

Three Draft Genome Sequences of White Spot Syndrome Virus from India

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ABSTRACT White spot syndrome virus (WSSV) is a pathogen causing significant economic losses to shrimp aquaculture worldwide. Previously, five genome sequences of the virus from farmed shrimp (Penaeus vannamei and Penaeus monodon) in India were reported, all originating from farms located on the east coast of the country. Here, we report three new and distinct WSSV genome sequences, two from shrimp (P. vannamei) farmed on the west coast of India and the third from the east coast.

White spot syndrome virus (WSSV) is a virulent pathogen infecting farmed shrimp that causes significant economic losses worldwide and has emerged as one of the most prevalent and widespread viruses of crustaceans ([1\)](#page-3-0).

WSSV-infected Pacific white shrimp (Penaeus [Litopenaeus] vannamei) were collected from shrimp farms in Maharashtra (west coast of India) in October 2016 and January 2019 (CWG3 and PG1, respectively) and from a shrimp farm in Tamil Nadu (east coast of India) in November 2018 (DBA1182). DNA was extracted from shrimp gut tissues using a cetyltrimethylammonium bromide (CTAB)-EDTA DNA extraction protocol ([2](#page-3-1)). Sequence libraries for samples CWG3 and PG1 were prepared using the NextSeq series midoutput kit (Illumina, San Diego, CA) and sequenced on an Illumina NextSeq 500 sequencer (2 \times 150 bp). A sequence library for DBA1182 was prepared using the NEBNext Ultra II DNA library preparation kit and sequenced on an Illumina HiSeq sequencer (2 \times 150 bp). Adapter sequences and low-quality bases were removed using fastp v0.20.0 ([3](#page-3-2)). For each sample, sequences were normalized using BBNorm, part of BBTools v38.03 ([4](#page-3-3)) (parameters used: ecc=t, bits=16, prefilter), prior to de novo assembly with SPAdes v3.13.1 [\(5\)](#page-3-4) using the only-assembler flag and k-mers of 21, 33, 55, 77, 99, and 127. Assembled contigs were compared to a reference WSSV genome sequence (WSSV-CN [GenBank accession number [AF332093.3](https://www.ncbi.nlm.nih.gov/nuccore/AF332093.3)]) using BLASTN v2.9.0+ [\(6](#page-3-5)) to identify contigs representing WSSV. Sequencing statistics, including the number of contigs generated, genome length, GC content, and genome coverage for each WSSV genome, are summarized in [Table 1.](#page-1-0) The genome sequences of two WSSV strains (WSSV-IN-LS and WSSV-IN-NS) previously reported from India [\(7](#page-3-6)) could not be found in the public databases and therefore were generated by processing the sequences downloaded from the NCBI Sequence Read Archive (SRA) (SRA accession numbers [SRR3233836](https://www.ncbi.nlm.nih.gov/sra/SRR3233836) and [SRR3233837](https://www.ncbi.nlm.nih.gov/sra/SRR3233837), respectively) using the methods described above. The WSSV-IN-LS sequence consists of 12 contigs with a total length of 299,240 bp (GC content of 40.97%, with $301 \times$ coverage), whereas WSSV-IN-NS has 3 contigs totaling 284,022 bp (GC content of 41.08%, with 1,119 \times coverage). A core genome sequence alignment containing parts of the genome present in all selected genome sequences was generated with Parsnp v1.2 [\(8\)](#page-3-7) using the WSSV reference genome and a selection of other WSSV genomes. A Bayesian phylogenetic tree was

Citation Kooloth Valappil R, Anand D, Kulkarni A, Mahapatra M, Kumar SH, Bedekar MK, Kollanoor RJ, Mulloorpeedikayil RG, Mohideenpitchai MM, Muthumariappan S, Panchavarnam S, Devaraj K, Bass D, van Aerle R. 2021. Three draft genome sequences of white spot syndrome virus from India. Microbiol Resour Announc 10:e00579-21. [https://doi.org/](https://doi.org/10.1128/MRA.00579-21) [10.1128/MRA.00579-21](https://doi.org/10.1128/MRA.00579-21).

Editor Jelle Matthijnssens, KU Leuven

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Received 10 June 2021 Accepted 5 August 2021 Published 26 August 2021

TABLE 1 Summary of sequencing statistics for each WSSV genome TABLE 1 Summary of sequencing statistics for each WSSV genome

FIG 1 Bayesian phylogeny of all 914 variable (single-nucleotide polymorphism [SNP]) positions in the core genome alignments (total alignment length, 225,757 bp) of 19 WSSV strains, including 3 new genomes sequenced for this study (WSSV-IN-DBA1182, WSSV-IN-CWG3, and WSSV-IN-PG1) (in rounded rectangles). Bayesian posterior probability (x) and maximum likelihood bootstrap (y) values are shown at the nodes (x/y) ; black circles show joint bipartition support of $>0.95/>95%$. Branch labels indicate the country of origin (IN, India; CN, China; AU, Australia; MEX, Mexico; EC, Ecuador; TW, Taiwan; BR, Brazil; TH, Thailand). All available genome sequences from India are shown in bold. GenBank accession numbers are provided in parentheses where available. *, the assemblies of WSSV-IN-LS and WSSV-IN-NS were generated during the course of this study and have been uploaded to figshare [\(https://doi.org/10.6084/m9.](https://doi.org/10.6084/m9.figshare.14252951)figshare.14252951 and [https://doi.org/10.6084/m9.](https://doi.org/10.6084/m9.figshare.14252966)figshare.14252966, respectively).

constructed based on the variable positions of only the core genome alignments using MrBayes v3.2.6 [\(9\)](#page-3-8) on the CIPRES server [\(10](#page-3-9)). Maximum likelihood bootstrap values were calculated in RAxML BlackBox [\(11\)](#page-3-10), also on the CIPRES server.

Phylogenetic analyses of the three new Indian WSSV genomes ([Fig. 1](#page-2-0)) showed that two of them (WSSV-IN-DBA1182 and WSSV-IN-CWG3) branched robustly with three other genotypes from India. The third genome (WSSV-IN-PG1) branched with a WSSV genome (WSSV-IN-LS) that we assembled from data generated in a previous study ([7](#page-3-6)) and the genome of a Mexican strain obtained from GenBank (accession number [KU216744](https://www.ncbi.nlm.nih.gov/nuccore/KU216744)). Two of the new strains sequenced in the present study (WSSV-IN-CWG3 and WSSV-IN-PG1) were from shrimp farmed on the west coast of India, whereas all other strains reported from India were from the east coast.

Data availability. This whole-genome shotgun project was deposited in DDBJ/ENA/ GenBank under BioProject accession number [PRJNA674024](https://www.ncbi.nlm.nih.gov/bioproject/PRJNA674024). The raw reads were deposited in the SRA under accession numbers [SRR12970169](https://trace.ncbi.nlm.nih.gov/Traces/sra/?run=SRR12970169), [SRR12970170](https://trace.ncbi.nlm.nih.gov/Traces/sra/?run=SRR12970170), and [SRR12970171.](https://trace.ncbi.nlm.nih.gov/Traces/sra/?run=SRR12970171) The assembled genome sequences for WSSV-IN-CWG3, WSSV-IN-PG1, and WSSV-IN-DBA1182 were deposited in GenBank under accession numbers [MW248108](https://www.ncbi.nlm.nih.gov/nuccore/MW248108), [MW248106,](https://www.ncbi.nlm.nih.gov/nuccore/MW248106) and [MW248107](https://www.ncbi.nlm.nih.gov/nuccore/MW248107), respectively. Genome assemblies for WSSV-IN-LS and WSSV-IN-NS are available from figshare ([https://doi.org/10.6084/m9.](https://doi.org/10.6084/m9.figshare.14252951)figshare.14252951 and [https://doi.org/10.6084/m9.](https://doi.org/10.6084/m9.figshare.14252966)figshare [.14252966,](https://doi.org/10.6084/m9.figshare.14252966) respectively).

ACKNOWLEDGMENTS

All authors were supported by the BBSRC/Newton Fund/DBT (Novel Molecular Approaches for Advancing Prediction and Mitigation of Disease Outbreaks in Aquaculture for Small Scale Farmers project [BT/IN/Indo-UK/BBSRC-Aqua/37/RJK/2015-16]) (grant BB/ N00504X/1). Indian authors (R.K.V., D.A., A.K., M.M., S.H.K., M.K.B., R.J.K., R.G.M., M.M.M., S.M., S.P., and K.D.) acknowledge the Department of Biotechnology, Government of India, and the Indian Council of Agricultural Research (ICAR) (New Delhi, India).

REFERENCES

- 1. Sánchez-Paz A. 2010. White spot syndrome virus: an overview on an emergent concern. Vet Res 41:43. [https://doi.org/10.1051/vetres/2010015.](https://doi.org/10.1051/vetres/2010015)
- 2. Kulkarni A, Deepika A, Sanath Kumar H, Bedekar MK, Chaput DL, Rajendran KV. 2017. Evaluation of genomic DNA extraction methods for gut microbiome analysis of Penaeus (Litopenaeus) vannamei by high throughput sequencing (HTS). 11th Indian Fisheries and Aquaculture Forum, Kochi, India, 21 to 24 November 2017, poster AH PO 36.
- 3. Chen S, Zhou Y, Chen Y, Gu J. 2018. fastp: an ultra-fast all-in-one FASTQ preprocessor. Bioinformatics 34:i884–i890. [https://doi.org/10.1093/bioinformatics/](https://doi.org/10.1093/bioinformatics/bty560) [bty560](https://doi.org/10.1093/bioinformatics/bty560).
- 4. Bushnell B. 2014. BBMap. <https://sourceforge.net/projects/bbmap>.
- 5. Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol 19: 455–477. <https://doi.org/10.1089/cmb.2012.0021>.
- 6. Camacho C, Coulouris G, Avagyan V, Ma N, Papadopoulos J, Bealer K, Madden TL. 2009. BLAST+: architecture and applications. BMC Bioinformatics 10:421. <https://doi.org/10.1186/1471-2105-10-421>.
- 7. Sivakumar S, Vimal S, Abdul Majeed S, Santhosh Kumar S, Taju G, Madan N, Rajkumar T, Thamizhvanan S, Shamsudheen KV, Scaria V, Sivasubbu S, Sahul Hameed AS. 2018. A new strain of white spot syndrome virus affecting Litopenaeus vannamei in Indian shrimp farms. J Fish Dis 41:1129–1146. [https://doi](https://doi.org/10.1111/jfd.12811) [.org/10.1111/jfd.12811](https://doi.org/10.1111/jfd.12811).
- 8. Treangen TJ, Ondov BD, Koren S, Phillippy AM. 2014. The Harvest suite for rapid core-genome alignment and visualization of thousands of intraspecific microbial genomes. Genome Biol 15:524. [https://doi.org/10.1186/s13059-014](https://doi.org/10.1186/s13059-014-0524-x) [-0524-x.](https://doi.org/10.1186/s13059-014-0524-x)
- 9. Ronquist F, Teslenko M, van der Mark P, Ayres DL, Darling A, Höhna S, Larget B, Liu L, Suchard MA, Huelsenbeck JP. 2012. MrBayes 3.2: efficient Bayesian phylogenetic inference and model choice across a large model space. Syst Biol 61:539–542. [https://doi.org/10.1093/sysbio/sys029.](https://doi.org/10.1093/sysbio/sys029)
- 10. Miller MA, Pfeiffer W, Schwartz T. 2010. Creating the CIPRES Science Gateway for inference of large phylogenetic trees, p 1-8. In 2010 Gateway Computing Environments Workshop (GCE). IEEE, Piscataway, NJ.
- 11. Stamatakis A. 2014. RAxML version 8: a tool for phylogenetic analysis and post-analysis of large phylogenies. Bioinformatics 30:1312–1313. [https://](https://doi.org/10.1093/bioinformatics/btu033) [doi.org/10.1093/bioinformatics/btu033.](https://doi.org/10.1093/bioinformatics/btu033)