

Segniliparus rugosus from the sputum of a child with cystic fibrosis in Ecuador: challenges in bacterial identification

J. Zurita¹, G. Sevillano¹, C. González¹ and Y. Lascano²

1) Biomedical Research Unit, Zurita & Zurita

Laboratorios and 2) Department of Paediatric Pneumology, Hospital Carlos Andrade Marin, Quito, Ecuador

Abstract

Using sequencing analyses of the 16S rRNA gene, we identified *Segniliparus rugosus* in an 8-year-old child with cystic fibrosis. We describe the difficulties we encountered in identifying this bacterium. To the best of our knowledge, this is the first reported case of *S. rugosus* in Ecuador.

© 2020 The Authors. Published by Elsevier Ltd.

Keywords: 16S rRNA, Cystic fibrosis, Ecuador, *Mycobacterium*, *Segniliparus rugosus*

Original Submission: 13 February 2020; **Accepted:** 10 March 2020

Article published online: 18 March 2020

Corresponding author. Zurita & Zurita Laboratories, Quito, Ecuador.
E-mail: jzurita@zuritalaboratorios.com

Segniliparus rugosus is a rapidly growing acid-fast bacillus (AFB) that was isolated from human samples in 2005 [1]. It has been identified as an emerging human pathogen that has been isolated from patients with and without cystic fibrosis (CF) [2–4].

We describe the case of an 8-year-old child who was diagnosed with CF 2 years ago. The patient had the F508del/H609R mutations. Unlike many CF patients, he did not have a history of chronic *Pseudomonas aeruginosa* infections; however, *Staphylococcus aureus* and *Burkholderia cepacia* had previously been isolated from his sputum. Although AFB cultures yielded *Mycobacterium terrae* growth on one occasion (2018), he did not require treatment. *Aspergillus fumigatus* was isolated from this patient in 2016, 2017 and 2018. He was then diagnosed

with allergic bronchopulmonary aspergillosis (ABPA) and was treated with corticosteroids and itraconazole.

On 8 November 2019, we received a sputum sample in our laboratory for bacteriological culture and testing for acid-fast staining. Because the sputum was 1+ AFB smear-positive, the sample was inoculated into Middlebrook 7H9 broth (Becton Dickinson, USA), using the Bactec MGIT 320 system (BD Diagnostics, USA), and Lowenstein–Jensen medium. On the fourth day of incubation, the Middlebrook broth was positive, and a subculture was performed on chocolate agar to test for rapidly growing non-tuberculosis bacteria (RGNTB). The bacteria grew on the chocolate agar at 35°C by the fourth day. Acid-fast staining tests of these colonies on the chocolate agar were positive. Therefore, we proceeded to perform PCR restriction fragment length polymorphism analysis (PRA) using the heat shock protein 65 (*hsp65*) gene [5], and sequencing of the RNA polymerase subunit β (*rpoB*) gene [6]. However, PCR of the *hsp65* gene did not produce amplicons, and *rpoB* gene sequencing identified *S. rotundus* with a low percentage identity (91.92%). We chose to sequence the 16S rRNA gene, which showed *S. rugosus* with 100% identity. Despite performing the recommended laboratory procedures, based on this AFB strain, the first molecular methods chosen were not appropriate in this case.

Antimicrobial susceptibility testing was performed using the Sensititre™ Rapid Myco (Thermo Scientific). The minimal inhibitory concentrations (MICs) were determined after incubation at 35°C for 4 days. The break points used for the RGNTB were determined according to the Clinical and Laboratory Standards Institute (CLSI) [7]. The clinical *S. rugosus* (Z&Z215) isolate was susceptible to all the antibiotics tested (Table 1).

S. rugosus may be confused with RGNTB due to its acid-fast staining properties. In our case, only the sequencing analyses of the 16S rRNA genes allowed for the accurate identification of *S. rugosus*.

TABLE 1. Antimicrobial susceptibility patterns of *Segniliparus rugosus* (Z&Z215)

Antibiotics	MIC ($\mu\text{g/mL}$)	Interpretation ^a
Trimethoprim/sulfamethoxazole	<0.25/4.75	S
Ciprofloxacin	<0.12	S
Moxifloxacin	<0.25	S
Cefoxitin	<4	S
Amikacin	<1	S
Doxycycline	<0.12	S
Tigecycline	0.06	S
Clarithromycin	<0.06	S
Linezolid	<1	S
Imipenem	<2	S
Cefepime	<1	S
Amoxicillin/clavulanic acid	<2/1	S
Ceftriaxone	<4	S
Minocycline	<1	S
Tobramycin	<1	S

MIC, minimal inhibitory concentration; S, susceptible.

^aAccording to the break points for rapid growth mycobacteria. Read on the 4th day of incubation.

With the presence of AFB in the sputum of CF patients, the *Segniliparus* genus should be considered as a potential pathogen in addition to the classical bacteria such as *Nocardia*, *Actinomyces* and *Mycobacterium*. The identification of this genus continues to be a challenge for microbiology laboratories. We need to learn more about the importance and impact of *Segniliparus* in CF patients. This is the first reported case of *S. rugosus* in a patient with CF in Ecuador.

Conflict of interest

The authors declare no conflict of interest.

References

- [1] Butler WR, Floyd MM, Brown JM, Toney SR, Daneshvar MI, Cooksey RC, et al. Novel mycolic acid-containing bacteria in the family Segniliparaceae fam. nov., including the genus *Segniliparus* gen. nov., with descriptions of *Segniliparus rotundus* sp. nov. and *Segniliparus rugosus* sp. nov. *Int J Syst Evol Microbiol* 2005;55:1615–24.
- [2] Lee JY, Chon GR, Jung TY, Sung H, Shim TS, Jo KW. A case of *Segniliparus rugosus* pulmonary infection in an immunocompetent patient with non-cystic fibrosis. *Tuberc Respir Dis (Seoul)* 2014;77:227–9.
- [3] Butler WR, Sheils CA, Brown-Elliott BA, Charles N, Colin AA, Gant MJ, et al. First isolations of *Segniliparus rugosus* from patients with cystic fibrosis. *J Clin Microbiol* 2007;45:3449–52.
- [4] Hansen T, van Kerckhof J, Jelfs P, Wainwright C, Ryan P, Coulter C. *Segniliparus rugosus* infection, Australia. *Emerg Infect Dis* 2009;15:611–3.
- [5] Chimara E, Ferrazoli L, Ueky SYM, Martins MC, Durham AM, Arbeit RD, et al. Reliable identification of mycobacterial species by PCR-restriction enzyme analysis (PRA)-hsp65 in a reference laboratory and elaboration of a sequence-based extended algorithm of PRA-hsp65 patterns. *BMC Microbiol* 2008;8:1–12.
- [6] Adékambi T, Colson P, Drancourt M. RpoB-based identification of nonpigmented and late-pigmenting rapidly growing mycobacteria. *J Clin Microbiol* 2003;41:5699–708.
- [7] CLSI. Susceptibility. Testing of mycobacteria, nocardiae, and other aerobic actinomycetes; approved standard. 2011. 2nd ed. Wayne P, editor. M24-A2.