



Draft Genome Sequence of Acinetobacter johnsonii MB44, Exhibiting Nematicidal Activity against Caenorhabditis elegans

Shijing Tian, Muhammad Ali, Li Xie, Lin Li

State Key Laboratory of Agricultural Microbiology, Huazhong Agricultural University, Wuhan, China

Acinetobacter johnsonii MB44 was isolated from a frost-plant-tissue sample, which showed noteworthy nematicidal activity against the model organism *Caenorhabditis elegans*. Here, we report the 3.4 Mb draft genome of *A. johnsonii* MB44, which will help in understanding the molecular mechanism of its ability to infect nematodes.

Received 31 December 2015 Accepted 5 January 2016 Published 18 February 2016

Citation Tian S, Ali M, Xie L, Li L. 2016. Draft genome sequence of Acinetobacter johnsonii MB44, exhibiting nematicidal activity against Caenorhabditis elegans. Genome Announc 4(1):e01772-15. doi:10.1128/genomeA.01772-15.

Copyright © 2016 Tian et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Lin Li, lilin@mail.hzau.edu.cn

B acterial species of the genus *Acinetobacter* are ubiquitous in nature and are usually found in the hospital environment; some of these species have been implicated in a variety of nosocomial infections (1, 2). The genus *Acinetobacter* consisted of 43 validly described species until 2015 (http://www.bacterio.net /acinetobacter.html). *Acinetobacter johnsonii*, which was first proposed and described by Bouvet and Gimont in 1986 (3), has rarely been reported to cause clinical infections (4, 5). We isolated *A. johnsonii* MB44 from a frost-plant-tissue sample in the process of screening for ice-nucleating bacteria in China (6). The strain showed remarkable nematicidal activity against *Caenorhabditis elegans* (unpublished data). To identify the potential nematodevirulent factors, we sequenced and annotated the genome of *A. johnsonii* MB44.

Genomic DNA was extracted using a bacterial DNA kit (GB-CBIO), and DNA quantity was determined with a Nanodrop spectrometer (Thermo Scientific, Wilmington, NC, USA). The genome sequencing of *A. johnsonii* MB44 was performed via Illumina HiSeq 2000 platform using the paired-end strategy (2×125 bp). A total of 8,593,104 reads with approximately 137-fold coverage were assembled via ABySS version 1.3.7 (7) using a k-mer size of 90. Through the data assembly, 75 contigs with a total length of 3,357,599 bp and an average G+C content of 41.37 were obtained, and the contig N_{50} was found to be 106,230 bp.

The annotation of the genome was performed by the NCBI Prokaryotic Genome Annotation Pipeline (http://www.ncbi.nlm .nih.gov/genome/annotation_prok/), which predicted 3,277 genes, including 3,064 coding sequences (CDS), 21 rRNAs, and 81 tRNAs. A total of 58.00% of CDSs could be assigned to the cluster of orthologous groups of proteins (COG) database, and 3 clusters of regularly interspaced short palindromic repeat (CRISPR) repeats were found using CRISPRFinder (8). The 16S rRNA gene sequence of *A. johnsonii* MB44 was 99.65% identical to the type strain *A. johnsonii* ATCC 17909^T. The average nucleotide identity (ANI) using the online calculator (http://enve-omics.ce.gatech .edu/ani/index) revealed a two-way ANI value of 95.65% between *A. johnsonii* MB44 and *A. johnsonii* ATCC 17909^T, which suggests that these two strains belong to same speices. We used the software MP3 (9) to predict virulent proteins in this genomic data and 108 potential virulent proteins were found. In addition, this genome encodes some proteins that are homologous to the known virulent proteins in *A. baumannii*, such as the outer membrane protein A, the phospholipase D, and penicillinbinding protein 7/8 (10). The genome of MB44 also contains genes involved in the biosynthesis of siderophore and capsular polysaccharide, which are important virulence factors associated with the killing of *C. elegans* (11, 12). The genome information of *A. johnsonii* MB44 may accelerate our understanding of the molecular mechanism of its ability to infect nematodes.

Nucleotide sequence accession numbers. This whole-genome shotgun project was deposited at DDBJ/EMBL/GenBank under the accession no. LBMO00000000. The version described in this paper is version LBMO01000000.

ACKNOWLEDGMENTS

This work was supported by a grant from the National Basic Research Program of China (973 Program, grant 2013CB127504) and grants from the National Natural Science Foundation of China (grants 31570123 and 31270158).

FUNDING INFORMATION

National Basic Research Program of China provided funding to Lin Li under grant number 2013CB127504. National Natural Science Foundation of China (NSFC) provided funding to Lin Li under grant number 31570123.

REFERENCES

- 1. Bergogne-Berezin E, Towner KJ. 1996. *Acinetobacter* spp. as nosocomial pathogens: microbiological, clinical, and epidemiological features. Clin Microbiol Rev 9:148–165.
- Wisplinghoff H, Paulus T, Lugenheim M, Stefanik D, Higgins PG, Edmond MB, Wenzel RP, Seifert H. 2012. Nosocomial bloodstream infections due to *Acinetobacter baumannii, Acinetobacter pittii* and *Acinetobacter nosocomialis* in the United States. J Infect 64:282–290. http:// dx.doi.org/10.1016/j.jinf.2011.12.008.
- 3. Bouvet PJM, Grimont PAD. 1986. Taxonomy of the genus Acinetobacter with the recognition of Acinetobacter baumannii sp. nov., Acinetobacter haemolyticus sp. nov., Acinetobacter johnsonii sp. nov., and Acinetobacter junii sp. nov. and emended descriptions of Acinetobacter calcoaceticus and

Acinetobacter lwoffii. Int J Syst Bacteriol 36:228–240. http://dx.doi.org/ 10.1099/00207713-36-2-228.

- Turton JF, Shah J, Ozongwu C, Pike R. 2010. Incidence of Acinetobacter species other than A. Baumannii among clinical isolates of Acinetobacter: evidence for emerging species. J Clin Microbiol 48:1445–1449. http:// dx.doi.org/10.1128/JCM.02467-09.
- Penzak SR, Gubbins PO, Stratton SL, Anaissie EJ. 2000. Investigation of an outbreak of gram-negative bacteremia among hematology-oncology outpatients. Infect Control Hosp Epidemiol 21:597–599. http:// dx.doi.org/10.1086/501810.
- Li Q, Yan Q, Chen J, He Y, Wang J, Zhang H, Yu Z, Li L. 2012. Molecular characterization of an ice nucleation protein variant (inaQ) from *Pseudomonas syringae* and the analysis of its transmembrane transport activity in *Escherichia coli*. Int J Biol Sci 8:1097–1108. http:// dx.doi.org/10.7150/ijbs.4524.
- Simpson JT, Wong K, Jackman SD, Schein JE, Jones SJ, Birol I. 2009. ABySS: a parallel assembler for short read sequence data. Genome Res 19:1117–1123. http://dx.doi.org/10.1101/gr.089532.108.

- Grissa I, Vergnaud G, Pourcel C. 2007. CRISPRFinder: a web tool to identify clustered regularly interspaced short palindromic repeats. Nucleic Acids Res 35:W52–W57. http://dx.doi.org/10.1093/nar/gkm360.
- Gupta A, Kapil R, Dhakan DB, Sharma VK. 2014. MP3: a software tool for the prediction of pathogenic proteins in genomic and metagenomic data. PLoS One 9:e93907. http://dx.doi.org/10.1371/ journal.pone.0093907.
- Cerqueira GM, Peleg AY. 2011. Insights into Acinetobacter baumannii pathogenicity. IUBMB Life 63:1055–1060. http://dx.doi.org/10.1002/ iub.533.
- Kirienko NV, Ausubel FM, Ruvkun G. 2015. Mitophagy confers resistance to siderophore-mediated killing by *Pseudomonas aeruginosa*. Proc Natl Acad Sci USA 112:1821–1826. http://dx.doi.org/10.1073/ pnas.1424954112.
- Bae T, Banger AK, Wallace A, Glass EM, Aslund F, Schneewind O, Missiakas DM. 2004. *Staphylococcus aureus* virulence genes identified by bursa *Aurealis mutagenesis* and nematode killing. Proc Natl Acad Sci USA 101:12312–12317. http://dx.doi.org/10.1073/pnas.0404728101.