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## Cinnamic aldehyde, an anti-inflammatory component in Du-Huo-Ji-Sheng-Tang, ameliorates arthritis in II collagenase and monosodium iodoacetate induced osteoarthritis rat models



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## ABSTRACT

*Background and aim:* Du-Huo-Ji-Sheng-Tang (DHJST) is a Chinese herbal formula used for arthralgia and arthritis treatment clinically. This study aims to evaluate the joint-protecting efficacy of DHJST and to identify the active constituents as the evaluation marker.

*Experimental procedure:* DHJST can be categorized into three recipes: Blood-tonifying-herbs Si-Wu-Tang (SWT), Wind-dampness-dispelling-herbs (WDH) and Qi-tonifying-herbs (TH). All formulas were used to explore the joint-protecting efficacies.

Results and conclusion: s: Firstly, DHJST could decrease the arthritis progression in the monosodiumiodoacetate-induced rat and cure arthritis in the type II collagenase-induced rat. Further, in lipopolysaccharide-stimulated RAW 264.7 cells, DHJST, TH and Cinnamomum cassia (CC), an ingredient in TH, were the most potent nitric oxide (NO) and prostaglandin E<sub>2</sub> (PGE<sub>2</sub>) inhibitors. The major components, cinnamic aldehyde, showed the strongest NO and PGE2 inhibition. Up-regulated inducible NO synthase (iNOS) and cyclooxygenase-2 were inhibited by DHIST, TH, CC, and cinnamic aldehyde. In interleukin-1β-stimulated primary chondrocytes, upregulated iNOS was inhibited by DHJST, TH, Cinnamomum cassia, and cinnamic aldehyde. Upregulated matrix metalloprotease-13 was only inhibited by DHIST and TH and Eucommia ulmoides (EU) extract. Results suggest that DHIST presented joint-protective and cure arthritis effects. TH presented equal joint-protective effects as DHJST. The major antiinflammatory ingredient in TH was Cinnamomum cassia in TH. And cinnamic aldehyde was the potent anti-inflammatory active compound in Cinnamomum cassia. Therefore, this study may facilitate the modern use of DHJST with TH as a simplified version but equally effective anti-osteoarthritic agents with cinnamic aldehyde as a quality control marker of DHIST and TH in osteoarthritis prevention or treatment. © 2023 Center for Food and Biomolecules, National Taiwan University. Production and hosting by Elsevier Taiwan LLC. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/ licenses/by-nc-nd/4.0/).

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### 1. Introduction

From the perspective of traditional Chinese medicine (TCM), arthralgia is due to wind, cold, and dampness in the joints as a blockage in the smooth flow of qi and blood within the body. Du-

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List of abbreviations		LPS	lipopolysaccharide
		LSD	least significant different
ANOVA	a one-way analysis variance	MIA	monosodium iodoacetate
CC	Cinnamomum cassia	MMP	matrix metalloproteinase
CIA	type II collagenase-induced arthritis	NO	nitric oxide
COX-2	cyclooxygenase-2	OA	osteoarthritis
DHJST	Du-Huo-Ji-Sheng-Tang	PG	Panax ginseng
DMEM	Dulbecco's modified Eagle's medium	PGE <sub>2</sub>	prostaglandin E <sub>2</sub>
ELISA	enzyme-linked immunosorbent assay	PRCs	primary chondrocytes
EU	Eucommia ulmoides	RIPA	radioimmunoprecipitation assay
FBS	fetal bovine serum	SDS-PAGE	sodium dodecylsulfate polyacrylamide gel
GU	Glycyrrhiza uralensis		electrophoresis
H&E	hematoxylin and eosin	SWT	Si-Wu-Tang
IC <sub>50</sub>	50% inhibition	TCM	traditional Chinese medicine;
IL	interleukin	TH	Qi-tonifying-herbs
iNOS	inducible NO synthase	WDH	Wind-dampness-dispelling-herbs

Huo-Ji-Sheng-Tang (DHJST) has been widely and safely used for over 1000 years. This formula, originally recorded in Bei-Ji-Qian-Jin-Yao-Fang (in about 652 A.D.), relieves arthralgia due to wind, cold, and dampness by three pharmacological actions: nourishing the blood, invigorating Qi, and expelling and removing wind and dampness. Today, it is still one of the most commonly prescribed Chinese herbal formulas to treat osteoarthritis (OA).<sup>1</sup> In 2002, the prescription rate of DHJST was 26.6%, ranked highest among a total of 32,050 Chinese herbal formulae reimbursed by National Health Insurance for OA treatment in Taiwan.<sup>2</sup>

Goal of arthritis treatment is to reduce pain, improve mobility and prevent further damages to the joints. However, effective disease modifying drugs for osteoarthritis treatment are still limited. Because inflammation plays an important role in the development of arthritis, therefore, anti-inflammation is one of the most frequently treatment strategy used to treat arthralgia and arthritis.<sup>3,4</sup> In vitro and in vivo studies have demonstrated that when inflammation is activated, the activated NF-KB in the macrophage induce interleukin (IL)-1 $\beta$  release, which in turn induced caspase-mediated chondrocyte damage and arthropathy.<sup>5–7</sup> TCM is becoming a promising option for arthritis treatment, as TCM and its components could ameliorate symptoms or prevent progression of arthritis through multiple targets, including inhibition of pro-inflammatory cytokines and cartilage degradation regulators.<sup>8,9</sup> So far, how this complex formula works as an anti-arthritic formula remained inadequately characterized. Reported action mechanism of DHJST and its component on joint protection include by inhibiting hypoxia or inflammation related factors.<sup>10–12</sup> Interestingly, the blood tonic components in DHJST, Angelica sinensis, Ligusticum chuanxiong, Paeonia lactiflora, and Rehmannia glutinosa, is actually a famous formula known as Si-Wu-Tang (SWT), but anti-inflammatory effect has not been reported in SWT.<sup>13</sup> Therefore, we studied DHIST by grouping the constituent herbs into one of the following categories, blood nourishing, Qi invigorating, and wind/dampness expelling and removing, and compared their anti-inflammatory activities using in vitro and in vivo methods. Deciphering the active anti-inflammatory components of DHJST may modernize the cultivation, manufacture, and clinical applications of this popular formula.

### 2. Materials and methods

#### 2.1. Preparation of extracts

DHJST is composed of the following 15 plant species, and the

(10 times the weight of the material) and refluxed for 2 h at  $65 \circ C$ . with this process being repeated twice. The twice-filtered solutions were combined and concentrated by rotary evaporation. The aquatic solution was freeze-dried to a powder.

## 2.2. HPLC analysis

The HPLC apparatus used was composed of an SCL-10Avp system controller, an LC-10ATvp liquid chromatographic pump, an SPD-M10A diode array detector, an SIL-10Avp auto-injector, a CTO-10A column oven, FCV-10Avp flow-channel selection valves (Shimadzu, Tokyo, Japan), and an ERC-3415 degasser (ERC, Altegolfsheim, Regensburg, Germany). The stationary phase was comprised of a Purospher® STAR RP-18e reverse-phase column (4 mm i.d.  $\times$  250 mm, 5  $\mu m$ , Merck, USA), and the mobile phase system included 0.05% trifluoroacetic acid: CH<sub>3</sub>CN gradient elution (5 min, 90:10; 15 min, 75:25; 25 min, 55:45; 35 min, 40:60; and 45 min, 0:100). The flow rate was 1 mL/min, and the column oven temperature was maintained at 40 °C. The ultraviolet wavelength was set to 280 nm for detecting the fingerprint chromatograms of the 50% ethanol extracts of DHJST, WDH, SWT, TH and CC.

prescription is as shown in Table 1. The herbs were grouped into

three compound recipes according to their use in clinical settings based on TCM theories: Recipe 1, Si-Wu-Tang (SWT): Rehmannia

glutinosa, Angelica sinensis, Paeonia lactiflora and Ligusticum

chuanxiong; Recipe 2, wind-dampness herbs (WDH): Achyranthes

bidentate, Angelica pubescens, Gentiana macrophylla, Taxillus chi-

nensis, Saposhnikovia divaricate, Asarum heteropoides and Poria co-

cos, and Recipe 3, tonifying herbs (TH): Eucommia ulmoides (EU),

Cinnamomum cassia (CC), Panax ginseng (PG) and Glycyrrhiza ura-

lensis (GU). Voucher specimens (DHJST-01 to DHJST-15) were

deposited in the School of Pharmacy, College of Pharmacy, Taipei Medical University (Taipei, Taiwan). DHIST, SWT, WDH and TH

decoction used in the experiment were extracted with 50% ethanol

### 2.3. Animals

Wistar Rats weighing 180-220 g were housed in a controlled environment at 21 °C with sufficient food and water and kept on an alternating 12 h dark and light cycle. Animal experiments were approved according to Ethical Regulations on Animal Research of Taipei Medical University (approval no: LAC-100-0043).

#### Table 1

Prescri	ption	of Du-	Huo-li	i-Sheng	g-Tang	and	herbal	traditional	use in	traditional	Chinese	medicine.

Traditional action	Name	Family	Scientific name	Part used	Dose (g)	Recipe
Blood-tonifying herb tonifies the blood in treatingblood deficiencies	地黃 Di Huang	Scrophulariaceae	Rehmannia glutinosa (Gaertn.) DC.	Root	2	SWT
	當歸 Dang Gui	Umbelliferae	Angelica sinensis (Oliv.) Diels	Root	2	SWT
	白芍 Shao Yao	Ranunculaceae	Paeonia lactiflora Pall.	Root	2	SWT
Blood-activating and stasis-resolving herb promotes blood flow to remove blood stasis	川芎 Chuan	Umbelliferae	Ligusticum chuanxiong Hort.	Rhizome	2	SWT
	Xiong 牛膝 Niu Xi	Amaranthaceae	Achyranthes bidentata Blume	Root	2	WDH
Wind/dampness-dispelling herb dispels wind and dampness from the body	獨活 Du Huo	Umbelliferae	Angelica pubescens Maxim.	Root	3	WDH
	秦艽 Oin Iiao	Gentianaceae	Gentiana macrophylla Pall.	Root	2	WDH
	桑寄生 Sang Ji Sheng	Loranthaceae	Taxillus chinensis (DC.) Danser	Twig	2	WDH
Wind/cold-dispersing herb eliminates wind and cold in superficial layers of the	防風 Fang Feng	Umbelliferae	Saposhnikovia divaricata (Turcz.) Schmidt	Root	2	WDH
body	細辛 Xi Xin	Aristolochiaceae	Asarum heterotropoides Fr. Schmidt var. mandshuricum Kitag	Root	2	WDH
Dampness-draining diuretic herb increases urine excretion and water discharge	茯苓 Fu Ling	Polyporaceae	Poria cocos (Schw.) Wolf	Indian Bread	2	WDH
Yang-tonifying herb tonifies yang qi in treating yang deficiency	杜仲 Du Zhong	Eucommiaceae	Eucommia ulmoides Oliv.	Bark	2	TH
Interior-warming herb warms the interior and expels internal cold	肉桂 Rou Gui	Lauraceae	Cinnamomum cassia (L.) J.Presl	Bark	2	TH
Qi-tonifying herb tonifies the healthy qi in treating qi deficiencies	人 Ren Shen	Araliaceae	Panax ginseng C. A. Mey.	Root	2	TH
	甘草 Gan Cao	Leguminosae	Glycyrrhiza uralensis Fisch. ex DC.	Root and rhizome	2	TH

## 2.4. Monosodium iodoacetate-induced arthritis (MIA) in Wistar rats model

Monosodium iodoacetate-induced arthritis (MIA) model was used to induce osteoarthritis in rats through cartilage digestion, inflammatory cell proliferation, matrix degradation, and cartilage destruction. Male Wistar rats were used in the experiments. divided into following three groups: control, DHJST low dose (25 mg/kg) and high dose (50 mg/kg), and TH groups, with 8 rats per group. MIA injections of 50 µL of 80 mg/mL on day 0 and 40 mg/ mL on day 6 were given into the left ankle in the control group and treatment groups. In addition, Treatment groups received medication (50 mg/kg) orally every day for 10 days since day 0. Control group received sham injection and served as the normal control (normal). Measurement of the efficacy of treatment included paw edema measurements using a plethysmometer (Ugo Basile, Comerio VA, Italy) on days 0, 1, and 3; hind-limb weight-bearing distribution ratio on days 0 and 5; and a hot-plate latent pain response test on days 0 and 10. This model was used to assay osteoarthritic protective effect of the treatment drug.

#### 2.5. Type II collagenase-induced arthritis (CIA) in Wistar rats model

Type II collagenase was used to digest cartilage to produce osteoarthritis joint before treatment. The experimental rats were divided into control group and two treatment groups (each group with 8 rats). The left knee of a Wistar rat was injected with type II collagenase (4 mg/kg) under anesthesia with Zoletil and xylazine. Rats' weight-bearing ratio had significantly changed on day 14 after the type II collagenase injection suggesting OA knee was successfully established. After the MIA mode has been established, treatment groups received 25 or 50 mg/kg DHJST every day orally for 21 days. Measurement of the efficacy of anti-osteoarthritic effect of the treatment drug include hind-limb weight-bearing distribution ratio on treatment days 0, day 7 and day 21 and histological examination. The model was used to assay the cure effect of the treatment drug.

#### 2.6. Assessment of change in hind limb weight distribution

Weight-bearing distributions of both hind limbs were measured, and the weight-bearing ratios of hind limbs were calculated. Measurements were observed by an incapacitance tester with a dual-channel weight averager (Linton Instrumentation, Norfolk, UK). The weight-bearing force measured by the hind limbs was averaged over a 3-s period. %weight distribution of left hind paw = weight on left hind limb/(weight on right hind limb + weight on left hind limb) X 100. The ratio of weight distribution for the MIA-treated group maintained significantly lower than that for the normal control group until day 21.

#### 2.7. Hot-plate latent pain responses

The hot-plate latent pain response test was performed as previously described and was assessed on days 0 and  $10.^{14}$  Briefly, in the thermal hyperalgesia analysis, animals were individually placed on a hot-plate instrument (Ugo Basile, Comerio, Varese, Italy). A radiant heat source was applied underneath the glass floor. The time between the placement of the animal on the platform and licking of the paws was recorded as the hotplate latency. The vehicle (soybean oil), DHJST, and TH were orally administered every day at 1–2 h before the hot-plate test (8 animals/group) for 10 days.



**Fig. 1.** Effects of DHJST on MIA-induced arthritis. Experimental flow chart (A). Effect of DHJST on MIA-induced paw edema swelling on days 1 and 3 (B). hind-limb weight-bearing on day 5 (C); hot-plate latent pain responses on day 10 (D). \*p < 0.05, compared to the control. n = 3.

#### 2.8. Histological examination

When the rats were sacrificed via anesthesia, joints from Type II collagenase and MIA-injected experimental animals were harvested and fixed in 10% (v/v) neutral buffered formalin for 24 h, paraffin-embedded, and cut into 5  $\mu$ m thicknesses for histopathological assessments. Hematoxylin and eosin (H&E) staining was performed on paraffin-embedded sections to evaluate the severity of cell infiltration, and cartilage damage. After staining, cartilage damage in the joint tissue were investigated by histological examination under a light microscope. The severity of arthritis was categorized no detectable changes, mild, or severe.

#### 2.9. Cell culture

The murine macrophage RAW 264.7 cell line was purchased from American Type Culture Collection (Rockville, MD, USA). Cells were cultivated in Dulbecco's modified Eagle's medium (DMEM) supplemented with 10% fetal bovine serum (FBS), 100 IU/mL penicillin, and 100  $\mu$ g/mL streptomycin (Gibco BRL, Grand Island, NY, USA) in a humidified incubator containing 5% CO<sub>2</sub> at 37 °C.

#### 2.10. Primary chondrocytes (PRCs) from rat cartilage

PRC culture was obtained as previously described.<sup>15</sup> Briefly, cartilage specimens cut into 1-mm<sup>3</sup> pieces were sequentially digested with pronase (10 mg/mL) and collagenase IV (5 mg/mL). A PRC monolayer culture was established in 60-mm Petri dishes at a concentration of  $6 \times 10^6$  cells/mL in DMEM with 10% FBS, 100 µg/mL streptomycin, and 100 µg/mL penicillin. PRCs were incubated in a humidified atmosphere of 95% air and 5% CO<sub>2</sub> at 37 °C. Experiments were performed with cells obtained from the 3rd passage.

#### 2.11. NO and PGE<sub>2</sub> assays

Anti-inflammatory drugs often possess iNOS and COX-2 inhibitory actions, therefore, inhibition of their products, NO and PGE<sub>2</sub> is frequently used to identify anti-inflammatory activity in the investigational medicinal herbs. RAW 264.7 cells were treated with lipopolysaccharide (LPS) 500 ng/mL. After 18 h of incubation with or without samples, NO in the culture medium were assessed spectrophotometrically at 530 nm after the Griess reaction. The NO inhibition percentage was calculated using the following equation: NO inhibition (%) = [1 (T/C)] 100%; where T and C represent the mean optical density of LPS-stimulated RAW 264.7 cells with and without samples, respectively. PRCs were stimulated with IL-1 $\beta$  (10 ng/mL) for 6 h. PGE<sub>2</sub> concentrations were determined with an enzyme-linked immunosorbent assay (ELISA) kit (Enzo Life Sciences, NY, USA).

## 2.12. Western blot analyses of iNOS, COX-2, and matrix metalloproteinase (MMP)-13 expressions

Whole-cell lysates from cells treated with or without a sample for the scheduled hours were prepared by washing with PBS and lysing with radioimmunoprecipitation assay (RIPA) buffer. Equal amounts of protein from cell lysates were boiled for 5 min in sodium dodecylsulfate polyacrylamide gel electrophoresis (SDS-PAGE) sample buffer, separated by 10% SDS-PAGE, transferred to nitrocellulose membranes, and visualized using a BCIP/NBT kit (Gibco BRL). GAPDH expression was used as an internal control to compare with iNOS, COX-2, and MMP-13 expressions.

#### 2.13. Statistical analyses

Results are shown as the mean  $\pm$  standard deviation of the mean (SD). Data were analyzed using SPSS 17.0 software (SPSS, Chicago, IL, USA). Group differences were statistically assessed by a one-way analysis variance (ANOVA), followed by Fisher's least significant different (LSD) test for comparison of the means. *p* value < 0.05 was considered statistically significant.

#### 3. Results

#### 3.1. Effects of DHJST on MIA-induced osteoarthritis

We first validated the joint-protective effect of DHJST with MIAinduced arthritic model. After MIA injection, paw edema swelling on days 1 and 3 (Fig. 1A), hind-limb weight-bearing on day 5 (Fig. 1B), and hot-plate latent pain responses on day 10 (Fig. 1C) were all beneficially and significantly attenuated by DHJST treatment as compared to the negative control group. These results supported that DHJST formulae possessed anti-inflammatory and joint-protective effects.

# 3.2. Effect of DHJST on weight bearing ratio and histopathology in type II collagenase-induced osteoarthritis (CIA) models

We further observed the joint-protective effect of DHJST on CIA model. As shown in Fig. 2, DHJST protected the joint from destruction as the treatment groups resumed more-equal weight-



**Fig. 2.** Effect of DHJST on CIA-induced arthritis. Experimental flow chart (A). Changes in hind limb weight bearing in rats were measured (B). The weight distribution ratio was calculated as the left hind-limb (collagenase injection side) weight divided by the right hind-limb (control side) weight. Values are the mean  $\pm$  SD. \*p < 0.05, compared to the control; #p < 0.05, compared to day 0. n = 3.

bearing distributions of both hind limbs in a dose dependent manner. Histopathological assessments revealed low grade inflammatory cell infiltration and thinning of cartilage in the joint of control group, while joints from both low dose (25 mg/kg) and high dose (50 mg/kg) treated groups showed no detectable changes as compare to the normal group. These results confirm that DHIST has anti-arthritic effect. Effect of DHJST on pathological changes of joints from CIA rats was shown as in Fig. 3. As DHIST anti-arthritic effect was confirmed, the complete formula was used as positive control in the following experiments.

## 3.3. HPLC fingerprint profiles of DHJST, SWT, WDH and TH

Chromatographic profiles of DHJST and 3 recipes were analysis by HPLC. The results showed that DHJST, TH and Cinnamomum cassia samples contained the same major components. The three major peaks included 2-methylcinnamic acid (1), cinnamic acid (2), and cinnamic aldehyde (3), were identified by comparison to standard compounds (Fig. 4).

#### 3.4. Effects of DHJST, SWT, WDH and TH and the constituent herbs on NO and PGE<sub>2</sub>

Furthermore, the anti-inflammation effect of DHJST and 3 recipes were evaluated by LPS-stimulated RAW 264.7 cells. All of the



Fig. 4. HPLC profiles of DHJST (A), SWT (B); TH (C); WDH (D); Cinnamomum cassia (E). Abbreviations: Peak (1) 2-methylcinnamic acid; Peak (2) cinnamic acid; Peak (3) cinnamic aldehvde.

tested samples showed no cytotoxicity. First, the NO inhibitory effect of DHJST (300 µg/mL) was compared with mixtures extracts of 3 recipes, WDH, SWT and TH (100 µg/mL each). These extracts showed no significant difference in NO inhibition (Fig. 5A). However, when each formula was examined individually, TH showed

(A)

**(B)** 

200 µm





Fig. 3. Effect of DHJST on pathologic changes in CIA-induced arthritis model. Representative images were presented. (A), normal with normal cartilage thickness (double head arrow); (B), control with cartilage thinning (double head arrow); (C), DHJST treated (25 mg/kg/day) with cartilage thickness similar to the negative control (double head arrow); (D), DHJST treated (50 mg/kg/day) with cartilage thickness restored (double head arrow).

200 µm



**Fig. 5.** Inhibitory Effect DHJST, SWT, WDH and TH on LPS-induced nitric oxide (NO) production in RAW 264.7 cells. (A), DHJST (300  $\mu$ g/mL) and combined extracts of mixture of SWT, WDH and TH (100  $\mu$ g/mL for each); (B), DHJST, SWT, WDH and TH. \*p < 0.05, compared to DHJST; (C), TH (100  $\mu$ g/mL), mixture of TH and SWT (50  $\mu$ g/mL for each); \*p < 0.05, compared to TH; #p < 0.05, compared to SWT and WDH; (D), effect of TH ingredients on NO inhibition; (E), effect of three major compounds in *Cinnamonum cassia* on NO inhibition. Abbreviations: PG, *Panax ginsen*; EU, *Eucommia ulmoides*; GU, *Glycyrrhiza. Uralensis*; CC, *Cinnamonum cassia*; (1), 2-methylcinnamic acid; (2), cinnamic acid; (3), cinnamic aldehyde. Values are the mean  $\pm$  SD. n = 3.

strongest NO inhibitory effects among all the tested formula (Fig. 5B). We also compared the NO inhibitory effect of twocombined recipes to TH alone. In order to understand the addition effects of 3 recipes, TH (50  $\mu$ g/mL), SWT (50  $\mu$ g/mL) or WDH (50  $\mu$ g/mL) were added to each other. The results showed that TH could enhanced the NO inhibitory effect of combined extract (Fig. 5C), while combination of SWT and WDH showed no significant change on NO inhibitory activity. These results suggested that the NO inhibitory activity was related to herbs in TH. Further analyzing the NO inhibitory activity of the four constituent herbs of TH, extract of *Cinnamomum cassia* showed the most potent NO inhibitory activity, and the potency of EU was in the second place (Fig. 5D), suggesting that *Cinnamomum cassia* could be the major anti-inflammatory herb in TH. Among the three compounds, cinnamic aldehyde (3) showed the strongest NO inhibition (Fig. 5E). The concentration of DHJST, TH, *Eucommia ulmoides, Cinnamomum* 

#### Table 2

The 50% inhibitory concentration ( $IC_{50}$ ) values of tested samples on nitric oxide (NO) and prostaglandin  $E_2$  (PGE<sub>2</sub>) production from lipopolysaccharide (LPS)-stimulated RAW 264.7 cells.

Tested sample	IC <sub>50</sub> (μg/mL)							
	NO			PGE <sub>2</sub>				
DHJST	123.94	±	1.45	131.73	±	5.42		
TH	44.35	±	4.44	75.74	±	0.74		
CC	18.45	±	3.03	9.49	±	0.00		
EU	66.60	±	4.31	41.88	±	7.63		
Cinnamic aldehyde	1.01	±	0.56	1.79	±	0.23		

Data are presented as the mean ( $\pm$ S.D.) of at least three independent experiments, each performed in triplicate. DHJST: Du-Huo-Ji-Sheng-Tang, TH: Tonifying herbs, CC: *Cinnamomum cassia*, EU: *Eucommia ulmoides*.

*cassia* and cinnamic aldehyde (3) required for 50% inhibition ( $IC_{50}$ ) of NO and PGE<sub>2</sub> induced by LPS is showed in Table 2. The above data supported the important anti-inflammatory role of TH in DHJST, and of *Cinnamomum cassia* in TH. Cinnamic aldehyde (3) was the active compound in them.

# 3.5. Effect of DHJST, TH, and component herbs and cinnamic aldehyde on macrophages stimulated with LPS

We then examined the modulatory effects of DHJST, TH, *Cinnamomum cassia*, and cinnamic aldehyde on iNOS and COX-2 expressions in LPS-stimulated RAW 264.7 cells (Fig. 6A–E). The upregulated iNOS and COX-2 were inhibited by DHJST, TH, *Cinnamomum cassia*, and cinnamic aldehyde, with cinnamic aldehyde showed the most potent inhibitory activities. EU did not decrease COX-2 expression, but decreased iNOS expression.

3.6. Effect of DHJST, TH, Cinnamomum cassia, Eucommia ulmoides and cinnamic aldehyde on chondrocytes stimulated with IL-1 $\beta$ 

We also measured these extracts modulating effects on iNOS, and MMP-13 expressions and PGE<sub>2</sub> levels in IL-1 $\beta$ -stimulated PRCs. The upregulated iNOS were inhibited by all of the materials tested, but inhibitory activities on MMP-13 were only observed in DHJST, TH, and *Eucommia ulmoides* (Fig. 7A–E). Inhibitory activities of the 50 µg/mL of tested materials on PGE<sub>2</sub> levels in IL-1 $\beta$ -stimulated PRCs were shown in Fig. 7F.

3.7. Comparison of joint protective effects of DHJST and TH on MIAinduced arthritis

Lastly, we first validated the joint-protective effect of TH with MIA-induced arthritic model. After MIA injection, paw edema swelling on days 1 and 3 (Fig. 8A), hind-limb weight-bearing on day 5 (Fig. 8B), and hot-plate latent pain responses on day 10 (Fig. 8C) were all beneficially and significantly attenuated by DHJST or TH treatment. These results supported that TH formulae possessed anti-inflammatory and joint-protective effects as well as DHJST.

#### 4. Discussion

In this study, we grouped the 15 constituent herbs of DHJST according to their clinical use in one of the following traditional anti-arthritis action mechanisms, blood nourishing, Qi invigorating, and wind/dampness expelling and removing, in order to illuminate the anti-inflammatory and cartilage protection components in such a complex formula. Our data first confirms that DHJST and TH is as effective as DHJST and not just ameliorates arthritic pain symptoms, but they also have chondroprotective effect as both agents



Fig. 6. Regulatory effect on lipopolysaccharide (LPS)-induced iNOS and COX-2 production in RAW 264.7 cells by DHJST (A), TH (B), *Cinnamomum cassia*; (C), *Eucommia ulmoides* (D), and cinnamic aldehyde (E). Abbreviations: CC, *Cinnamomum cassia*; EU, *Eucommia ulmoides*.



**Fig. 7.** Regulatory effect on IL-1 $\beta$ -induced iNOS and MMP-13 in PRCs by DHJST (A), TH (B), *Cinnamomum cassia* (C), *Eucommia ulmoides* (D), and cinnamic aldehyde (E). Effect of 50 µg/mL of DHJST, TH, *Cinnamomum cassia* and *Eucommia ulmoides* on PGE<sub>2</sub> level (F), data represented as the mean (±S.D.) of at least three independent experiments, each performed in triplicate. Abbreviations: CC, *Cinnamomum cassia*; EU, *Eucommia ulmoides*.

significantly improved weight-bearing ability, attenuated inflammatory cells infiltration and cartilage degradation in MIA and CIA joints. Our results also suggested that TH, the Qi invigorating group of herbs, contained the anti-inflammatory and joint protective components in DHJST. *Cinnamomum cassia* was the effective antiinflammatory herb in TH, while cinnamic aldehyde was the major anti-inflammatory active compound in *Cinnamomum cassia*, as the IC<sub>50</sub> values of tested samples on LPS-induced NO and PGE<sub>2</sub> productions decreased as the extract constituents reduced in complexity.

In TCM, *Cinnamomum cassia* is commonly used for treating colds, fevers, headaches, myalgia, arthralgia, and amenorrhea because of its anti-inflammatory and analgesic activities. Several compounds extracted from this herb showed iNOS and COX-2 suppressive effects.<sup>16</sup> The major anti-inflammatory compound identified in this experiment was cinnamic aldehyde. Through inhibition of NF-κB activation, cinnamic aldehyde was found to have

anti-inflammatory and anti-oxidative activities on *Helicobacter pylori*-induced IL-8 secretion, and high glucose-induced damage in cultured dorsal root ganglion neurons.<sup>17,18</sup> Taken together, these results support our findings that cinnamic aldehyde could be used as an anti-inflammatory quality and quantity marker for DHJST.

Preclinical studies have provided sufficient evidences to support that iNOS, COX-2, MMP-13, and PGE<sub>2</sub> are all key targets for protecting joints from degradation during inflammation caused by inducers like IL-1 $\beta$ .<sup>19,20</sup> MMP-13 is over-expressed in the joints and articular cartilage in patients with OA, and is a new target of treatment. In fact, MMP-13 inhibitory activity in activated chondrocytes was reported in two TH constituent herbs, EU and *Panax ginseng*.<sup>21–23</sup> In our study, MMP-13 in IL-1 $\beta$  induced chondrocytes was inhibited by DHJST and TH extracts, and *Eucommia ulmoides*, but not by *Cinnamomum cassia*. We did not perform further study on *P. ginseng* as this herb showed only moderate anti-inflammatory activity on LPS induced macrophages. The results not only revealed how multiple ingredients act on multiple targets to produce the beneficial effects of DHIST for arthritis treatment, but they also support the notion that the pharmacological effects of combinatory formula could be more than the sum of each individual component. Osteoarthritis is a chronic painful joint disease. Cytokines as TNF- $\beta$ , IL-1a, IL-6 and IL-17 present in the OA joint can activate proinflammatory cytokines, there should have innervating nociceptors and leading to pain. For examples, TNF- $\beta$  and IL-1 $\alpha$  can stimulate TRPV1 expression and make joints feel hot and painful; IL-17 can increase TRPV4 expression and enhance the sensitivity to mechanical pain.<sup>24</sup> In this study, both DHJST and TH can inhibit the pain of osteoarthritic animal models. Therefore, in addition to DHJST and TH could reduce the production of PGE<sub>2</sub> in chondrocytes, there should be have other cytokine and chemokines targets, such as TNF- $\alpha$ , IL-1 $\beta$  and IL-17 et al. In the future, we will continue to explore whether DHIST and TH can reduce the expression of cytokines and chemokines in chondrocytes, synovial cells and other cells in the joints.

Du Huo (*A. pubescens*) is a widely used medicinal herb for arthritic treatment because of its anti-inflammatory, analgesic, and antioxidant properties through bioactive coumarin compounds.<sup>25–28</sup> Columbianectin, one of the coumarins obtainable from *A. pubescens*, has been reported to inhibit IL-1 $\beta$ , IL-6, IL-8, and TNF- $\alpha$  level in activated mast cells.<sup>28–30</sup> Du Huo is also a major herb in DHJST under the theory of TCM. However, because as a constituent in WDH group and the inhibitory activity of WDH on NO production by LSP induced RAW 264.7 cells was modest among the three compound recipes, Du Huo was not further explored in the current study either. However, due to the anti-inflammatory nature of Du Huo and columbianectin, investigation is worthwhile to illuminate whether this herb and its phytoconstituents are beneficial in the treatment of inflammatory arthritis in future clinical practice.

Drugs or therapies that could modify pathological structure and alleviate symptoms in the model animals may lead to the development of disease modifying drug for human arthritic disorders including osteoarthritis. Inflammatory reaction promotes cartilage destruction in animal models of arthritis such as the two models used in current study. CIA models have been extensively used in rheumatoid arthritis research. MIA model is frequently used as an acute osteoarthritis model because the chemical induce rapid cartilage degradation. Our results confirm the anti-osteoarthritic and anti-inflammatory effects of DHJST and the simplified formula TH by providing evidences from moderating inflammatory cytokine levels, weight-bearing distribution, and histopathological changes.

### 5. Conclusion

In our study, DHIST reduced paw swelling, weight bearing distribution, analgesic latency ratio of.collagenase type II- and MIAinduced arthritis in rats. The anti-inflammatory and joint protective mechanisms of DHJST were through inhibition of iNOS, COX-2, and MMP-13 expressions and PGE<sub>2</sub> production. Among the three simplified formulae we tested, TH, the Qi tonic, showed similar anti-inflammatory joint protective mechanism and effect as DHJST. A TCM prescription usually include at the same time the jun (or sovereign), chen (or minister), zuo (or assistant) and shi (or messenger) ingredient drugs. As Cinnamomum cassia showed the most potent anti-inflammatory effect among the 15 constituent herbs, it could be the sovereign medicine. Eucommia ulmoides could more inhibit MMP-13 expression than Cinnamomum cassia, therefore we suggested Eucommia ulmoides could be the minister medicine in DHIST or TH to help the action of Cinnamomum cassia. Cinnamic aldehyde is the active compound in Cinnamomum cassia



**Fig. 8.** Effects of DHJST and TH on MIA-induced paw edema swelling on days 1 and 3 (A). hind-limb weight-bearing on day 5 (B); hot-plate latent pain responses on day 10 (C). \*p < 0.05, compared to the control. n = 3.

and is suitable to be used as a quality control marker of DHJST. In the future, incorporation of the TCM network pharmacological approach with empirical evidence from traditional clinical experience and mechanistic studies will enhance the modern use of DHJST or the simplified version TH as effective anti-arthritic agents. Further toxicity and clinical studies are also warranted to establish more detailed data on the crude extracts and pure compounds. These results also can be used to guide future research into the effectiveness of DHJST with more convenient preparations in treating human arthritis with inflammatory symptoms.

#### Declaration of competing interest

The author reports no conflicts of interest in this report.

#### References

- 1. Zhang W, Wang S, Zhang R, et al. Evidence of Chinese herbal medicine Duhuo Jisheng decoction for knee osteoarthritis: a systematic review of randomized clinical trials. BMJ Open. 2016;6(1), e008973.
- 2. Chen FP, Chang CM, Hwang SJ, Chen YC, Chen FJ. Chinese herbal prescriptions for osteoarthritis in Taiwan: analysis of National Health Insurance dataset. BMC Compl Alternative Med. 2014;14:91.
- Smolen JS, Steiner G. Therapeutic strategies for rheumatoid arthritis. *Nat Rev Drug Discov*. 2003;2(6):473–488.
- 4. Srivastava A. Inflammation is key to hemophilic arthropathy. Blood 126(19): 2175-2176..
- 5. Park JU, Kim SJ, Na CS, et al. Chondroprotective and anti-inflammatory effects of ChondroT, a new complex herbal medication. BMC Compl Alternative Med. 2016;16:213.
- 6. Schwager J, Richard N, Fowler A, Seifert N, Raederstorff D. Carnosol and related substances modulate chemokine and cytokine production in macrophages and chondrocytes. Molecules. 2016;21(4):465.
- 7. Nirmal P, Koppikar S, Bhondave P, et al. Influence of six medicinal herbs on collagenase-induced osteoarthritis in rats. Am J Chin Med. 2013;41(6): 1407-1425.
- 8. Chen B, Zhan H, Marszalek I, et al. Traditional Chinese medications for knee osteoarthritis pain: a meta-analysis of randomized controlled trials. Am J Chin Med. 2016:44(4):677-703.
- 9 Li L Liu H Shi W et al Insights into the action mechanisms of traditional Chinese medicine in osteoarthritis. Evid Based Complement Alternat Med. 2017, 5190986.
- 10. Chen CW, Sun J, Li YM, Shen PA, Chen YO, Action mechanisms of du-huo-iisheng-tang on cartilage degradation in a rabbit model of osteoarthritis. Evid Based Complement Alternat Med 2011 571479
- 11. Chen Y, Li J, Li Q, et al. Du-Huo-Ji-Sheng-Tang attenuates inflammation of TNF-Tg mice related to promoting lymphatic drainage function. Evid Based Complement Alternat Med. 2016, 7067691.
- 12. Park E, Kum S, Wang C, Park SY, Kim BS, Schuller-Levis G. Anti-inflammatory activity of herbal medicines: inhibition of nitric oxide production and tumor necrosis factor-alpha secretion in an activated macrophage-like cell line. Am J Chin Med. 2005:33(3):415-424.
- 13. Dai Y, But PP, Chan YP, Matsuda H, Kubo M. Antipruritic and antiinflammatory effects of aqueous extract from Si-Wu-Tang. Biol Pharm Bull. 2002:25(9): 1175-1178
- 14. Chen LG, Jan YS, Tsai PW, et al. Anti-inflammatory and antinociceptive

constituents of Atractvlodes japonica Koidzumi, I Agric Food Chem, 2016:64(11): 2254-2262.

- 15. Chien TY, Huang SK, Lee CJ, Tsai PW, Wang CC. Antinociceptive and antiinflammatory effects of zerumbone against mono-iodoacetate-induced arthritis. Int J Mol Sci. 2016;17(2):249.
- 16. Rao PV, Gan SH. Cinnamon: a multifaceted medicinal plant. Evid Based Complement Alternat Med. 2014, 642942.
- 17. Muhammad JS, Zaidi SF, Shaharyar S, et al. Anti-inflammatory effect of cinnamaldehyde in Helicobacter pylori induced gastric inflammation. Biol Pharm Bull. 2015;38(1):109-115.
- 18. Yang D, Liang XC, Shi Y, et al. Anti-oxidative and anti-inflammatory effects of cinnamaldehyde on protecting high glucose-induced damage in cultured dorsal root ganglion neurons of rats. Chin J Integr Med. 2016;22(1):19-27.
- 19. Cho H. Walker A. Williams I. Hasty KA. Study of osteoarthritis treatment with anti-inflammatory drugs: cyclooxygenase-2 inhibitor and steroids. BioMed Res Int. 2015. 595273.
- 20. Wang JY, Yuan Y, Chen XJ, et al. Extract from Eucommia ulmoides Oliv. ameliorates arthritis via regulation of inflammation, synoviocyte proliferation and osteoclastogenesis in vitro and in vivo. J Ethnopharmacol. 2016;194:609-616.
- 21. Wang KT, Chen LG, Tseng SH, Huang JS, Hsieh MS, Wang CC. Anti-inflammatory effects of resveratrol and oligostilbenes from Vitis thunbergii var. taiwaniana against lipopolysaccharide-induced arthritis. J Agric Food Chem. 2011;59(8): 3649-3656.
- 22. Lee JH, Lim H, Shehzad O, Kim YS, Kim HP. Ginsenosides from Korean red ginseng inhibit matrix metalloproteinase-13 expression in articular chondrocytes and prevent cartilage degradation. Eur J Pharmacol. 2014;724: 145-151
- 23. Lee JH, Shehzad O, Ko SK, Kim YS, Kim HP. Matrix metalloproteinase-13 downregulation and potential cartilage protective action of the Korean Red Ginseng preparation. J Ginseng Res. 2015;39(1):54–60. 24. Miller RE, Miller RJ, Malfait AM. Osteoarthritis joint pain: the cytokine
- connection. Cytokine. 2014;70(2):185-193.
- 25. Chen YF, Tsai HY, Wu TS. Anti-inflammatory and analgesic activities from roots of Angelica pubescens. Planta Med. 1995;61(1):2-8.
- 26. Li X, Wang J, Gao L. Anti-inflammatory and analgesic activity of R.A.P. (Radix Angelicae Pubescentis) ethanol extracts. Afr J Tradit, Complementary Altern Med. 2013;10(3):422-426.
- 27. Yang YF, Xu W, Song W, Ye M, Yang XW. Transport of twelve coumarins from Angelicae Pubescentis radix across a MDCK-pHaMDR cell monolayer-an in vitro model for blood-brain barrier permeability. Molecules. 2015;20(7): 11719-11732
- 28. Ding XF, Feng X, Dong YF, et al. Studies on chemical constituents of the roots of Angelica pubescens. Zhong Yao Cai. 2008;31(4):516-518.
- 29. Jeong HJ, Na HJ, Kim SJ, et al. Anti-inflammatory effect of columbianetin on activated human mast cells. Biol Pharm Bull. 2009;32(6):1027-1031.
- 30. Chang YX, Zhang QH, Li J, et al. Simultaneous determination of scopoletin, psoralen, bergapten, xanthotoxin, columbianetin acetate, imperatorin, osthole and isoimperatorin in rat plasma by LC-MS/MS for pharmacokinetic studies following oral administration of Radix Angelicae Pubescentis extract. J Pharm Biomed Anal. 2013;77:71-75.