



Genetic Determinants of Glycated Hemoglobin in Type 1 Diabetes

Anna Syreeni,^{1,2,3} Niina Sandholm,^{1,2,3} Jingjing Cao,⁴ Iiro Toppila,^{1,2,3} David M. Maahs,⁵ Marian J. Rewers,⁶ Janet K. Snell-Bergeon,⁶ Tina Costacou,⁷ Trevor J. Orchard,⁷ M. Luiza Caramori,⁸ Michael Mauer,⁹ Barbara E.K. Klein,¹⁰ Ronald Klein,¹⁰ Erkka Valo,^{1,2,3} Maija Parkkonen,^{1,2,3} Carol Forsblom,^{1,2,3} Valma Harjutsalo,^{1,2,3,11} Andrew D. Paterson,⁴ for the DCCT/EDIC Research Group,* and Per-Henrik Groop,^{1,2,3,12} on behalf of the FinnDiane Study Group*

Diabetes 2019;68:858–867 | <https://doi.org/10.2337/db18-0573>

Glycated hemoglobin (HbA_{1c}) is an important measure of glycemia in diabetes. HbA_{1c} is influenced by environmental and genetic factors both in people with and in people without diabetes. We performed a genome-wide association study (GWAS) for HbA_{1c} in a Finnish type 1 diabetes (T1D) cohort, FinnDiane. Top results were examined for replication in T1D cohorts DCCT/EDIC, WESDR, CACTI, EDC, and RASS, and a meta-analysis was performed. Three SNPs in high linkage disequilibrium on chromosome 13 near relaxin family peptide receptor 2 (*RXFP2*) were associated with HbA_{1c} in FinnDiane at genome-wide significance ($P < 5 \times 10^{-8}$). The minor alleles of rs2085277 and rs1360072 were associated with higher HbA_{1c} also in the meta-analysis with RASS ($P < 5 \times 10^{-8}$), where these variants had minor allele frequencies $\geq 1\%$. Furthermore, these SNPs were associated with HbA_{1c} in an East Asian population without diabetes ($P \leq 0.013$). A weighted genetic risk score created from 55 HbA_{1c}-associated variants from the literature was associated with HbA_{1c} in FinnDiane but explained only a small amount of variation. Understanding

the genetic basis of glycemic control and HbA_{1c} may lead to better prevention of diabetes complications.

Glycated hemoglobin (HbA_{1c}) reflects the level of blood glycemia and is commonly used to monitor glycemic control in diabetes and to diagnose type 2 diabetes. Genome-wide association studies (GWAS) have revealed >60 genetic variants to be associated with HbA_{1c} in individuals without diabetes (1–6). A recent GWAS of multiple ethnic groups confirmed 18 variants previously associated with HbA_{1c} and added 42 new loci (6). Some of the variants affect biological pathways maintaining glucose homeostasis. For example, single nucleotide polymorphisms (SNPs) in or near glucokinase (*GCK*) and melatonin receptor 1B (*MTNR1B*) are known to be associated with HbA_{1c}, fasting plasma glucose, and β -cell function (3). HbA_{1c} is further influenced by genes and pathways that affect erythrocytes. In particular, SNPs in hemochromatosis (*HFE*) and hexokinase 1 (*HK1*) (3) have an effect on HbA_{1c}

¹Folkhälsan Institute of Genetics, Folkhälsan Research Center, Helsinki, Finland

²Abdominal Center, Nephrology, University of Helsinki and Helsinki University Hospital, Helsinki, Finland

³Research Program for Clinical and Molecular Metabolism, Faculty of Medicine, University of Helsinki, Helsinki, Finland

⁴Program in Genetics and Genome Biology, The Hospital for Sick Children, Toronto, Ontario, Canada

⁵Division of Endocrinology and Diabetes, Stanford Diabetes Research Center, Stanford University School of Medicine, Stanford, CA

⁶Barbara Davis Center for Childhood Diabetes, University of Colorado Anschutz Medical Campus, Aurora, CO

⁷Department of Epidemiology, Graduate School of Public Health, University of Pittsburgh, Pittsburgh, PA

⁸Division of Diabetes, Endocrinology and Metabolism, Department of Medicine, University of Minnesota, Minneapolis, MN

⁹Departments of Pediatrics and Medicine, University of Minnesota, Minneapolis, MN

¹⁰Department of Ophthalmology and Visual Sciences, University of Wisconsin, Madison, WI

¹¹Chronic Disease Prevention Unit, National Institute for Health and Welfare, Helsinki, Finland

¹²Department of Diabetes, Central Clinical School, Monash University, Melbourne, Victoria, Australia

Corresponding author: Per-Henrik Groop, per-henrik.groop@helsinki.fi

Received 30 May 2018 and accepted 14 January 2019

This article contains Supplementary Data online at <http://diabetes.diabetesjournals.org/lookup/suppl/doi:10.2337/db18-0573/-/DC1>.

*A complete list of the DCCT/EDIC Research Group and the FinnDiane Study Group can be found in the Supplementary Data online.

© 2019 by the American Diabetes Association. Readers may use this article as long as the work is properly cited, the use is educational and not for profit, and the work is not altered. More information is available at <http://www.diabetesjournals.org/content/license>.

through nonglycemic pathways that affect erythrocyte physiology and survival.

HbA_{1c} reflects the glycemic load over ~3 months (7), and traits such as hemoglobinopathies, alterations in intracellular glucose metabolism, or defects of glucose transport into erythrocytes affect HbA_{1c}. In addition, diet, drugs, and insulin administration affect HbA_{1c} in individuals with type 1 diabetes (T1D) who have little to no endogenous insulin production and, thus, rely on an external source of insulin to ensure glucose uptake in cells. Despite a strong environmental stimulus, HbA_{1c} levels are correlated between both dizygotic twins and monozygotic twins concordant or discordant for T1D (8). Heritability estimates for HbA_{1c} in individuals without diabetes range from 27% to 62% (8,9), thus providing evidence for a genetic component for the trait.

To date, one GWAS on HbA_{1c} has been conducted in individuals with T1D in the Diabetes Control and Complications Trial (DCCT) (10). The authors found that rs1358030 near sortilin-related VPS10 domain-containing receptor 1 (*SORCS1*) was associated with HbA_{1c} ($P < 2.2 \times 10^{-9}$). The current study aims to define more genetic variants associated with HbA_{1c} using GWAS of 4,622 individuals with T1D from the Finnish Diabetic Nephropathy (FinnDiane) Study and attempts to replicate GWAS results in five additional cohorts including, altogether, 3,097 individuals with T1D.

RESEARCH DESIGN AND METHODS

Phenotypes

HbA_{1c}

FinnDiane is a nationwide, comprehensive multicenter study launched in 1997. Local ethics committees approved the study protocol, and participants gave informed consent before participation. The study was performed in accordance with the Declaration of Helsinki. HbA_{1c} was measured with standardized laboratory methods locally from a blood sample taken during the visits to the FinnDiane study centers. Additional HbA_{1c} values were collected from laboratory records both before and after the FinnDiane visit. A shift from high-performance liquid chromatography method to immunoassays was seen in many laboratories during the HbA_{1c} collection time (1 January 1991–1 June 2016). In Finland, the HbA_{1c} measurements show a high correlation with the DCCT reference method (11). HbA_{1c} measurements from <1 year of diabetes diagnosis and the measurements after end-stage renal disease (ESRD) diagnosis were excluded. A median of 19 HbA_{1c} measurements per person (range 1–129) were collected (Supplementary Fig. 1). Phenotype in the GWAS was an intrapersonal mean HbA_{1c}, and it roughly followed normal distribution (Supplementary Fig. 2). In the replication cohorts, a mean of natural log-transformed HbA_{1c} was used. Similar phenotypic values were subsequently calculated in FinnDiane to enable comparison of effect sizes (β) in the meta-analysis.

Clinical Parameters

At the FinnDiane visit, each patient underwent a thorough clinical investigation as previously described (12). The GWAS covariates age and diabetes duration were calculated at the intrapersonal mean date of HbA_{1c} measurements. Micro- or macroalbuminuria was defined based on the urinary albumin excretion rate (AER) in two out of three consecutive timed overnight or 24-h urine collections as previously described (12). ESRD was defined as ongoing dialysis or the patient having received a kidney transplant. ESRD diagnosis year was verified from hospital records. Hematological measurements were taken from whole blood samples of 429 individuals using flow cytometry in the Helsinki University Central Hospital Laboratory.

Statistical Methods and Analyses

Patient Selection

Individuals with age at diabetes onset of <40 years, insulin treatment initiated within 1 year of the diabetes diagnosis, and one or more HbA_{1c} measurements were selected from FinnDiane. Relatedness of individuals was calculated with KING 1.40 (13). Among first-degree family members, the individual with the least number of HbA_{1c} measurements ($n\text{HbA}_{1c}$) was excluded, and altogether 4,622 individuals were included in the GWAS.

After also excluding more distantly related individuals according to the estimated genetic relationship matrix (≥ 0.05), we calculated heritability in 4,319 individuals. Narrow-sense heritability of HbA_{1c} (h^2) was the proportion of the phenotypic variance explained by the additive effects of the genotyped SNPs with or without covariates sex, age, duration, genotyping batch, and $n\text{HbA}_{1c}$ (GCTA, version 1.24.4) (14).

Power Calculations

Power calculations were performed with Quanto, version 1.2.4 (<http://biostats.usc.edu/Quanto.html>), using α -level of 5×10^{-8} (genome-wide significance). In the GWAS, we had 94% power to detect variants with a minor allele frequency (MAF) $\geq 5\%$ and an effect size (β) 0.40 (non-transformed HbA_{1c} [%]) [Supplementary Table 1]. In the replication cohort ($n = 3,097$), we had >57% and 99% power (two-sided $\alpha = 0.05$, $\beta = 0.30$) to replicate variants with MAFs 1–5% or MAFs $\geq 5\%$, respectively.

Genotyping and GWAS

SNP genotyping was performed with HumanCoreExome-12 v. 1.0, -12 v. 1.1, or -24 v. 1.0 BeadChip (Illumina, San Diego, CA). SNPs with low genotyping call rate, deviation from Hardy-Weinberg equilibrium, MAF <1%, Mendelian inconsistency, allele frequency difference >20% (MAF $\geq 5\%$), or allele frequency difference >5% (MAF <5%) with 1000 Genomes European (EUR) population were filtered.

After quality control, phased haplotypes (SHAPEIT v2.r837) (15) of 316,899 SNPs were used for imputation with 1000 Genomes EUR phase 3, version 5 (16), using

Minimac3/Minimac3-omp v1.0.14 (17). A total of 6,019 individuals passed quality control when samples with genotyping rate <0.95 or extreme heterozygosity, sample mix-ups, and genetic outliers were removed.

GWAS was performed with dosages of 8,354,674 variants with imputation quality $r^2 \geq 0.7$ with PLINK v1.9b3.26 (18) using linear regression with covariates sex, age, diabetes duration, $n\text{HbA}_{1c}$, genotyping batch, and 10 first principal components that were calculated with the EIGENSTRAT, version 3.0 (19). A genomic inflation factor λ was 1.003 (Supplementary Fig. 3).

Altogether, 1,489 and 1,863 individuals were genotyped with the TaqMan technology and ABI Prism 7000/7900HT Sequence Detection Systems and SDS 2.3 software (Life Technologies, Foster City, CA) to verify the imputed genotypes of rs2085277 and rs17076364, respectively. Of the TaqMan-genotyped individuals, 1,259 of 1,489 (84.6%) for rs2085277 and 1,545 of 1,863 (82.9%) for rs17076364 were included in the HbA_{1c} GWAS.

Longitudinal HbA_{1c}

Longitudinal HbA_{1c} data were studied with linear mixed models (nlme package in R, version 3.3.3) in 4,153 individuals with three or more HbA_{1c} measurements. The top SNPs were analyzed with continuous time including diabetes duration (at first HbA_{1c}), sex, age, genotyping batch, and 10 first principal components as covariates assuming random intercept and slope. The best model (lowest Akaike information criterion) had an autocorrelation structure of order 1 with continuous time covariate and a combined covariance structure with exponential variances for fitted model values and separate variances for HbA_{1c} measurements from different decades of diabetes duration.

Phenotypic Comparisons

Groups were compared with Student *t* test, Welch *t* test, or Mann-Whitey *U* test in R. Histograms were used for evaluating normality. Linear regression was used to study phenotypic associations with covariates.

Replication

T1D Cohorts. Replication of the top SNPs was conducted in DCCT/Epidemiology of Diabetes Interventions and Complications (EDIC), the Wisconsin Epidemiologic Study of Diabetic Retinopathy (WESDR), the Coronary Artery Calcification in Type 1 Diabetes (CACTI) study, the Pittsburgh Epidemiology of Diabetes Complications (EDC) study, and the Renin Angiotensin System Study (RASS) comprising altogether 3,097 individuals with T1D (Fig. 1 and Supplementary Table 2). Individuals with conventional ($n = 667$) or intensive ($n = 637$) treatment of diabetes during DCCT were analyzed separately. CACTI was also divided (clinic, $n = 265$, and nonclinic, $n = 253$). SNPs with imputation quality $r^2 \geq 0.6$ and MAF $\geq 1\%$ in the individual replication cohorts were included in the meta-analysis. These SNPs were replicated in one to seven cohorts comprising 253–3,097 individuals. A mean of natural log-transformed HbA_{1c} was the phenotype used in the replication cohorts, and to get comparable effect

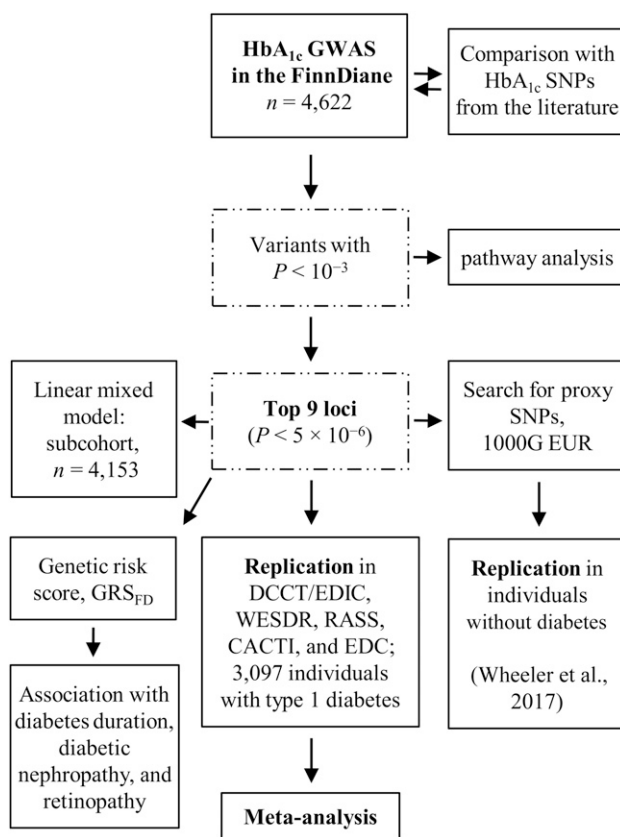


Figure 1—Outline of the GWAS for HbA_{1c} in the FinnDiane cohort. 1000G, 1000 Genomes.

sizes (β) the association of the top SNPs and similarly transformed HbA_{1c} was calculated in the discovery cohort. Thereafter, a fixed-effects meta-analysis based on β s and SEs was performed with GWAMA (20).

Populations Without Diabetes and Literature Search.

The top nine independent variants were compared with publicly available data from GWAS meta-analyses of up to 123,517 individuals without diabetes of European ancestry (6). For variants not included in the GWAS by Wheeler et al. (6), we searched for proxy variants ($r^2 \geq 0.3$) in the 1000 Genomes EUR population using LDlink (21) (<https://analysistools.nci.nih.gov/LDlink>). Seven out of nine SNPs or proxy variants had HbA_{1c} association data available. In addition, replication of three genome-wide significant SNPs on chromosome 13 was performed in the GWAS data from individuals with East Asian ancestry ($n = 23,461$), since these SNPs were not available in the Europeans, likely due to their low MAFs. Thereafter, we compared 86 previously reported variants for HbA_{1c} with our GWAS results: 11 variants from the DCCT GWAS (10) and 75 variants from GWAS of individuals without diabetes.

Genetic Risk Scores

Genome-wide significant autosomal SNPs in the largest HbA_{1c} GWAS (6), which had MAF $\geq 1\%$ and $P < 0.01$

also in the Europeans, were selected for genetic risk score (GRS). GRS-ALL comprised all 55 SNPs, and like Wheeler et al. (6), we calculated separate GRS for erythrocytic SNPs ($n = 19$) (GRS-E) and glycemic SNPs ($n = 18$) (GRS-G). GRS were the sum of expected number of risk alleles weighted by the effect size (β) of the SNP in the Europeans, multiplied by the number of SNPs and divided by the sum of β s. Association of the GRS with HbA_{1c} was studied using linear regression with covariates age, sex, diabetes duration, and n HbA_{1c}.

After the FinnDiane GWAS, a weighted GRS, GRS_{FD}, was created from nine noncorrelated SNPs showing significant or suggestive association with HbA_{1c} ($P < 5 \times 10^{-6}$). GRS_{FD} was tested for associations with diabetes duration, diabetic nephropathy (DN), and laser-treated diabetic retinopathy.

In Silico Analyses

The closest genes were searched (Ensembl transcripts downloaded from <https://genome.ucsc.edu/cgi-bin/hgTables> [July 2017]) for variants with $P < 0.001$ in the GWAS. A total of 9,742 SNPs were mapped to 4,470 unique transcripts and were then connected with the HUGO Gene IDs. A total of 1,175 genes were located <150 kb from the GWAS SNPs. PANTHER database (22) (<http://www.pantherdb.org>) was used to identify the underlying biological pathways, and STRING, version 10.5 (<https://string-db.org>), was used to study protein interactions.

For the nearest genes of the top variants, gene expression (Expression Atlas [<http://www.ebi.ac.uk/gxa/home>]) and protein expression (ProteomicsDB [<https://www.proteomicsdb.org>]) were studied. The ENCODE data were accessed through UCSC Genome Browser (<http://www.genome.ucsc.edu>) and RegulomeDB v1.1 (23) (<http://regulome.stanford.edu/> [July 2017]). Expression quantitative trait loci (eQTL) of the top variants were studied using the Genotype-Tissue Expression project portal (GTEx V7) (<https://www.gtexportal.org/home/>). PhenoScanner V2 database (24) (<http://www.phenoscaner.medschl.cam.ac.uk/phenoscaner>) was used to find associations with metabolites.

RESULTS

Mean \pm SD HbA_{1c} was $8.5 \pm 1.2\%$ (69.5 ± 13.1 mmol/mol) in FinnDiane (Table 1). Narrow-sense heritability of HbA_{1c} was 17.0% ($P = 0.002$) (unadjusted) or 15.3% ($P = 0.003$) when adjusted for covariates. Minor alleles of three SNPs on chromosome 13q12.3 showed genome-wide significant association with higher HbA_{1c} values in the GWAS (lead SNP rs2085277: $\beta = 0.42$ [95% CI 0.27, 0.56], $P = 1.52 \times 10^{-8}$) (Fig. 2). The three SNPs rs2085277, rs1360072, and rs17076364 are in high linkage disequilibrium (LD) in the 1000 Genomes EUR population and are located between β 3-glucosyltransferase (*B3GLCT*) (SNPs 232–261 kb downstream) and 146–175 kb upstream of the

Table 1—Baseline demographics of the FinnDiane cohort

<i>n</i>	4,622
Sex (% male)	51.0
Age (years)	38.8 \pm 12.4
HbA _{1c} , % (mmol/mol)	8.5 \pm 1.2 (69.5 \pm 13.1)
Age at diabetes diagnosis (years)	13.9 (8.9–21.6)
Duration of diabetes (years)	21.9 (12.4–31.0)
Number of HbA _{1c} measurements	19 (9–32)
HbA _{1c} collection time (years)*	25.4 (22.4–25.4)
HbA _{1c} measurements (n/year)	0.9 (0.4–1.4)
HbA _{1c} variability, CV (%)	8.9 (6.8–11.9)
BMI (kg/m ²)	25.1 \pm 3.7
Insulin (IU)	50.7 \pm 21.3
Insulin dose (IU/kg)	0.70 \pm 0.26
SBP (mmHg)	134 \pm 19
DBP (mmHg)	79 \pm 10
Triglycerides (mmol/L)	1.02 (0.76–1.45)
DN status, <i>n</i> (%)	
Normal AER	2,794 (60.5)
Microalbuminuria	610 (13.2)
Macroalbuminuria	746 (16.1)
ESRD	227 (4.9)
Undefined	245 (5.3)
Retinopathy, <i>n</i> (%)†	1,530 (34.6)
Hard event, <i>n</i> (%)	447 (9.8)

Data are mean \pm SD or median (interquartile range) unless otherwise indicated. Coefficient of variation (CV) is defined as $CV = SD/mean \times 100$ and calculated from T1D subjects with three or more HbA_{1c} measurements. Cardiovascular hard events were myocardial infarction, cardiac bypass surgery, stroke, peripheral vascular disease, or amputation at baseline. HbA_{1c} measurements between 1991 or 1 year after diabetes diagnosis year and 1 June 2016 or before the ESRD year were collected. DBP, diastolic blood pressure; SBP, systolic blood pressure. *HbA_{1c} collection time is calculated as years between the beginning of the HbA_{1c} collection (1 January 1991 or 1 year after diabetes diagnosis year) and the end of the collection (1 June 2016 or ESRD year). †Laser-treated diabetic retinopathy.

closest gene, relaxin family peptide receptor 2 (*RXFP2*) (Supplementary Fig. 4).

Additional genotyping confirmed the imputed rs2085277 and rs17076364 genotypes (Supplementary Table 3). The lead SNP rs2085277 was not significantly associated with insulin dose (IU/kg), coefficient of variation of HbA_{1c} (Supplementary Table 4), or hematocrit or other hematological parameters that were measured in a small subcohort of 429 individuals with T1D (Supplementary Table 5). Altogether, 68 variants had $P < 5 \times 10^{-6}$ in the GWAS. Of these, five tightly correlated SNPs from chr13 locus and eight other independent SNPs with $P < 5 \times 10^{-6}$ were selected for replication and subsequent analyses. Notably, these variants were associated with HbA_{1c} also when longitudinal HbA_{1c} measurements were analyzed in a subcohort of 4,153 individuals with three or more HbA_{1c} measurements (Supplementary Table 6).

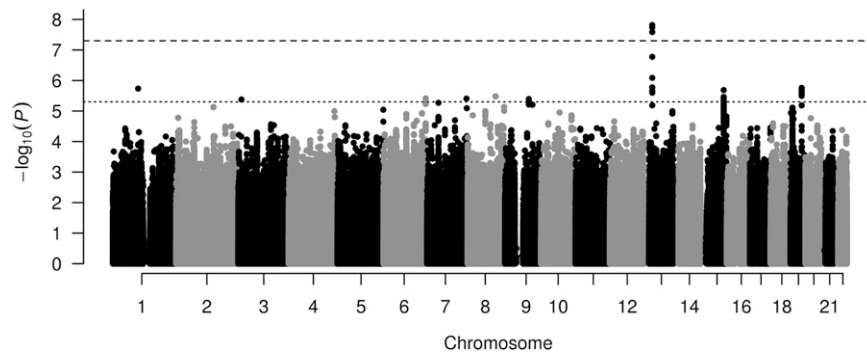


Figure 2—Manhattan plot of the FinnDiane HbA_{1c} GWAS results shows genome-wide significant ($P < 5.0 \times 10^{-8}$) variants on chromosome 13. The lead SNP is rs2085277 ($P = 1.52 \times 10^{-8}$). y-axis shows the $-\log_{10} P$ value, and x-axis tells the chromosomal position of the variants. The two horizontal lines denote the thresholds for genome-wide significant P value of 5×10^{-8} (upper dashed line) and for a suggestive association with P value of 5×10^{-6} (lower dotted line).

Replication

T1D Cohorts

The five T1D cohorts all used the mean of the natural log-transformed HbA_{1c} values as the end phenotype. Two SNPs showed significant replication with $\ln(\text{HbA}_{1c})$: rs373883321 in CACTI_{clinic} (chr6:165,403,975, $P = 0.006$) and rs10459356 near the lead SNP rs2085277 on chromosome 13 in RASS ($P = 0.042$) (Supplementary Table 7). In a meta-analysis with the discovery cohort, variants rs2085277 and rs1360072 in high LD on chromosome 13 were significantly associated with $\ln(\text{HbA}_{1c})$ with $\beta = 0.046$ (95% CI 0.030, 0.063) and $P = 3.7 \times 10^{-8}$ and $P = 4.2 \times 10^{-8}$, respectively (Table 2 and Supplementary Table 8).

Of note, the MAFs for rs2085277 and rs1360072 on chromosome 13 were 4% in FinnDiane and 1% in RASS but only 0.2%–0.5% in the DCCT/EDIC and WESDR cohorts that were thus not included in the meta-analysis for these SNPs (Supplementary Fig. 5). Because of low imputation quality of the chr13 top variants ($r^2 < 0.60$), CACTI and EDC cohorts were also excluded from the meta-analysis of these SNPs. Low MAFs reduced the statistical power to replicate the findings, but in RASS the effect was in the same direction as in FinnDiane: lead SNP rs2085277, $\beta = 0.059$ [95% CI $-0.079, 0.198$] for $\ln(\text{HbA}_{1c})$, $P = 0.402$. The MAF for rs2085277 is similar to our study in the 1000 Genomes populations: 5.6% in FIN (Finnish in Finland), 0.5% in GBR (British in England and Scotland),

Table 2—Top loci in the FinnDiane HbA_{1c} GWAS and results from replication in other T1D cohorts

Variant	Position	Closest gene	FinnDiane ($n = 4,622$)				T1D replication cohorts		Meta-analysis P
			EA/OA	EAF (%)	β (95% CI)*	P	n cohorts; subjects	EAF (%)	
rs7514675	1:99,782,478	LPPR4	G/A	10.1	0.23 (0.13, 0.33)	2.7×10^{-6}	7; 3,097	12.4	1.5×10^{-2}
rs144918527	3:12,740,810	RAF1	T/TTG	53.2	0.13 (0.08, 0.19)	4.4×10^{-6}	5; 2,248	51.2	2.5×10^{-4}
rs373883321	6:165,403,975	MEAT6	T/-	34.4	0.13 (0.07, 0.18)	3.9×10^{-6}	5; 2,248	36.8	2.1×10^{-2}
rs478414	7:154,526,318	DPP6	C/T	56.0	0.13 (0.07, 0.18)	3.1×10^{-6}	7; 3,097	50.5	8.3×10^{-3}
rs113241624	8:109,658,463	TMEM74	A/G	1.2	0.60 (0.35, 0.85)	3.2×10^{-6}	1; 596	1.0	8.5×10^{-5}
rs4744017	9:93,615,717	SYK	A/G	47.9	0.12 (0.07, 0.17)	8.8×10^{-6}	7; 3,097	46.8	8.3×10^{-4}
rs2085277	13:32,167,717	RXFP2	T/A	4.1	0.42 (0.27, 0.56)	1.5×10^{-8}	1; 253	1.0	3.7×10^{-8}
rs552894079	15:89,840,012	FANCI	A/G	97.6	0.40 (0.23, 0.56)	8.1×10^{-6}	NA	NA	NA
rs2599441	19:43,989,088	PHLDB3	G/A	72.8	0.14 (0.08, 0.19)	2.0×10^{-6}	7; 3,097	76.2	3.2×10^{-3}

EA, effect allele; EAF, effect allele frequency; OA, other allele. * β is the effect size for the mean HbA_{1c} phenotype in the discovery cohort. Replication cohorts are two DCCT/EDIC cohorts (conventional [$n = 667$] and intensive [$n = 637$] treatment of HbA_{1c} in the DCCT separately), WESDR ($n = 596$), RASS ($n = 253$), CACTI divided into two cohorts (nonclinic [$n = 253$] and clinic [$n = 265$]), and EDC ($n = 253$). Since the mean of natural log-transformed HbA_{1c} was the phenotype used in the T1D replication cohorts, top SNP association with $\ln(\text{HbA}_{1c})$ was further calculated in the discovery cohort to get comparable β s and SEs for the meta-analysis. Meta-analysis P value is from fixed-effects meta-analysis of FinnDiane and all replication cohorts restricted to variants with MAF $\geq 1\%$ in the individual replication cohorts. Results for the variants rs1360072 and rs17076364 on chromosome 13 locus were similar to the top variant, rs2085277: FinnDiane, $P = 1.8 \times 10^{-8}$ and $P = 2.6 \times 10^{-8}$, and meta-analysis, $P = 4.2 \times 10^{-8}$ and $P = 6.5 \times 10^{-8}$, respectively. rs113241624 on chromosome 8 had MAF $\geq 1\%$ only in WESDR and rs2085277 on chromosome 13 only in RASS. SNP rs552894079 was not available in the T1D replication cohorts.

and 0% (monomorphic) in the CEU (Northern Europeans from Utah). The variant showed a markedly higher frequency in East Asians and South Asians with MAFs of 38.5% and 9.2%, respectively.

Cohorts Without Diabetes

The association of the thirteen SNPs with HbA_{1c} was then queried in GWAS meta-analysis data on multiple cohorts without diabetes. As expected due to low MAF in Europeans, no data were available for the chr13 top SNPs or for any proxy SNPs with $r^2 \geq 0.3$ in the European cohorts. Therefore, replication of the chr13 variants was sought in the meta-analysis results of individuals of Asian ancestry. The SNPs were missing from the South Asian cohorts, but the chr13 top SNPs rs2085277 and rs1360072 were associated with HbA_{1c} (%) in East Asians with $\beta = 0.02$ (95% CI 0.006, 0.03), $P = 0.005$, and $\beta = 0.02$ (0.003, 0.03), $P = 0.013$, respectively. (Table 3). None of the other top SNPs or the proxy variants were associated with HbA_{1c} in the Europeans (Supplementary Table 9).

In Silico Analyses

Interestingly, minor T allele for rs2085277 was associated with higher levels of ketone bodies acetoacetate and 3-hydroxybutyrate (25) (PhenoScanner) ($P = 0.003$ and $P = 0.020$, respectively). No direct ENCODE data were available for rs2085277, and only minimal transcription factor-binding evidence (RegulomeDB scores 4–6) was seen for other top SNPs in the chr13 locus. Although none of the chr13 top variants were included in the GTEx database, many SNPs near rs2085277 affect the expression of *RXFP2* in adrenal gland, which suggests that rs2085277 might also affect *RXFP2* expression.

RXFP2 belongs to the positive regulation of cAMP biosynthetic process pathway (Gene Ontology term in the STRING protein network analysis [Fig. 3]). Interestingly, when the search was restricted to databases as opposed to all interaction sources (databases, text mining, experiments, coexpression, neighborhood, gene fusion, and co-occurrence),

gastric inhibitory polypeptide receptor (GIPR) and glucagon (GCG)—proteins relevant for glucose homeostasis and diabetes—occurred in the same protein interaction network with *RXFP2*.

Genes involved in cell-cell adhesion processes and developmental processes were overrepresented in GWAS results (PANTHER, $P < 0.05$, corrected for multiple testing [Supplementary Table 10]). PANTHER pathway analysis pointed out cadherin signaling, endogenous cannabinoid signaling, and Wnt signaling pathways ($P < 0.05$, corrected for 160 independent classes).

Duration of Diabetes and Chronic Diabetes Complications

Our GWAS top variants had a consistent effect on HbA_{1c} irrespective of diabetes duration. In all 10-year diabetes duration bins, the GRS_{FD} (Supplementary Table 11 and Supplementary Fig. 6) was significantly associated with HbA_{1c} (Supplementary Table 12). We confirmed this constant association between GRS_{FD} and HbA_{1c} in a small group of individuals ($n = 271$) having HbA_{1c} measurements from three consecutive diabetes duration bins: 1–9.9, 10–19.9, and 20–29.9 years.

Subjects with T1D with DN had higher mean HbA_{1c} compared with individuals with normal AER ($8.9 \pm 1.3\%$ vs. $8.2 \pm 1.0\%$, $P < 10^{-16}$). Similarly, individuals with diabetic retinopathy had higher HbA_{1c} compared with non-laser-treated subjects with at least 15 years of diabetes duration ($8.8 \pm 1.2\%$ vs. $8.4 \pm 1.2\%$, $P < 10^{-16}$). As expected, individuals with DN had higher GRS_{FD} (6.5 vs. 6.4, $P = 0.038$), while the subjects with diabetic retinopathy did not have higher GRS_{FD} ($P = 0.205$).

Previous HbA_{1c} Studies

rs1358030 was previously associated with HbA_{1c} in the GWAS of 1,307 individuals with T1D (10). However, this association was not replicated in FinnDiane, although the MAFs were similar (MAF_{DCCT} = 36% vs. MAF_{FinnDiane} = 40%), imputation quality was $r^2 = 0.99$, and the effect was

Table 3—Three genome-wide significant variants on chromosome 13: LD structures and HbA_{1c} association in a population without diabetes

Variant	EA < OA	MAF (%)*				r^2 †				HbA _{1c} association in a Finnish T1D cohort		HbA _{1c} association in an East Asian population without diabetes	
		FD	EAS	SAS	EUR	FD	EAS	SAS	EUR	β (95% CI)	P	β (95% CI)	P
rs2085277	T < A	4.2	38.5	9.2	1.2					0.42 (0.27, 0.56)	1.5×10^{-8}	0.02 (0.006, 0.03)	0.005
rs1360072	C < T	4.1	34.6	9.0	1.2	1.0	0.75	0.95	1.0	0.41 (0.27, 0.56)	1.8×10^{-8}	0.02 (0.003, 0.03)	0.013
rs17076364	T < C	3.1	2.5	0.2	1.2	0.97	0.04	0.02	1.0	0.45 (0.29, 0.61)	2.6×10^{-8}	NA	NA
<i>n</i>		4,622	504	503	489	4,622	504	503	489	4,622		23,461	

EA is the effect allele in the FinnDiane GWAS and OA is the other allele. *MAF frequency in the FinnDiane cohort (FD) and in East Asian (EAS), South Asian (SAS), and EUR populations in the 1000 Genomes phase 3 data. †LD (r^2) with rs2085277 in the FinnDiane and in the 1000 Genomes phase 3 populations. Position of the SNPs in the human genome build GRCh37 (hg19) was rs2085277, chr13:32,167,717; rs1360072, chr13:32,138,331; and rs17076364: chr13:32,163,399. HbA_{1c} association statistics for these variants were not available for the European population without diabetes ($n = 123,517$) or South Asian population without diabetes ($n = 9,908$) (Wheeler et al. [6]).

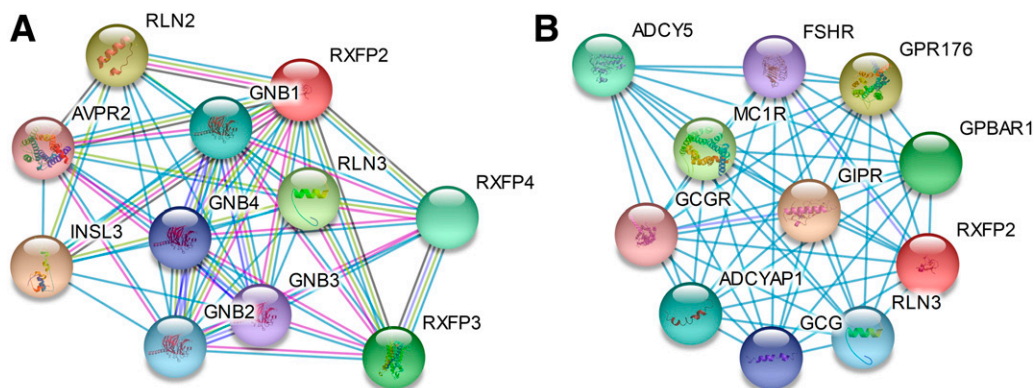


Figure 3—Protein networks around RXFP2 (red). **A:** Two most enriched Gene Ontology (GO) biological pathways were “cellular response to glucagon stimulus” (GNB1, GNB4, GNB2, and GNB3) and “positive regulation of cAMP biosynthetic process” (RXFP2, RLN2 [relaxin], AVPR2, and INSL3), with false discovery rates 4.6×10^{-5} and 2.8×10^{-4} , respectively. Protein network was generated with STRING, version 10.5, selecting all active interaction sources, minimum required interaction score 0.90 (highest confidence), and no more than 10 interactions. **B:** Only database interaction sources were selected with minimum required interaction score 0.90 (highest confidence), and no more than 10 interactions. Six out of 11 proteins in this network belong to “positive regulation of cAMP biosynthetic process” (GO biological process), which was the most enriched pathway in this network, connecting proteins RXFP2, GIPR, GCG, ADCYAP1, MC1R, and FSHR (false discovery rate 1.3×10^{-8}). Of note is that gastric inhibitory polypeptide receptor (GIPR) and glucagon (GCG) present in this protein network have a high relevance in glucose homeostasis and diabetes.

in the same direction ($\beta = 0.034$ [95% CI $-0.02, 0.09$], $P = 0.192$) (Supplementary Table 13).

Of the SNPs identified in populations without diabetes, four variants were associated with HbA_{1c} in FinnDiane: the minor A allele of the missense variant rs1800562 in *HFE* was associated with lower HbA_{1c} ($P = 0.007$) with higher effects size than in the population without diabetes (Fig. 4). The HbA_{1c} values for the genotypes AA ($n = 3$), AG ($n = 250$), and GG ($n = 4,369$) were $7.6 \pm 1.1\%$, $8.3 \pm 1.2\%$, and $8.5 \pm 1.2\%$, respectively. Variants rs906216 and rs7072268 in *HK1* and rs12621844 near *FOXN2* were also directionally consistently associated with HbA_{1c} in FinnDiane ($P < 0.05$) (Supplementary Tables 13 and 14). Of note, 16 of 19 SNPs classified as erythrocytic variants by Wheeler et al. (6) showed similar direction of effect in FinnDiane compared with the original study (binomial test for enrichment, $P = 0.002$ [Fig. 4]).

GRS

Next, we tested the combined genetic effect by creating GRS from significant variants found in the largest HbA_{1c} GWAS (6) (Supplementary Fig. 7). A GRS-ALL comprising 55 SNPs was associated with HbA_{1c} in FinnDiane ($\beta = 0.010$ [95% CI $0.002, 0.018$], $P = 0.014$) and, along with covariates, explained 2.5% of the variation in HbA_{1c}. However, age, diabetes duration, sex, and n HbA_{1c} alone explained 2.4% of the variation, whereas GRS-ALL explained only 0.1% of the variation in HbA_{1c}. A similar amount of HbA_{1c} variance was also explained by GRS-E including only 19 erythrocytic SNPs ($\beta = 0.021$ [95% CI $0.005, 0.037$], $P = 0.01$). In contrast, GRS-G created from the 18 glycemic variants was not associated with HbA_{1c} ($\beta = 0.004$ [95% CI $-0.01, 0.018$], $P = 0.573$).

DISCUSSION

We identified a locus on chromosome 13 that show genome-wide significant association with HbA_{1c} in 4,622 Finnish individuals with T1D. The three associated SNPs in the locus are located <30 kb from each other and are in high LD, therefore indicating the same association signal. Notably, the association with HbA_{1c} at rs2085277 and rs1360072 remained genome-wide significant in a meta-analysis including individuals with T1D from RASS. In DCCT/EDIC and WESDR, the MAFs for the chr13 top SNPs were $\leq 0.5\%$. The low MAFs ($\leq 1\%$) might explain the lack of significant replication in the T1D cohorts, since the power to detect significant association with $MAF \leq 1\%$ is limited. On the contrary, both SNP associations were replicated ($P \leq 0.013$) in the East Asian population without diabetes, where the SNPs were present with high allelic frequency of 38.5% and 34.5% for rs2085277 and rs1360072, respectively.

rs2085277 is an intergenic SNP surrounded by *B3GLCT* (261 kb 5') and *RXFP2* (146 kb 3'). More distant *B3GLCT* is a glucosyltransferase that adds glucose molecules to multiple proteins. *RXFP2* seems a more plausible candidate gene, since many SNPs in close vicinity to rs2085277 (although not in LD) have *cis*-eQTLs for *RXFP2* in adrenal gland. *RXFP2* is a receptor for insulin-like peptide 3 (INSL3) and relaxin, and *RXFP2* is expressed at low levels in many tissues. ProteomicsDB shows *RXFP2* expression in adrenal glands and in pancreatic islets. Although the pharmacology and biology of relaxin receptors are relatively well-known (26), we are not aware of any previous studies on *RXFP2* in relation to glucose homeostasis. Of note, protein interaction analysis restricted to databases shows that *RXFP2* is linked to the glucagon

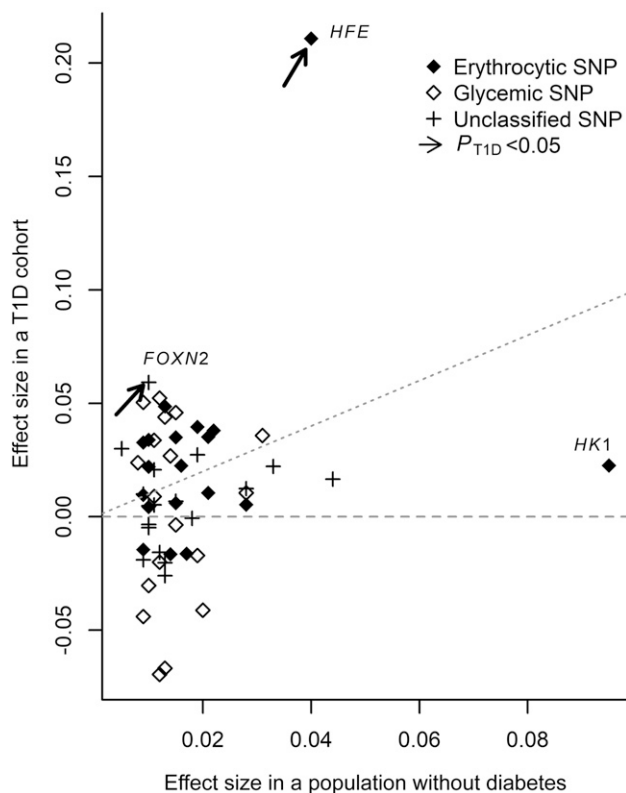


Figure 4—Effect sizes (β s) of 55 variants associated with HbA_{1c} (%) in the original study by Wheeler et al. (6) (European population without diabetes on x-axis) and in FinnDiane (T1D cohort on y-axis). The SNPs were classified into three groups (erythrocytic SNPs, glycemic SNPs, or unclassified SNPs) in the study by Wheeler et al. based on the literature search of known associations of the SNPs in earlier published GWAS for glycemic or erythrocytic traits. Horizontal dashed line denotes the effect size zero ($\beta = 0$) in FinnDiane indicating that variants with $\beta > 0$ have the effect on HbA_{1c} in the same direction as in the population without diabetes. SNPs above the diagonal dotted line have a stronger effect on HbA_{1c} in the T1D cohort. Two SNPs with HbA_{1c} association $P < 0.05$ in FinnDiane are indicated by arrows. Gene names are shown for these variants and for *HK1* because this particular SNP (rs4745982) in *HK1* was not associated with HbA_{1c} in FinnDiane. Instead, two other SNPs in *HK1* (rs906216 and rs7072268 from Paré et al. [1]) showed significant association with HbA_{1c} in FinnDiane. *FOXN2*, forkhead box protein N2; *HFE*, hemochromatosis; *HK1*, hexokinase.

response pathway and to relevant proteins for glucose homeostasis, like GIPR and GCG. Interestingly, adenylate cyclase 5 (ADCY5), which has previously been linked to HbA_{1c} and T2D (6,27), is found in the same protein interaction network. However, there is yet no experimental evidence that would link these proteins. Thus, the biological mechanisms by which RXFP2 possibly affects HbA_{1c} remain to be elucidated.

RXFP2 stimulates adenylate cyclase to increase intracellular second messenger cAMP concentrations. The main ligand for RXFP2 is relaxin hormone, which is naturally increased in pregnancy, and it has both vasodilative and antifibrotic actions (26). Interestingly, relaxin might act along with insulin to control blood glucose homeostasis

(28). This was also suggested by a study in mice, where relaxin treatment ameliorated the insulin sensitivity of mice on a high-fat diet (29). The main receptor for human relaxin is relaxin family peptide receptor 1 (RXFP1) but, to a smaller extent, relaxin is also a ligand for RXFP2.

Our main phenotype was mean HbA_{1c}, which was based on a median of 19 measurements per individual. One limitation of the study is that some individuals had only one HbA_{1c} measurement. A single HbA_{1c} measurement is not optimal for T1D studies, since HbA_{1c} values tend to vary within a person over time. When a smaller subcohort of FinnDiane patients with three or more HbA_{1c} measurements was studied with another method, linear mixed models, all top variants were consistently associated with HbA_{1c}, although the effect sizes tended to be smaller. This was maybe due to a smaller cohort ($n = 4,153$) and deviation from the linear trend over time (individual slopes) in the HbA_{1c} values.

In diabetes, HbA_{1c} values are used as an indicator of glycemic control achieved with insulin treatment or with drugs improving insulin sensitivity, the latter mainly used in patients with type 2 diabetes. The heritability estimate for HbA_{1c} in our study was modest but significant ($h^2 = 15\%$) and confirmed that HbA_{1c} in T1D can hold a heritable component (30). In the HbA_{1c} GWAS in DCCT, a variant near *SORCS1* was found. We were, however, unable to replicate the finding, although the effect was in the same direction. The lack of replication may be due to the winner's curse, which causes the effect in the discovery cohort to be overestimated, and therefore the power for replicating the true smaller genetic effect is lower. Here, we had 100% power to replicate rs1358030 with a $\beta = 0.045$ [for $\ln(\text{HbA}_{1c})$] and only 62% power with a $\beta = 0.003$ corrected for winner's curse with a bootstrap method at two-sided $\alpha = 0.05$ (31). On the other hand, four variants in or near *HFE*, *HK1*, and *FOXN2* originally found in cohorts without diabetes were also associated with HbA_{1c} ($P < 0.05$) in FinnDiane.

Variants associated with HbA_{1c} have mostly been discovered in large GWAS meta-analyses and have relatively small effect sizes. Therefore, the power to replicate these variants in our study was only $\leq 33\%$. However, a GRS comprising 55 SNPs associated with HbA_{1c} in our T1D cohort, although it explained only a small part of the variation of HbA_{1c}. Interestingly, when divided into erythrocytic and glycemic risk scores, GRS-E including only 19 variants classified as erythrocytic SNPs was also associated with HbA_{1c}, while no association was seen for the glycemic risk score. This is expected, as the erythrocyte biology can be expected to be similar in individuals with and without diabetes, but the mechanisms affecting the level of glycemia are presumably different in individuals with T1D compared with the individuals without diabetes. As discussed by Wheeler et al. (6), the genetic factors primarily affecting erythrocyte biology may result in falsely high or low HbA_{1c} levels no longer fully reflecting the level of glycemia and, thus, in false interpretation of HbA_{1c}.

Allele frequencies of risk variants may differ between populations, and therefore it is important to conduct genetic analyses in different populations. Finns represent a population that has been genetically isolated from the rest of Europe, thus presenting an interesting opportunity to explore genetic variants affecting HbA_{1c} in T1D. Indeed, the lead SNP was observed with a markedly higher MAF in the Finnish than in non-Finnish Europeans.

To conclude, we identified a locus on chromosome 13 near *RXFP2* that is associated with HbA_{1c} in individuals with T1D. Further studies are needed to find out whether relaxin hormone mediates the effect of *RXFP2* on HbA_{1c}. Since effect sizes are expected to be smaller in the population without diabetes, additional replication of the top locus as well as the search for additional genetic factors affecting HbA_{1c} in other T1D cohorts is warranted. As HbA_{1c} has a major influence on the development of diabetes complications, it is important to continue the search for genetic variants affecting HbA_{1c}.

Acknowledgments. The authors thank the staff and all participants in the FinnDiane, DCCT/EDIC, CACTI, EDC, RASS, and WESDR study cohorts. Data on glycemic traits have been contributed by MAGIC (Meta-Analyses of Glucose and Insulin-related traits Consortium) investigators and have been downloaded from www.magicinvestigators.org.

Funding. The FinnDiane Study and the work reported here were supported by grants from the Folkhälsan Research Foundation, Wilhelm och Else Stockmann Foundation, Novo Nordisk Foundation, Liv och Hälsa Society, Helsinki University Central Hospital Research Funds (EVO) and Academy of Finland (299200 and 275614), European Foundation for the Study of Diabetes (to N.S.), and the Orion Research Foundation (to A.S.). JDRF supported the genotyping of the FinnDiane subjects (grant 17-2013-7). CACTI was performed at the Barbara Davis Center for Childhood Diabetes in Denver, CO; the Colorado Heart Imaging Center in Denver, CO; and the University of Colorado Hospital. Support for CACTI was provided by the National Heart, Lung, and Blood Institute, National Institutes of Health (NIH), grants R01-HL-61753, R01-HL-079611, and HL-113029, and JDRF grants 17-2013-313 and 17-2013-9. The study was performed at the Clinical Translational Research Center at the University of Colorado Denver supported by NIH M01-RR-000051 and NIH/National Center for Advancing Translational Sciences Colorado CTSA grant UL1-TR-001082. RASS was partly supported by research grants from the NIH, the National Institute of Diabetes and Digestive and Kidney Diseases (DK-51975), and the Canadian Institutes of Health Research (DCT 14281). RASS was also supported in part by a grant from the National Center for Research Resources of the NIH to the University of Minnesota General Clinical Research Center (M01-RR00400). WESDR was supported by grants from Research to Prevent Blindness and by National Eye Institute grant EY016378 (B.E.K.K. and R.K.).

Duality of Interest. RASS was partly supported by research grants from Merck (in the U.S.) and Merck Frosst (in Canada). P.-H.G. has received investigator-initiated research grants from Eli Lilly and Roche; is an advisory board member for AbbVie, Astellas, AstraZeneca, Boehringer Ingelheim, Cebix, Eli Lilly, Janssen, Medscape, Merck Sharp & Dohme (MSD), Mundipharma, Nestle, Novartis, Novo Nordisk, and Sanofi; and has received lecture fees from AstraZeneca, Boehringer Ingelheim, Eli Lilly, Elo Water, Genzyme, MSD, Medscape, Novartis, Novo Nordisk, PeerVoice, and Sanofi. No other potential conflicts of interest relevant to this article were reported.

Author Contributions. A.S. contributed to study design, researched FinnDiane data, performed meta-analysis, and wrote and edited the manuscript. N.S. prepared genotype data, contributed to study design, and wrote and reviewed the manuscript. J.C. prepared and analyzed data from the T1D replication cohorts. I.T. prepared the genotype data, contributed to data

analysis, and reviewed the manuscript. D.M.M. and M.J.R. contributed to the CACTI study. J.K.S.-B. contributed to the CACTI study and reviewed the manuscript. T.C. contributed to the EDC study and reviewed and edited the manuscript. T.J.O. contributed to the EDC study and reviewed the manuscript. M.L.C. and M.M. contributed to RASS and reviewed the manuscript. B.E.K.K. and R.K. contributed to the WESDR study and reviewed the manuscript. E.V. reviewed the manuscript. M.P. contributed to the genome-wide genotyping and TaqMan SNP genotyping. C.F. and V.H. contributed to the collection of clinical data (FinnDiane) and reviewed the manuscript. A.D.P. contributed to study design and reviewed and edited the manuscript. P.-H.G. contributed to study design and reviewed and edited the manuscript. P.-H.G. is the guarantor of this work and, as such, had full access to all the data in the study and takes responsibility for the integrity of the data and the accuracy of the data analysis.

Prior Presentation. Parts of this study were presented in abstract form at the 54th Annual Meeting of the European Association for the Study of Diabetes, Berlin, Germany, 1–5 October 2018.

References

- Paré G, Chasman DI, Parker AN, et al. Novel association of HK1 with glycated hemoglobin in a non-diabetic population: a genome-wide evaluation of 14,618 participants in the Women's Genome Health Study. *PLoS Genet* 2008;4:e1000312
- Franklin CS, Aulchenko YS, Huffman JE, et al. The TCF7L2 diabetes risk variant is associated with HbA_{1c} levels: a genome-wide association meta-analysis. *Ann Hum Genet* 2010;74:471–478
- Soranzo N, Sanna S, Wheeler E, et al.; WTCCC. Common variants at 10 genomic loci influence hemoglobin A_{1c} levels via glycemic and nonglycemic pathways. *Diabetes* 2010;59:3229–3239
- An P, Miljkovic I, Thyagarajan B, et al. Genome-wide association study identifies common loci influencing circulating glycated hemoglobin (HbA_{1c}) levels in non-diabetic subjects: the Long Life Family Study (LLFS). *Metabolism* 2014;63:461–468
- Chen P, Takeuchi F, Lee JY, et al.; CHARGE Hematology Working Group. Multiple nonglycemic genomic loci are newly associated with blood level of glycated hemoglobin in East Asians. *Diabetes* 2014;63:2551–2562
- Wheeler E, Leong A, Liu CT, et al.; EPIC-CVD Consortium; EPIC-InterAct Consortium; Lifelines Cohort Study. Impact of common genetic determinants of hemoglobin A_{1c} on type 2 diabetes risk and diagnosis in ancestrally diverse populations: a transethnic genome-wide meta-analysis. *PLoS Med* 2017;14:e1002383
- Nathan DM, Turgeon H, Regan S. Relationship between glycated haemoglobin levels and mean glucose levels over time. *Diabetologia* 2007;50:2239–2244
- Snieider H, Sawtell PA, Ross L, Walker J, Spector TD, Leslie RD. HbA_{1c} levels are genetically determined even in type 1 diabetes: evidence from healthy and diabetic twins. *Diabetes* 2001;50:2858–2863
- Meigs JB, Panhuysen CI, Myers RH, Wilson PW, Cupples LA. A genome-wide scan for loci linked to plasma levels of glucose and HbA_{1c} in a community-based sample of Caucasian pedigrees: the Framingham Offspring Study. *Diabetes* 2002;51:833–840
- Paterson AD, Waggott D, Boright AP, et al.; MAGIC (Meta-Analyses of Glucose and Insulin-related traits Consortium); Diabetes Control and Complications Trial/Epidemiology of Diabetes Interventions and Complications Research Group. A genome-wide association study identifies a novel major locus for glycemic control in type 1 diabetes, as measured by both A1C and glucose. *Diabetes* 2010;59:539–549
- Penttilä IM, Halonen T, Punnonen K, Tiikkainen U. Best use of the recommended IFCC reference method, material and values in HbA_{1c} analyses. *Scand J Clin Lab Invest* 2005;65:453–462
- Syreeni A, El-Osta A, Forsblom C, et al.; FinnDiane Study Group. Genetic examination of SETD7 and SUV39H1/H2 methyltransferases and the risk of diabetes complications in patients with type 1 diabetes. *Diabetes* 2011;60:3073–3080

13. Manichaikul A, Mychaleckyj JC, Rich SS, Daly K, Sale M, Chen WM. Robust relationship inference in genome-wide association studies. *Bioinformatics* 2010; 26:2867–2873
14. Yang J, Lee SH, Goddard ME, Visscher PM. GCTA: a tool for genome-wide complex trait analysis. *Am J Hum Genet* 2011;88:76–82
15. Delaneau O, Marchini J, Zagury JF. A linear complexity phasing method for thousands of genomes. *Nat Methods* 2011;9:179–181
16. 1000 Genomes Project Consortium, Auton A, Brooks LD, et al. A global reference for human genetic variation. *Nature* 2015;526:68–74
17. Das S, Forer L, Schönerr S, et al. Next-generation genotype imputation service and methods. *Nat Genet* 2016;48:1284–1287
18. Purcell S, Neale B, Todd-Brown K, et al. PLINK: a tool set for whole-genome association and population-based linkage analyses. *Am J Hum Genet* 2007;81: 559–575
19. Price AL, Patterson NJ, Plenge RM, Weinblatt ME, Shadick NA, Reich D. Principal components analysis corrects for stratification in genome-wide association studies. *Nat Genet* 2006;38:904–909
20. Mägi R, Morris AP. GWAMA: software for genome-wide association meta-analysis. *BMC Bioinformatics* 2010;11:288
21. Machiela MJ, Chanock SJ. LDlink: a web-based application for exploring population-specific haplotype structure and linking correlated alleles of possible functional variants. *Bioinformatics* 2015;31:3555–3557
22. Mi H, Huang X, Muruganujan A, et al. PANTHER version 11: Expanded annotation data from gene ontology and reactome pathways, and data analysis tool enhancements. *Nucleic Acids Res* 2017;45:D183–D189
23. Boyle AP, Hong EL, Hariharan M, et al. Annotation of functional variation in personal genomes using RegulomeDB. *Genome Res* 2012;22:1790–1797
24. Staley JR, Blackshaw J, Kamat MA, et al. PhenoScanner: a database of human genotype-phenotype associations. *Bioinformatics* 2016;32:3207–3209
25. Kettunen J, Demirkan A, Würtz P, et al. Genome-wide study for circulating metabolites identifies 62 loci and reveals novel systemic effects of LPA. *Nat Commun* 2016;7:11122
26. Halls ML, Bathgate RA, Sutton SW, Dschietzig TB, Summers RJ. International Union of Basic and Clinical Pharmacology. XCV. Recent advances in the understanding of the pharmacology and biological roles of relaxin family peptide receptors 1-4, the receptors for relaxin family peptides. *Pharmacol Rev* 2015;67: 389–440
27. Dupuis J, Langenberg C, Prokopenko I, et al.; DIAGRAM Consortium; GIANT Consortium; Global BPgen Consortium; Anders Hamsten on behalf of Procardis Consortium; MAGIC investigators. New genetic loci implicated in fasting glucose homeostasis and their impact on type 2 diabetes risk. *Nat Genet* 2010;42:105–116
28. Bani D, Pini A, Yue SK. Relaxin, insulin and diabetes: an intriguing connection. *Curr Diabetes Rev* 2012;8:329–335
29. Bonner JS, Lantier L, Hocking KM, et al. Relaxin treatment reverses insulin resistance in mice fed a high-fat diet. *Diabetes* 2013;62:3251–3260
30. Florez JC. A genome-wide association study of treated A1C: a genetic needle in an environmental haystack? *Diabetes* 2010;59:332–334
31. Sun L, Dimitromanolakis A, Faye LL, Paterson AD, Waggott D, Bull SB; DCCT/EDIC Research Group. BR-squared: a practical solution to the winner's curse in genome-wide scans. *Hum Genet* 2011;129:545–552