Isolation and Identification of Myricitrin, an Antioxidant Flavonoid, from Daebong Persimmon Peel

In-Wook Hwang¹ and Shin-Kyo Chung²

¹Department of Food Science and Nutrition, Dong-A University, Busan 49315, Korea ²School of Food Science and Biotechnology, Kyungpook National University, Daegu 41566, Korea

ABSTRACT: In this study, the antioxidant flavonoid, myricitrin, was isolated and identified from Daebong persimmon peel. The persimmon peel extract was successively fractionated with *n*-hexane, ethyl acetate, and *n*-butanol. The ethyl acetate fraction had the strongest antioxidant activities among the solvent fractions and was further fractionated by silica gel and octadecylsilane column chromatography, and preparative high performance liquid chromatography. Three antioxidant compounds were finally isolated, and compound 2 was identified as myricitrin by liquid chromatography/mass spectrometry and ¹H and ¹³C nuclear magnetic resonance. Myricitrin had the strongest antioxidant activities by ferric ion reducing antioxidant power and α, α -diphenyl-2-picrylhydrazyl radical scavenging assays. These results suggested that myricitrin was found as a major antioxidant flavonoid responsible for the strong antioxidant activities of Daebong persimmon peels.

Keywords: antioxidant, myricitrin, persimmon peel, LC/MS, NMR

INTRODUCTION

Free radicals and oxidative stress have been associated with many diseases including atherosclerosis, diabetes mellitus, neurodegenerative diseases, and virus infections (1). Flavonoids are capable of preventing free radical and oxidative stress, thus stopping oxidative damage (2,3). Consequently, flavonoids have become of interest for contemporary health care.

In this respect, persimmon (Diospyros kaki), having various biological effects including antioxidant, anti-diabetic, and anti-cancer (4,5), is one of the useful resources to obtain the natural antioxidant flavonoids. It contains abundant phenolic acids, vitamins, and flavonoids including catechin, epicatechin, and gallocatechin (6). Among the persimmon varieties, Daebong persimmon (Diospyros kaki L. cv. Hachiya) is one of the astringent-type varieties consumed as dried product after removing peels. Consequently, a large amount of persimmon peels are discarded at the production areas. Some researchers had reported persimmon health-benefits include hypocholesterolemic (7), anti-tumor (8), and anti-diabetic effects (9). We also reported the physicochemical properties and pretreatment of Daebong persimmon peel for functional food materials (10), and studied the enzymatic hydrolysis process of Daebong persimmon peels for vinegar fermentation (11).

In this study, we isolated the antioxidant compounds from Daebong persimmon peel by antioxidant activity-guided fractionation and identified the chemical structure by instrumental analyses, including high performance liquid chromatography (HPLC), liquid chromatography/mass spectrometry (LC/MS), ¹H and ¹³C nuclear magnetic resonance (NMR).

MATERIALS AND METHODS

Preparation of Daebong persimmon peel powder and chemicals

Daebong persimmon peels were collected at the production area (Gwangyang, Korea). Persimmon peels were dried in the shade and ground in a roller mill (C.W. Brabender Instruments, Inc., South Hackensack, NJ, USA) at 1,000 rpm with a 0.5-mm screen. α , α -Diphenyl-2-picrylhydrazyl (DPPH), 2,4,6-tris(2-pyridyl)-1,3,5-triazine (TPTZ), gallic acid, and Trolox were purchased from Sigma-Aldrich Co. (St. Louis, MO, USA). All organic solvents were of analytical grade (Merck KGaA, Darmstadt, Germany), except for HPLC, LC/MS (J.T. Baker Chemical

Copyright © 2018 by The Korean Society of Food Science and Nutrition. All rights Reserved.

Received 18 July 2018; Accepted 24 August 2018; Published online 31 December 2018

Correspondence to Shin-Kyo Chung, Tel: +82-53-950-5778, E-mail: kchung@knu.ac.kr

This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (http://creativecommons.org/licenses/by-nc/4.0) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

Co., Phillipsburg, NJ, USA), and NMR (Cambridge Isotope Laboratories, Andover, MA, USA).

Extraction and isolation of antioxidant compounds

The isolation scheme of antioxidant from Daebong persimmon peel is shown in Fig. 1. Briefly, persimmon peel powders were extracted with 80% ethanol under reflux for 12 h. This ethanol extract was filtered and concentrated to remove ethanol using rotary evaporation. The resultant extract was successively fractionated with nhexane, ethyl acetate, and *n*-butanol. The *n*-hexane, ethyl acetate, and *n*-butanol extracts were separately evaporated, whereas the aqueous layer was lyophilized to dryness. Among them, greater antioxidant activity was observed in the ethyl acetate fraction compared with other fractions. The ethyl acetate fraction was subjected to silica gel column chromatography, and was eluted with solvents of increasing polarities. Fraction 2 was further isolated using octadecylsilane (ODS) column chromatography. Finally, fraction 2 was purified by preparative HPLC, and compounds 1, 2, and 3 were obtained. Isolated compounds were analyzed using an HPLC (1260 Infinity Quaternary LC System, Agilent Technologies, Palo Alto, CA, USA) equipped with a ultraviolet (UV) detector at 280 nm (C18 column, Develosil ODS-HG-5, 4.6×150 mm, Nomura Chemical Co., Ltd., Seto, Japan).

Instrumental analyses

UV/visible (Vis) analysis was conducted to confirm the polyphenol type of fractions using UV/Vis spectrophotometer (UV 1601 PC, Shimadzu Co., Kyoto, Japan) in the wavelength range of 200~600 nm. LC/MS analysis

was conducted to identify the molecular weight of isolated compounds. Determination was performed with a 6410 Triple Quadrupole LC/MS system (Agilent Technologies). Electro spray ionization-MS was performed in the positive ion mode (600° C, 20 psi). Data were collected using a mass scan from 0 to 460 m/z, at 1.5 s per scan. ¹H and ¹³C NMR spectra of isolated compounds were recorded on a NMR spectrometer (400 MHz for ¹H NMR and 100 MHz for ¹³C NMR; Bruker Avance Digital 400, Bruker, Karlsruhe, Germany). Chemical shift (δ) for ¹H and ¹³C NMR are recorded in parts per million (ppm) relative to solvent signals (methanol- d_4 : δ_H 3.30 and δ_C 49.0) as internal standards.

Antioxidant activities

To test the antioxidant activities, DPPH radical scavenging activity (12) and ferric ion reducing antioxidant power (FRAP) assays (13) were conducted. The DPPH radical scavenging activities of the extracts and fractions were determined by measuring the decrease in absorbance of methanolic DPPH solution at 517 nm in the presence of the sample. The initial concentration of DPPH was 0.1 mM and the reading was taken after allowing the solution to stand for 30 min. In cases where the absorbance decreased too much (when the solution turned yellow) before the 30 min period, the sample was appropriately diluted. The results are expressed in μ M gallic acid equivalents (μ M GAE).

The FRAP assay was performed using TPTZ solution. The working solution was prepared by mixing 25 mL of acetate buffer (pH 3.6), 2.5 mL of TPTZ solution (10 mM), and 2.5 mL of FeCl₃· $6H_2O$ solution (20 mM), and



Fig. 1. Isolation schemes of antioxidant from Daebong persimmon peels.

warmed at 37°C before every determination. Then, 175 μ L of this working solution was mixed with 25 μ L of the sample solution. After incubation at 37°C for 30 min in the dark, the absorbance of the sample solution was measured at 590 nm. Results are expressed in μ M trolox equivalents (μ M TE). All of the tests were performed in triplicate.

Statistical analysis

The values were expressed as the mean \pm standard deviation. The results were evaluated through analysis of variance with Duncan's multiple range test (*P*<0.05) using Statistical Analysis System software version 9.3 (SAS Institute Inc., Cary, NC, USA).

RESULTS AND DISCUSSION

Isolation of antioxidant compounds

Isolation process of antioxidant compounds is shown in Fig. 1. The extraction yield was 17.5 g of extract per 100 g of persimmon peel powder. The ethanol extract was fractionated with *n*-hexane, ethyl acetate, and *n*-butanol

by their polarity. The yields of fractions including *n*-hexane fraction, ethyl acetate fraction, *n*-butanol fraction, and aqueous fraction were 365 mg, 1.45 g, 3.24 g, and 12.16 g, respectively. Among them, the ethyl acetate fraction showed the strongest antioxidant activity. Thus, antioxidant compounds of ethyl acetate fraction were isolated in further experiments. The ethyl acetate fraction was separated with chloroform/methanol by step-wise elution using a silica gel column chromatography, and three fractions were obtained. Among them, fraction 2 showed the strongest antioxidant activity and the highest amount (975.8 mg). Thus, fraction 2 was separated by ODS column chromatography, and fraction 2-1 (466.2 mg) and 2-2 (505.6 mg) were obtained. Finally, prep-HPLC was conducted to completely separate with Fr. 2-2.

As a result, fraction 2-2 was separated to three compounds (compound 1, 35.2 mg; compound 2, 180.6 mg; compound 3, 274.3 mg). In HPLC chromatogram (Fig. 2), it was supposed that compound 2 was pure.

Identification of isolated compound

To identify the chemical structure of compound 2, UV/Vis absorption spectrum, LC/MS, 1 H and 13 C NMR



Fig. 2. High performance liquid chromatography (HPLC) chromatograms of compound 2 from Daebong persimmon peels. Fr. 2-2, ODS column chromatography fractions; Compound 2, prep-HPLC fraction from fraction 2-2.



Fig. 3. Liquid chromatography/mass spectrometry spectrum of compound 2 from Daebong persimmon peel.

analyses were conducted. The instrumental data are shown in Fig. 3 and described as follows:

Compound 2 (myricitrin, myricetin 3-O-rhamnoside; 180.6 mg). UV, λ_{max} nm 257, 360 (MeOH); LC/MS, *m/z* 303, 465 (M+H)⁺; ¹H NMR (in CD₃OD), δ 6.19 (1H, *d*, 1.8 Hz, H-6), 6.35 (1H, *d*, 2.3 Hz, H-8), 6.94 (2H, *s*, H-2', 6'), 5.30 (1H, *d*, 1.8 Hz, H-1"), 4.21 (1H, *dd*, 3.2, 1.8 Hz, H-2"), 3.76~3.78 (1H, *dd*, 9.4, 3.4 Hz, H-3"), 3.31~ 3.34 (1H, *m*, H-4"), 3.48~3.54 (1H, *m*, H-5"), 0.94~0.96 (3H, *m*, H-6"); ¹³C NMR (in CD₃OD), δ 159.2 (C-2), 136.1 (C-3), 179.5 (C-4), 163.1 (C-5), 99.7 (C-6), 164.0 (C-7), 94.6 (C-8), 158.4 (C-9), 105.6 (C-10), 121.7 (C-1'), 109.6 (C-2', 6'), 146.7 (C-3', 5'), 137.7 (C-4'), 103.5 (C-1"), 71.7 (C-2"), 72.0 (C-3"), 73.2 (C-4"), 71.9 (C-5"), 17.5 (C-6").

UV/Vis absorption spectra of compound 2 showed specific absorption peaks at 257 and 360 nm (data not shown). This result indicated that compound 2 could be assumed as a flavonol (14,15). Molecular mass of compound 2 could be assumed as a myricitrin (Fig. 3) (C₂₁H₂₀O₁₂, myricetin 3-O-rhamnoside, m/z 303, 465 $[M+H]^+$, consistent with previous studies (16-18). The fragment ions at m/z 303 might be the aglycone ion from myricitrin whose rhamnosyl group was cleaved $([M-(C_6H_{12}O_5)+H]^+)$. Collision induction dissociation of the clusters led to the fragment at m/z 465, $[M+H]^+$, which, in turn, gave rise to the aglycone myricetin after rhamnose loss. Considering this result and previous research (19), electrospray proved to be the most efficient mass ionization method for the identification of biologically active substances.

The data of ¹H and ¹³C NMR indicated 21 carbon signals and the typical flavonol α -L-rhamnopyranoside pattern; δ 5.30 (1H, *d*; H-1"), δ 4.21 (1H, *dd*; H-2"), δ 3.76 \sim 3.78 (1H, *dd*; H-3"), δ 3.31 \sim 3.34 (1H, *m*; H-4"), δ 3.48 \sim 3.54 (1H, *m*; H-5"), and δ 0.94 \sim 0.96 (3H, *m*; H-6"). Moreover, the data showed chemical shift values of a sugar moiety and a singlet of two protons at δ 6.94 (2H, *s*; H-2' and 6') with pair of doublets at δ 6.19 (1H, *d*; H-6) and δ 6.35 (1H, *d*; H-8). It demonstrated the presence of myricetin structure and *O*-glycosidic linkage due to the rhamnopyranoside at C-3 position (δ 136.1). These NMR data were in good agreement with those of previous research (18,20). Thus, compound 2 was identified as a myricetin 3-*O*-rhamnoside, commonly known as a myricitrin.

The chemical structure of the compound 2 (myricitrin) is shown in Fig. 4. It has hydroxyl groups at 5 and 7 positions, and three continuous hydroxyl groups at 3', 4', and 5' positions. Rhamnose was attached at C-3 position, and it has three hydroxyl groups. Flavonoid glycosides such as astragalin, isoquercitrin, and myricitrin are contained in persimmon leaves (21). In addition, myricitrin was also isolated from *Nymphaea lotus* (22), *Chryso*-



Fig. 4. Chemical structure of compound 2 from Daebong persimmon peels.

balanus icaco (23), and Polygonum aviculare (24).

Antioxidant activities of isolated compounds

Antioxidant activities of isolated compounds were determined by DPPH radical scavenging activity and FRAP assay. The values of DPPH and FRAP were ranged from 105.18 $\sim\!165.75~\mu\text{M}$ GAE and 682.79 $\sim\!1,\!609.56~\mu\text{M}$ TE, respectively (Table 1). Compound 1 and 3 had similar activities and showed no significant differences. Compound 2, identified as a myricitrin, showed the strongest antioxidant activities. Thus, myricitrin is a compound that has the greatest impact on the antioxidant activities of Daebong persimmon peel. Myricitrin is rhamnose glycoside of myricetin contained in various plants, and it has been reported that has strong antioxidant activity as radical scavenger and metal-ion chelator (25). In addition, myricitrin has been reported to inhibit the aldose reductase activity (18) and the oxidation of low-density lipoprotein (26), and has anti-malarial effect (27). It also showed lower IC₅₀ values for \cdot OH and \cdot O₂⁻ than ascorbic acid in radical scavenging activity by electron spin resonance measurement (20). The reason of these activities is that flavonoids having hydroxyl groups in the B-

 Table 1. Antioxidant activities of isolated compounds from

 Daebong persimmon peels

Samples (1 mg/mL)	DPPH (µM GAE)	FRAP (µM TE)
Compound 1 Compound 2	138.27±3.46 ^b 165.75±1.57 ^a	703.09±90.18 ^b 1,609.56±90.88 ^a
Compound 3	105.18±5.03 ^c	682.79±40.87°

DPPH, α,α -diphenyl-2-picrylhydrazyl radical scavenging activity; FRAP, ferric ion reducing antioxidant power; GAE, gallic acid equivalent; TE, trolox equivalent.

Different letters (a-c) in a column are significantly different at P<0.05 by Duncan's multiple range test.

ring can act as strong radical scavengers through formation of a hydrogen bond with the semiquinone radical of the B-ring (28). In a previous study, the pyrogallol substituents in the B-ring of myricitrin also showed stronger inhibition effects of low-density lipoprotein oxidation induced by copper ion and α, α' -azobis-(2-amidinopropane dihydrochloride than the O-dihydroxy-substituted flavonoid (20). In terms of the importance of the B-ring, other researchers reported that myricetin reacts 6 times more quickly than quercetin to the galvinoxyl radical and concluded that flavonoid reactivity depends highly on the substitution pattern of OH groups in the B-ring (29). Moreover, flavonol glycosides showed stronger antioxidant activity than that of aglycones due to the number and localization of hydroxyl groups as well as hydrogen bonds (30). The role of the hydrogen bonds is to repair the damage in the structure of these flavonols when they donate hydroxyl hydrogens to free radicals. It is of crucial importance for human health because it can prevent the formation of new and perhaps more potent free radicals (31). Thus, myricitrin-rich persimmon peel can be utilized as a natural and strong antioxidant food material.

AUTHOR DISCLOSURE STATEMENT

The authors declare no conflict of interest.

REFERENCES

- Halliwell B, Gutteridge JMC. 2006. Free radicals in biology and medicine. 4th ed. Oxford University Press, New York, NY, USA. p 199.
- 2. Niki E. 2011. Do free radicals play causal role in atherosclerosis? Low density lipoprotein oxidation and vitamin E revisited. *J Clin Biochem Nutr* 48: 3-7.
- 3. Zhang Z, Liao L, Moore J, Wu T, Wang Z. 2009. Antioxidant phenolic compounds from walnut kernels (*Juglans regia* L.). *Food Chem* 113: 160-165.
- 4. Choi JH, Lee EY, Kim GJ, Park IH, Kim JS, Choi GB, Jung SG, Ham YS. 2006. Physicochemical properties and physiological activities of Ulsan sweet persimmon peel-flesh according to cultivars. *J Korean Soc Appl Biol Chem* 49: 309-314.
- Kim SK, Lee GD, Chung SK. 2003. Monitoring on fermentation of persimmon vinegar from persimmon peel. *Korean J Food Sci Technol* 35: 642-647.
- Nakatsubo F, Enokita K, Murakami K, Yonemori K, Sugiura A, Utsunomiya N, Subhadrabandhu S. 2002. Chemical structures of the condensed tannins in the fruits of *Diospyros* species. J Wood Sci 48: 414-418.
- Gorinstein S, Kulasek GW, Bartnikowska E, Leontowicz M, Zemser M, Morawiec M, Trakhtenberg S. 1998. The influence of persimmon peel and persimmon pulp on the lipid metabolism and antioxidant activity of rats fed cholesterol. J Nutr Biochem 9: 223-227.
- 8. Kawase M, Motohashi N, Satoh K, Sakagami H, Nakashima H, Tani S, Shirataki Y, Kurihara T, Spengler G, Wolfard K,

Molnár J. 2003. Biological activity of persimmon (*Diospyros kaki*) peel extracts. *Phytother Res* 17: 495-500.

- Lee SO, Chung SK, Lee IS. 2006. The antidiabetic effect of dietary persimmon (*Diospyros kaki* L. cv. Sangjudungsi) peel in streptozotocin-induced diabetic rats. *J Food Sci* 71: S293-S298.
- Hwang IW, Jeong MC, Chung SK. 2011. The physicochemical properties and the antioxidant activities of persimmon peel powders with different particle sizes. J Korean Soc Appl Biol Chem 54: 442-426.
- Hwang IW, Chung SK, Jeong MC, Chung HS, Zheng HZ. 2013. Optimization of enzymatic hydrolysis of persimmon peels for vinegar fermentation. *J Korean Soc Appl Biol Chem* 56: 435-440.
- 12. Blois MS. 1958. Antioxidant determinations by the use of a stable free radical. *Nature* 181: 1199-1200.
- Benzie IFF, Strain JJ. 1996. The ferric reducing ability of plasma (FRAP) as a measure of "antioxidant power": the FRAP assay. *Anal Biochem* 239: 70-76.
- Cockell CS, Knowland J. 1999. Ultraviolet radiation screening compounds. *Biol Rev Camb Philos Soc* 74: 311-345.
- Anouar EH, Gierschner J, Duroux JL, Trouillas P. 2012. UV/ Visible spectra of natural polyphenols: a time-dependent density functional theory study. *Food Chem* 131: 79-89.
- Yokomizo A, Moriwaki M. 2005. Transepithelial permeability of myricitrin and its degradation by simulated digestion in human intestinal Caco-2 cell monolayer. *Biosci Biotechnol Biochem* 69: 1774-1776.
- Shimizu R, Shimabayashi H, Moriwaki M. 2006. Enzymatic production of highly soluble myricitrin glycosides using βgalactosidase. *Biosci Biotechnol Biochem* 70: 940-948.
- Mok SY, Lee S. 2013. Identification of flavonoids and flavonoid rhamnosides from *Rhododendron mucronulatum* for. *albiflorum* and their inhibitory activities against aldose reductase. *Food chem* 136: 969-974.
- de Oliveira FMG, Romão W, Kuster RM. 2018. Identification of phenolic compounds in *Eugenia uniflora* leaves by FTICR MS in association with different ionization sources. *Anal Methods* 10: 1647-1655.
- 20. Chung SK, Kim YC, Takaya Y, Terashima K, Niwa M. 2004. Novel flavonol glycoside, 7-O-methyl mearnsitrin, from *Sageretia theezans* and its antioxidant effect. *J Agric Food Chem* 52: 4664-4668.
- Moon SH, Park KY. 1995. Antimutagenic effects of boiled water extract and tannin from persimmon leaves. J Korean Soc Food Nutr 24: 880-886.
- Elegami AA, Bates C, Gray AI, Mackay SP, Skellern GG, Waigh RD. 2003. Two very unusual macrocyclic flavonoids from the water lily *Nymphaea lotus*. *Phytochemistry* 63: 727-731.
- Barbosa WLR, Peres A, Gallori S, Vincieri FF. 2006. Determination of myricetin derivatives in *Chrysobalanus icaco L*. (Chrysobalanaceae). *Rev Bras Farmacogn* 16: 333-337.
- 24. Xu F, Guan H, Li G, Liu H. 2009. LC method for analysis of three flavonols in rat plasma and urine after oral administration of *Polygonum aviculare* extract. *Chromatographia* 69: 1251.
- Luo XD, Basile MJ, Kennelly EJ. 2002. Polyphenolic antioxidants from the fruits of *Chrysophyllum cainito* L. (star apple). *J Agric Food Chem* 50: 1379-1382.
- Yokomizo A, Moriwaki M. 2005. Myricitrin degraded by simulated digestion inhibits oxidation of human low-density lipoprotein. *Biosci Biotechnol Biochem* 69: 693-699.
- Liu Y, Murakami N, Ji H, Abreu P, Zhang S. 2007. Antimalarial flavonol glycosides from *Euphorbia hirta*. *Pharm Biol* 45: 278-281.
- Guo Q, Zhao B, Shen S, Hou J, Hu J, Xin W. 1999. ESR study on the structure-antioxidant activity relationship of tea ca-

techins and their epimers. Biochim Biophys Acta 1427: 13-23.

- 29. McPhail DB, Hartley RC, Gardner PT, Duthie GG. 2003. Kinetic and stoichiometric assessment of the antioxidant activity of flavonoids by electron spin resonance spectroscopy. *J Agric Food Chem* 51: 1684-1690.
- 30. Chen ZY, Chan PT, Ho KY, Fung KP, Wang J. 1996. Antioxi-

dant activity of natural flavonoids is governed by number and location of their aromatic hydroxyl groups. *Chem Phys Lipids* 79: 157-163.

31. Leopoldini M, Marino T, Russo N, Toscano M. 2004. Antioxidant properties of phenolic compounds: H-atom versus electron transfer mechanism. *J Phys Chem A* 108: 4916-4922.