



# Draft Genome Sequence of *Pseudomonas* sp. Strain RGM 3321, a Phyllosphere Endophyte from *Fragaria chiloensis* subsp. *chiloensis* f. *patagonica*

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**ABSTRACT** *Pseudomonas* sp. strain RGM 3321 is a phyllosphere endophyte from *Fragaria chiloensis* subsp. *chiloensis* f. *patagonica* that harbors genes associated with plant growth promotion pathways, as well as genes typically found in plant pathogens.

*Fragaria chiloensis* subsp. *chiloensis* f. *patagonica* is the wild form of the Chilean strawberry, which grows in the mountains and in coastal environments (1). In January 2021, a sample of this species was removed from forest soil in the Yungay Precordillera, Ñuble, Chile (−37.05905, −71.64625). The phyllosphere was surface sterilized by submerging the tissues in 70% ethanol (1 min), 1.5% NaClO (3 min), and 96% ethanol (1 min) and finally rinsed three times. The tissues were subsequently ground in physiological solution, and volumes of 100  $\mu$ L of serial dilutions were inoculated on King's B (KB) agar medium supplemented with 25  $\mu$ g/mL nystatin and 50  $\mu$ g/mL cycloheximide. Plates were incubated at 25°C for 48 h. A UV-fluorescent colony was streaked on KB agar and incubated under the same conditions. This step was repeated twice to obtain axenic cultures. The isolate was deposited in the Chilean Collection of Microbial Genetic Resources (CChRGM), under the code RGM 3321. Strain RGM 3321 grown in yeast extract-malt extract-dextrose broth supplemented with 1% L-tryptophan produced 106.47  $\mu$ g/mL indole acetic acid (IAA) and grown on NBRIP agar displayed a phosphate solubilization index of 2.4, suggesting potential plant growth-promoting traits (2–4).

Two genomic DNA libraries were constructed using the Nextera XT library preparation kit (Illumina, USA) and sequenced on an Illumina HiSeq/NovaSeq platform using a 250-bp paired-end protocol at MicrobesNG (UK). Whole-genome sequencing reads were adapter trimmed using Trimmomatic v0.30 with a sliding window quality cutoff value of Q15 (5, 6). *De novo* assembly was performed using SPAdes v3.7 (7). Contigs were annotated using the NCBI Prokaryotic Genome Annotation Pipeline (PGAP) v6.0 (8). The assembled genome has a size of 6,380,675 nucleotides (nt), distributed in 121 contigs (the largest contig was 609,627 nt), with mean coverage of 82.91 $\times$ , an  $N_{50}$  value of 171,869 nt, and a G+C content of 58.41%; 5,669 genes and 56 tRNAs were predicted from the annotation. On the EzBioCloud webserver (9), RGM 3321 shared the greatest 16S rRNA gene similarity with the phytopathogenic species of the *Pseudomonas syringae* group (10), i.e., *Pseudomonas* KCTC 12500<sup>T</sup> (99.93%) and *Pseudomonas congelans* DSM 14939<sup>T</sup> (99.86%) and *Pseudomonas cerasi* 58<sup>T</sup> (99.86%).

BLAST searches were performed using SequenceServer v2.0.0 (11). Strain RGM 3321 harbors one copy of a gene for 1-aminocyclopropane-1-carboxylate (ACC) deaminase (*acdS*), which is involved in the degradation of ACC, a precursor of ethylene in plants (12, 13). The strain contains genes for a quinoprotein glucose dehydrogenase (*gcd*) and the pyrroloquinoline quinone (*pqq*) operon (14), suggesting a mechanism for phosphate

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**TABLE 1** Genome sequence features of *Pseudomonas* sp. strain RGM 3321

Trait and protein name (gene name) or BGC	RGM 3321 protein code	Identity to reference protein (%)	UniProt accession no. for reference protein	Strain encoding the BGC	Similarity to reference BGC (%) <sup>a</sup>
Trait and protein name					
Phosphate solubilization					
Quinoprotein glucose dehydrogenase ( <i>gcd</i> )	RGM3321_17085	69.27	<a href="#">A0A0B6F0P5</a>		
Coenzyme PQQ synthesis protein A ( <i>pqqA</i> )	RGM3321_09145	95.83	<a href="#">Q3K5R0</a>		
Coenzyme PQQ synthesis protein B ( <i>pqqB</i> )	RGM3321_09140	91.42	<a href="#">C3K348</a>		
Pyroloquinoline-quinone synthase ( <i>pqqC</i> )	RGM3321_09135	92.83	<a href="#">Q88QV6</a>		
PqqA binding protein ( <i>pqqD</i> )	RGM3321_09130	68.97	<a href="#">Q4K4U9</a>		
PqqA peptide cyclase ( <i>pqqE</i> )	RGM3321_09125	89.10	<a href="#">Q4K4U8</a>		
Coenzyme PQQ synthesis protein F ( <i>pqqF</i> )	RGM3321_09150	44.84	<a href="#">P55174</a>		
IAA production					
Tryptophan 2-monooxygenase ( <i>iaaM</i> ); IAM pathway	RGM3321_01470	94.61	<a href="#">P06617</a>		
Indoleacetamide hydrolase ( <i>iaaH</i> ); IAM pathway	RGM3321_01475	93.02	<a href="#">P06618</a>		
Indole-3-pyruvate decarboxylase ( <i>ipdC</i> ); indole-3-pyruvate pathway	RGM3321_13685	26.95	<a href="#">A0A5E6Q147</a>		
Nitrilase ( <i>nit</i> ); IAN pathway	RGM3321_22075	28.52	<a href="#">K9NKH3</a>		
Aldehyde dehydrogenase family protein ( <i>aldA</i> ); IAN pathway	RGM3321_15565	94.16	<a href="#">Q88BC5</a>		
Aldehyde dehydrogenase family protein ( <i>aldB</i> ); IAN pathway	RGM3321_11565	97.57	<a href="#">Q88BC5</a>		
Aldehyde dehydrogenase family protein ( <i>aldB</i> ); IAN pathway	RGM3321_06435	45.50	<a href="#">Q88BC5</a>		
ACC deaminase activity					
ACC deaminase ( <i>acdS</i> )	RGM3321_08860	88.76	<a href="#">Q51813</a>		
Leucine-responsive regulatory protein ( <i>acdR</i> )	RGM3321_08855	80.47	<a href="#">K9NP20</a>		
T3SS					
Hypersensitivity response secretion protein HrpJ ( <i>hrpJ</i> )	RGM3321_26170	98.77	<a href="#">Q05395</a>		
Lipoprotein ( <i>hrcJ</i> )	RGM3321_26075	90.30	<a href="#">G3XDC8</a>		
Type III secretion protein HrcR ( <i>hrcR</i> )	RGM3321_26130	93.00	<a href="#">Q887B8</a>		
Type III secretion protein HrcS ( <i>hrcS</i> )	RGM3321_26125	94.32	<a href="#">G3XDB8</a>		
Type III secretion protein HrcT ( <i>hrcT</i> )	RGM3321_26120	84.09	<a href="#">G3XDD0</a>		
Type III secretion protein HrcU ( <i>hrcU</i> )	RGM3321_26115	85.52	<a href="#">Q887B9</a>		
Hypersensitivity response secretion protein HrpI ( <i>hrpI</i> )	RGM3321_26165	98.99	<a href="#">P35655</a>		
T3SS ATPase SctN ( <i>sctN</i> )	RGM3321_26155	99.33	<a href="#">Q52371</a>		
V-type ATP synthase subunit E ( <i>hrpE</i> )	RGM3321_26085	76.17	<a href="#">Q887C4</a>		
Type III secretion protein HrcQb ( <i>hrcQb</i> )	RGM3321_26135	93.98	<a href="#">Q60235</a>		
T3SS regulator					
RNA polymerase sigma factor HrpL ( <i>hrpL</i> )	RGM3321_26175	95.70	<a href="#">P37929</a>		
Hrp pilus protein HrpA1 ( <i>hrpA</i> )	RGM3321_26060	99.07	<a href="#">Q52420</a>		
T3SS key effector					
Type III effector HopAA1 ( <i>hopAA1</i> )	RGM3321_26015	74.74	<a href="#">G3XDB9</a>		
Type III effector AvrE1 ( <i>avrE1</i> )	RGM3321_26040	67.83	<a href="#">Q887C9</a>		
Type III effector HopM1 ( <i>hopM1</i> )	RGM3321_26030	93.87	<a href="#">Q4ZX82</a>		
BGC					
Pyoverdine				<i>Pseudomonas protegens</i> Pf-5	22
Syringafactin				<i>Pseudomonas syringae</i> pv. tomato DC3000	66
Syringolin A				<i>Pseudomonas syringae</i> pv. syringae B301 D-R	100
Syringomycin				<i>Pseudomonas syringae</i> pv. syringae B728a	100

<sup>a</sup>Similarity of the BGC found in the genome of RGM 3321 to the reference BGC used by antiSMASH.

solubilization in soil via gluconic acid synthesis (15, 16). The identification of *iaaM* and *iaaH* genes suggests production of IAA via the indole-3-acetamide (IAM) and/or indole-3-acetonitrile (IAN) pathways (17, 18). Genes encoding a nitrile hydratase (*nthAB*) were also found, suggesting the transformation of IAN into an IAM intermediate in the IAN pathway (19; Table 1).

Fourteen biosynthetic gene clusters (BGCs) were predicted from the RGM 3321 genome using antiSMASH 6.0 (20), four of which, including the pyoverdine (21), syringomycin (22), syringolin A (23), and syringafactin (24) BGCs, are present in plant-pathogenic bacteria (Table 1). In addition, RGM 3321 contains a type III secretion system (T3SS) gene and additional auxiliary genes (25), including an homolog of the RNA polymerase sigma factor

(*hrpL*), which is a master regulator of the T3SS that interacts with a conserved *hrp* box motif and promotes the expression of effectors and other virulence factors (26), including HopAA1, which specifically enhances the epiphytic bacterial survival/growth in plants (27).

The draft genome of *Pseudomonas* sp. strain RGM 3321 expands our knowledge about the bacterial diversity of Chilean wild plants, revealing plant growth-promoting genes and additional open reading frames associated with plant virulence factors. All tools were run with default parameters unless otherwise specified.

**Data availability.** This whole-genome shotgun project has been deposited at DDBJ/ENA/GenBank under the accession number [JALHBBH0000000000](https://doi.org/10.1101/2021.03.03.43389). The version described in this paper is version [JALHBBH0000000000.1](https://doi.org/10.1101/2021.03.03.43389). The raw data are available under SRA accession numbers [SRR18554678](https://doi.org/10.1101/2021.03.03.43389), [SRR18554679](https://doi.org/10.1101/2021.03.03.43389), and [SRR18554680](https://doi.org/10.1101/2021.03.03.43389). All project data are available under BioProject accession number [PRJNA820724](https://doi.org/10.1101/2021.03.03.43389).

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