# **Ecology and Evolution**



# Leaf traits in Chilean matorral: sclerophylly within, among, and beyond matorral, and its environmental determinants

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#### Keywords

Chile, ET<sub>0</sub>, leaf toughness, leaf traits, matorral, sclerophylly, soil nutrients, soil phosphorus.

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#### **Abstract**

Studies of leaf traits often focus on tradeoffs between growth and resource conservation, but little is known about variation in the mechanical traits that influence resource conservation. This study investigates how leaf mechanical traits vary across matorral vegetation in central Chile, how they correlate with environmental factors, and how these trends compare at a broader geographic scale. Leaf toughness, strength, stiffness, and associated traits were measured in five matorral types in central Chile, and relationships with soil N and P and climate variables were assessed. Trends with soil and climate were then analyzed across shrubland and woodland in Chile, Western Australia, and New Caledonia. Chilean species varied in leaf mechanics and associated traits, both within and among matorral types, with more species in sclerophyll matorral having strong, tough, and stiff leaves than in arid and littoral matorral. Overall, leaves with high leaf dry mass per area were stiffer, tougher, stronger, thicker, denser, with more fiber, lignin, phenolics and fiber per unit protein and less protein: tannin activity and N and P per mass, forming a broad sclerophylly syndrome. Mechanical traits of matorral species were not correlated with soil N or P, or predictably with climate variables, except flexural stiffness (EIW) which correlated positively with annual reference evapotranspiration (ET<sub>0</sub>). However, soil P made strong independent contributions to variation in leaf mechanics across shrublands and woodlands of Chile, Western Australia, and New Caledonia, either separately (strength) or together with ET<sub>0</sub> (toughness) explaining 46-90% of variation. Hence ET<sub>0</sub> was predictive of EI<sub>W</sub> in Chilean matorral, whereas soil P was highly predictive of variation in leaf strength, and combined with ET<sub>0</sub> was highly predictive of toughness, at a broader geographic scale. The biological basis of these relationships, however, may be complex.

#### Introduction

Both climate and soil nutrition appear to be strong selective forces on leaf texture, reflecting tradeoffs between growth and resource conservation (Wright et al. 2005; Ordoñez et al. 2009). Sclerophylly, a leaf textural form (hard, tough, stiff, leathery leaves: Schimper 1903) at the slow-return end of the suggested leaf economics spectrum (Wright et al. 2005), is common in mediterranean climates (Schimper 1903; Turner 1994). In particular, sclerophylly has been suggested to be an adaptation to seasonal drought, with, for example, stiff cell walls facilitating turgor maintenance (Oertli et al. 1990), and

sclerenchyma bundle sheath extensions and thick cuticle and leaves contributing to water conservation (Heide-Jørgensen 1990). Arguably, under the drought resistance hypothesis, sclerophylly (a leaf-level textural trait) is not adaptive *per se* but rather a consequence of anatomical traits that are adaptations to seasonal drought.

Sclerophylly is also common in vegetation on lownutrient soils across a range of climates, including moist tropical (Loveless 1961; Sobrado and Medina 1980; Grubb 1986; Specht and Rundel 1990; Choong et al. 1992; Turner 1994; Lamont et al. 2002). Some anatomical features may enhance nutrient conservation on low-nutrient soils, for example, reduced leaching through a thick

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cuticle, but sclerophylly may provide direct benefits by conferring protection, particularly against herbivores, since enhanced LLS (leaf life span) is advantageous in conditions where it takes longer to maximize returns on investment (Turner 1994; Westoby et al. 2002). Indeed, protection from herbivores and consequent resource conservation may be cost-effective in a wide range of stressful environments (Turner 1994). A fourth alternative is that sclerophylly is just a consequence of constraints imposed by severe environments, for example, more limited synthesis of protein than carbohydrate on low-nutrient soils (Salleo and Nardini 2000). Most probably, sclerophylly is a complex and variable syndrome of traits, to which all these factors may contribute.

Leaf trait studies usually focus on the growth-resource conservation tradeoff (Díaz et al. 2004; Wright et al. 2005; Ordoñez et al. 2009), with less emphasis on specific mechanical traits that contribute to resource conservation (but see Choong et al. 1992; Read et al. 2005, 2006; Onoda et al. 2011; Méndez-Alonzo et al. 2013). Furthermore, there are few datasets that allow detailed comparison of the mechanical traits that contribute to sclerophylly, and how they differ in contrasting environments, and indeed of any leaf economic traits in relation to soil variables across large spatial scales (Ordoñez et al. 2009; Maire et al. 2015). Comparative studies of mechanical traits are of value in resolving the relative importance of adaptive and "nonadaptive" traits and hence better understand the forces leading to the prevalence of sclerophylly in certain environments and the consequences for growth-resource conservation tradeoffs. In particular, it is not clear how sclerophylly varies in different environments, that is, whether leaves have similar mechanics, morphology, anatomy, and chemistry, relate to environmental conditions in similar ways (e.g., Wright and Westoby 2002; Méndez-Alonzo et al. 2013), and have similar influences on, for example, plant-animal interactions, nutrient cycling, and carbon economy (Díaz et al. 2004; Peeters et al. 2007; Méndez-Alonzo et al. 2013).

The matorral vegetation of Chile provides a useful contribution to this discussion since distinct forms of vegetation occur across a range of mediterranean-type climates and soils, including, but not limited to, sclerophyll communities (Rundel 1981; Armesto et al. 2007). Furthermore, it contrasts with sclerophyll vegetation in some regions by having more fertile soil (Lamont 1994), so is predicted by the nutrient conservation hypothesis to have softer leaves than, for example, the heath vegetation of Western Australia. Here we address the general hypothesis that leaf mechanical traits such as strength, toughness, and stiffness increase along gradients of water deficit and/or soil nutrient deficiency. We investigate (1) how leaf toughness, strength, and stiffness, and associated traits,

vary among differing matorral vegetation of central Chile, and (2) the potential influence of soil nutrients and climate factors on leaf trait syndromes. We then compare these data with those recorded in sclerophyll vegetation in mediterranean climates of southwest Australia and tropical climates of New Caledonia, over a range of soil fertility, testing (3) the relative contribution of soil nutrients versus climate variables to variation in mechanical traits at this broader geographic scale.

### **Materials and Methods**

# **Study region**

Mediterranean-type vegetation, or matorral, occurs in central Chile at 30–36°S (Armesto et al. 2007). Evergreen sclerophyll shrubland is the most common form, particularly on the slopes of the Coastal Cordillera and the lower slopes of the Andes, but matorral varies considerably, occurring across varying climates and soils (Armesto et al. 2007). Mean annual maximum temperatures for the matorral zone are *c*. 20–25°C, with extremes moderated near the coast (Rundel 1981). Annual rainfall is <200–700 mm, mostly during the coolest months (April—September), with a long dry spring—summer of up to 6 months (Armesto et al. 2007). In some regions, fog provides an important source of moisture, particularly in summer (Armesto et al. 2007; Negret et al. 2013).

### **Study sites**

Five types of matorral were selected, following the main types described by Rundel (1981), across contrasting environments from the coast to the lower mountains of the Andes at c. 31-34°S (Table 1, Appendix). They were sampled in September (austral spring) 2008, including only woody dicots and subshrubs and excluding succulents. Littoral matorral was sampled on coastal bluffs at two sites near Zapallar in shrubland dominated by malacophyllous shrubs, including summer-deciduous species, interspersed with Puva spp. (Bromeliaceae) and cacti. Lowland sclerophyll matorral was sampled at two sites c. 2 km from the sea at Cachagua, dominated by evergreen sclerophyll shrubs of c. 1-5 m high with small trees to c. 10 m high. Both the Zapallar and Cachagua sites commonly experience morning fog. Mid-elevation sclerophyll matorral was studied at Reserva Nacional Río Clarillo in open sclerophyll shrubland, including vegetation along drainage lines (Fig. 1). Montane sclerophyll matorral of low shrubs with infrequent tall shrubs/small trees was sampled at Santuario de la Naturaleza Yerba Loca. Arid matorral of short shrubs with sparse small trees and frequent cacti was studied at Reserva Nacional Las Chinchillas.

**Table 1.** Environmental comparisons of the Chilean matorral vegetation types. The soil data are means  $\pm$  standard errors. See text for sources of climate data. Results of ANOVA are presented, based on log-transformed data.

	Arid matorral	Littoral matorral	Lowland sclerophyll matorral	Mid-elevation sclerophyll matorral	Montane sclerophyll matorral
Study area	Las Chinchillas	Zapallar	Cachagua	Río Clarillo	Yerba Loca
Lat/long	31°30.7′S, 71°6.4′W	32°33.1′S 71°28.1′W	32°35.6′S, 71°25.1′W	33°43.7′S, 70° 28.4′W	33°20.3′S, 70°20.1′W
Distance to sea (km)	44	0	2	108	120
Elevation (m asl)	550-700	5–30	100–180	900–950	1700-1780
AMT (°C)	14.3	15.7	15.7	14.2	12.7
AP (mm)	212	357	357	497	445 (at 2500 m asl)
PDQ (mm)	2	4	2	13	20
ET <sub>0</sub> (mm)	1103	926	921	1118	1245
Soil properties					
pH**	$7.8\pm0.3^{a}$	$7.2\pm0.4^{ab}$	$6.3 \pm 0.3^{b}$	$6.3 \pm 0.1^{b}$	$7.3\pm0.1^{ab}$
LOI (%)***	$4.2 \pm 0.4^{a}$	$4.7 \pm 0.4^{a}$	$8.5\pm0.8^{b}$	$4.1 \pm 0.7^{a}$	$9.6 \pm 0.4^{b}$
Nitrogen (mg g <sup>-1</sup> )**	$0.61\pm0.09^{ab}$	$0.83\pm0.15^{ab}$	$1.37\pm0.26^{a}$	$0.34\pm0.13^{b}$	$0.86\pm0.04^{ab}$
Phosphorus (mg g <sup>-1</sup> )***	$0.74\pm0.06^a$	$0.54\pm0.05^a$	$0.62\pm0.09^a$	$0.20\pm0.02^{b}$	$0.67\pm0.06^a$

AMT, annual mean temperature; AP, annual precipitation; PDQ, precipitation of the driest quarter; ET<sub>0</sub>, annual reference evapotranspiration; LOI, loss on ignition, an estimate of soil organic content.

Asterisks indicate P-values: \*\*P < 0.01; \*\*\*P < 0.001. Shared alphabet letters indicate no significant difference. Latitudes and longitudes are given for a single collection area within the study site. Precipitation variables for Zapallar and Cachagua do not take supplementary humidity due to fog into account.



**Figure 1.** Mid-elevation materral at Reserva Nacional Río Clarillo, south-central Chile.

Climate data (annual precipitation and annual mean temperature) were obtained for study areas from local meteorological stations, with monthly precipitation estimated by WorldClim v. 1.4 (Hijmans et al. 2005; 30 arc seconds resolution) with DIVA-GIS v. 7 (Hijmans et al. 2012). Penman–Monteith reference evapotranspiration rate (ET<sub>0</sub>) was estimated by the 10-arcmin IWMI World Water Climate Atlas (http://www.iwmi.cgiar.org/). Soil samples were collected at three to five semi-randomly located points in each matorral. Each sample comprised two to five subsamples, taken at least 5 m apart, from the top 10 cm of soil, and was air-dried

and sieved to 2 mm. Soil organic concentration was measured by LOI (loss on ignition) of a sample combusted at 550°C for 2 h. Soil pH was measured using a 1:5 mass: volume mixture. Total N and P were measured as for leaves (see below), on samples ground to 0.2 mm.

# Sample collection

Each maternal type was sampled at five locations across at least 1 km, sampling each species at each location when possible. Healthy, mature sunlit leaves were collected from up to five plants of the 10-25 most common shrubs and trees. A total of 62 species was collected, with 13 sampled in more than one type of matorral. We aimed to sample leaves that were c. 6-12 months old, produced during the last growing season, and having experienced winter. However, leaves of summer-deciduous species were c. 3 months old, produced during winter (e.g., Aljaro et al. 1972), and the oldest leaves of some subshrubs were possibly also produced during winter. Hence, although all leaves were mature, some differed in age and exposure to seasons. Mechanical and morphological traits and water content were measured on fully hydrated leaves within 12-48 h of collection. Leaves used for chemical analyses were air-dried, then oven-dried at 40°C. Potential LLS (age to senescence) was estimated in three shoots per plant, based on twig morphology and color, bud scars, and senescing leaves.

# Leaf morphological and mechanical traits

One to five leaves per plant were weighed fresh, without petioles. Lamina thickness was measured at three random locations, avoiding the midrib, with a micrometer or calipers depending on leaf morphology. Leaf area was measured by image analysis (Mix Image analysis v. 3, Monash University, Victoria, Australia). Leaves were then ovendried at 40°C and reweighed to calculate water content, LMA (leaf dry mass per area) which is a commonly used index of sclerophylly (Groom and Lamont 1999), and tissue density.

Relatively little is known about how mechanical traits derived from differing test types (punch, shear, tear and bending) vary within and among vegetation types, including in sclerophyll vegetation. Hence, we investigated multiple leaf traits, derived from the main test types (e.g., Read and Sanson 2003). Leaf strength, toughness, and stiffness were measured following Read and Sanson (2003), using a portable custom-made force tester (Read et al. 2005, 2006). Strength was measured as maximum force to fracture using a punch test (0.5 mm diameter, sharp-sided punch), with five random punches across the lamina, avoiding the midrib. Toughness was measured as work to fracture, calculated as the area under the forcedisplacement curve generated by punch, shear, and tear tests. Shear tests (20° blade approach angle) were undertaken on a strip cut from one side of the leaf, avoiding the midrib and margins. The flexural Young's modulus (E, a "material property") and flexural stiffness (EI, a "structural property" that includes the effect of leaf thickness, where I is the second moment of area) were measured on a leaf strip using a three-point bending test with a span:depth ratio of 70. EIW was calculated as flexural stiffness per unit strip width. Tear strength and toughness were measured on notched leaf strips with an aspect ratio >10 (Vincent 1990), mounted in clamps, and secured with cyanoacrylate glue. Results for fracture tests were also expressed per unit leaf thickness ("material properties"), termed "specific work" and "specific strength" (Read and Sanson 2003). Leaf thickness was measured at the point of fracture, with a micrometer or calipers. Tearing and bending tests could not be undertaken in some small-leaved species (Appendix).

#### Leaf chemistry

Leaves were freeze-dried, then ground to a powder. Foliar N was measured by a Leco CHN-200 auto-analyser (Leco Corp., St Joseph, MI) and P by the molybdenum-blue colorimetric method (Grimshaw et al. 1989) after digestion by the sulfuric-peroxide procedure (Grimshaw 1987). N and P were expressed per unit dry mass ( $N_{mass}$ ,  $P_{mass}$ )

and per unit mass of water ( $N_{water}$ ,  $P_{water}$ ), the latter allowing an estimate of protoplasmic concentration. "Total phenolics" were extracted in acetone (Cork and Krockenberger 1991) and assayed by the Prussian blue method (Price and Butler 1977) with concentration expressed as GAE (gallic acid equivalents) per leaf dry mass. Tannin activity was estimated as the amount of bovine serum albumen bound by the phenolic extract (Asquith and Butler 1985), expressed as mass of protein precipitated per unit leaf dry mass. Total cell wall, measured as NDF (neutral detergent fiber), and ADL (acid detergent lignin) were quantified following Van Soest et al. (1991). The Loveless sclerophylly index (SI) was calculated as NDF per unit protein ( $N_{mass} \times 6.25$ ).

#### **Data analysis**

Differences among sites were tested by ANOVA with Tukey's post hoc comparisons. Pearson correlation was used to test associations among leaf traits, averaging values of species sampled in multiple vegetation types. Log transformations were used when necessary to improve data normality, reduce the influence of outliers, and improve linearity for correlations and regressions. PCA (Principal components analysis) was used to summarize data (species' averages) into main components, first using the full data set of species averages for each site, then averaging those species sampled in multiple vegetation types. Only traits measured in all species were included, that is, excluding traits from tearing and bending tests. Since there was little difference in results, PCA of the full data set is presented to show within-species variation across sites. Spearman correlation was used to test associations of leaf traits with environmental variables.

Comparisons were made with mediterranean-climate shrubland and woodland in southwest Western Australia (Read et al. 2005) and tropical moist shrubland on ultramafic soils and dry forest (excluding lianes) in New Caledonia (Read et al. 2006), these studies having used identical methods to measure leaf mechanics and providing contrasts in climate and soil fertility. For example, shrubland study sites in southwest Western Australia and New Caledonia have lower soil phosphorus concentrations (<0.2 mg g<sup>-1</sup>) than in Chilean matorral ( $\geq$ 0.2 mg g<sup>-1</sup>), and the New Caledonian sites have higher annual rainfall (>900 mm) than the Australian and Chilean sites (<500 mm).

ANCOVA was used to test the relationship of work to punch with LMA among regions, across an LMA range of 100–300 g m<sup>-2</sup> (where there was most overlap among regions). Trait variation in evergreen species was summarized across regions with PCA, excluding the littoral matorral in Chile because of likely influences of salinity

and wind exposure. Pearson correlation was used to test associations of selected leaf traits with environmental variables across regions. Of the mechanical traits, only punch tests were included, since shear tests could not always avoid the midrib in small-leaved species, so were less comparable across species, and stiffness and tear tests could not be undertaken in small-leaved species. Hierarchical partitioning (Chevan and Sutherland 1991) was used to calculate independent contributions ( $I_{\rm HP}$ ) of environmental variables to explained variance in mechanical traits and LMA, with statistical significance determined by randomization techniques (Walsh and Mac Nally 2013). Linear regression was then used to determine the variance explained individually or together of significant variables. Hierarchical partitioning was undertaken with hier.part (Walsh and Mac Nally 2013) in R 2.14.1 (R Development Core Team, 2011). SYSTAT v. 13 (Systat Software Inc., San Jose, CA) was used for other analyses, with  $\alpha = 0.05$ for hypothesis testing.

#### Results

#### Soil nutrients

Soils were generally neutral or weakly acidic, with low organic content, and low levels of P and particularly N (Table 1). There were some differences among vegetation types, notably in higher LOI at the lowland and montane sclerophyll matorral, and low N and P at mid-elevation sclerophyll matorral (Table 1).

## Plant and leaf traits across species and sites

Deciduous and semi-deciduous species occurred in all types of matorral, but more were recorded in arid matorral (Table 2). Some evergreen species, particularly subshrubs, had leaves only on soft young stems, suggesting LLS of <6 months. Overall, 1+ years (≥1 year, but <2 years) (43% of species) was the modal LLS, with 38% of species having LLS <1 year, and only 20% of species having LLS of 2−3 years. A high proportion of species in littoral matorral had short-lived leaves, with fewest in mid-elevation sclerophyll matorral (Table 2).

# Foliar nutrients, cell wall and phenolics

Foliar  $N_{mass}$  and  $P_{mass}$  varied six- to ninefold among species, with a trend of lowest concentrations in sclerophyll matorral (Table 2).  $N_{water}$  and  $P_{water}$  differed among vegetation types, with low values in littoral matorral (Table 2). N:P ratios varied fourfold among species, with one-third having N:P ratios <10 (particularly in littoral matorral), suggesting likely N limitation (Güsewell 2004).

There was little clear evidence of P limitation (N:P > 20: Güsewell 2004), with the highest N:P of 20–21 recorded in only four species. C:N ranged from 10 in some deciduous species to c. 60 in numerous evergreen species, with higher values in mid-elevation sclerophyll matorral than in arid and littoral matorral (Table 2).

Neutral detergent fiber varied *c*. fivefold among species, but did not differ among vegetation types (Table 2). ADL varied 10-fold among species, with no vegetation differences, but the trend was for higher values in sclerophyll matorral (Table 2). Total phenolics varied 16-fold among species, with no differences among vegetation types, although the trend was also for higher values in sclerophyll matorral (Table 2). Tannin activity (protein precipitation), with very high variability among species (>300-fold), was higher in sclerophyll matorral than littoral matorral (Table 2). The ratio of protein: tannin activity, an estimate of protein availability to herbivores, also varied 300-fold among species, with a trend of lowest values in sclerophyll matorral (Table 2).

# Leaf morphology, sclerophylly indices and mechanics

Water content varied 12-fold among species, highest in littoral matorral (Table 2). Leaf size varied from <20 mm² to >4000 mm², with smaller leaves in arid matorral than lowland and mid-elevation sclerophyll matorral (Table 2). LMA varied from 24 g m² to >300 g m², lower in littoral matorral than in sclerophyll matorral (Table 2). Of its components, tissue density in littoral matorral was less than half that of sclerophyll matorral, but leaf thickness did not differ among vegetation types (Table 2). SI varied *c.* 20-fold among species, but not among vegetation types, although the trend was for highest values in sclerophyll matorral (Table 2).

There was little significant difference in mechanical traits among vegetation types (Table 2), although there was 16- to 950-fold variation among species. Punch strength varied 41-fold among species and was c. twofold higher in mid-elevation sclerophyll matorral than littoral matorral, with a trend for higher values in sclerophyll matorral (Table 2). Specific punch strength ranged 24fold, higher in lowland and mid-elevation sclerophyll matorral than in littoral matorral (Table 2). Flexural stiffness, EI<sub>W</sub>, was higher in montane sclerophyll matorral than in littoral matorral (Table 2) and varied 950-fold among species. No other mechanical traits differed among vegetation types, despite 20- to 70-fold variation among species (Table 2). However, examination of mechanical data showed wide variation within matorral types, particularly in coastal and mid-elevation sclerophyll matorral, but with sclerophyll matorral typically showing a greater

**Table 2.** Comparisons of leaf traits among the Chilean matorral vegetation types. The data are means of species' means  $\pm$  standard errors. The results of ANOVA are given, with *post hoc* Tukey's tests (shared alphabet letters indicate no significant difference among sites).

Trait	Arid matorral	Littoral matorral	Lowland sclerophyll matorral	Mid-elevation sclerophyll matorral	Montane sclerophyll matorral	F	Р
Deciduous (% of species)	40	20	8	9	10		_
Leaf life span (% <1 year)	50	78	40	19	34		
Leaf chemistry and morphology							
N <sub>mass</sub> (%) <sub>L</sub>	$2.48\pm0.23^{a}$	$2.35\pm0.21^{a}$	$1.80\pm0.20^{a}$	$1.75\pm0.23^{a}$	$2.01\pm0.35^{a}$	2.6	0.045
N <sub>water</sub> (mg g <sup>-1</sup> water) <sub>L</sub>	$11.51 \pm 0.60^{ac}$	$4.31\pm0.30^{b}$	$9.28\pm0.67^{c}$	$11.80\pm0.69^a$	$14.80\pm1.21^{a}$	31.5	< 0.001
$P_{mass}$ (mg g <sup>-1</sup> ) <sub>L</sub>	$2.10\pm0.25^{ab}$	$3.25\pm0.41^{b}$	$1.84\pm0.19^{a}$	$1.39\pm0.15^{a}$	$1.85\pm0.23^{a}$	6.4	< 0.001
P <sub>water</sub> (mg g <sup>-1</sup> water) <sub>L</sub>	$1.02\pm0.12^{ac}$	$0.61\pm0.09^{b}$	$0.97\pm0.09^{c}$	$0.98\pm0.06^{ac}$	$1.47\pm0.22^{a}$	6.8	< 0.001
Nitrogen: phosphorus	$13.8 \pm 2.0^{a}$	$8.4\pm0.8^{b}$	$10.9\pm0.7^{ab}$	$13.1\pm0.6^{a}$	$12.7\pm1.6^{ab}$	3.7	0.009
Carbon: nitrogen	$19.7\pm3.0^{a}$	$19.7\pm2.0^{a}$	$29.9\pm3.2^{ab}$	$34.0\pm3.1^{b}$	$27.6 \pm 3.2^{ab}$	3.3	0.016
NDF (%) L	$21.4 \pm 2.9$	$20.7\pm1.7$	$25.4 \pm 2.0$	$24.1 \pm 2.1$	$20.4 \pm 2.1$	8.0	0.553
ADL (%) L	$4.8\pm0.8$	$4.1 \pm 0.6$	$8.2 \pm 1.4$	$8.0 \pm 1.2$	$7.8\pm1.3$	2.1	0.089
Total phenolics (g GAE. 100 g <sup>-1</sup> ) <sub>L</sub>	$2.9\pm0.9$	$2.6\pm0.8$	$4.4 \pm 0.6$	$4.6\pm0.6$	$4.6 \pm 1.0$	2.0	0.105
Protein precipitation (mg g <sup>-1</sup> ) <sub>L</sub>	$145\pm113^{ab}$	$76 \pm 55^{b}$	$223\pm51^a$	$241\pm53^a$	$229\pm74^a$	4.4	0.003
Protein: protein precipitation (g $g^{-1}$ ) <sub>L</sub>	6.6 ± 1.5	6.6 ± 1.3	3.3 ± 0.9	3.1 ± 0.8	3.0 ± 1.3	2.3	0.063
Water (g g <sup>-1</sup> ) <sub>L</sub>	$2.19\pm0.17^{a}$	$5.58 \pm 0.46^{b}$	$2.18 \pm 0.31^{a}$	$1.46\pm0.14^{a}$	$1.39\pm0.23^{a}$	18.6	< 0.001
Leaf size (mm²) <sub>L</sub>	$308\pm99^a$	$729\pm194^{ab}$	$1163 \pm 201^{b}$	$959\pm197^{\rm b}$	$481\pm99^{ab}$	4.6	0.002
Thickness (mm) <sub>L</sub>	$0.36\pm0.08$	$0.35\pm0.06$	$0.29\pm0.02$	$0.34\pm0.03$	$0.41\pm0.04$	1.5	0.222
Tissue density (mg mm <sup>-3</sup> ) <sub>L</sub>	$0.350\pm0.027^{a}$	$0.206\pm0.017^{c}$	$0.421\pm0.025^{ab}$	$0.473\pm0.022^{b}$	$0.480\pm0.028^{b}$	14.6	< 0.001
LMA (g $m^{-2}$ ) <sub>L</sub>	$124\pm29^{ab}$	$63 \pm 8^{b}$	$127\pm14^a$	$159\pm14^a$	$196\pm22^a$	7.0	< 0.001
SI (g g <sup>-1</sup> ) <sub>L</sub>	$1.52\pm0.24$	$1.69 \pm 0.24$	$3.29\pm0.53$	$3.15\pm0.47$	$2.16\pm0.37$	1.5	0.200
Leaf mechanics							
Work to shear (J m $^{-1}$ ) $_{\rm L}$	$0.144\pm0.023$	$0.106\pm0.022$	$0.151\pm0.020$	$0.200\pm0.045$	$0.173\pm0.035$	8.0	0.524
Specific work to shear (kJ $m^{-2}$ ) L	$0.456\pm0.078$	$0.343\pm0.033$	$0.498\pm0.050$	$0.576\pm0.107$	$0.419\pm0.078$	8.0	0.512
Punch strength (MN m $^{-2}$ ) $_{\rm L}$	$4.43\pm0.56^{ab}$	$2.94\pm0.38^{a}$	$5.86\pm0.75^{ab}$	$6.67\pm0.93^{b}$	$5.34\pm0.67^{ab}$	3.0	0.025
Specific punch strength (GN $m^{-2}$ $m^{-1}$ ) <sub>L</sub>	$18.6 \pm 4.2^{ab}$	12.5 ± 1.5 <sup>a</sup>	$23.3 \pm 2.2^{b}$	$22.6 \pm 2.6^{b}$	$14.9 \pm 1.6^{ab}$	4.3	0.004
Work to punch (kJ m $^{-2}$ ) $_{\rm L}$	$1.22\pm0.25$	$0.93\pm0.19$	$1.27\pm0.18$	$1.60\pm0.27$	$1.63\pm0.24$	1.5	0.202
Specific work to punch $(MJ m^{-2} m^{-1})_L$	$4.23 \pm 0.76$	3.44 ± 0.32	$4.79 \pm 0.46$	5.10 ± 0.66	$4.41 \pm 0.57$	1.0	0.418
Tear strength (MN m <sup>-2</sup> ) <sub>L</sub>	$1.46 \pm 0.62$	$0.68\pm0.10$	$1.44 \pm 0.16$	$1.52\pm0.31$	$1.03\pm0.17$	2.2	0.078
Work to tear (J m <sup>-1</sup> ) <sub>L</sub>	$0.349 \pm 0.018$	$0.193\pm0.063$	$0.244\pm0.041$	$0.303\pm0.054$	$0.240\pm0.053$	0.7	0.580
Specific work to tear (kJ m $^{-2}$ ) $_{\rm L}$	$1.25\pm0.34$	$0.58\pm0.11$	$0.82\pm0.11$	$0.78\pm0.11$	$0.57\pm0.14$	1.4	0.264
$E  (MN  m^{-2})_{L}$	$177\pm60$	$75 \pm 6$	$232\pm37$	$241\pm58$	$164\pm42$	1.7	0.144
$El_{W}$ (mN m <sup>2</sup> m <sup>-1</sup> ) <sub>L</sub>	$1.01\pm0.65^{ab}$	$0.17\pm0.05^{b}$	$1.06\pm0.34^{ab}$	$1.82\pm0.69^{ab}$	$2.01\pm0.58^{a}$	2.8	0.035

L, log-transformed for analysis. NDF, neutral detergent fiber; ADL, acid detergent lignin; GAE, gallic acid equivalents; LMA, leaf dry mass per area; SI, Loveless sclerophylly index; E, Young's modulus; El<sub>W</sub>, flexural stiffness.

Significant P-values are shown in bold.

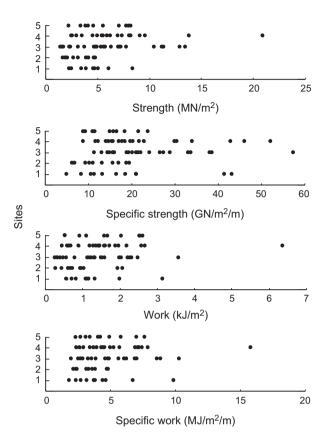
proportion of high values of each mechanical property than arid and littoral matorral (Fig. 2 for punch variables). Relatively few differences in conclusions were reached when deciduous and semi-deciduous species were excluded from analyses, and these were generally minor (Appendix S1).

Leaf traits differed in magnitude among LLS classes, with the exception of  $N_{water}$  and  $P_{water}$  (Table 3). Overall, short-lived leaves had higher  $N_{mass}$  and  $P_{mass}$ , high protein: tannin activity, and lower NDF, ADL, phenolics, tannin activity, LMA, strength, specific strength, toughness,

specific toughness, and stiffness (Table 3). Thus, they are potentially more nutritious and less defended, at least by these carbon-based defenses, than longer-lived leaves.

# Correlations among leaf traits and with environmental variables across species

All mechanical traits were strongly positively intercorrelated ( $P \le 0.001$ ), except for a weak correlation of  $EI_{\rm W}$  with specific work to tear (r = 0.30, P = 0.034) and no correlation with specific punch strength (r = 0.22,



**Figure 2.** Comparison of mechanical variables from punch tests among matorral types in Chile. Dots represent average values of individual species. (1) Arid matorral; (2) littoral matorral; (3) lowland sclerophyll matorral; (4) mid-elevation sclerophyll matorral; (5) montane sclerophyll matorral.

P = 0.125) (Appendix S2). For simplicity, tearing variables and E will not be considered further. The sclerophylly indices, LMA and SI, were positively correlated (r = 0.67, P < 0.001). All mechanical traits except specific punch strength were strongly positively correlated with LMA, particularly EI<sub>W</sub>, and with SI (Table 4). All nonmechanical traits except N:P and leaf size correlated with LMA (Table 4). Notably, while N<sub>mass</sub> and P<sub>mass</sub> correlated strongly and negatively with LMA (r = -0.78, -0.68, respectively; P < 0.001), N<sub>water</sub> and P<sub>water</sub> correlated less strongly and positively with LMA (r = 0.38, P = 0.002; r = 0.36, P = 0.004, respectively). Water content and protein: tannin activity also correlated negatively with LMA, but other nonmechanical traits correlated positively (Table 4). Notably, investment in putative chemical defenses (total phenolics and tannin activity) as well as physical defenses (strength, toughness, etc.) increased as mass investment per leaf area increased (Fig. 3).

N<sub>mass</sub> and P<sub>mass</sub> were strongly negatively correlated with mechanical traits, and with C:N, NDF, ADL, total phenolics, tannin activity, leaf thickness, and tissue density, and

positively with protein: protein precipitation and water content (Table 4). They did not correlate with  $N_{water}$  and  $P_{water}$  (P > 0.05), and the latter traits correlated with few other traits (Table 3). N:P did not correlate with any mechanical trait (P > 0.05).

These and other correlations are summarized in the PCA plot (Fig. 4A), with 63% of variation among species explained by the first two components. The traits contributing most to the first component (explaining 50% of the variation) were structural strength and toughness (work to punch and to shear), SI, LMA, and C:N (positively), and N<sub>mass</sub> and water content (negatively) (component loadings ≥0.85) (Fig. 4B). Clustering was consistent with ANOVA trends, but showed more clearly the relationship between the sclerophyll matorral and other forms of matorral (Fig. 4A). In particular, littoral matorral showed little overlap with inland matorral, but overlapped substantially with lowland sclerophyll matorral, which occurred near the coast and also experiences fogs. Second, the three forms of sclerophyll matorral showed considerable trait similarity, but with wider variation in the lowland sclerophyll matorral (Fig. 4A). Arid matorral species had traits intermediate between those of littoral and inland matorral. It was also evident that, overall, leaves investing highly in mass per area were, on average, thicker, denser, stronger, tougher, with more fiber and lignin, more fiber per unit protein, more phenolics, less protein per unit tannin activity, and less N, P, and water per dry mass (Fig. 4B).

Some differences were recorded within species sampled from multiple vegetation types, such as slightly higher water content in populations in littoral matorral than at other sites. However, generally there was little difference between sampled populations, as summarized in the PCA (Fig. 4A).

Soil and climate variables were not intercorrelated (see Table 1 for climate variables tested), except that P<sub>soil</sub> correlated negatively with AP: ET<sub>0</sub> (Spearman correlation,  $r_{\rm S} = -0.98$ , P = 0.005; n = 5 sites) and AMT (annual mean temperature) negatively with ET<sub>0</sub> ( $r_S = -0.98$ , P = 0.005; n = 5). There was no correlation of LMA, punch traits, or  $EI_{W}$  with  $N_{soil}$  or  $P_{soil}$  ( $r_{S} = -0.11$  to 0.08, P = 0.32-0.99; *n* species = 61-75). LMA correlated negatively with AMT ( $r_S = -0.43, P < 0.001; n \text{ species} = 74$ ) and positively with moisture variables ( $r_S = 0.31-0.34$ , P = <0.001-0.008; n species = 74) except AP: ET<sub>0</sub>, that is, generally opposite to the direction expected if there was a causal relationship, except for ET<sub>0</sub>. Of the punch variables, only work to punch and punch strength correlated with any climate traits: weakly and negatively with AMT  $(r_S = -0.24, P = 0.040; n \text{ species} = 75)$  for the former and positively with AP ( $r_S = 0.23$ , n = 75: P = 0.047; nspecies = 75) for the latter, again, generally opposite to the

**Table 3.** Variation in selected leaf traits among leaf life span classes of Chilean matorral species. Potential leaf life spans of each species (across all types of matorral) were categorized as <1 year, 1+ years ( $\geq$ 1 year but <2 years) and 2–3 years. The values presented are means of species' means  $\pm$  standard errors. All data were log-transformed ( $_{L}$ ) for analysis.

	Leaf life span (year				
Leaf trait	<1	1+	2–3	F	Р
N <sub>mass</sub> (%) <sub>L</sub>	$2.79 \pm 0.19^{a}$	1.56 ± 0.09 <sup>b</sup>	1.00 ± 0.05 <sup>c</sup>	54.0	<0.001
N <sub>water</sub> (mg g <sup>-1</sup> water) <sub>L</sub>	$10.5\pm1.2$	$10.7\pm0.7$	$10.6 \pm 0.6$	0.5	0.578
$P_{\text{mass}} (\text{mg g}^{-1})_{\text{L}}$	$2.72\pm0.11^{a}$	$1.51\pm0.13^{b}$	$0.98\pm0.06^{c}$	43.3	< 0.001
P <sub>water</sub> (mg g <sup>-1</sup> water) <sub>L</sub>	$1.00 \pm 0.10$	$1.04 \pm 0.11$	$1.05\pm0.07$	0.3	0.757
NDF (%) L	$18.4 \pm 1.5^{a}$	$23.0\pm1.3^{b}$	$34.6 \pm 0.3^{c}$	14.6	< 0.001
ADL (%) L	$4.8\pm0.9^{a}$	$8.3\pm1.2^{b}$	$8.7\pm1.4^{b}$	7.5	< 0.001
Total phenolics (g GAE. 100 g <sup>-1</sup> ) <sub>L</sub>	$2.4\pm0.3^{a}$	$4.8\pm0.6^{b}$	$5.7\pm0.6^{b}$	12.2	< 0.001
Protein precipitation (mg g <sup>-1</sup> ) <sub>L</sub>	$69 \pm 22^a$	$252\pm53^b$	$339\pm65^b$	9.2	< 0.001
Protein: protein precipitation (g $g^{-1}$ ) L	$6.4\pm0.8^{a}$	$3.0\pm0.7^{b}$	$0.6 \pm 0.2^{c}$	17.5	< 0.001
LMA (g m <sup>-2</sup> ) <sub>L</sub>	$79\pm9^a$	$168\pm13^{b}$	$209\pm18^{b}$	31.7	< 0.001
Punch strength (MN m <sup>-2</sup> ) <sub>L</sub>	$3.0\pm1.3^{a}$	$5.9\pm1.2^{\rm b}$	$14.1 \pm 1.7^{c}$	42.3	< 0.001
Specific punch strength (GN m <sup>-2</sup> m <sup>-1</sup> ) <sub>L</sub>	$15 \pm 3^{a}$	$19\pm3^a$	$39\pm4^{b}$	10.8	< 0.001
Work to punch (kJ m <sup>-2</sup> ) <sub>L</sub>	$0.7\pm0.4^{a}$	$1.6\pm0.4^{b}$	$3.6\pm0.6^{c}$	29.9	< 0.001
Specific work to punch (MJ m <sup>-2</sup> m <sup>-1</sup> ) <sub>L</sub>	$3.4\pm0.8^a$	$4.9\pm0.8^{b}$	$9.5\pm1.1^{c}$	15.5	< 0.001
El <sub>W</sub> (mN m <sup>2</sup> m <sup>-1</sup> ) <sub>L</sub>	$0.3\pm0.4^a$	$1.3\pm0.4^b$	$3.7\pm0.6^c$	20.5	<0.001

NDF, neutral detergent fiber; ADL, acid detergent lignin; GAE, gallic acid equivalents; LMA, leaf dry mass per area; *El*<sub>W</sub>, flexural stiffness. Results are given for ANOVA (significant *P*-values are shown in bold) with *post hoc* Tukey's tests where appropriate (shared alphabet letters indicate no significant difference). Note that the leaves in the two upper size classes were the same age at the time of sampling, but that leaves in the shorter life span class were younger (<1 year old) although mature at the time of sampling.

**Table 4.** Pearson correlations (r) of LMA, SI, and leaf nutrients with other measured leaf traits in Chilean material species. Values for species measured at multiple sites were averaged (n = 51-63).

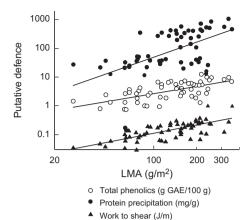
	LMA <sub>L</sub>	SI <sub>L</sub>	N <sub>mass L</sub>	P <sub>mass L</sub>	N <sub>water L</sub>	P <sub>water L</sub>
Nitrogen: phosphorus	0.04	-0.13	0.11	-0.51***	0.42**	-0.32*
Carbon: nitrogen	0.76***	0.88***	-0.96***	-0.81***	0.03	0.04
NDF L	0.40**	0.89***	-0.63***	-0.47***	0.01	0.19
ADL L	0.45***	0.70***	-0.56***	-0.45***	0.13	0.19
Total phenolics L	0.58***	0.56***	-0.66***	-0.59***	0.12	0.10
Protein precipitation L	0.50***	0.48***	-0.54***	-0.54***	0.25	0.16
Protein: protein precipitation L	-0.65***	-0.64***	0.73***	0.60***	-0.14	-0.18
Water L	-0.85***	-0.66***	0.71***	0.73***	-0.66***	-0.53***
Leaf size L	0.03	0.29*	-0.21	-0.05	-0.05	0.11
Thickness L	0.78***	0.42**	-0.58***	-0.42**	-0.09	-0.02
Tissue density L	0.72***	0.56***	-0.58***	-0.61***	0.70***	0.58***
Punch strength L	0.75***	0.78***	-0.74***	-0.66**	0.26*	0.24
Specific punch strength L	0.21	0.54***	-0.38**	-0.36**	0.27*	0.25
Work to punch L	0.80***	0.71***	-0.73***	-0.60***	0.12	0.14
Specific work to punch L	0.45***	0.63***	-0.53***	-0.43***	0.14	0.18
El <sub>W L</sub>	0.88***	0.62***	-0.69***	-0.56***	0.24	0.25

L, log-transformed for analysis. NDF, neutral detergent fiber; ADL, acid detergent lignin; LMA, leaf dry mass per area; SI, Loveless sclerophylly index;  $EI_{W}$ , flexural stiffness.

Asterisks indicate *P*-values: \*P < 0.05; \*\*P < 0.01; \*\*\*P < 0.001.

direction expected if causal.  $EI_W$  also correlated negatively with AMT ( $r_S = -0.36$ , P = 0.005; n species = 61) and positively with PDQ (precipitation of the driest quarter) ( $r_S = 0.27$ , P = 0.037; n species = 61), and notably, posi-

tively with ET<sub>0</sub> ( $r_S = 0.31$ , P = 0.016; n species = 61). Work to shear, specific work to shear, and PCA Component 1 did not correlate with any environmental variables (P > 0.05).

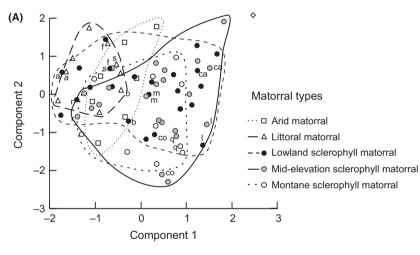


**Figure 3.** The relationship between leaf dry mass per area and putative defenses (total phenolics, tannin activity (protein precipitation), and work to shear) of evergreen species across matorral types in Chile. Work to shear was chosen from the suite of measured mechanical traits to allow comparison with relationships measured by Read et al. (2009). Species sampled at multiple vegetation types are included as a single data point (averaged values). GAE, gallic acid equivalents.

# Leaf trait relationships with soil and climate across Southern Hemisphere regions

Trends in leaf traits recorded among Chilean species were largely also recorded across the full dataset of the Southern Hemisphere regions, despite deciduous species being absent from the vegetation of both the non-Chile study areas. LMA was strongly positively correlated (log–log) with most mechanical traits (excluding tearing tests which were not undertaken in Western Australia) across the full dataset across regions (r = 0.41-0.85, P = <0.001-0.009). The exception was with specific punch strength (r = 0.29, P = 0.078). Strongest associations were with structural traits ( $EI_{\rm W}$ , followed by strength and work,  $r^2 = 0.52-0.72$ ).

For evergreen species, the relationship of work to punch with LMA differed significantly among regions (ANCOVA: F = 13.3; P < 0.001), with lower values of work for a given LMA in Chilean matorral than in other regions (P = <0.001-0.004) (Fig. 5). Second, across all evergreen species, LMA and all punch variables were



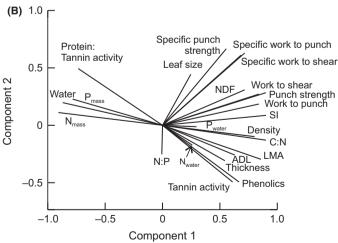


Figure 4. Results of principal components analysis of species across matorral types in Chile, based on measured leaf traits. Traits from tearing and bending tests were not included because they could not be measured across all species. (A) Score plot, with spatial position of species reflecting similarity in leaf traits. Circles, sclerophyll matorral; triangles, littoral matorral; squares, arid matorral; diamond, Pouteria splendens at Los Molles. Species sampled from multiple sites were included as individual data points (two could not be included because data were unavailable for some traits): a, Ageratina glechonophylla; ca, Cryptocarya alba; co, Colliguaja odorifera; f, Flourensia thurifera; I, Lithraea caustica; m, Maytenus boaria; r, Retanilla trinervia; q, Quillaja saponaria; s, Lepechinia salviae. Transformation of leaf traits is the same as listed in Table 2. (B) Component loadings plot, showing the relationships of traits with each other and the principal components. NDF, neutral detergent fiber; ADL, acid detergent lignin; LMA, leaf dry mass per area; SI, Loveless sclerophylly index.

strongly negatively correlated with P<sub>mass</sub> and N<sub>mass</sub> (loglog relationship) (Table 5, Fig. 6), as in Chilean matorral. Notably, they also correlated negatively, but more weakly, with Pwater, except specific punch strength (Table 5), in contrast to the Chilean matorral. Only specific punch strength correlated with N<sub>water</sub> across regions, but positively (Table 5). These and other correlations are summarized by the PCA (Fig. 7A). The first component explained 51% of the total variance, with SI, work to punch, Pmass, LMA, and punch strength contributing most strongly (component loadings >0.85), and Nwater and specific punch strength contributing most to the second component explaining 14% of the variance (component loadings >0.70) (Fig. 7B). Overall, leaves of evergreen species in West Australian woodland and shrubland and New Caledonian maquis were tougher, stronger, with higher SI and LMA, lower N<sub>mass</sub> and P<sub>mass</sub>, and lower protein: tannin activity than plants in Chilean matorral and New Caledonian dry forest (Fig. 7A and B).

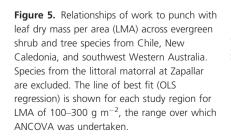
Stronger correlations were recorded with environmental variables across regions than across Chilean matorral. LMA and mechanical traits were strongly negatively correlated with Psoil and positively with ETo and PDQ (Table 5), the latter in the opposite direction to that expected if causal. When values were averaged for sites, most mechanical traits and LMA correlated strongly and negatively with  $P_{soil}$  across regions (r = -0.86 to -0.68; P = 0.002-0.035) (Fig. 8), and positively with  $ET_0$  (r = 0.68-0.87; P = 0.003-0.044) (all variables logtransformed). In addition, the first component of the PCA correlated negatively with  $P_{soil}$  (r = -0.87, P = 0.003) and positively with ET<sub>0</sub> (r = 0.78, P = 0.021) (environmental variables log-transformed). Only P<sub>soil</sub> and ET<sub>0</sub> made significant independent contributions to variation in punch variables among regions (Table 6), with  $P_{soil}$  contributing singly (strength and specific strength) or with  $ET_0$  (work and specific work) up to 90% of the variation in the dependent variables (Table 6). High but nonsignificant independent contributions were made by  $P_{soil}$  and  $ET_0$  to LMA (Table 6).

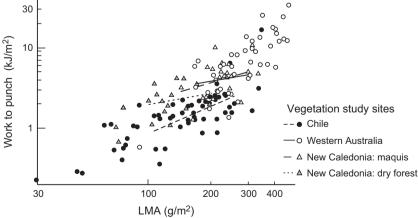
**Table 5.** Relationships of leaf dry mass per area (LMA) and mechanical traits (punch variables) with nutrient concentration in leaf and soil, and with climate variables, in evergreen species from shrubland and woodland in three regions: southwest Western Australia, New Caledonia, and Chile (data from this paper, excluding the littoral Zapallar site). The data are Pearson r-values (n = 140, across nine soils and seven climates).

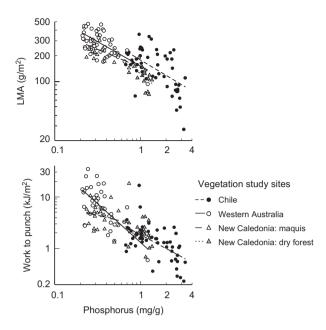
	LMA L	Strength <sub>L</sub>	Specific strength <sub>L</sub>	Work L	Specific work <sub>L</sub>
P <sub>mass L</sub>	-0.74***	-0.64***	-0.27**	-0.77***	-0.63***
P <sub>water L</sub>	-0.46***	-0.34***	0.03	-0.54***	-0.37***
N <sub>mass L</sub>	-0.73***	-0.60***	-0.26**	-0.63***	-0.48***
N <sub>water L</sub>	-0.17	0.01	0.29**	-0.13	0.06
P <sub>soil L</sub>	-0.57***	-0.53***	-0.24**	-0.63***	-0.53***
N <sub>soil L</sub>	-0.27**	-0.13	0.08	-0.17*	-0.05
AMT <sub>L</sub>	-0.06	0.14	0.19*	0.16	0.24**
AP L	0.07	0.13	0.07	0.18*	0.17
PDQ L	0.31***	0.35***	0.20*	0.45***	0.43***
ET <sub>o L</sub>	0.52***	0.51***	0.27**	0.66***	0.60***
AP:	-0.22	-0.17	-0.04	-0.23	-0.16
ET <sub>O L</sub>					

L, log-transformed for analysis. AMT, annual mean temperature; AP, annual precipitation; PDQ, precipitation of the driest quarter; ET<sub>0</sub>, annual reference evapotranspiration. Climate data were obtained as described for Chile study sites, except for New Caledonia where AMT was estimated by WorldClim v. 1.4 (Hijmans et al. 2005; 30 arc seconds resolution) with DIVA-GIS v. 7 (Hijmans et al. 2012), and AP from local stations and isohyet maps (Read et al. 2006) (Appendix S3).

Asterisks indicate *P*-values: \*P < 0.05; \*\*P < 0.01; \*\*\*P < 0.001.







**Figure 6.** Relationships of leaf dry mass per area and work to punch with foliar P concentration (P<sub>mass</sub>) across evergreen shrub and tree species from Chile, New Caledonia, and southwest Western Australia. Species from the littoral matorral at Zapallar are excluded. The line of best fit (OLS regression) is shown for each study region.

#### Discussion

#### Variation in matorral leaf form

Leaf form varied considerably among Chilean matorral species, including within types of matorral (see also Rundel 1981). Some species were summer-deciduous, with LLS of c. 3–4 months (e.g., Mooney and Kummerow 1971), but most were evergreen, although with a LLS range of <1 year to c. 3 years. A small number of evergreen sclerophylls, such as *Kageneckia angustifolia* (Cavieres et al. 2007), *Quillaja saponaria*, and *Colliguaja odorifera*, show significant leaf loss during summer (Rundel 1981). Differences in rooting depth and physiology among species are likely to be associated with variation in leaf habit (Giliberto and Estay 1978; Montenegro et al. 1979) and may contribute to the wide variability in leaf mechanics recorded within matorral types.

There was particularly strong variation in structural mechanical traits, from leaves that were soft, often short-lived (including deciduous species), with high nutrient concentrations, to leaves that were strong, stiff, and tough, having, on average, high LMA, C:N, SI, and tissue density, and low N, P, and water concentration. Species with high LMA tended to be tough with high tannin activity and levels of total phenolics. Hence, additive investment in C-based mechanical and chemical defenses

may be cost-effective on these low-nutrient soils where plant carbohydrates may be in surplus, as suggested in other sclerophyll vegetation (Read et al. 2009). N limitation was common, but 70% of species had N:P ratios of 10–20, suggesting growth may be limited by factors other than N or P (Güsewell 2004). This differs considerably from sclerophyll vegetation in Australia (Read et al. 2000, 2005) and in maquis in New Caledonia (Read et al. 2006), where N:P ratios are generally high, with low P<sub>mass</sub>, suggesting strong P limitation (Güsewell 2004).

### Variation in sclerophylly among types of matorral

Environmental severity was high in the arid matorral (aridity), the mid-elevation sclerophyll matorral (aridity and low-nutrient soils), and the montane sclerophyll matorral (low winter temperatures and aridity). In particular, the sclerophyll matorral vegetation types contained many species with strong, tough, and stiff leaves, both at the structural level and per unit leaf thickness, with high LMA and tissue density, and with notably low levels of protein: tannin activity, and high levels of phenolics, tannin activity, and ADL. Hence, they were potentially well protected against biotic and abiotic stresses. These associated leaf traits form a broad sclerophyll syndrome that has been reported elsewhere, consistent with low soil fertility and/or seasonal dryness (Read et al. 2005, 2006).

However, despite the relatively severe conditions, there was considerable within-vegetation trait variability, as noted in other studies (Ordonez et al. 2009). In the arid matorral, this was largely due to a high representation of soft-leaved deciduous species, as well as strong and tough evergreen species. Similarly, in the sclerophyll matorral, trait variability appeared largely related to the range of LLS among species, particularly in the lowland sclerophyll matorral due to environmental amelioration (moisture, N, and temperature) related to fog (e.g., Weathers et al. 2000). It is expected that if traits were weighted by species abundance, much stronger patterns in leaf traits would be evident among the matorral types. For example, toughleaved species such as Lithraea caustica (Anacardiaceae) and Cryptocarya alba (Lauraceae) dominated the lowmid-elevation sclerophyll matorral. Nevertheless, it is clear that a range of plant strategies, exemplified by the range of leaf trait combinations observed within sites, are successful in these low-resource environments, probably reflecting a range of variation in biomass partitioning, physiology, and phenology (Giliberto and Estay 1978; Montenegro et al. 1979; Rundel 1981), as well as withinsite environmental heterogeneity.

The littoral matorral must also experience stress from exposure to coastal winds and salinity. Leaf traits

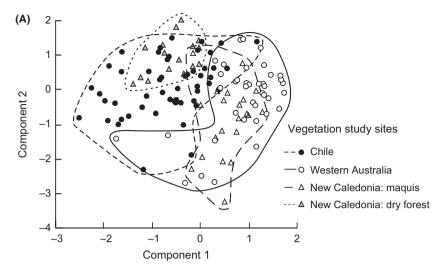
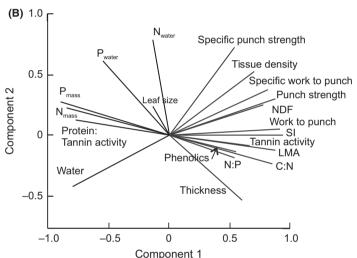


Figure 7. Results of principal components analysis of evergreen shrub and tree species from Chile, New Caledonia, and southwest Western Australia. The same leaf traits were used in this analysis as shown in Figure 4, except that only punch data were used of the mechanical traits, and excluding acid detergent lignin (not measured in earlier studies), and the littoral matorral at Zapallar. (A) Score plot. with spatial position of species reflecting similarity in leaf traits. Average trait values were used for species sampled from multiple sites. Transformation of leaf traits is the same as listed in Table 2. (B) Component loadings plot, showing the relationships of traits with each other and the principal components. NDF, neutral detergent fiber; LMA, leaf dry mass per area; SI, Loveless sclerophylly index.



contrasted greatly with those of sclerophyll matorral species, with low values of most mechanical traits, LMA, and tissue density, as well as other traits associated with defense. These differences are probably largely due to physiological and morphological responses to the overriding stress imposed by the saline windy environment, but may in part be indirect effects, related to the apparently shorter LLS of many species in this vegetation.

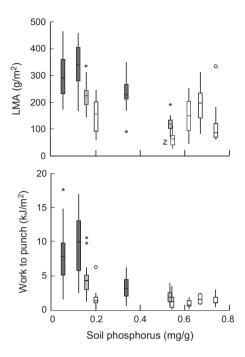
Environmental complexity (including effects of fog) and low statistical power limited explanation of variation in leaf traits among types of matorral. There was strong negative correlation of LMA, SI, and mechanical traits with  $P_{mass}$  and  $N_{mass}$  across species. However, this trend is in part due to dilution of foliar nutrients per unit mass by cell wall in scleromorphic species, as suggested by lack of trends in  $P_{water}$  and  $N_{water}$  (see also Read et al. 2005, 2006), and there were no significant correlations of LMA or mechanical traits with  $N_{soil}$  or  $P_{soil}$ . There was also lit-

tle correlation with climate variables in a direction likely to indicate causality, although some components of scleromorphy may enhance frost resistance (Larcher 2005) and contribute to negative correlations of LMA, work, and  $EI_{\rm W}$  with AMT. Nevertheless, the positive correlation of  $EI_{\rm W}$  and LMA with ET $_{\rm 0}$  suggests water availability has a strong influence on variation in leaf structure among these types of matorral.

# Associations of mechanical traits with soil and climate across Southern Hemisphere regions

Summer-deciduous species were more common in Chilean matorral (see also Lamont 1994) and LLSs of evergreen species were shorter on average than those in some sclerophyll communities in southwest Western Australia, for example, mode of 3 years (J. Read, unpubl. data).

Leaves of evergreen Chilean matorral species were on average softer, with lower LMA, higher nutrient content per mass, and less defended than those of southwest Aus-



**Figure 8.** Relationships of leaf dry mass per area and work to punch with total soil P ( $P_{soil}$ ) in evergreen shrub and tree species from Chile, New Caledonia, and southwest Western Australia. Box plots are shown for each vegetation. The littoral matorral at Zapallar is included, indicated by "z". Chile sites, white; New Caledonia sites (maquis and dry forest), light gray; Western Australia sites (on dolerite and laterite soils and gray sand), dark gray.

tralia or maquis in New Caledonia. LMA and mechanical traits of Chilean matorral species rarely reached the high values seen in species from the latter vegetation types. Soil total P and ET<sub>0</sub> were the only environmental variables that made a significant independent contribution to variance in mechanical traits of evergreen species among regions, even given potential N limitation in some Chilean species. Total N is not necessarily a good indicator of soil N availability, possibly contributing to its often poor relationships with leaf traits, whereas total P may provide a fairly robust index of P fertility (Ordoñez et al. 2009). Notably, Pwater, an estimate of protoplasmic P concentration, was also negatively correlated with LMA and most mechanical traits across these vegetation types. Interestingly, ET<sub>0</sub> contributed independently only to work and specific work, rather than to strength and specific strength, suggesting a different anatomical basis of these mechanical traits.

Overall, somewhat similar tradeoffs between leaf traits associated with growth versus resource conservation were suggested across these shrublands and woodlands to those noted in global comparisons of leaf traits across a much wider variety of vegetation types (e.g., Wright et al. 2005; Ordoñez et al. 2009; Onoda et al. 2011). Our study suggests important independent roles of moisture availability (ET<sub>0</sub>) and soil P in variation in leaf texture (LMA and leaf mechanics, the effects differing among mechanical traits) across shrubland and woodland at a broad geographic scale, with the combination of ET<sub>0</sub> and soil P explaining 87–90% of variation in work to shear and specific work to shear. Important roles of soil P and moisture availability have been shown recently in

**Table 6.** Independent contributions of soil nutrient concentrations and climate variables to mechanical traits (punch variables) and LMA in evergreen species from shrubland and woodland in three regions: southwest Western Australia, New Caledonia, and Chile (data from this paper, excluding the littoral Zapallar site). The data presented are  $I_{HP}$ , the independent contribution of predictor variables to explained variance based on hierarchical partitioning, with *Z*-scores derived by randomization techniques and statistical significance based on the upper 95 percentile of the normal distribution ( $Z \ge 1.65$ , shown in bold type). The data used in the analysis were site averages (n = 9 sites). *F*- and  $r^2$ -values are presented from regression of significant predictor variables; in the case of LMA, the two variables with high but nonsignificant *Z*-values were included.

	$LMA_L$		Strength <sub>i</sub>	L	Specific :	strength <sub>L</sub>	Work L		Specific v	work <sub>L</sub>
	I <sub>HP</sub>	Z	I <sub>HP</sub>	Z	I <sub>HP</sub>	Z	I <sub>HP</sub>	Z	I <sub>HP</sub>	Z
P <sub>soil L</sub>	0.30	1.38	0.43	2.47	0.35	2.00	0.35	1.80	0.33	1.98
N <sub>soil L</sub>	0.07	-0.46	0.08	-0.26	0.10	-0.16	0.06	-0.57	0.06	-0.76
AMT <sub>I</sub>	0.11	-0.20	0.03	-0.94	0.17	0.46	0.02	-0.84	0.06	-0.69
AP <sub>I</sub>	0.07	-0.50	0.04	-0.78	0.08	-0.27	0.05	-0.76	0.06	-0.53
PDQ <sub>1</sub>	0.14	0.13	0.13	0.26	0.10	-0.33	0.16	0.34	0.16	0.19
ET <sub>O I</sub>	0.28	1.45	0.27	1.28	0.19	0.61	0.34	2.70	0.32	2.06
F	7.5*		19.3**		5.9*		27.1**		19.7**	
$r^2$	0.72		0.73		0.46		0.90		0.87	

L, log-transformed for analysis; LMA, leaf dry mass per area; AMT, annual mean temperature; AP, annual precipitation; PDQ, precipitation of the driest quarter;  $ET_0$ , annual reference evapotranspiration.

Asterisks indicate *P*-values for regressions: \*P < 0.05; \*\*P < 0.01.

photosynthetic and associated leaf traits at a global scale (Maire et al. 2015). Notably, precipitation:  $ET_0$  (Maire et al. 2015) was not correlated with LMA and mechanical traits in our study, probably due to high variability in rainfall seasonality across the small number of sites.

If sclerophylly is predominantly a consequence of evolved responses to low availability of either nutrients or water, different anatomical traits may contribute to the variation in leaf textures among differing environments, depending on the degree to which each factor is limiting, or has an additive or interactive effect. In contrast, if sclerophylly has evolved predominantly by enhancing protection against damage in suboptimal environments, the suites of anatomical features contributing to texture may be more similar among differing environments, although differing environments may influence the efficiency of various forms of protection. If instead sclerophylly is a variable syndrome of traits, to which multiple factors contribute, identifying the relative contribution of direct v. indirect adaptations v. nonadaptive factors is likely to be difficult. Nevertheless, the simple comparison of work to punch with LMA suggests a difference in the anatomical or material basis of sclerophylly in the Chilean species compared with those from the other Southern Hemisphere regions, and possibly a differing adaptive basis. Studies of leaf anatomy (Kummerow 1973; Read et al. 2000) and tissue-level mechanical traits (Méndez-Alonzo et al. 2013) across these regions should assist in addressing these questions.

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## **Conflict of Interest**

None declared.

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# **Supporting Information**

Additional Supporting Information may be found in the online version of this article:

**Appendix S1.** Comparisons of leaf traits of evergreen species among matorral vegetation types.

**Appendix S2.** Pearson correlation matrix of mechanical traits from shear, punch, tearing and bending tests across Chilean matorral species.

**Appendix S3.** Environmental data from New Caledonia (NC) and Western Australia (WA) used in meta analysis of leaf traits.

**Appendix:** Species sampled in matorral in central Chile, and their life form and leaf habit. Nomenclature was taken from "The International Plant Names Index" (http://www.ipni.org/) and "The Plant List" (http://www.theplantlist.org/) (accessed 16/12/2015) with additional leaf habit (e, evergreen; d, deciduous; sd, semi-deciduous) and plant habit information from Hoffmann (1978), Montenegro et al. (1979), and Ginocchio and Montenegro (1992). "Subshrub" is used here to refer to weakly woody shrubs. The final column indicates the mechanical tests undertaken: p, punching; s, shearing; t, tearing; b, bending. Adesmia microphylla (Fabaceae) occurred in most matorral sites but was only included at Las Chinchillas because of difficulty conducting mechanical tests on the tiny leaflets.

Anish madamak Lan Chimakillan			
Arid matorral: Las Chinchillas	a al	ale en els	_
Adesmia microphylla (Fabaceae)	sd	shrub	р
Bridgesia incisifolia (Sapindaceae)	d	shrub	ps
Cordia decandra (Boraginaceae)	е	shrub	psb
Flourensia thurifera (Asteraceae)	d	shrub	all
Krameria cistoidea (Krameriaceae)	е	shrub	р
Llagunoa glandulosa (Sapindaceae)	е	shrub	р
Porlieria chilensis (Zygophyllaceae)	е	shrub, small tree	р
Proustia ilicifolia (Asteraceae)	е	shrub, small tree	all
Proustia cuneifolia	d	shrub	psb
Senna arnottiana (Caesalpiniaceae)	е	shrub	psb
Littoral matorral: Zapallar			
Ageratina glechonophylla (Asteraceae)	е	subshrub	all
Baccharis vernalis (Asteraceae)	е	shrub	pst
Bahia ambrosioides (Asteraceae)	е	subshrub	ps
Flourensia thurifera	d	shrub	all
Fuchsia lycioides (Onagraceae)	sd	shrub	all
Haplopappus foliosus (Asteraceae)	е	subshrub	pst
Lepechinia salviae (Lamiaceae)	е	subshrub	all
Lobelia polyphylla (Campanulaceae)	е	shrub	all
Mathewsia foliosa (Brassicaceae)	е	subshrub	ps
Senecio sinuatilobus (Asteraceae)	е	subshrub	all
Lowland sclerophyll matorral: Cachagua			
Adenopeltis serrata (Euphorbiaceae)	е	shrub	all
Aextoxicon punctatum (Aextoxicaceae)	е	tree	all
Ageratina glechonophylla	е	subshrub	all
Aristeguietia salvia (Asteraceae)	е	shrub	all
Azara celastrina (Salicaceae)	е	small tree	all
Baccharis vernalis (Asteraceae)	е	shrub	ps
Beilschmiedia miersii (Lauraceae)	е	tree	all
*			

(Continued).

# Appendix. Continued.

Calceolaria morisii (Calceolariaceae)         e         stree           Citronella mucronata (Cardioptenideceae)         e         tree           Colliguaja dordine (Euphorbiaceae)         e         shrub           Cryptocarya alba (Lauraceae)         e         small tree           Escallonia revolta (Escalloniaceae)         e         small tree           Fuchsia yelioides         sd         shrub           Lepechinia salviae         e         subshrub           Lepechinia salviae         e         stree           Myreugenia (Castageae)         e         stree           Myreugenia correifola (Myrtaceae)         e         stree           Myreugenia correifola (Myrtaceae)         e         stree           Retarulla trineria (Bhamnaceae)         d         stree           Retarulla trineria (Bhamnaceae)         e         strub           Retarulla trineria (Grassplariaceae)         e         small tree           Schinus politicis (Saleocarpaceaea)         e         small tree <th>11</th> <th></th> <th></th> <th></th>	11			
Colliguaja donfriera (Euplinothiaceae)         e         shrub           Cryptocarya alba (Lauraceae)         e         small tree           Fuchsia Iyrioides         sd         shrub           Lepechinia salviae         e         subshrub           Lithriaes caustica (Anacrdiaceae)         e         tree           Lobeia excesa (Campanulaceae)         e         tree           Maytenus boaria (Celastraceae)         e         tree           Myrecugenia correifolia (Myrtaceae)         e         tree           Myrecugenia obtusa         e         small tree           Peumus boldus (Monimiaceae)         e         tree           Ribes punctatum (Grossubriaceae)         e         shrub           Schinus latifolius (Anacardiaceae)         e         small tree           Sema candolleara (Caesalpiniaceae)         e         shrub           Schinus latifolius (Anacardiaceae)         e         small tree           Sema cardolleara (Caesalpiniaceae)         e         small tree           Senius actifolius (Anacardiaceae)         e         small tree           Selius (Shalaceae)         e         small tree           Selius (Shalaceae)         e         small tree           Cestrum parqui (Solanaceae)         e	alceolaria morisii (Calceolariaceae)	е	subshrub	all
Cryptocaya alba (Lauraceae)         e         small tree           Excallonia revolutz (Escalloniaceae)         e         smult           Lepechina salviae         e         subshrub           Lepechina salviae         e         tree           Lobelia excelsa (Campanulaceae)         e         shrub           Maytenus boaria (Celastraceae)         e         tree           Myrecugenia correifolia (Myrtaceae)         e         tree           Myrecugenia obtusa         e         small tree           Peumus boblux (Monimiaceae)         e         tree           Retarilla trinervia (Rhamnaceae)         d         shrub           Ribes punctatum (Grossulariaceae)         e         shrub           Schinus laitofius (Anacardiaceae)         e         shrub           Schinus laitofius (Anacardiaceae)         e         shrub           Schinus (Selatolius (Salariaceae)         e         shrub           Schinus (Saltofius (Salariaceae)         e         small tree           Schinus (Saltofius (Salariaceae)         e         small tree           Baccharis Sp. (Asteraceae)         e         shrub           Berberis Chileriss (Eleocarpaceae)         e         shrub           Berberis Chileriss (Eleocarpaceae)         e <td>itronella mucronata (Cardiopteridaceae)</td> <td>е</td> <td>tree</td> <td>all</td>	itronella mucronata (Cardiopteridaceae)	е	tree	all
Eicallonia revoluta (Escalloniaceae)         e         small tree           Fuchsia lycioides         sd         shrub           Lepechinia salviae         e         subshrub           Lepechinia salviae         e         tree           Lobelia excesa (Campanulaceae)         e         tree           Maytenus boana (Celastraceae)         e         tree           Myrceugenia correifolia (Myrtaceae)         e         small tree           Myrceugenia obtusa         e         small tree           Peumus boldus (Monimiaceae)         d         shrub           Ribes punctatum (Grossulariaceae)         e         small tree           Ribes punctatum (Grossulariaceae)         e         shrub           Schirus latifolias (Anacardiaceae)         e         shrub           Schirus latifolias (Shaccaeae)         e         small tree           Schirus latifolias (Shaccaeae)         e         small tree           Azara petiolaris (Salicaceae)         e         small tree           Berberis Oliterias (Salicaceae)         e         small tree           Berberis Oliterias (Beleacapaeae)         e         smulture           Colliquaja dolorifea         e         shrub           Colliquaja parqui (Solanaceae)         e	olliguaja odorifera (Euphorbiaceae)	е	shrub	all
Fuchsia ylcioides         sd         shrub           Lepechinia salviae         e         subshrub           Libraea caustica (Anacardiaceae)         e         tree           Lobelia excelsa (Campanulaceae)         e         shrub           Mayreus genia Celastraceae)         e         tree           Myreugenia obtusa         e         tree           Peuruns boldus (Monimiaceae)         e         tree           Retarilla trinervia (Rhamnaceae)         d         shrub           Rebes punctatum (Grossulariaceae)         e         small tree           Schinus Iattifolius (Anacardiaceae)         e         small tree           Schinus Scalicaceaeae         e         small tree           Schinus Scalicaceaeae         e         small tree           Baccharis Sp. (Asteraceae)         e         shrub           Berberis Schieras (Berberidaceae)         e         shrub           Colliquaja odorifera         e         shrub           Culliquaja odorifera         e         shrub	ryptocarya alba (Lauraceae)	е	tree	all
Lepech Ín a salviae         e         tree           Lithraea caustica (Anacardiaceae)         e         tree           Lobelia excèsa (Campanulaceae)         e         tree           Mayterus boaria (Celastraceae)         e         tree           Myrceugenia cotreifolía (Myrtaceae)         e         tree           Myrceugenia obtusa         e         tree           Peumus bolous (Monimiaceae)         d         shrub           Réastallia trineria (Rharmaceae)         d         shrub           Schinus latifolius (Anacardiaceae)         e         small tree           Senna candolieana (Caesalpiniaceae)         e         small tree           Baccharia S(Salicaceae)         e         small tree           Cestrum parqui (Solanaceae)         e         shrub           Colliquia	scallonia revoluta (Escalloniaceae)	е	small tree	all
Lithraea caustica (Anaçardiaceae)         e         shrub           Lobelia excelsa (Campanulaceae)         e         shrub           Maytenus boaria (Celastraceae)         e         tree           Myrceugenia Correlfolia (Myrtaceae)         e         small tree           Myrceugenia obtus         e         tree           Myrceugenia Obtus         e         tree           Retarilla trinervia (Rhamnaceae)         d         shrub           Schinus latifolius (Anacardiaceae)         e         shrub           Schinus latifolius (Anacardiaceae)         e         small tree           Schinus latifolius (Anacardiaceae)         e         small tree           Schinus faceafoliaris (Saliacaceae)         e         small tree           Schinus faceafoliaris (Saliacaceae)         e         small tree           Baccharis Sp. (Asteraceae)         e         small tree           Baccharis Sp. (Asteraceae)         e         shrub           Berberis Chilenisis (Berberidaceae)         e         shrub           Colliquia odorfiera         e         shrub           Colliquia odorfiera         e         shrub           Colliquia idorfiera         e         shrub           Escallonia illinita         e         tree <td>uchsia lycioides</td> <td>sd</td> <td>shrub</td> <td>all</td>	uchsia lycioides	sd	shrub	all
Lobelia excelsa (Campanulaceae)         e         tree           Maytenus boaria (Celastraceae)         e         tree           Myrceugenia correifola (Myrtaceae)         e         small tree           Peumus bodus (Monimiscae)         e         shrub           Retanilla trineria (Rharmaceae)         d         shrub           Ribes punctatum (Grossulariaceae)         e         shrub           Schinus latifolis (Anacardiaceae)         e         small tree           Sema candolleana (Caesalpiniaceae)         e         small tree           Mid-elevation sclerophyll matoral: Rio Clarillo         small tree           Aristotelia chilensis (Baecarpaceae)         e         small tree           Azara petiolaris (Salicaceae)         e         small tree           Baccharis Sp. (Asteraceae)         e         small tree           Berberis chilensis (Berberidaceae)         e         shrub           Cestrum parqui (Solanaceae)         e         shrub           Cestrum parqui (Solanaceae)         e         shrub           Cestrum parqui (Solanaceae)         e         shrub           Colliquaja odorifera         e         shrub           Cupitaria illinita         e         shrub           Litria pulverulenta (Escalloniaceae)	epechinia salviae	е	subshrub	all
Maytenus boaria (Celastraceae)         e         tree           Myrecugenia correliola (Myrtaceae)         e         tree           Myrecugenia obtusa         e         small tree           Peurnus boldus (Monimaceae)         e         tree           Retanilla trinervia (Rhamnaceae)         d         shrub           Ribes punctatum (Grossulariaceae)         e         shrub           Schinus latifolius (Anacardiaceae)         e         small tree           Schinus latifolius (Anacardiaceae)         e         shrub (small tree)           Senna candolleana (Cassalpiniaceae)         e         shrub (small tree)           Azia petiolaris (Elabeocarpaceae)         e         small tree           Azia petiolaris (Selicaceaee)         e         small tree           Baccharis sp. (Asteraceae)         e         shrub           Berberis Chiliensis (Berberidaceae)         e         shrub           Colliquaja odorifera         e         shrub           Colliquaja odorifera         e         tree           Colliquaja odorifera         e         tree           Escallonia julture (Iscalloniaceae)         e         tree           Escallonia jultinita         e         tree           Haplopappus sp. (Asteraceae)         e	thraea caustica (Anacardiaceae)	е	tree	all
Myrceugenia correifolia (Myrtaceae)         e         small tree           Myrceugenia obtusa         e         small tree           Peumus bofus (Monimiaceae)         e         tree           Retanila trinervia (Rhamaceae)         d         shrub           Schinus latifolius (Anacardiaceae)         e         small tree           Senna candolleana (Caesalpiniaceae)         e         small tree           Mid-elevation sclerophyll matorral: Rio Clarillo         small tree           Aristotelia chilensis (Elaeocarpaceae)         e         small tree           Azara petiolaris (Salicaceae)         e         small tree           Baccharis Sp. (Asteraceae)         e         shrub           Berberis chilensis (Berberidaceae)         e         shrub           Cestrum parqui (Solanaceae)         e         tree           Culliqua admitere         e         shrub           Escallonia pullevulenta (Escalloniaceae)         e         tree           Escallonia illinita         e	obelia excelsa (Campanulaceae)	е	shrub	all
Myrceugenia obtusa         e         tree           Peurmus boldus (Monimiaceae)         e         tree           Retanila trinervia (Rhamnaceae)         d         shrub           Ribes punctatum (Grossulariaceae)         e         shrub           Schinus latifolius (Anacardiaceae)         e         shrub (small tree           Schinus latifolius (Anacardiaceae)         e         shrub (small tree           Mid-elevation sclerophyll matorral: Rio Clarillo         strub (small tree)           Mistelevation sclerophyll matorral: Rio Clarillo         small tree           Aziza petioliaris (Salicaceae)         e         small tree           Aziza petioliaris (Salicaceae)         e         small tree           Baccharis (Selberiasis (Berberiasis (Berberias) (Berbaris) (Berbarias) (Berbaris) (B	laytenus boaria (Celastraceae)	е	tree	all
Peumus boldus (Monimiaceae)         e         tree           Retarilla trinervia (Rhamnaceae)         d         shrub           Ribes punctarum (Grossulariaceae)         e         shrub           Schinus latifolius (Anacardiaceae)         e         small tree           Senna candolleana (Caesalpiniaceae)         e         small tree           Midi-elevation sclerophyll matorral: Rio Clarillo         re         small tree           Azara petiolaris (Saliaceaee)         e         small tree           Azara petiolaris (Saliaceaee)         e         small tree           Berberis chilensis (Berberidaceae)         e         shrub           Berberis chilensis (Berberidaceae)         e         shrub           Cestrum parqui (Solanaceae)         e         shrub           Cestrum parqui (Solanaceae)         e         shrub           Cestrum parqui (Solanaceae)         e         shrub           Culliquia adorifera         e         shrub           Cyptocarya alba         e         tree           Drimys winteri (Winteraceae)         e         small tree           Escallonia illinita         e         shrub (small tree)           Haplopappus sp. (Asteraceae)         e         small tree           Escallonia illinita	lyrceugenia correifolia (Myrtaceae)	е	tree	all
Retanilla trinervia (Rhamnaceae)         d         shrub           Ribes punctatum (Grossulariaceae)         e         shrub           Schinus latifolius (Anacardiaceae)         e         small tree           Senna candolleana (Caesalpiniaceae)         e         small tree           Mid-levation sclerophyll matorral: Rio Clarillo         v         small tree           Azara petiolaris (Salicaceae)         e         small tree           Baccharis sp. (Asteraceae)         e         small tree           Baccharis sp. (Asteraceae)         e         shrub           Berberis chilensis (Berberidaceae)         e         shrub           Cestrum parqui (Solanaceae)         e         shrub           Culiguaja dodorfera         e         tree           Colliquaja odorifera         e         tree           Scallonia julinitar (Escalloniaceae)         e         small tree           Escallonia illinita (Escalloniaceae)         e         shrub (small tree)           Lithrace acustica         e         small tree           Luma chequen (Myrtaceae	lyrceugenia obtusa	е	small tree	pst
Ribes punctatum (Grossulariaceae)         e         shrub           Schinus latifolius (Anacardiaceae)         e         small tree           Senna candolleana (Caesalpiniaceae)         e         shrub (small tree)           Mid-elevation sclerophyll matorral: Rio Clarillo         small tree           Aristotelia chillensis (Elaeccaep)         e         small tree           Baccharis Sp. (Asteraceae)         e         shrub           Berberis (Selicaceae)         e         shrub           Berberis (Chilensis (Berberidaceae)         e         shrub           Cestrum parqui (Solanaceae)         e         shrub           Colliguaja odorifera         e         shrub           Colliguaja odorifera         e         shrub           Cyptocarya alba         tree         shrub           Drimys winteri (Winteraceae)         e         tree           Escallonia pulverulenta (Escalloniaceae)         e         small tree           Escallonia illinita         e         shrub           Haplopappus sp. (Asteraceae)         e         small tree           Lithraea caustica         e         tree           Luma chequen (Myrtaceae)         e         small tree           Persea lingue (Lauraceae)         e         tree		е	tree	all
Schinus latifolius (Anacardiaceae)         e         small tree           Senna candolleana (Caesalpiniaceae)         e         shrub (small tree)           Mid-elevation sclerophyll matorral: Rio Clarillo           Azistotelia chilensis (Elaeocarpaceae)         e         small tree           Azara petiolaris (Salicaceae)         e         small tree           Baccharis Sp. (Asteraceae)         e         shrub           Berberis chilensis (Berberidaceae)         e         shrub           Cestrum parqui (Solanaceae)         e         shrub           Cestrum parqui (Solanaceae)         e         shrub           Costilugia a dorifera         e         shrub           Cyptocarya alba         e         tree           Escallonia pulverulenta (Escalloniaceae)         e         small tree           Escallonia pulverulenta (Escalloniaceae)         e         small tree           Escallonia illinita         e         shrub           Haplopappus sp. (Asteraceae)         e         shrub           Lithraea caustica         e         shrub           Lithraea caustica         e         tree           Luma cheque (Myrtaceae)         e         small tree           Maytenus boaria         e         tree	etanilla trinervia (Rhamnaceae)	d	shrub	all
Senna candolleana (Caesalpiniaceae)         e         shrub (small tree)           Mid-elevation sclerophyll matorral: Rio Clarillo         X-ristotelia chilensis (Elaeocarpaceae)         e         small tree           Azara petiolaris (Salicaceae)         e         small tree           Baccharis sp.(Asteraceae)         e         shrub           Berberis chilensis (Berberidaceae)         e         shrub           Cestrum parqui (Solnanceae)         e         shrub           Colliguaja odorifera         e         tree           Cryptocarya alba         e         tree           Drimys winteri (Winterceae)         e         tree           Escallonia pulverulenta (Escalloniaceae)         e         small tree           Escallonia illinita         e         shrub (small tree)           Haplopappus sp. (Asteraceae)         e         small tree           Escallonia illinita         e         shrub (small tree)           Haplopappus sp. (Asteraceae)         e         small tree           Lithraea caustica         e         tree           Luma chequen (Myrtaceae)         e         tree           Persea lingue (Lauraceae)         e         tree           Posala glandulosa (Fabaceae)         e         tree <t< td=""><td>bes punctatum (Grossulariaceae)</td><td>е</td><td>shrub</td><td>all</td></t<>	bes punctatum (Grossulariaceae)	е	shrub	all
Mid-elevation sclerophyll matorral: Río Clarillo           Aristotelia chilensis (Elaeocarpaceae)         e         small tree           Azara petiolaris (Salicaceae)         e         small tree           Baccharis sp. (Asteraceae)         e         shrub           Berberis chilensis (Berberidaceae)         e         shrub           Cestrum parqui (Solanaceae)         e         shrub           Colliguaja odonfera         e         tree           Cryptocarya alba         e         tree           Drimys winteri (Winteraceae)         e         tree           Escallonia illinita         e         small tree           Escallonia illinita         e         shrub (small tree)           Haplopappus sp. (Asteraceae)         e         tree           Lithraea caustica         e         small tree           Luma chequen (Myrtaceae)         e         tree           Maytenus boaria         e         tree           Persea lingue (Lauraceae)         e/sd         shrub           Posoralea glandulosa (Fabaceae)         e/sd         shrub           Soralea glandulosa (Fabaceae)         e         tree           Quillaja saponaria (Quillajaceae)         e         shrub           Schinus polygama (Anacardia	chinus latifolius (Anacardiaceae)	е	small tree	all
Aristotelia chilensis (Elaeocarpaceae) Azara petiolaris (Salicaceae) Berberis chilensis (Berberidaceae) Berberiaceae, Berberidaceae, Berberid	enna candolleana (Caesalpiniaceae)	е	shrub (small tree)	all
Aristotelia chilensis (Elaocarpaceae)       e       small tree         Azara petiolaris (Salicaceae)       e       small tree         Baccharis sp. (Asteraceae)       e       shrub         Berberis chilensis (Berberidaceae)       e       shrub         Cestrum parqui (Solanaceae)       e       shrub         Colliguaja odorifera       e       shrub         Cryptocarya alba       e       tree         Drimys winteri (Winteraceae)       e       small tree         Escallonia pulverulenta (Escalloniaceae)       e       small tree         Escallonia illinita       e       shrub (small tree)         Haplopappus sp. (Asteraceae)       e       shrub (small tree)         Lithraea caustica       e       tree         Lithraea caustica       e       small tree         Lithraea caustica       e       tree         Lithraea caustica       e       tree         Lithraea caustica       e       small tree         Lithraea caustica       e       tree         Lithraea caustica       e       tree         Lithraea caustica       e       small tree         Lithraea caustica       e       tree         Lithraea caustica       e	lid-elevation sclerophyll matorral: Río Clarillo			
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				all
Solanum ligustrinum (Solanaceae) e shrub	olanum ligustrinum (Solanaceae)		shrub	all
Trevoa quinquenervia (Rhamnaceae) d shrub, small tree	revoa quinquenervia (Rhamnaceae)			pst
Los Molles				
Pouteria splendens (Sapotaceae) <sup>1</sup> e shrub	outeria splendens (Sapotaceae)'	е	shrub	all

<sup>&</sup>lt;sup>1</sup>Sampled in matorral at Los Molles (c. 32° 13′ S, 71° 31′ W, 70 m asl) and only included in trait correlation analyses.