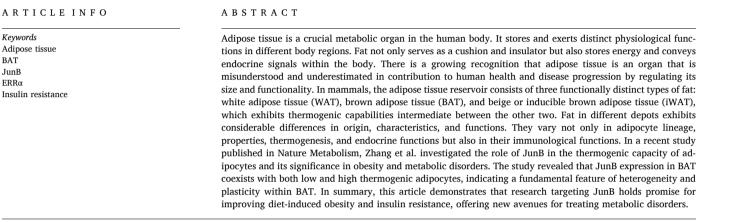
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Depletion of JunB increases adipocyte thermogenic capacity and ameliorates diet-induced insulin resistance



WAT functions as the primary energy storage site in the body. It synthesizes triglycerides for long-term storage and releases free fatty acids to provide energy when needed [1]. In comparison to BAT, WAT is primarily consists of large adipocytes with a single lipid droplet and fewer mitochondria [2]. Depending on their anatomical location in the body, WAT can be categorized into two types: visceral WAT (VAT) and subcutaneous WAT (SAT) [3]. Lineage studies in mice have shown that VAT and SAT have different origins, but their developmental origins in humans are not yet fully elucidated [4]. VAT encompasses adipose depots such as the omentum, mesentery, retroperitoneum, gonadal, and pericardial WAT. The accumulation of VAT and the resulting inflammation increase the risk of metabolic syndrome, insulin resistance, and cardiovascular diseases [5-7].

It is well-known that newborns possess active BAT to maintain normal body temperature, which regresses with age. However, functional BAT expressing significant levels of uncoupling protein 1 (UCP1) can be found in healthy adults in areas such as the supraclavicular region and other upper trunk sites [8,9]. Various factors such as cold exposure, feeding, low BMI, sympathetic nervous system (SNS) activity, cancer, and burn injury, positively correlate with BAT content in adults [10]. Unlike WAT, BAT contains multiple small lipid droplets and abundant mitochondria [11]. Within these mitochondria, UCP1 located on the inner mitochondrial membrane can be activated under stimuli such as cold exposure, generating heat and promoting systemic energy metabolism [12-14]. For instance, cold exposure activates BAT via the

sympathetic nervous system, leading to increased glucose uptake, accelerated plasma triglyceride clearance, and suppression of triglyceride concentrations, thereby reducing the risk of obesity and cardiovascular diseases [15]. Consequently, BAT plays a pivotal role in thermoregulation and energy metabolism through non-shivering thermogenesis [10]. However, despite the anti-obesity effects of BAT activation induced by cold stimuli, its activation declines significantly with age [16]. Thus, there are currently no reports of BAT being utilized as a pharmacological target for therapeutic intervention.

Increasing evidence suggests the existence of distinct subpopulations of brown adipocytes with different characteristics within BAT, demonstrating high heterogeneity in terms of thermogenesis, insulin sensitivity, and secretory functions. For instance, both high and low thermogenic fat cells coexist within BAT [17,18]. Nevertheless, little is known about the molecular mechanisms regulating thermogenic adipocyte heterogeneity and its significance in obesity and metabolic disorders.

Beige adipose tissue represents a phenotypic transition of WAT into a BAT-like phenotype [19]. The induction of WAT browning is primarily stimulated by cold exposure or dietary factors and transmitted through the release of norepinephrine (NE) from sympathetic nerve terminals [20]. Similar to BAT, beige adipose tissue contains multilocular lipid droplets and exhibits abundant mitochondria, thus possessing the capability to generate heat and promote systemic energy metabolism upon stimulation.

https://doi.org/10.1016/j.metop.2024.100277

Received 19 February 2024; Accepted 20 February 2024 Available online 22 February 2024

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Keywords

BAT

JunB

ERRo

Adipose tissue

Insulin resistance





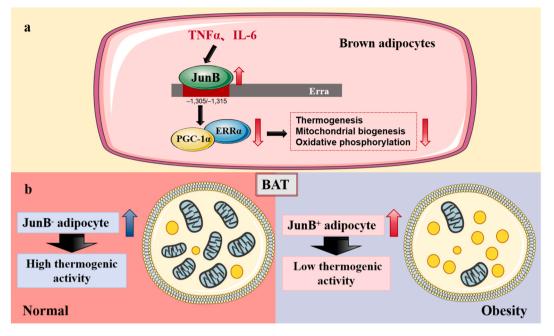


Fig. 1. Mechanisms of JunB in thermogenic capacity of brown adipose tissue. (For interpretation of the references to colour in this figure legend, the reader is referred to the Web version of this article.)

In a recent study published in *Nature Metabolism*, Zhang et al. identified an atypical subset of brown adipocytes that express the AP1 transcription factor JunB, which increases in abundance during obesity and is induced by inflammation. These JunB-expressing adipocytes can be found throughout various depots of BAT, exhibiting significantly lower lipid droplets and thermogenic capacity compared to high thermogenic adipocytes. Acting as a transcriptional regulatory factor, JunB integrates inflammatory and metabolic signals, modulating the interconversion and energy metabolism of low and high thermogenic adipocytes by binding to the ERR α promoter and inhibiting the PGC-1 α -ERR α pathway [21]. Specific knockout of JunB in thermogenic adicytes or the ablation of JunB expression in adipocytes improves energy metabolism, ameliorating energy balance and preventing/treating obesity and insulin resistance.

In conclusion, this study unveils an atypical subset of thermogenic adipocytes that responds to obesity and inflammation by expressing JunB. The abnormal induction of JunB affects the heterogeneity of thermogenic adipocytes and compromises BAT function by generating more low thermogenic cells. As an AP1 transcription factor, JunB regulates the interconversion of low and high thermogenic adipocytes through the PGC-1 α -ERR α pathway, thereby altering adipose thermogenic capacity and heterogeneity by integrating inflammatory and metabolic signals (see Fig. 1). These findings provide insights into the biological characteristics of brown adipocytes and lay a solid foundation for the development of therapeutic interventions targeting obesity, inflammation, and related diseases. Further research is warranted to better understand the biological functions and mechanisms of JunB and other AP1 members in various types of adipose tissue.

CRediT authorship contribution statement

Qian Zhou: Conceptualization, Writing – original draft, Writing – review & editing. **Suzhen Chen:** Conceptualization, Supervision, Validation, Writing – review & editing. **Junli Liu:** Conceptualization, Supervision, Validation, Writing – review & editing.

References

- Vishvanath L, Gupta RK. Contribution of adipogenesis to healthy adipose tissue expansion in obesity. J Clin Invest 2019;129:4022–31. https://doi.org/10.1172/ JCI129191.
- [2] Zwick RK, Guerrero-Juarez CF, Horsley V, Plikus MV. Anatomical, physiological, and functional Diversity of adipose tissue. Cell Metabol 2018;27:68–83. https:// doi.org/10.1016/j.cmet.2017.12.002.
- [3] Fruhbeck G. Overview of adipose tissue and its role in obesity and metabolic disorders. Methods Mol Biol 2008;456:1–22. https://doi.org/10.1007/978-1-59745-245-8_1.
- [4] Chau YY, Bandiera R, Serrels A, Martinez-Estrada OM, Qing W, Lee M, Slight J, Thornburn A, Berry R, McHaffie S, et al. Visceral and subcutaneous fat have different origins and evidence supports a mesothelial source. Nat Cell Biol 2014;16: 367–75. https://doi.org/10.1038/ncb2922.
- [5] Reyes-Farias M, Fos-Domenech J, Serra D, Herrero L, Sanchez-Infantes D. White adipose tissue dysfunction in obesity and aging. Biochem Pharmacol 2021;192: 114723. https://doi.org/10.1016/j.bcp.2021.114723.
- [6] Ritchie SA, Connell JM. The link between abdominal obesity, metabolic syndrome and cardiovascular disease. Nutr Metabol Cardiovasc Dis 2007;17:319–26. https:// doi.org/10.1016/j.numecd.2006.07.005.
- [7] Appleton SL, Seaborn CJ, Visvanathan R, Hill CL, Gill TK, Taylor AW, Adams RJ, North West Adelaide Health Study T. Diabetes and cardiovascular disease outcomes in the metabolically healthy obese phenotype: a cohort study. Diabetes Care 2013;36:2388–94. https://doi.org/10.2337/dc12-1971.
- [8] Virtanen KA, Lidell ME, Orava J, Heglind M, Westergren R, Niemi T, Taittonen M, Laine J, Savisto NJ, Enerback S, Nuutila P. Functional brown adipose tissue in healthy adults. N Engl J Med 2009;360:1518–25. https://doi.org/10.1056/ NEJMoa0808949.
- [9] Cypess AM, Lehman S, Williams G, Tal I, Rodman D, Goldfine AB, Kuo FC, Palmer EL, Tseng YH, Doria A, et al. Identification and importance of brown adipose tissue in adult humans. N Engl J Med 2009;360:1509–17. https://doi.org/ 10.1056/NEJMoa0810780.
- [10] Marlatt KL, Ravussin E. Brown adipose tissue: an update on recent findings. Current Obesity Reports 2017;6:389–96. https://doi.org/10.1007/s13679-017-0283-
- [11] Wang W, Seale P. Control of brown and beige fat development. Nat Rev Mol Cell Biol 2016;17:691–702. https://doi.org/10.1038/nrm.2016.96.
- [12] Fedorenko A, Lishko PV, Kirichok Y. Mechanism of fatty-acid-dependent UCP1 uncoupling in brown fat mitochondria. Cell 2012;151:400–13. https://doi.org/ 10.1016/j.cell.2012.09.010.
- [13] Ricquier D. Uncoupling protein 1 of brown adipocytes, the only uncoupler: a historical perspective. Front Endocrinol 2011;2:85. https://doi.org/10.3389/ fendo.2011.00085.
- [14] Ricquier D, Kader JC. Mitochondrial protein alteration in active brown fat: a soidum dodecyl sulfate-polyacrylamide gel electrophoretic study. Biochem Biophys Res Commun 1976;73:577–83. https://doi.org/10.1016/0006-291x(76)90849-4.
- [15] Bartelt A, Bruns OT, Reimer R, Hohenberg H, Ittrich H, Peldschus K, Kaul MG, Tromsdorf UJ, Weller H, Waurisch C, et al. Brown adipose tissue activity controls triglyceride clearance. Nat Med 2011;17:200–5. https://doi.org/10.1038/ nm.2297.

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- [16] Yoneshiro T, Aita S, Matsushita M, Okamatsu-Ogura Y, Kameya T, Kawai Y, Miyagawa M, Tsujisaki M, Saito M. Age-related decrease in cold-activated brown adipose tissue and accumulation of body fat in healthy humans. Obesity 2011;19: 1755–60. https://doi.org/10.1038/oby.2011.125.
- [17] Sun W, Dong H, Balaz M, Slyper M, Drokhlyansky E, Colleluori G, Giordano A, Kovanicova Z, Stefanicka P, Balazova L, et al. snRNA-seq reveals a subpopulation of adipocytes that regulates thermogenesis. Nature 2020;587:98–102. https://doi. org/10.1038/s41586-020-2856-x.
- [18] Song A, Dai W, Jang MJ, Medrano L, Li Z, Zhao H, Shao M, Tan J, Li A, Ning T, et al. Low- and high-thermogenic brown adipocyte subpopulations coexist in murine adipose tissue. J Clin Invest 2020;130:247–57. https://doi.org/10.1172/ JCI129167.
- [19] Whitehead A, Krause FN, Moran A, MacCannell ADV, Scragg JL, McNally BD, Boateng E, Murfitt SA, Virtue S, Wright J, et al. Brown and beige adipose tissue regulate systemic metabolism through a metabolite interorgan signaling axis. Nat Commun 2021;12:1905. https://doi.org/10.1038/s41467-021-22272-3.
- [20] Villarroya F, Cereijo R, Gavalda-Navarro A, Villarroya J, Giralt M. Inflammation of brown/beige adipose tissues in obesity and metabolic disease. J Intern Med 2018; 284:492–504. https://doi.org/10.1111/joim.12803.
- [21] Zhang X, Ding X, Wang C, Le Q, Wu D, Song A, Huang G, Luo L, Luo Y, Yang X, et al. Depletion of JunB increases adipocyte thermogenic capacity and ameliorates diet-induced insulin resistance. Nat Metab 2024;6:78–93. https://doi.org/ 10.1038/s42255-023-00945-1.

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