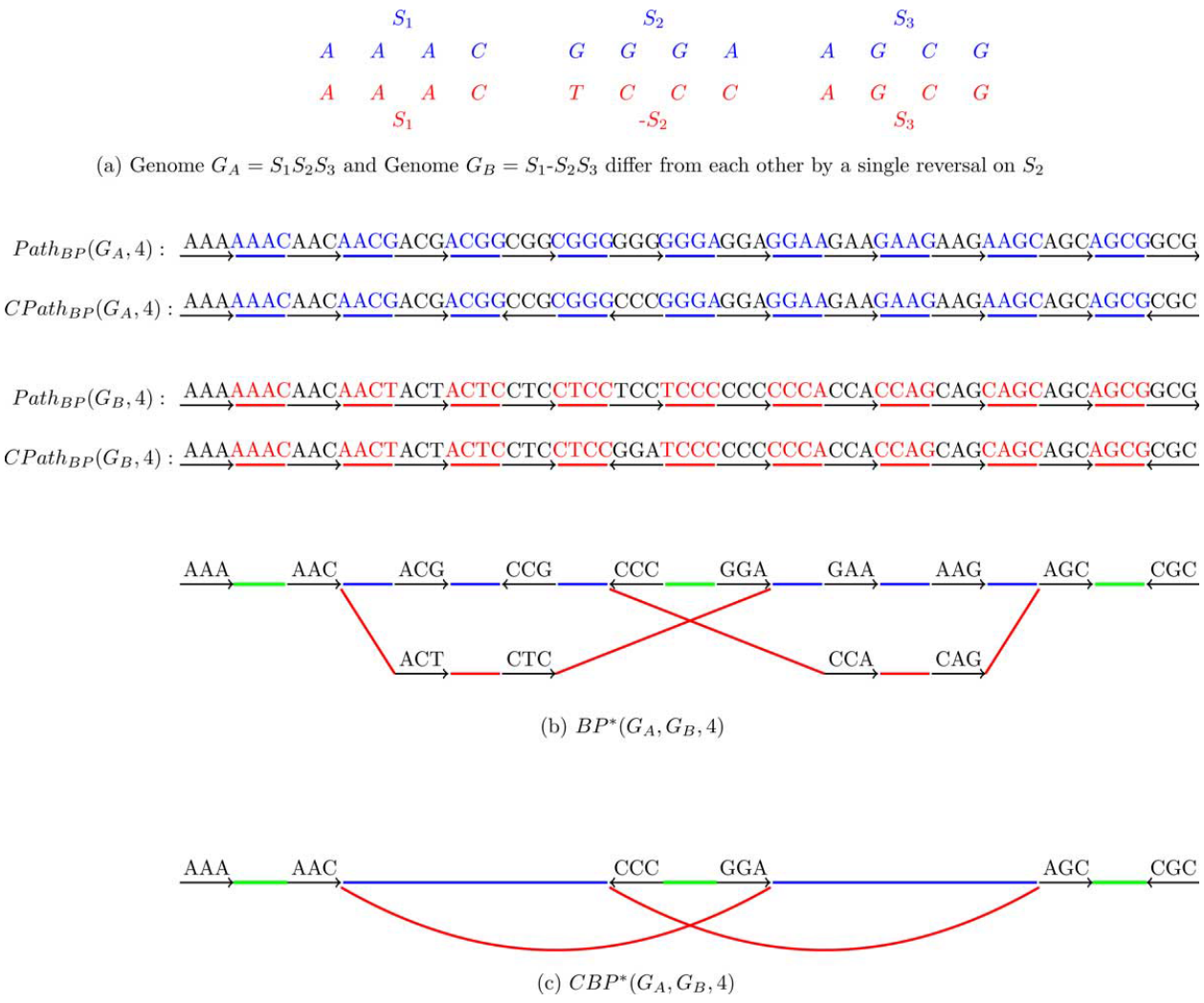


Figure 11 The double-stranded breakpoint graph *BP*(G_A, G_B, 4)* and the condensed double-stranded breakpoint graph *CBP*(G_A, G_B, 4)* of two genomes G_A and G_B. Each of 3 synteny blocks in both BP*(G_A, G_B, 4) and *CBP** (G_A, G_B, 4) is represented as a path (starting and ending in black edges and having a green edge in the middle).

Both $BP^*(G_A, G_B, k)$ and $DB^*(G_A, G_B, k)$ can be further condensed as described in Table 2 resulting in the condensed double-stranded breakpoint graph $CBP^*(G_A, G_B, k)$ and the *condensed double-stranded de Bruijn graph* $CDB^*(G_A, G_B, k)$, respectively (Figure 11(c) and Figure 12(c)).

As before, $BP^*(G_A, G_B, k)$ ($CBP^*(G_A, G_B, k)$) is obtained from $DB^*(G_A, G_B, k)$ ($CDB^*(G_A, G_B, k)$) by collapsing all black edges, while $DB^*(G_A, G_B, k)$ (CDB^* (G_A, G_B, k)) is obtained from $BP^*(G_A, G_B, k)$ ($CBP^*(G_A, G_B, k)$) by expanding all vertices into black edges (and connecting black edges according to the labels on the colored edges).

While the above notions of the double-stranded breakpoint graph and the double-stranded de Bruijn graph were defined for 2 genomes, they naturally generalize to any number of genomes.

Conclusion

We described the connection between the breakpoint graph and the de Bruijn graph that reveals that these constructions (that have been treated as two different data structures for over two decades) are essentially identical. We believe that the explicit description of this connection will contribute to a dialog between two previously separated bioinformatics communities studying genome rearrangements and genome assembly. It may also clarify the connection between the breakpoint graph, the de Bruijn graph, and the string graph introduced by Myers [30], another powerful paradigm for genome assembly. As hinted by Pop [31], the string graphs are functionally equivalent to the de Bruijn graphs, e.g., the comparison of Figure 1 in [30] and Figure 2 in [13]) suggests that the string graph is a special case of the ABruijn graph.

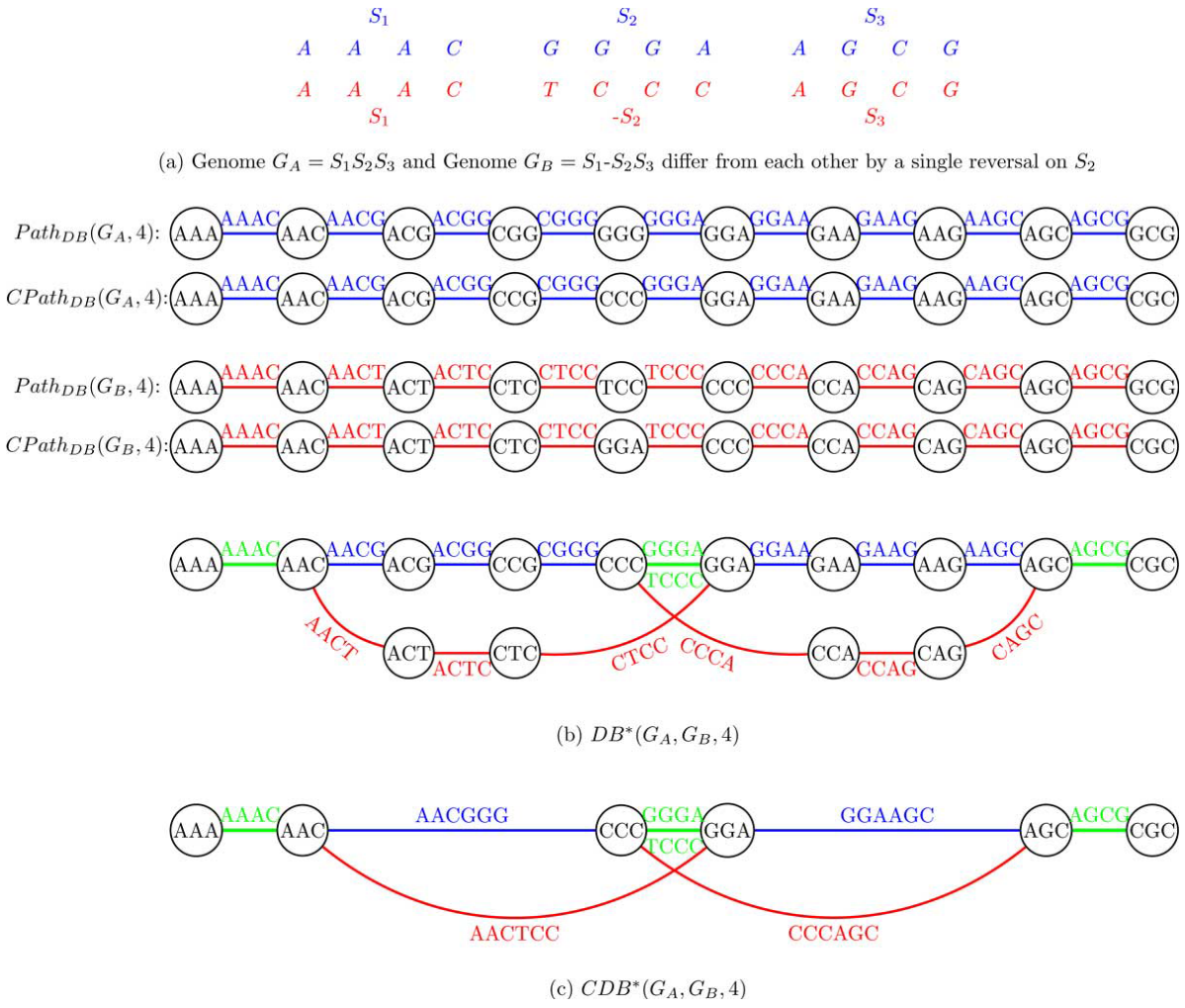


Figure 12 The double-stranded de Bruijn graph *DB*(G_A, G_B, 4)* and the condensed double-stranded de Bruijn graph *CDB*(G_A, G_B, 4)* of genomes G_A and G_B.

Simpson and Durbin [32] further suggested that the de Bruijn graph and the string graph constructions on all the k-mers of a genome (using parameter $\tau = k - 1$ for the string graph) are equivalent. However, the explicit description of this equivalence is still missing and we hope that the proposed A-Bruijn graph framework can be further extended to cover the string graphs as well.

Competing interests
The authors declare that they have no competing interests.

Authors' contributions
PAP conceived the study. YL, SN and PAP performed the analysis. YL and PAP wrote the paper with help from SN. All authors read and approved the final manuscript.

Acknowledgements
YL was supported by the Swiss National Science Foundation (grant no. 146708). SN and PAP were supported by the Government of the Russian Federation (grant no. 11.G34.31.0018).

Declarations
Publication costs for this article were funded by St. Petersburg Academic University.
This article has been published as part of *BMC Genomics* Volume 15 Supplement 6, 2014: Proceedings of the Twelfth Annual Research in Computational Molecular Biology (RECOMB) Satellite Workshop on Comparative Genomics. The full contents of the supplement are available online at http://www.biomedcentral.com/bmcgenomics/supplements/15/S6.

Authors' details
[1]Department of Computer Science and Engineering, University of California, San Diego, 9500 Gilman Dr, CA 92093 La Jolla, USA. [2]Algorithmic Biology Laboratory, St. Petersburg Academic University, St. Petersburg, Russia. [3]St. Petersburg State University, St. Petersburg, Russia.

Published: 17 October 2014

References
1. Pevzner PA: **l-tuple DNA sequencing: computer analysis.** *J Biomol Struct Dyn* 1989, **7**:63-73.
2. Idury RM, Waterman MS: **A new algorithm for DNA sequence assembly.** *J Comput Biol* 1995, **2(2)**:291-306.

3. Pevzner PA, Tang H, Waterman MS: **An Eulerian path approach to DNA fragment assembly.** *Proc Nat'l Acad Sci USA* 2001, **98(17)**:9748.
4. Zerbino DR, Birney E: **Velvet: algorithms for de novo short read assembly using de Bruijn graphs.** *Genome Research* 2008, **18(5)**:821-829.
5. Chaisson MJ, Pevzner PA: **Short read fragment assembly of bacterial genomes.** *Genome Research* 2008, **18(2)**:324-330.
6. Simpson JT, Wong K, Jackman SD, Schein JE, Jones SJM, Birol I: **Abyss: a parallel assembler for short read sequence data.** *Genome Research* 2009, **19(6)**:1117-1123.
7. Peng Y, Leung H, Yiu S, Chin F: **IDBA - a practical iterative de Bruijn graph de novo assembler.** *Proc 14th Int'l Conf Comput Mol Biol (RECOMB'10) Lecture Notes in Comp Sci* 2010, **6044**:426-440.
8. Butler J, MacCallum I, Kleber M, *et al*: **ALLPATHS: de novo assembly of whole-genome shotgun microreads.** *Genome Research* 2008, **18(5)**:810-820.
9. Boisvert S, Laviolette F, Corbeil J: **Ray: simultaneous assembly of reads from a mix of high-throughput sequencing technologies.** *J Comput Biol* 2010, **17(11)**:1519-1533.
10. Li R, Zhu H, Ruan J, *et al*: **De novo assembly of human genomes with massively parallel short read sequencing.** *Genome Research* 2010, **20(2)**:265-272.
11. Chitsaz H, Yee-Greenbaum JL, Tesler G, *et al*: **Efficient de novo assembly of single-cell bacterial genomes from short-read data sets.** *Nature biotechnology* 2011.
12. Bankevich A, Nurk S, *et al*: **SPAdes: A new genome assembly algorithm and its applications to single-cell sequencing.** *J Comput Biol* 2012, **19(5)**:455-477.
13. Pevzner PA, Tang H, Tesler G: **De novo repeat classification and fragment assembly.** *Genome Research* 2004, **14(9)**:1786-1796.
14. Böcker S: **Sequencing from compomers: Using mass spectrometry for dna de novo sequencing of 200+ nt.** *J Comput Biol* 2004, **11(6)**:1110-1134.
15. Pham SK, Pevzner PA: **DRIMM-Synteny: decomposing genomes into evolutionary conserved segments.** *Bioinformatics* 2010, **26(20)**:2509-2516.
16. Minkin I, Patel A, Kolmogorov M, Vyahhi N, Pham S: **Sibelia: a scalable and comprehensive synteny block generation tool for closely related microbial genomes.** *Proc 13th Workshop Algs in Bioinf (WABI'13) Lecture Notes in Comp Sci* 2013, **8126**:215-229.
17. Raphael B, Zhi D, Tang H, Pevzner PA: **A novel method for multiple alignment of sequences with repeated and shuffled elements.** *Genome Research* 2004, **14(11)**:2336-2346.
18. Bafna V, Pevzner PA: **Genome rearrangements and sorting by reversals.** *Proc 34th Ann IEEE Symp Foundations of Comput Sci (FOCS'93)* 1993, 148-157.
19. Fertin G, Labarre A, Rusu I, Tannier E, Vialette S: **Combinatorics of Genome Rearrangements.** MIT Press, Inc.
20. Iqbal Z, Caccamo M, Turner I, Flicek P, McVean G: **De novo assembly and genotyping of variants using colored de bruijn graphs.** *Nature genetics* 2012, **44(2)**:226-232.
21. Boussau B, Daubin V: **Genomes as documents of evolutionary history.** *Trends in ecology & evolution* 2010, **25(4)**:224-232.
22. Bailey JA, Baertsch R, Kent WJ, Haussler D, Eichler EE: **Hotspots of mammalian chromosomal evolution.** *Genome Biology* 2004, **5(4)**:23.
23. Zhao H, Bourque G: **Recovering genome rearrangements in the mammalian phylogeny.** *Genome Research* 2009, **19(5)**:934-942.
24. Alekseyev MA, Pevzner PA: **Comparative genomics reveals birth and death of fragile regions in mammalian evolution.** *Genome Biology* 2010, **11(11)**:117.
25. Nurk S, Pevzner PA: **Sparcle: using colored de bruijn graphs for analysing genome variations, unpublished manuscript.**.
26. Guzman GI, Utrilla J, Monk JM, Brunk E, Ebrahim A, Nurk S, Palsson BO, Feist AM: **Model-driven discovery of 'underground' isozyme functions in escherichia coli, unpublished manuscript.**.
27. Alekseyev MA, Pevzner PA: **Breakpoint graphs and ancestral genome reconstructions.** *Genome Research* 2009, **19(5)**:943-957.
28. Compeau PEC, Pevzner PA: **Bioinformatics Algorithms: An Active-Learning Approach**.
29. Medvedev P, Georgiou K, Myers G, Brudno M: **Computability of models for sequence assembly.** *Proc 7th Workshop Algs in Bioinf (WABI'07) Lecture Notes in Comp Sci* 2007, **4645**:289-301.
30. Myers EW: **The fragment assembly string graph.** *Bioinformatics* 2005, **21(suppl 2)**:79-85.
31. Pop M: **Genome assembly reborn: recent computational challenges.** *Briefings in bioinformatics* 2009, **10(4)**:354-366.
32. Simpson JT, Durbin R: **Efficient construction of an assembly string graph using the fm-index.** *Bioinformatics* 2010, **26(12)**:367-373.
33. Pevzner PA, Tesler G: **Genome rearrangements in mammalian evolution: lessons from human and mouse genomes.** *Genome Research* 2003, **13(1)**:37-45.

doi:10.1186/1471-2164-15-S6-S6
Cite this article as: Lin *et al.*: What is the difference between the breakpoint graph and the de Bruijn graph? *BMC Genomics* 2014 **15**(Suppl 6):S6.