Submit a Manuscript: https://www.f6publishing.com

World J Transplant 2022 March 18; 12(3): 27-41

DOI: 10.5500/wjt.v12.i3.27 ISSN 2220-3230 (online)

REVIEW

Innovative immunosuppression in kidney transplantation: A challenge for unmet needs

Maurizio Salvadori, Aris Tsalouchos

Specialty type: Transplantation

Provenance and peer review:

Invited article; Externally peer reviewed.

Peer-review model: Single blind

Peer-review report's scientific quality classification

Grade A (Excellent): 0 Grade B (Very good): B, B Grade C (Good): 0 Grade D (Fair): 0 Grade E (Poor): 0

P-Reviewer: Kaewput W, Thailand; Krishnan N, United Kingdom

Received: October 22, 2021 Peer-review started: October 22,

First decision: January 22, 2022 Revised: January 27, 2022 Accepted: March 6, 2022 Article in press: March 6, 2022 Published online: March 18, 2022



Maurizio Salvadori, Department of Renal Transplantation, Careggi University Hospital, Florence 50139, Italy

Aris Tsalouchos, Division of Nephrology, Santa Maria Annunziata Hospital, Florence 50012,

Corresponding author: Maurizio Salvadori, MD, Professor, Department of Renal Transplantation, Careggi University Hospital, Largo Palagi 1, Florence 50139, Italy. maurizio.salvadori1@gmail.com

Abstract

Due to the optimal results obtained in kidney transplantation and to the lack of interest of the industries, new innovative drugs in kidney transplantation are difficult to be encountered. The best strategy to find the new drugs recently developed or under development is to search in the sections of kidney transplantation still not completely covered by the drugs on the market. These unmet needs are the prevention of delayed graft function (DGF), the protection of the graft over the long time and the desensitization of preformed anti human leukocyte antigen antibodies and the treatment of the acute antibody-mediated rejection. These needs are particularly relevant due to the expansion of some kind of kidney transplantation as transplantation from non-heart beating donor and in the case of antibody-incompatible grafts. The first are particularly exposed to DGF, the latter need a safe desensitization and a safe treatments of the antibody mediated rejections that often occur. Particular caution is needed in treating these drugs. First, they are described in very recent studies and the follow-up of their effect is of course rather short. Second, some of these drugs are still in an early phase of study, even if in well-conducted randomized controlled trials. Particular caution and a careful check need to be used in trials launched 2 or 3 years ago. Indeed, is always necessary to verify whether the study is still going on or whether and why the study itself was abandoned.

Key Words: New drugs; Unmet needs in kidney transplantation; Delayed graft function; Long-term outcomes; Kidney inflammation; Anti-human leukocyte antigen antibodies

©The Author(s) 2022. Published by Baishideng Publishing Group Inc. All rights reserved.

Core Tip: Finding new innovative drugs for kidney transplantation is not easy but looking for unmet needs it is possible to find new interesting drugs and opportunities to use in kidney transplantation. Many of these drugs are just at the beginning of their process toward the approval and should be careful checked until the finish of their path. Principal unmet needs are treatment and prevention of delayed graft function, improve the long-term outcomes, desensitization and treatment of acute antibody-mediated rejection. Finding new drugs in these fields results extremely important to face new kind of transplantation as transplant from non-heart beating donor and transplant in ABO incompatibles pairs.

Citation: Salvadori M, Tsalouchos A. Innovative immunosuppression in kidney transplantation: A challenge for unmet needs. World J Transplant 2022; 12(3): 27-41

URL: https://www.wjgnet.com/2220-3230/full/v12/i3/27.htm

DOI: https://dx.doi.org/10.5500/wjt.v12.i3.27

INTRODUCTION

Little progress has been made over the past decade in the development of new therapeutic measures in clinical kidney transplantation, chiefly because of a lack of interest by industries and providers and because most centers have reached optimal outcomes with the drugs used today[1]. However, a strategy may be adopted to identify new immunosuppressant drugs in kidney transplantation.

New immunosuppressant drugs may be found looking for identified unmet therapeutic needs.

These new drugs may also be adopted as new immunosuppressive treatments or new strategies for special kidney transplantation scenarios such as ABO incompatibility, non-heart-beating donor (NHBD) transplantation and transplantation from high-risk donors.

Drugs for unmet therapeutic needs

These drugs may be categorized as follows: (1) Therapy for ischemia-reperfusion injury (IRI) that results in delayed graft function (DGF); (2) Therapy to preserve optimal kidney function over the long-term; and (3) Therapy for desensitization and antibody-mediated rejection (ABMR).

THERAPY FOR DGF

DGF refers to acute kidney injury (AKI) occurring in the first week of transplantation that cannot not be ascribed to acute rejection[2].

DGF is associated with increased immune activation, complement activation and release of damageassociated molecular patterns, such as hypomethylated DNA, hyaluronic acid, heparin sulfate, fibrinogen and heat shock proteins. Consequently, nuclear factor κB is activated and induces inflammatory cytokines such as interleukin (IL)-1, IL-6, tumoral necrosis factor alpha and interferon beta[3].

Due to this complex mechanism, although several drugs to treat DGF have been tried, many of them failed to prove their effectiveness. Indeed, DGF has also been called the graveyard of drugs for transplantation.

However, new drugs have recently emerged and they are still in randomized controlled trials (RCTs) to control DGF.

Anti-apoptotic strategies

Apoptosis plays an important role in shaping DGF. Indeed, the pro-apoptotic gene p53 is activated by hypoxia and induces cell cycle arrest and apoptosis[4].

QPI-1002 also known as 15 NP, is a short interfering RNA that inhibits the expression of p53. The results of a phase I/II clinical trial in kidney transplant recipients demonstrated beneficial effects on IRI/DGF in humans[5]. Additionally, two studies reported good results in mice[6,7]. However, the RCT was terminated in 2018 without positive results because of a lack of documented efficacy.

Pegylated carboxyhemoglobin

Carbon monoxide (CO) is involved in regulating endothelial cell survival and proliferation. It also plays roles in protecting against DGF through IRI, vessel relaxation and inhibition of proinflammatory responses[8-10]. The infusion of pegylated carboxyhemoglobin delivers CO to organs. CO is a very powerful anti-apoptotic substance and has anti-inflammatory effects. In animal studies, CO is extremely effective in both cold and warm ischemia.

The use of pegylated carboxyhemoglobin is currently the object of a phase 2/3 study to analyze the efficacy and safety of SANGUINATE for reducing the DGF rate in patients receiving a kidney transplant [11,12]. In a recent study by Thuillier et al[13], 3 oxygen transporters, HBOC-201, BbV and M101, were tested in organ preservation[13-15].

Relaxin

In DGF, relaxin (RLX) has an anti-inflammatory effect by reducing the expression of intracellular adhesion molecule 1, inducing the expression of Notch 1 in macrophages and reducing neutrophil adhesion through increased synthesis of nitric oxide [16-18]. Additionally, RLX causes vasodilatation through increased NO production and inhibition of endothelin 1 production[19]. Two studies[18,20] documented improved renal function, histologic improvement in damaged tissue after DGF, and a reduced number of apoptotic cells.

Hepatocyte growth factor

ANG-3777, formerly BB3, is a hepatocyte growth factor mimetic that binds to its transmembrane tyrosine kinase receptor, cMET[21]. In preclinical studies, ANG-3777 was renoprotective in a variety of animal models of AKI, exerting anti-inflammatory and regenerative effects and preventing tubular cell apoptosis, epithelial to mesenchymal transition and fibrosis[22,23]. In a randomized, placebo-controlled phase 2 trial on oliguric patients after kidney transplantation, patients treated with ANG-3777 had a larger increase in urine output, a greater reduction in C reactive protein and neutrophil gelatinaseassociated lipocalin and a higher estimated glomerular filtration rate (eGFR)[24]. More recently, Vincenti et al[25] started the Graft Improvement Following Transplant (GIFT) trial, which is a phase 3 trial on the hepatocyte growth factor mimetic ANG-3777 in kidney transplant recipients with DGF. The aim of GIFT is to generate data to advance the treatment of DGF. In addition, the authors stress that a significant factor is that ANG-3777 may also be effective when administered after AKI-related DGF.

Complement inhibition

Complement activation plays a significant role in IRI, which causes and precedes DGF. The most studied among the complement inhibitor drugs to minimize DGF has been Mirocept (APT 070), which inhibits C3/C5 convertases and C1 esterase inhibitors.

Mirocept, still in a phase 1 trial (ISRCTN49958194)[26], is a potent membrane-localizing complement inhibitor and may be administered ex vivo to the donor kidney prior to transplantation. However, a recent dose finding study in animals[27] documented that a high dose of Mirocept might be needed to achieve adequate complement inhibition. More promising results have been obtained with C1 esterase inhibition.

This drug may also be administered as a donor pretreatment strategy in high-risk recipients (NCT02435732)[28], but the trial results are still unknown. Better results have been obtained by administering C1 esterase inhibitors to recipients of kidneys from high-risk donors or in the case of donation after circulatory death (DCD)[29-31]. A recent study from Huang et al[32] studied the three-year outcomes of patients treated with C1 esterase inhibitors to avoid DGF in a randomized controlled study. The study found that the treatment was associated with a lower incidence of graft failure.

Table 1 summarizes representative drugs in the categories described above used to prevent DGF and their targets.

Improving perfusion techniques

Improving perfusion techniques is not drugs in the sense of the word but rather a different strategy to prevent IRI and DGF by improving kidney perfusion at the time of kidney transplantation.

In a recently published study, Urbanellis et al[33] documented that continuous normothermic ex vivo kidney perfusion significantly improved early kidney function compared with hypothermic anoxic machine perfusion and static cold storage (SCS) in a porcine kidney auto-transplantation model.

A more interesting study was performed by Niemann et al[34]. The authors documented that reducing the body temperature by 2 °C of the deceased donor achieved a significant reduction in DGF rates and that the effect was more significant in the extended criteria donors.

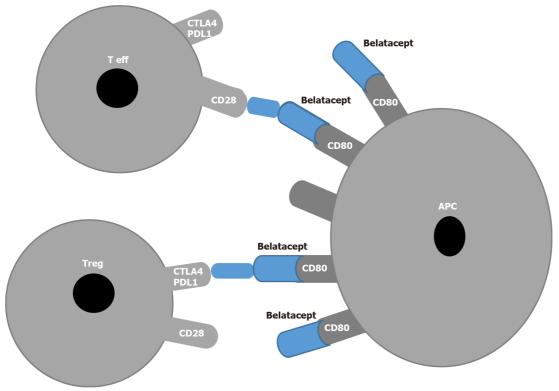
Finally, in a recent review[35], it was documented that active oxygenation during hypothermic machine perfusion is the most beneficial in cases involving the use of DCD kidneys when applied starting from kidney procurement until transplantation. Active oxygenation improves preservation and subsequent early graft function.

THERAPY TO PRESERVE RENAL FUNCTION

These drugs may be divided into the following categories: (1) Therapy to avoid nephrotoxicity, usually by elimination of calcineurin inhibitors (CNIs); (2) Therapy to control inflammation and fibrosis (principally when inflammation overlaps fibrosis); and (3) Therapy to prevent donor-specific antibodies (DSAs) and treat chronic ABMR (cABMR).

Table 1 Therapies targeting delayed graft function in kidney transplantation		
Drug	Molecular target	Mechanism of action
15NP or QPI-1002	p53	Inhibition of apoptosis
Pegylated carboxyhemo- globin	Cytochrome C oxidase; cytochrome P450; HMGB-1; P38 MAPK pathway	Inhibition of oxidative injury, inflammation, and apoptosis
Relaxin	ICAM-1; neutrophil adhesion	Vasodilatation; inhibition of apoptosis
ANG-3777 (BB3)	Tyrosine kinase receptor cMET	Antinflammation; inhibition of epithelial to mesenchymal transition
Mirocept (APT 070)	Inhibition of C3/C5 convertase	Inhibition of complement activation
C1 esterase inhibitor	C1 esterase	Inhibition of complement activation

HMGB-1: High mobility group protein box-1; MAPK: Mitogen-activated protein kinases; ICAM 1: Intercellular adhesion molecule 1.



DOI: 10.5500/wjt.v12.i3.27 Copyright © The Author(s) 2022.

Figure 1 Block of co-stimulation with Belatacept. APC: Antigen presenting cell; T effector; T reg: Regulatory T cells; PDL1: Programmed cell death receptor ligand 1; CTLA4: Cytotoxic T-lymphocyte-associated antigen 4.

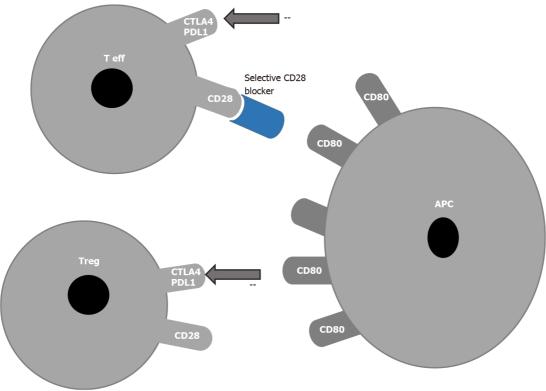
Therapy to avoid nephrotoxicity induced by CNIs

Until recently and even today, the two main strategies for a CNI-free regimen have been as follows: Mammalian target of rapamycin inhibitor-based immunosuppression; belatacept based immunosup-

Several studies have documented the efficacy of everolimus therapy in conjunction with low-dose CNIs[36-39]. The study by Pascual et al[36] "the Advancing renal TRANSplant eFficacy and safety Outcomes with eveRoliMus based regimen (TRANSFORM)" was a randomized open label, two-arm study with 2037 de novo kidney transplant recipients recruited in 186 centers worldwide. Everolimus efficacy was demonstrated, but the administration of low-dose tacrolimus (TAC) was needed.

The complete withdrawal of CNIs is difficult to achieve and is only appropriate for low-risk patients and donors and for living donors, and in the absence of DSAs[40].

The use of belatacept or other agents blocking the costimulatory pathways is the other method to avoid CNIs.



DOI: 10.5500/wjt.v12.i3.27 **Copyright** © The Author(s) 2022.

Figure 2 Block of co-stimulation with anti CD28. APC: Antigen presenting cell; T eff: T effector; T reg: Regulatory T cells; PDL1: Programmed cell death receptor ligand 1; CTLA4: Cytotoxic T-lymphocyte-associated antigen 4.

The blockade of CD28/cytotoxic T-lymphocyte-associated antigen 4 (CTLA-4) on T effector lymphocytes and CD80/CD86 on antigen presenting cells (APCs) was the first pathway to be targeted in the trials BENEFIT and BENEFIT-EXT[41,42]. Independent of well-preserved kidney function, the use of belatacept in a subset of patients was associated with an increased number of severe rejections[43, 44] and an increased number of opportunistic infections[45], including cytomegalovirus[46]. In addition a correlation between the incidence of post-lymphoproliferative disease and Epstein-Barr virus seronegative patients in the belatacept group was found[47].

These drawbacks are related to the fact that belatacept, which binds to CD80 and CD86 on APCs, blocks not only the T effectors that represent the positive signal but also the regulatory T (Tregs) that constitute the inhibitory signal (Figure 1).

In 2015, a report showed that the blockade of CD28 on effector T cells without inhibition of Treg cells prolonged survival in a nonhuman primate kidney transplant model. In this way, effector cells can be inhibited without inhibiting Tregs because selective CD28 blockade allows inhibitory signals *via* CTLA-4 and programmed cell death ligand-1 to remain intact while blocking T cell activation by CD28[48] (Figure 2).

Selective targeting of the CD28 antigen on T cells might be a more effective immunosuppressive therapy than belatacept, since this blockade leaves the inhibitory signal of CTLA-4 intact and may preserve Treg functions[49-51].

Currently, two monovalent antibodies, FR104 and lulizumab-pegol are under development for clinical application. These antibodies have antagonistic activity against CD28 alone[52,53]. To date, an RCT has been conducted at the University of California to modulate Tregs with combinatorial treatment with CD28 and IL-6 receptor antagonists[54] (Figure 3). The addition of an IL-6 receptor antagonist (tocilizumab) aims to further stimulate Treg cells and exert an anti-inflammatory effect. In the CTOT24 trial, after induction with thymoglobulin, steroids are administered from the beginning, lulizumab is started at the beginning and then continued weekly through day 77, belatacept is started on day 84 and administered every 4 wk, tocilizumab is started at the beginning and continued every 2 wk through day 168, and everolimus is started on day 14 and administered twice daily.

A different way to block costimulation is to block the interaction between CD40 and CD40 L. A first attempt was made to block the CD 40 receptor, but the studies were interrupted because of a number of thromboembolic complications[55,56]. This was because CD40 L is also expressed on platelets, which causes thromboembolic complications.

In 2014, Okimura *et al*[57] reported that ASKP 1240, a fully human antibody targeting human CD40, had a potent immunosuppressive effect that did not interfere with platelets.

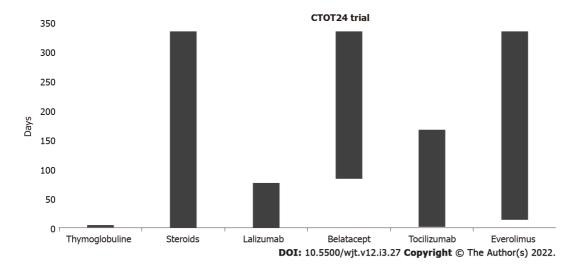


Figure 3 CTOT24 trial.

Recently, in a phase 1b study, the safety and efficacy of bleselumab, a fully human anti-CD40 monoclonal antibody, was documented by Vincenti et al [58]. The results were confirmed by a phase 2, randomized, open label, noninferiority study by Harland et al [59].

Novartis claimed to have developed another anti-CD40 monoclonal antibody (CFZ-533, Iscalimab). The antibody was characterized by several studies [60,61]. The antibody is the object of an RCT in de novo renal transplantation[62] to demonstrate comparable efficacy to and better renal function than TAC in *de novo* CNI-free kidney transplantation.

Until recently, it was believed that the main cause of kidney injury over time after transplantation was primarily due to CNI nephrotoxicity.

The first study questioning this opinion was the DeKAF study by Gaston et al [63] . The study documented that the decline in kidney function was not only due to CNI nephrotoxicity but also due primarily to the presence in the recipient of DSAs and the consequent activation of the humoral response[64]. Indeed, long-term graft survival was lower in patients with DSAs in the serum and C4d, a marker of immune response activation on the glomerular capillary wall. The role of DSAs and ABMR was further documented by Sellarés et al [65] and Lefaucheur et al [66]. A separate study documented that both *de novo* and pre-existing DSAs caused ABMR and reduced graft survival [67].

A more recent study by Stegall et al [68] examined 575 surveillance biopsies of kidney transplants from living donors on low-dose TAC therapy and found that 82% of patients whose grafts survived 10 years were affected by inflammatory lesions not related to CNI toxicity or to immunological mechanisms.

Preserving renal function requires other therapies in addition to safely reducing or withdrawing CNIs.

Therapy to control inflammation and fibrosis not related to immunological causes

Several factors, such as hyperuricemia, glucose intolerance, arterial hypertension, dyslipidemia and infection, may induce an inflammatory state in kidney transplant patients[69]. In addition, chronic hypoxia mediated by IL-1 and IL-6, angiotensin II and transforming growth factor beta may result in the accumulation of extracellular matrix, which can lead to interstitial fibrosis. In particular, several studies [70-72] document that IL-6 leads to allograft injury by acute inflammation, adaptive cellular/humoral responses, innate immunity and fibrosis. All these studies indicate that IL-6 is a mainstay in inducing inflammation and allograft injury.

Several drugs have been proposed to control the graft inflammatory state, including low-dose aspirin, statins, renin-angiotensin inhibitors, and xanthine-oxidase inhibitors, but no prospective trial with these drugs has been conducted in kidney transplantation. The only drug object of an RCT is the IL-6R inhibitor.

Currently, available agents for IL-6 signaling inhibition include monoclonal antibodies against IL-6 or IL-6R and Janus kinase inhibitors. The most often studied is tocilizumab, an IL-6R blocker. In a study conducted by Chandran et al [73], IL-6 blockade with tocilizumab increased Tregs and reduced T effector cytokines in renal graft inflammation. Tocilizumab-treated patients showed an improved tubulointerstitial Banff score and an increased Treg frequency.

Therapy to control chronic humoral rejection

Important advances have been made in the treatment of ABMR, but less effective treatments are available to control cABMR, which is a slowly progressing disease in which grafts are primarily injured

by de novo DSAs[74].

Until recently, attempts to treat cABMR had been limited to a combination of plasmapheresis and intravenous immunoglobulins (IVIGs)[75] and rituximab (RTX)[76,77]. Recently, proteasome inhibitors such as bortezomib[78] and carfilzomib[79] have also been studied, but these drugs were not as effective as anticipated.

In addition, complement inhibitors such as C1 inhibitors (C1-INH) and eculizumab, failed to control cABMR[80,81] probably because antibodies may injure the endothelium in a complement-independent pathway. Better results have been obtained with the use of IL-6R or IL-6 inhibitors.

In a previous study, Shin *et al*[82] documented the efficacy of tocilizumab in blocking monocyte activation in an *in vitro* model, to inhibit the inflammatory cascade induced by alloantibodies. In a more recent study, Shin *et al*[83] documented a beneficial effect of tocilizumab on cABMR owing to a reduction in antibody production by B cells.

Similarly, Choi *et al*[84] documented a reduction in DSAs and cABMR and stabilization of renal function in patients with cABMR, DSAs and transplant glomerulopathy treated with tocilizumab. A phase 4 RCT in patients with cABMR was recently designed[85].

Clazakizumab is a humanized monoclonal antibody directed against IL-6. In a study by Dobere *et al* [86], clazakizumab reduced DSAs and demonstrated beneficial effects on cABMR and renal function.

THERAPY FOR DESENSITIZATION AND ACUTE ABMR

Desensitization and treatment of ABMR are the two faces of the same coin. It has already been discussed how DSAs play a relevant role in inducing AKI and graft failure. DSAs may already be present before transplantation, or they may appear *de novo* after kidney transplantation. In both conditions, they may cause ABMR.

Desensitization is the treatment to reduce or, when possible, completely eradicate DSAs before or at the time of transplantation. Treatment of ABMR includes powerful drugs aimed at controlling this severe complication.

To better understand the mechanism of action of these drugs, Figure 4 represents how DSAs are formed and where the immunosuppressant drugs may act[87]. Naïve CD4+ T cells recognize the antigen presented by APCs. Activated CD4+ cells process antigens, which are presented to naïve B cells. Costimulatory molecules mediate the presentation through CD80/86 and CD28. B cell maturation and development into B-memory cells and plasma cells (PCs) is regulated by cytokines (principally IL-6 and IL-21), B cell activating factor (BAFF) and a proliferation-inducing ligand that interact with B cell maturation antigen. PCs produce antibodies that bind to donor-specific human leukocyte antigen (HLA) molecules, activate complement and initiate injury leading to ABMR. Agents capable of interfering with this complex system are numerous and act at different levels.

Several studies and reviews have described the drugs used in desensitization and in the treatment of ABMR[88-93].

Novel agents will be discussed in this chapter. New agents acting on costimulatory signals have already been discussed [48,49,57,59]. Similarly, anti-IL-6/IL-6R agents have been discussed [83-86].

Obintuzumab is a type 2 anti-CD20 antibody that induces more robust B cell depletion than RTX. To date, the drug has been evaluated in a phase 1b study to induce desensitization[94].

Belimumab belongs to the anti BAFF family. The drug is effective in treating systemic lupus erythematosus[95] but less effective in treating ABMR[96] due to possible infective complications. Proteasome inhibitors such as bortezomib and carfilzomib act on PCs, but are not as effective as anticipated. Carfilzomib has been studied in desensitization in a nonhuman primate model[97].

Drugs acting directly on PCs target CD38. Several studies or case reports have documented the efficacy of daratuzumab in the treatment of ABMR[98-100]. Isatuximab is effective on PCs and other immune cells, such as Tregs and Bregs. This fact may limit its applicability in the treatment of ABMR [101].

Inebilizumab is a humanized anti-CD19 monoclonal antibody approved for neuromyelitis optica [102].

An RCT with inebilizumab for pretransplant desensitization[103] was suspended due to the coronavirus disease pandemic.

Finally, another fully human monoclonal antibody, anti-CD38, is the object of an RCT for the treatment of ABMR[104].

In ABMR, the activation of the complement cascade is triggered by ligation of the C1 complex to HLA antigens that are bound by DSAs. Several drugs are capable of blocking complement activation (Figure 5). The C1 complex is activated upon antibody binding. The humanized monoclonal antibody BIVV009 (sutinlimab) targets its enzymatic subcomponent C1 s and this therapy blocks C4 and C2 cleavage and the formation of C3 convertase.

A phase 1 study with this drug[105] was concluded, and Eskandary *et al*[80] studied 10 kidney transplant recipients with ABMR. Repeated biopsies documented a reduction in C4d deposition even if DSA levels and microvascular inflammation were unchanged.

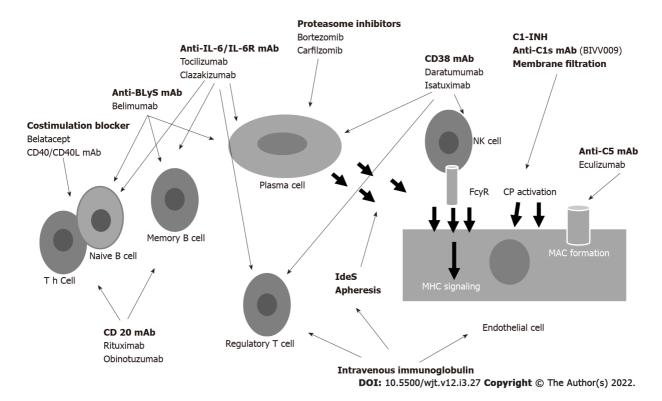


Figure 4 Drugs acting at different levels to control the antibody formation. BLyS: B Lymphocyte stimulating factor; mAb: Monoclonal antibody; C1-INH: C1 inhibitors; NK: Natural killer; Cp: Complement; FcyR: FcyReceptor; MAC: Membrane attacking complex; MHC: Major histocompatibility complex; IL: Interleukin.

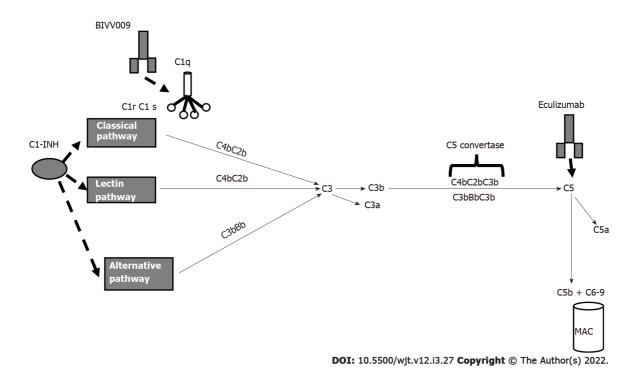


Figure 5 Principal drugs affecting complement. C1-INH: C1 inhibitor; MAC: Membrane attacking complex.

C1-INH regulates several pathways that contribute to complement activation and cause ABMR. In 2015, in a phase I/II placebo-controlled trial, Vo et al [106] reported the efficacy of C1-INH in the prevention of ABMR in HLA-sensitized patients. Later, Montgomery et al[107] in a randomized controlled pilot study, documented the efficacy of C1-INH in controlling ABMR. More recently, two more studies are ongoing to document the efficacy of human plasma C1 esterase inhibition as an addition to the standard of care for the treatment of ABMR[108,109].

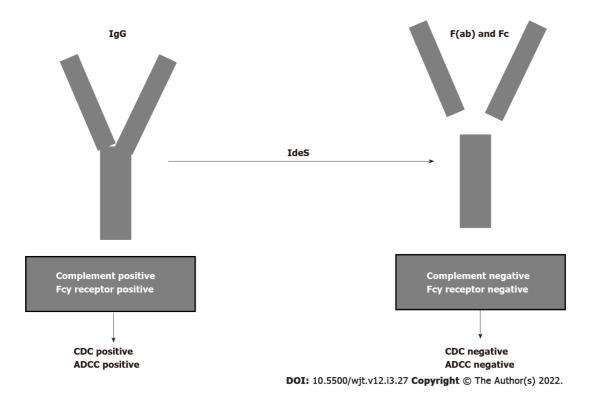


Figure 6 Cleaving intact immunoglobulin G by imlifidase. CDC: Complement dependent cytotoxicity; ADCC: Antibody dependent cell cytotoxicity; F(ab): Fragment ab; Fc: Fragment c; Ides: Imlifidase; IgG: Immunoglobulin G.

The humanized monoclonal antibody eculizumab binds to C5 with high affinity and prevents C5 convertase-mediated cleavage to C5a and C5b. In the past, several studies documented the efficacy of eculizumab in treating ABMR[110-112]. Recently, other studies documented the efficacy of eculizumab in treating and preventing ABMR[113,114]. Antibody removal is another therapeutic technique that may be applied primarily to desensitize patients with preformed DSAs before transplantation. Until recently, antibody removal and/or inhibition have been performed by plasmapheresis and IVIGs. Recently, it was documented that imlifidase (IdeS), a recombinant cysteine protease derived from Streptococcus pyogenes, rapidly cleaves IgG in the lower hinge region to a Fab fragment and a dimeric Fc fragment [115] (Figure 6). In addition to eliminating HLA antibodies, Ge et al [116] demonstrated that IdeS is a potent inhibitor of antibody-dependent cell cytotoxicity. A drawback of IdeS treatment is antibody recurrence after the interruption of the treatment. Incorporation of plasmapheresis and RTX to this treatment may overcome this drawback.

An international phase 2 trial was conducted in five transplant centers[117] for desensitization of cross-match-positive, highly sensitized kidney transplant recipients. Antibody rebound occurred 3-14 d after lipopolysaccharide administration, but graft survival at six months was 88.9%. The study conclusion was that IdeS converted positive cross matches to negative cross matches and achieved the transplantation of high-sensitized patients with optimal results at 6 mo.

In a more recent study, Kjellman et al [118] documented that lipifidase treatment administered to 39 cross-match-positive patients accomplished a 3-year graft survival of 93% with an ABMR incidence of 38% in the first month post-transplantation.

CONCLUSION

Lack of interest by industries and optimal outcomes reached by the drugs used to date has resulted in little progress in finding new drugs. However, examining unmet needs in the field of kidney transplantation may help us to find new drugs. Needs not optimally covered by current drugs are control of DGF, improvement of the long-term immunosuppression with graft outcomes reduced by chronic damage and the control of desensitization and ABMR. The control of these needs is of outmost importance, considering the expanding numbers of new kinds of kidney transplantation as transplantation from older donors and from NHBDs and transplantation from antibody-incompatible

In the first kind, controlling or reducing DGF is essential; in the latter kind, the reduction of antibodies against HLA is essential.

DGF may be controlled either with optimal management of the donor before or during kidney removal or with drugs attempting to target one of the multiple pathways involved in causing the IRI that is conducive to DGF.

New drugs are also emerging to control or reduce the antibody serum level. Several steps are involved in antibody generation and for each of those steps new drugs will be found.

In addition, drugs are able to reduce the nephrotoxicity induced by the long-term use of CNIs and to control kidney inflammation that may contribute to a worse graft outcome.

The majority of these drugs have been very recently found and are still in RCTs. Therefore, trials with novel agents require a careful approach and these new agents in transplantation face many challenges, but may provide a hopeful pipeline in this issue.

FOOTNOTES

Author contributions: Salvadori M and Tsalouchos A contributed equally to the manuscript; Salvadori M designed the study, performed the last revision and provided answers to the reviewers; Tsalouchos A collected the data from literature; Salvadori M and Tsalouchos A analyzed the collected data and wrote the manuscript.

Conflict-of-interest statement: Salvadori M and Tsalouchos A do not have any conflict of interest in relation to the manuscript, as in the attached form.

Open-Access: This article is an open-access article that was selected by an in-house editor and fully peer-reviewed by external reviewers. It is distributed in accordance with the Creative Commons Attribution NonCommercial (CC BY-NC 4.0) license, which permits others to distribute, remix, adapt, build upon this work non-commercially, and license their derivative works on different terms, provided the original work is properly cited and the use is noncommercial. See: https://creativecommons.org/Licenses/by-nc/4.0/

Country/Territory of origin: Italy

ORCID number: Maurizio Salvadori 0000-0003-1503-2681; Aris Tsalouchos 0000-0002-8565-4059.

S-Editor: Wang JJ L-Editor: A P-Editor: Wang JJ

REFERENCES

- Viklicky O, Novotny M, Hruba P. Future developments in kidney transplantation. Curr Opin Organ Transplant 2020; 25: 92-98 [PMID: 31833964 DOI: 10.1097/MOT.00000000000000722]
- Siedlecki A, Irish W, Brennan DC. Delayed graft function in the kidney transplant. Am J Transplant 2011; 11: 2279-2296 [PMID: 21929642 DOI: 10.1111/j.1600-6143.2011.03754.x]
- Rosin DL, Okusa MD. Dangers within: DAMP responses to damage and cell death in kidney disease. J Am Soc Nephrol 2011; **22**: 416-425 [PMID: 21335516 DOI: 10.1681/ASN.2010040430]
- 4 Moll UM, Zaika A. Nuclear and mitochondrial apoptotic pathways of p53. FEBS Lett 2001; 493: 65-69 [PMID: 11286997 DOI: 10.1016/s0014-5793(01)02284-0]
- Orlando G, Remuzzi G, Williams DF. Kidney Transplantation, Bioengineering and Regeneration. In: Chandran S, Vincenti F. Novel Drugs in Kidney Transplantation in Kidney Transplantation. New York: Elsevier, 2017: 277-290
- Tchervenkov J, Squiers E, Stratta R, Odenheimer D, Rothenstein D; QPI-1002 DGF Study Group. QPI-1002, a siRNA Targeting p53: Improvement in Outcomes Following Acute Kidney Injury (AKI): Cardiac Surgery to AKI Donors. Am J Transplant 2017: 17
- Schutt R, Case J, Barrick B, Toll A, Schaffer R, Fisher J, Marsh C. Living kidney transplant: the influence of intraoperative hemodynamics on delayed graft function. 2017 American Transplant Congress; 2017 Apr 29-May 3; Chicago, USA [DOI: 10.13140/RG.2.2.35308.97920]
- Ruan Y, Wang L, Zhao Y, Yao Y, Chen S, Li J, Guo H, Ming C, Gong F, Chen G. Carbon monoxide potently prevents ischemia-induced high-mobility group box 1 translocation and release and protects against lethal renal ischemiareperfusion injury. Kidney Int 2014; 86: 525-537 [PMID: 24694987 DOI: 10.1038/ki.2014.80]
- 9 Kohmoto J, Nakao A, Stolz DB, Kaizu T, Tsung A, Ikeda A, Shimizu H, Takahashi T, Tomiyama K, Sugimoto R, Choi AM, Billiar TR, Murase N, McCurry KR. Carbon monoxide protects rat lung transplants from ischemia-reperfusion injury via a mechanism involving p38 MAPK pathway. Am J Transplant 2007; 7: 2279-2290 [PMID: 17711551 DOI: 10.1111/j.1600-6143.2007.01940.x]
- 10 Hanto DW, Maki T, Yoon MH, Csizmadia E, Chin BY, Gallo D, Konduru B, Kuramitsu K, Smith NR, Berssenbrugge A, Attanasio C, Thomas M, Wegiel B, Otterbein LE. Intraoperative administration of inhaled carbon monoxide reduces delayed graft function in kidney allografts in Swine. Am J Transplant 2010; 10: 2421-2430 [PMID: 20977633 DOI: 10.1111/j.1600-6143.2010.03289.x
- Abuchowski A. SANGUINATE (PEGylated Carboxyhemoglobin Bovine): Mechanism of Action and Clinical Update.

- Artif Organs 2017; 41: 346-350 [PMID: 28397407 DOI: 10.1111/aor.12934]
- 12 Pharmaceuticals P. Efficacy and Safety of SANGUINATETM for Reduction of Delayed Graft Function in Patients Receiving a Kidney Transplant. [accessed 2015 Jul 3]. In: ClinicalTrials.gov [Internet]. Bethesda (MD): U.S. National Library of Medicine. Available from: https://clinicaltrials.gov/ct2/show/NCT02490202 ClinicalTrials.gov Identifier:
- 13 Thuillier R, Delpy E, Matillon X, Kaminski J, Kasil A, Soussi D, Danion J, Sauvageon Y, Rod X, Donatini G, Barrou B, Badet L, Zal F, Hauet T. Preventing acute kidney injury during transplantation: the application of novel oxygen carriers. Expert Opin Investig Drugs 2019; 28: 643-657 [PMID: 31165652 DOI: 10.1080/13543784.2019.1628217]
- Steichen C, Giraud S, Bon D, Barrou B, Badet L, Salamé E, Kerforne T, Allain G, Roumy J, Jayle C, Hannaert P, Hauet T, Thuillier R. Barriers and Advances in Kidney Preservation. Biomed Res Int 2018; 2018: 9206257 [PMID: 30643824 DOI: 10.1155/2018/9206257]
- Kaminski J, Hannaert P, Kasil A, Thuillier R, Leize E, Delpy E, Steichen C, Goujon JM, Zal F, Hauet T. Efficacy of the natural oxygen transporter HEMO2 life® in cold preservation in a preclinical porcine model of donation after cardiac death. Transpl Int 2019; 32: 985-996 [PMID: 30924562 DOI: 10.1111/tri.13434]
- Jakubauskiene L, Jakubauskas M, Leber B, Strupas K, Stiegler P, Schemmer P. Relaxin Positively Influences Ischemia-Reperfusion Injury in Solid Organ Transplantation: A Comprehensive Review. Int J Mol Sci 2020; 21 [PMID: 31963613 DOI: 10.3390/ijms21020631]
- Kageyama S, Nakamura K, Ke B, Busuttil RW, Kupiec-Weglinski JW. Serelaxin induces Notch1 signaling and alleviates hepatocellular damage in orthotopic liver transplantation. Am J Transplant 2018; 18: 1755-1763 [PMID: 29464890 DOI: 10.1111/ajt.14706
- Alexiou K, Wilbring M, Matschke K, Dschietzig T. Relaxin protects rat lungs from ischemia-reperfusion injury via inducible NO synthase: role of ERK-1/2, PI3K, and forkhead transcription factor FKHRL1. PLoS One 2013; 8: e75592 [PMID: 24098703 DOI: 10.1371/journal.pone.0075592]
- Yoshida T, Kumagai H, Kohsaka T, Ikegaya N. Relaxin protects against renal ischemia-reperfusion injury. Am J Physiol Renal Physiol 2013; 305: F1169-F1176 [PMID: 23946288 DOI: 10.1152/ajprenal.00654.2012]
- Collino M, Rogazzo M, Pini A, Benetti E, Rosa AC, Chiazza F, Fantozzi R, Bani D, Masini E. Acute treatment with relaxin protects the kidney against ischaemia/reperfusion injury. J Cell Mol Med 2013; 17: 1494-1505 [PMID: 24079335 DOI: 10.1111/jcmm.12120]
- Weidner KM, Sachs M, Birchmeier W. The Met receptor tyrosine kinase transduces motility, proliferation, and morphogenic signals of scatter factor/hepatocyte growth factor in epithelial cells. J Cell Biol 1993; 121: 145-154 [PMID: 8384622 DOI: 10.1083/jcb.121.1.145]
- Benoit SW, Devarajan P. Acute kidney injury: emerging pharmacotherapies in current clinical trials. Pediatr Nephrol 2018; **33**: 779-787 [PMID: 28601936 DOI: 10.1007/s00467-017-3695-3]
- Zhou D, Tan RJ, Lin L, Zhou L, Liu Y. Activation of hepatocyte growth factor receptor, c-met, in renal tubules is required 23 for renoprotection after acute kidney injury. Kidney Int 2013; 84: 509-520 [PMID: 23715119 DOI: 10.1038/ki.2013.102]
- Bromberg JS, Weir MR, Gaber AO, Yamin MA, Goldberg ID, Mayne TJ, Cal W, Cooper M. Renal Function Improvement Following ANG-3777 Treatment in Patients at High Risk for Delayed Graft Function After Kidney Transplantation. Transplantation 2021; 105: 443-450 [PMID: 32265417 DOI: 10.1097/TP.00000000000003255]
- Vincenti F, Kim J, Gouveia D, Pelle G, Mayne TJ, Neylan JF. Phase 3 trial Design of the Hepatocyte Growth Factor Mimetic ANG-3777 in Renal Transplant Recipients With Delayed Graft Function. Kidney Int Rep 2021; 6: 296-303 [PMID: 33615054 DOI: 10.1016/j.ekir.2020.11.001]
- Kassimatis T, Qasem A, Douiri A, Ryan EG, Rebollo-Mesa I, Nichols LL, Greenlaw R, Olsburgh J, Smith RA, Sacks SH, Drage M. A double-blind randomised controlled investigation into the efficacy of Mirococept (APT070) for preventing ischaemia reperfusion injury in the kidney allograft (EMPIRIKAL): study protocol for a randomised controlled trial. Trials 2017; 18: 255 [PMID: 28587616 DOI: 10.1186/s13063-017-1972-x]
- 27 Kassimatis T, Greenlaw R, Hunter JP, Douiri A, Flach C, Rebollo-Mesa I, Nichols LL, Qasem A, Danzi G, Olsburgh J, Drage M, Friend PJ, Neri F, Karegli J, Horsfield C, Smith RA, Sacks SH. Ex vivo delivery of Mirococept: A dose-finding study in pig kidney after showing a low dose is insufficient to reduce delayed graft function in human kidney. Am J Transplant 2021; 21: 1012-1026 [PMID: 33225626 DOI: 10.1111/ajt.16265]
- 28 University of Winsconsin. Cinrize as a Donor Pre-treatment Strategy in Kidney Recipients of KDPI>60%. [accessed 2015 May 6]. In: ClinicalTrials.gov [Internet]. Bethesda (MD): U.S. National Library of Medicine. Available from: https://clinicaltrials.gov/ct2/show/NCT02435732 ClinicalTrials.gov Identifier: NCT02435732
- Jordan SC, Choi J, Aubert O, Haas M, Loupy A, Huang E, Peng A, Kim I, Louie S, Ammerman N, Najjar R, Puliyanda D, Vo A. A phase I/II, double-blind, placebo-controlled study assessing safety and efficacy of C1 esterase inhibitor for prevention of delayed graft function in deceased donor kidney transplant recipients. Am J Transplant 2018; 18: 2955-2964 [PMID: 29637714 DOI: 10.1111/ajt.14767]
- Berger M, Lefaucheur C, Jordan SC. Update on C1 Esterase Inhibitor in Human Solid Organ Transplantation. Transplantation 2019; 103: 1763-1775 [PMID: 30946220 DOI: 10.1097/TP.0000000000002717]
- Jordan S. C11NH Inhibitor Preoperative and Post Kidney Transplant to Prevent DGF & IRI (C11NHDGF). [accessed 2014 May 9]. In: ClinicalTrials.gov [Internet]. Bethesda (MD): U.S. National Library of Medicine. Available from: https://clinicaltrials.gov/ct2/show/NCT02134314 ClinicalTrials.gov Identifier: NCT02134314
- Huang E, Vo A, Choi J, Ammerman N, Lim K, Sethi S, Kim I, Kumar S, Najjar R, Peng A, Jordan SC. Three-Year Outcomes of a Randomized, Double-Blind, Placebo-Controlled Study Assessing Safety and Efficacy of C1 Esterase Inhibitor for Prevention of Delayed Graft Function in Deceased Donor Kidney Transplant Recipients. Clin J Am Soc Nephrol 2020; **15**: 109-116 [PMID: 31843975 DOI: 10.2215/CJN.04840419]
- Urbanellis P, Hamar M, Kaths JM, Kollmann D, Linares I, Mazilescu L, Ganesh S, Wiebe A, Yip PM, John R, Konvalinka A, Mucsi I, Ghanekar A, Bagli DJ, Robinson LA, Selzner M. Normothermic Ex Vivo Kidney Perfusion Improves Early DCD Graft Function Compared With Hypothermic Machine Perfusion and Static Cold Storage.

- Transplantation 2020; 104: 947-955 [PMID: 31815900 DOI: 10.1097/TP.00000000000000066]
- Niemann CU, Feiner J, Swain S, Bunting S, Friedman M, Crutchfield M, Broglio K, Hirose R, Roberts JP, Malinoski D. Therapeutic Hypothermia in Deceased Organ Donors and Kidney-Graft Function. N Engl J Med 2015; 373: 405-414 [PMID: 26222557 DOI: 10.1056/NEJMoa1501969]
- Ngo K, Castillo P, Laine RA, Sun Q. Effects of Menadione on Survival, Feeding, and Tunneling Activity of the Formosan Subterranean Termite. Insects 2021; 12 [PMID: 34940197 DOI: 10.3390/biomedicines9080993]
- 36 Pascual J, Berger SP, Witzke O, Tedesco H, Mulgaonkar S, Qazi Y, Chadban S, Oppenheimer F, Sommerer C, Oberbauer R, Watarai Y, Legendre C, Citterio F, Henry M, Srinivas TR, Luo WL, Marti A, Bernhardt P, Vincenti F; TRANSFORM Investigators. Everolimus with Reduced Calcineurin Inhibitor Exposure in Renal Transplantation. J Am Soc Nephrol 2018; 29: 1979-1991 [PMID: 29752413 DOI: 10.1681/ASN.2018010009]
- Aala A, Brennan DC. Transformation in Immunosuppression: Are We Ready for it? J Am Soc Nephrol 2018; 29: 1791-1792 [PMID: 29884733 DOI: 10.1681/ASN.2018050491]
- Zeng J, Zhong Q, Feng X, Li L, Feng S, Fan Y, Song T, Huang Z, Wang X, Lin T. Conversion From Calcineurin Inhibitors to Mammalian Target of Rapamycin Inhibitors in Kidney Transplant Recipients: A Systematic Review and Meta-Analysis of Randomized Controlled Trials. Front Immunol 2021; 12: 663602 [PMID: 34539621 DOI: 10.3389/fimmu.2021.663602]
- Montero N, Quero M, Melilli E, Pérez-Sáez MJ, Redondo-Pachón D, Bestard O, Crespo M, Cruzado JM, Pascual J. Mammalian Target of Rapamycin Inhibitors Combined With Calcineurin Inhibitors as Initial Immunosuppression in Renal Transplantation: A Meta-analysis. Transplantation 2019; 103: 2031-2056 [PMID: 31343574 DOI: 10.1097/TP.00000000000027691
- Wojciechowski D, Wiseman A. Long-Term Immunosuppression Management: Opportunities and Uncertainties. Clin J Am Soc Nephrol 2021; 16: 1264-1271 [PMID: 33853841 DOI: 10.2215/CJN.15040920]
- 41 Pestana JO, Grinyo JM, Vanrenterghem Y, Becker T, Campistol JM, Florman S, Garcia VD, Kamar N, Lang P, Manfro RC, Massari P, Rial MD, Schnitzler MA, Vitko S, Duan T, Block A, Harler MB, Durrbach A. Three-year outcomes from BENEFIT-EXT: a phase III study of belatacept vs cyclosporine in recipients of extended criteria donor kidneys. Am J Transplant 2012; 12: 630-639 [PMID: 22300431 DOI: 10.1111/j.1600-6143.2011.03914.x]
- Rostaing L, Vincenti F, Grinyó J, Rice KM, Bresnahan B, Steinberg S, Gang S, Gaite LE, Moal MC, Mondragón-Ramirez GA, Kothari J, Pupim L, Larsen CP. Long-term belatacept exposure maintains efficacy and safety at 5 years: results from the long-term extension of the BENEFIT study. Am J Transplant 2013; 13: 2875-2883 [PMID: 24047110 DOI: 10.1111/ait.124601
- Mathews DV, Dong Y, Higginbotham LB, Kim SC, Breeden CP, Stobert EA, Jenkins J, Tso JY, Larsen CP, Adams AB. CD122 signaling in CD8+ memory T cells drives costimulation-independent rejection. J Clin Invest 2018; 128: 4557-4572 [PMID: 30222140 DOI: 10.1172/JCI95914]
- Kumar D, Raynaud M, Chang J, Reeve J, Yakubu I, Kamal L, Levy M, Bhati C, Kimball P, King A, Massey D, Halloran P, Gupta G. Impact of Belatacept Conversion on Renal Function, Histology, and Gene Expression in Kidney Transplant Patients With Chronic Active Antibody-mediated Rejection. Transplantation 2021; 105: 660-667 [PMID: 32510913 DOI: 10.1097/TP.00000000000032781
- 45 Bertrand D, Terrec F, Etienne I, Chavarot N, Sberro R, Gatault P, Garrouste C, Bouvier N, Grall-Jezequel A, Jaureguy M, Caillard S, Thervet E, Colosio C, Golbin L, Rerolle JP, Thierry A, Sayegh J, Janbon B, Malvezzi P, Jouve T, Rostaing L, Noble J. Opportunistic Infections and Efficacy Following Conversion to Belatacept-Based Therapy after Kidney Transplantation: A French Multicenter Cohort. J Clin Med 2020; 9 [PMID: 33126667 DOI: 10.3390/jcm9113479]
- Chavarot N, Divard G, Scemla A, Amrouche L, Aubert O, Leruez-Ville M, Timsit MO, Tinel C, Zuber J, Legendre C, Anglicheau D, Sberro-Soussan R. Increased incidence and unusual presentations of CMV disease in kidney transplant recipients after conversion to belatacept. Am J Transplant 2021; 21: 2448-2458 [PMID: 33283406 DOI: 10.1111/ajt.16430]
- 47 Ville S, Poirier N, Branchereau J, Charpy V, Pengam S, Nerriere-Daguin V, Le Bas-Bernardet S, Coulon F, Mary C, Chenouard A, Hervouet J, Minault D, Nedellec S, Renaudin K, Vanhove B, Blancho G. Anti-CD28 Antibody and Belatacept Exert Differential Effects on Mechanisms of Renal Allograft Rejection. J Am Soc Nephrol 2016; 27: 3577-3588 [PMID: 27160407 DOI: 10.1681/ASN.2015070774]
- 48 van der Zwan M, Hesselink DA, van den Hoogen MWF, Baan CC. Costimulation Blockade in Kidney Transplant Recipients. Drugs 2020; 80: 33-46 [PMID: 31749062 DOI: 10.1007/s40265-019-01226-6]
- El Hennawy H, Safar O, Al Faifi AS, El Nazer W, Kamal A, Mahedy A, Zaitoun M, Fahmy AE. Belatacept rescue therapy of CNI-induced nephrotoxicity, meta-analysis. Transplant Rev (Orlando) 2021; 35: 100653 [PMID: 34597943 DOI: 10.1016/j.trre.2021.100653]
- Poirier N, Blancho G, Vanhove B. A more selective costimulatory blockade of the CD28-B7 pathway. Transpl Int 2011; **24**: 2-11 [PMID: 20964725 DOI: 10.1111/j.1432-2277.2010.01176.x]
- 51 Liu D, Krummey SM, Badell IR, Wagener M, Schneeweis LA, Stetsko DK, Suchard SJ, Nadler SG, Ford ML. 2B4 (CD244) induced by selective CD28 blockade functionally regulates allograft-specific CD8+ T cell responses. J Exp Med 2014; **211**: 297-311 [PMID: 24493803 DOI: 10.1084/jem.20130902]
- Shi R, Honczarenko M, Zhang S, Fleener C, Mora J, Lee SK, Wang R, Liu X, Shevell DE, Yang Z, Wang H, Murthy B. Pharmacokinetic, Pharmacodynamic, and Safety Profile of a Novel Anti-CD28 Domain Antibody Antagonist in Healthy Subjects. J Clin Pharmacol 2017; 57: 161-172 [PMID: 27402064 DOI: 10.1002/jcph.791]
- Poirier N, Blancho G, Hiance M, Mary C, Van Assche T, Lempoels J, Ramael S, Wang W, Thepenier V, Braudeau C, Salabert N, Josien R, Anderson I, Gourley I, Soulillou JP, Coquoz D, Vanhove B. First-in-Human Study in Healthy Subjects with FR104, a Pegylated Monoclonal Antibody Fragment Antagonist of CD28. J Immunol 2016; 197: 4593-4602 [PMID: 27849166 DOI: 10.4049/jimmunol.1601538]
- National Institute of Allergy and Infectious Diseases. Treg modulation with CD28 and IL-6 receptor antagonists. [accessed 2019 Aug 26]. In: ClinicalTrials.gov [Internet]. Bethesda (MD): U.S. National Library of Medicine. Available from: https://clinicaltrials.gov/ct2/show/NCT04066114 ClinicalTrials.gov Identifier: NCT04066114

- 55 Henn V, Slupsky JR, Gräfe M, Anagnostopoulos I, Förster R, Müller-Berghaus G, Kroczek RA. CD40 ligand on activated platelets triggers an inflammatory reaction of endothelial cells. Nature 1998; 391: 591-594 [PMID: 9468137 DOI: 10.1038/353931
- Kawai T, Andrews D, Colvin RB, Sachs DH, Cosimi AB. Thromboembolic complications after treatment with monoclonal antibody against CD40 ligand. Nat Med 2000; 6: 114 [PMID: 10655073 DOI: 10.1038/72162]
- Okimura K, Maeta K, Kobayashi N, Goto M, Kano N, Ishihara T, Ishikawa T, Tsumura H, Ueno A, Miyao Y, Sakuma S, 57 Kinugasa F, Takahashi N, Miura T. Characterization of ASKP1240, a fully human antibody targeting human CD40 with potent immunosuppressive effects. Am J Transplant 2014; 14: 1290-1299 [PMID: 24731050 DOI: 10.1111/ajt.12678]
- Vincenti F, Klintmalm G, Yang H, Ram Peddi V, Blahunka P, Conkle A, Santos V, Holman J. A randomized, phase 1b study of the pharmacokinetics, pharmacodynamics, safety, and tolerability of bleselumab, a fully human, anti-CD40 monoclonal antibody, in kidney transplantation. Am J Transplant 2020; 20: 172-180 [PMID: 31397943 DOI: 10.1111/ajt.15560]
- Harland RC, Klintmalm G, Jensik S, Yang H, Bromberg J, Holman J, Kumar MSA, Santos V, Larson TJ, Wang X. Efficacy and safety of bleselumab in kidney transplant recipients: A phase 2, randomized, open-label, noninferiority study. Am J Transplant 2020; 20: 159-171 [PMID: 31509331 DOI: 10.1111/ajt.15591]
- Ristov J, Espie P, Ulrich P, Sickert D, Flandre T, Dimitrova M, Müller-Ristig D, Weider D, Robert G, Schmutz P, Greutmann B, Cordoba-Castro F, Schneider MA, Warncke M, Kolbinger F, Cote S, Heusser C, Bruns C, Rush JS. Characterization of the in vitro and in vivo properties of CFZ533, a blocking and non-depleting anti-CD40 monoclonal antibody. Am J Transplant 2018; 18: 2895-2904 [PMID: 29665205 DOI: 10.1111/ajt.14872]
- Espié P, He Y, Koo P, Sickert D, Dupuy C, Chokoté E, Schuler R, Mergentaler H, Ristov J, Milojevic J, Verles A, Groenewegen A, Auger A, Avrameas A, Rotte M, Colin L, Tomek CS, Hernandez-Illas M, Rush JS, Gergely P. First-inhuman clinical trial to assess pharmacokinetics, pharmacodynamics, safety, and tolerability of iscalimab, an anti-CD40 monoclonal antibody. Am J Transplant 2020; 20: 463-473 [PMID: 31647605 DOI: 10.1111/ajt.15661]
- Novartis. CCFZ533X2201- PoC Study in de novo renal transplantation. [accessed 2014 Aug 15]. In: ClinicalTrials.gov [Internet]. Bethesda (MD): U.S. National Library of Medicine. Available from: https://clinicaltrials.gov/ct2/show/NCT02217410 ClinicalTrials.gov Identifier: NCT02217410
- Gaston RS, Cecka JM, Kasiske BL, Fieberg AM, Leduc R, Cosio FC, Gourishankar S, Grande J, Halloran P, Hunsicker L, Mannon R, Rush D, Matas AJ. Evidence for antibody-mediated injury as a major determinant of late kidney allograft failure. Transplantation 2010; 90: 68-74 [PMID: 20463643 DOI: 10.1097/TP.0b013e3181e065de]
- Ye HZ, Tran HK, Van Voorhis T. Accurate Electronic Excitation Energies in Full-Valence Active Space via Bootstrap Embedding. J Chem Theory Comput 2021; 17: 3335-3347 [PMID: 33957050 DOI: 10.1111/ctr.14337]
- Sellarés J, de Freitas DG, Mengel M, Reeve J, Einecke G, Sis B, Hidalgo LG, Famulski K, Matas A, Halloran PF. Understanding the causes of kidney transplant failure: the dominant role of antibody-mediated rejection and nonadherence. Am J Transplant 2012; 12: 388-399 [PMID: 22081892 DOI: 10.1111/j.1600-6143.2011.03840.x]
- Lefaucheur C, Suberbielle-Boissel C, Hill GS, Nochy D, Andrade J, Antoine C, Gautreau C, Charron D, Glotz D. Clinical relevance of preformed HLA donor-specific antibodies in kidney transplantation. Contrib Nephrol 2009; 162: 1-12 [PMID: 19001809 DOI: 10.1159/000170788]
- Sinha BK. Role of free radicals in etoposide (VP-16,213) action. Basic Life Sci 1988; 49: 765-768 [PMID: 2855002 DOI: 10.1681/ASN.20160707971
- Stegall MD, Cornell LD, Park WD, Smith BH, Cosio FG. Renal Allograft Histology at 10 Years After Transplantation in the Tacrolimus Era: Evidence of Pervasive Chronic Injury. Am J Transplant 2018; 18: 180-188 [PMID: 28710896 DOI: 10.1111/ajt.14431]
- Ponticelli C, Campise MR. The inflammatory state is a risk factor for cardiovascular disease and graft fibrosis in kidney transplantation. Kidney Int 2021; 100: 536-545 [PMID: 33932457 DOI: 10.1016/j.kint.2021.04.016]
- Miller CL, Madsen JC. IL-6 Directed Therapy in Transplantation. Curr Transplant Rep 2021; 1-14 [PMID: 34099967 DOI: 10.1007/s40472-021-00331-4]
- Jordan SC, Choi J, Kim I, Wu G, Toyoda M, Shin B, Vo A. Interleukin-6, A Cytokine Critical to Mediation of Inflammation, Autoimmunity and Allograft Rejection: Therapeutic Implications of IL-6 Receptor Blockade. Transplantation 2017; 101: 32-44 [PMID: 27547870 DOI: 10.1097/TP.0000000000001452]
- Jordan SC, Ammerman N, Choi J, Kumar S, Huang E, Toyoda M, Kim I, Wu G, Vo A. Interleukin-6: An Important Mediator of Allograft Injury. Transplantation 2020; 104: 2497-2506 [PMID: 32235253 DOI: 10.1097/TP.000000000003249]
- Chandran S, Leung J, Hu C, Laszik ZG, Tang Q, Vincenti FG. Interleukin-6 blockade with tocilizumab increases Tregs and reduces T effector cytokines in renal graft inflammation: A randomized controlled trial. Am J Transplant 2021; 21: 2543-2554 [PMID: 33331082 DOI: 10.1111/ajt.16459]
- 74 Kim MY, Brennan DC. Therapies for Chronic Allograft Rejection. Front Pharmacol 2021; 12: 651222 [PMID: 33935762 DOI: 10.3389/fphar.2021.651222]
- Redfield RR, Ellis TM, Zhong W, Scalea JR, Zens TJ, Mandelbrot D, Muth BL, Panzer S, Samaniego M, Kaufman DB, Astor BC, Djamali A. Current outcomes of chronic active antibody mediated rejection - A large single center retrospective review using the updated BANFF 2013 criteria. Hum Immunol 2016; 77: 346-352 [PMID: 26867813 DOI: 10.1016/i.humimm.2016.01.0181
- Chung BH, Kim Y, Jeong HS, Hong YA, Choi BS, Park CW, Choi YJ, Kim YS, Yang CW. Clinical outcome in patients with chronic antibody-mediated rejection treated with and without rituximab and intravenous immunoglobulin combination therapy. Transpl Immunol 2014; 31: 140-144 [PMID: 25179826 DOI: 10.1016/j.trim.2014.08.005]
- Piñeiro GJ, De Sousa-Amorim E, Solé M, Ríos J, Lozano M, Cofán F, Ventura-Aguiar P, Cucchiari D, Revuelta I, Cid J, Palou E, Campistol JM, Oppenheimer F, Rovira J, Diekmann F. Rituximab, plasma exchange and immunoglobulins: an ineffective treatment for chronic active antibody-mediated rejection. BMC Nephrol 2018; 19: 261 [PMID: 30309322 DOI: 10.1186/s12882-018-1057-4]
- Eskandary F, Regele H, Baumann L, Bond G, Kozakowski N, Wahrmann M, Hidalgo LG, Haslacher H, Kaltenecker CC,

- Aretin MB, Oberbauer R, Posch M, Staudenherz A, Handisurya A, Reeve J, Halloran PF, Böhmig GA. A Randomized Trial of Bortezomib in Late Antibody-Mediated Kidney Transplant Rejection. J Am Soc Nephrol 2018; 29: 591-605 [PMID: 29242250 DOI: 10.1681/ASN.2017070818]
- Tremblay S, Driscoll JJ, Rike-Shields A, Hildeman DA, Alloway RR, Girnita AL, Brailey PA, Woodle ES. A prospective, iterative, adaptive trial of carfilzomib-based desensitization. Am J Transplant 2020; 20: 411-421 [PMID: 31550069 DOI: 10.1111/ajt.15613]
- Eskandary F, Jilma B, Mühlbacher J, Wahrmann M, Regele H, Kozakowski N, Firbas C, Panicker S, Parry GC, Gilbert JC, Halloran PF, Böhmig GA. Anti-C1s monoclonal antibody BIVV009 in late antibody-mediated kidney allograft rejection-results from a first-in-patient phase 1 trial. Am J Transplant 2018; 18: 916-926 [PMID: 28980446 DOI:
- Schinstock CA, Bentall AJ, Smith BH, Cornell LD, Everly M, Gandhi MJ, Stegall MD. Long-term outcomes of eculizumab-treated positive crossmatch recipients: Allograft survival, histologic findings, and natural history of the donorspecific antibodies. Am J Transplant 2019; 19: 1671-1683 [PMID: 30412654 DOI: 10.1111/ajt.15175]
- Shin BH, Ge S, Mirocha J, Jordan SC, Toyoda M. Tocilizumab (Anti-IL-6R) Suppressed TNFα Production by Human Monocytes in an In Vitro Model of Anti-HLA Antibody-Induced Antibody-Dependent Cellular Cytotoxicity. Transplant Direct 2017; 3: e139 [PMID: 28361123 DOI: 10.1097/TXD.00000000000000653]
- Shin BH, Everly MJ, Zhang H, Choi J, Vo A, Zhang X, Huang E, Jordan SC, Toyoda M. Impact of Tocilizumab (Anti-IL-6R) Treatment on Immunoglobulins and Anti-HLA Antibodies in Kidney Transplant Patients With Chronic Antibodymediated Rejection. Transplantation 2020; 104: 856-863 [PMID: 31385933 DOI: 10.1097/TP.00000000000002895]
- Choi J, Aubert O, Vo A, Loupy A, Haas M, Puliyanda D, Kim I, Louie S, Kang A, Peng A, Kahwaji J, Reinsmoen N, Toyoda M, Jordan SC. Assessment of Tocilizumab (Anti-Interleukin-6 Receptor Monoclonal) as a Potential Treatment for Chronic Antibody-Mediated Rejection and Transplant Glomerulopathy in HLA-Sensitized Renal Allograft Recipients. Am J Transplant 2017; 17: 2381-2389 [PMID: 28199785 DOI: 10.1111/ajt.14228]
- 85 Baid-Agrawal S. Tocilizumab in chronic antibody-mediated rejection in kidney transplant recipients (INTERCEPT). [accessed 2020 Sep 24]. In: Clinical Trials.gov [Internet]. Bethesda (MD): U.S. National Library of Medicine. Available from: https://clinicaltrials.gov/ct2/show/NCT04561986 ClinicalTrials.gov Identifier: NCT04561986
- Doberer K, Duerr M, Halloran PF, Eskandary F, Budde K, Regele H, Reeve J, Borski A, Kozakowski N, Reindl-Schwaighofer R, Waiser J, Lachmann N, Schranz S, Firbas C, Mühlbacher J, Gelbenegger G, Perkmann T, Wahrmann M, Kainz A, Ristl R, Halleck F, Bond G, Chong E, Jilma B, Böhmig GA. A Randomized Clinical Trial of Anti-IL-6 Antibody Clazakizumab in Late Antibody-Mediated Kidney Transplant Rejection. J Am Soc Nephrol 2021; 32: 708-722 [PMID: 33443079 DOI: 10.1681/ASN.2020071106]
- Jordan SC, Ammerman N, Choi J, Huang E, Peng A, Sethi S, Najjar R, Kim I, Toyoda M, Kumar S, Lim K, Vo A. The role of novel therapeutic approaches for prevention of allosensitization and antibody-mediated rejection. Am J Transplant 2020; **20** Suppl 4: 42-56 [PMID: 32538536 DOI: 10.1111/ajt.15913]
- Schinstock CA, Mannon RB, Budde K, Chong AS, Haas M, Knechtle S, Lefaucheur C, Montgomery RA, Nickerson P, Tullius SG, Ahn C, Askar M, Crespo M, Chadban SJ, Feng S, Jordan SC, Man K, Mengel M, Morris RE, O'Doherty I, Ozdemir BH, Seron D, Tambur AR, Tanabe K, Taupin JL, O'Connell PJ. Recommended Treatment for Antibodymediated Rejection After Kidney Transplantation: The 2019 Expert Consensus From the Transplantion Society Working Group. Transplantation 2020; 104: 911-922 [PMID: 31895348 DOI: 10.1097/TP.000000000000003095]
- Nickerson PW. What have we learned about how to prevent and treat antibody-mediated rejection in kidney transplantation? Am J Transplant 2020; 20 Suppl 4: 12-22 [PMID: 32538535 DOI: 10.1111/ajt.15859]
- Valenzuela NM, Reed EF. Antibody-mediated rejection across solid organ transplants: manifestations, mechanisms, and therapies. J Clin Invest 2017; **127**: 2492-2504 [PMID: 28604384 DOI: 10.1172/JCI90597]
- Böhmig GA, Wahrmann M, Eskandary F, Rostaing L. Novel Approaches to Block Complement. Transplantation 2018; **102**: 1837-1843 [PMID: 29870459 DOI: 10.1097/TP.00000000000002267]
- Tatapudi VS, Montgomery RA. Therapeutic Modulation of the Complement System in Kidney Transplantation: Clinical Indications and Emerging Drug Leads. Front Immunol 2019; 10: 2306 [PMID: 31632397 DOI: 10.3389/fimmu.2019.02306]
- **Bhalla A**, Alachkar N, Alasfar S. Complement-Based Therapy in the Management of Antibody-Mediated Rejection. Adv Chronic Kidney Dis 2020; 27: 138-148 [PMID: 32553246 DOI: 10.1053/j.ackd.2019.12.002]
- Redfield RR, Jordan SC, Busque S, Vincenti F, Woodle ES, Desai N, Reed EF, Tremblay S, Zachary AA, Vo AA, Formica R, Schindler T, Tran H, Looney C, Jamois C, Green C, Morimoto A, Rajwanshi R, Schroeder A, Cascino MD, Brunetta P, Borie D. Safety, pharmacokinetics, and pharmacodynamic activity of obinutuzumab, a type 2 anti-CD20 monoclonal antibody for the desensitization of candidates for renal transplant. Am J Transplant 2019; 19: 3035-3045 [PMID: 31257724 DOI: 10.1111/ajt.15514]
- Levy RA, Gonzalez-Rivera T, Khamashta M, Fox NL, Jones-Leone A, Rubin B, Burriss SW, Gairy K, Maurik AV, Roth DA. 10 Years of belimumab experience: What have we learnt? Lupus 2021; 30: 1705-1721 [PMID: 34238087 DOI: 10.1177/09612033211028653]
- Banham GD, Flint SM, Torpey N, Lyons PA, Shanahan DN, Gibson A, Watson CJE, O'Sullivan AM, Chadwick JA, Foster KE, Jones RB, Devey LR, Richards A, Erwig LP, Savage CO, Smith KGC, Henderson RB, Clatworthy MR. Belimumab in kidney transplantation: an experimental medicine, randomised, placebo-controlled phase 2 trial. Lancet 2018; **391**: 2619-2630 [PMID: 29910042 DOI: 10.1016/S0140-6736(18)30984-X]
- Schroder PM, Schmitz R, Fitch ZW, Ezekian B, Yoon J, Choi AY, Manook M, Barbas A, Leopardi F, Song M, Farris AB, Collins B, Kwun J, Knechtle SJ. Preoperative carfilzomib and lulizumab based desensitization prolongs graft survival in a sensitized non-human primate model. Kidney Int 2021; 99: 161-172 [PMID: 32898569 DOI: 10.1016/j.kint.2020.08.020]
- Doberer K, Kläger J, Gualdoni GA, Mayer KA, Eskandary F, Farkash EA, Agis H, Reiter T, Reindl-Schwaighofer R, Wahrmann M, Cohen G, Haslacher H, Bond G, Simonitsch-Klupp I, Halloran PF, Böhmig GA. CD38 Antibody Daratumumab for the Treatment of Chronic Active Antibody-mediated Kidney Allograft Rejection. Transplantation 2021;



- 105: 451-457 [PMID: 32235256 DOI: 10.1097/TP.0000000000003247]
- Joher N, Matignon M, Grimbert P. HLA Desensitization in Solid Organ Transplantation: Anti-CD38 to Across the Immunological Barriers. Front Immunol 2021; 12: 688301 [PMID: 34093594 DOI: 10.3389/fimmu.2021.688301]
- Spica D, Junker T, Dickenmann M, Schaub S, Steiger J, Rüfli T, Halter J, Hopfer H, Holbro A, Hirt-Minkowski P. Daratumumab for Treatment of Antibody-Mediated Rejection after ABO-Incompatible Kidney Transplantation. Case Rep Nephrol Dial 2019; 9: 149-157 [PMID: 31828078 DOI: 10.1159/000503951]
- Feng X, Zhang L, Acharya C, An G, Wen K, Qiu L, Munshi NC, Tai YT, Anderson KC. Targeting CD38 Suppresses Induction and Function of T Regulatory Cells to Mitigate Immunosuppression in Multiple Myeloma. Clin Cancer Res 2017; **23**: 4290-4300 [PMID: 28249894 DOI: 10.1158/1078-0432.CCR-16-3192]
- Frampton JE. Inebilizumab: First Approval. Drugs 2020; 80: 1259-1264 [PMID: 32729016 DOI: 10.1007/s40265-020-01370-4]
- Viela Bio. Safety and Tolerability of Inebilizumab, VIB4920, or the Combination in Highly Sensitized Candidates Awaiting Kidney Transplantation from a Deceased Donor. [accessed 2019 Nov 22]. In: ClinicalTrials.gov [Internet]. Bethesda (MD): U.S. National Library of Medicine. Available from: https://clinicaltrials.gov/ct2/show/NCT04174677 ClinicalTrials.gov Identifier: NCT04174677
- Eskandary F. Felzartamab in Late Antibody-Mediated Rejection (MOR202). [accessed 2021 Aug 25]. In: ClinicalTrials.gov [Internet]. Bethesda (MD): U.S. National Library of Medicine. Available from: https://clinicaltrials.gov/ct2/show/NCT05021484 ClinicalTrials.gov Identifier: NCT05021484
- Sanofi. Safety, Tolerability and Activity of BIVV009 in Healthy Volunteers and Patients With Complement Mediated Disorders (BIVV009-01). [accessed 2015 Jul 20]. In: ClinicalTrials.gov [Internet]. Bethesda (MD): U.S. National Library of Medicine. Available from: https://clinicaltrials.gov/ct2/show/NCT02502903 ClinicalTrials.gov Identifier:
- Vo AA, Zeevi A, Choi J, Cisneros K, Toyoda M, Kahwaji J, Peng A, Villicana R, Puliyanda D, Reinsmoen N, Haas M, Jordan SC. A phase I/II placebo-controlled trial of C1-inhibitor for prevention of antibody-mediated rejection in HLA sensitized patients. Transplantation 2015; 99: 299-308 [PMID: 25606785 DOI: 10.1097/TP.0000000000000592]
- Montgomery RA, Orandi BJ, Racusen L, Jackson AM, Garonzik-Wang JM, Shah T, Woodle ES, Sommerer C, Fitts D, Rockich K, Zhang P, Uknis ME, Plasma-Derived C1 Esterase Inhibitor for Acute Antibody-Mediated Rejection Following Kidney Transplantation: Results of a Randomized Double-Blind Placebo-Controlled Pilot Study. Am J Transplant 2016; 16: 3468-3478 [PMID: 27184779 DOI: 10.1111/ajt.13871]
- CSL Behring. Efficacy and Safety of Human Plasma-derived C1-esterase Inhibitor as add-on to Standard of Care for the Treatment of Refractory Antibody Mediated Rejection (AMR) in Adult Renal Transplant Recipients. [accessed 2017 Jul 19]. In: ClinicalTrials.gov [Internet]. Bethesda (MD): U.S. National Library of Medicine. Available from: https://clinicaltrials.gov/ct2/show/NCT03221842 ClinicalTrials.gov Identifier: NCT03221842
- Takeda. A Multicenter Study to Evaluate the Efficacy and Safety of Cinryze® for the Treatment of Acute Antibodymediated Rejection in Participants With Kidney Transplant. [accessed 2015 Sep 11]. In: ClinicalTrials.gov [Internet]. Bethesda (MD): U.S. National Library of Medicine. Available from: https://clinicaltrials.gov/ct2/show/NCT02547220 ClinicalTrials.gov Identifier: NCT02547220
- 110 Locke JE, Magro CM, Singer AL, Segev DL, Haas M, Hillel AT, King KE, Kraus E, Lees LM, Melancon JK, Stewart ZA, Warren DS, Zachary AA, Montgomery RA. The use of antibody to complement protein C5 for salvage treatment of severe antibody-mediated rejection. Am J Transplant 2009; 9: 231-235 [PMID: 18976298 DOI: 10.1111/j.1600-6143.2008.02451.x]
- 111 Kocak B, Arpali E, Demiralp E, Yelken B, Karatas C, Gorcin S, Gorgulu N, Uzunalan M, Turkmen A, Kalayoglu M. Eculizumab for salvage treatment of refractory antibody-mediated rejection in kidney transplant patients: case reports. Transplant Proc 2013; 45: 1022-1025 [PMID: 23622614 DOI: 10.1016/j.transproceed.2013.02.062]
- Ghirardo G, Benetti E, Poli F, Vidal E, Della Vella M, Cozzi E, Murer L. Plasmapheresis-resistant acute humoral rejection successfully treated with anti-C5 antibody. Pediatr Transplant 2014; 18: E1-E5 [PMID: 24266980 DOI: 10.1111/petr.12187
- Tan EK, Bentall A, Dean PG, Shaheen MF, Stegall MD, Schinstock CA. Use of Eculizumab for Active Antibodymediated Rejection That Occurs Early Post-kidney Transplantation: A Consecutive Series of 15 Cases. Transplantation 2019; **103**: 2397-2404 [PMID: 30801549 DOI: 10.1097/TP.0000000000002639]
- Glotz D, Russ G, Rostaing L, Legendre C, Tufveson G, Chadban S, Grinyó J, Mamode N, Rigotti P, Couzi L, Büchler M, Sandrini S, Dain B, Garfield M, Ogawa M, Richard T, Marks WH; C10-002 Study Group. Safety and efficacy of eculizumab for the prevention of antibody-mediated rejection after deceased-donor kidney transplantation in patients with preformed donor-specific antibodies. Am J Transplant 2019; 19: 2865-2875 [PMID: 31012541 DOI: 10.1111/ajt.15397]
- Jordan SC, Lorant T, Choi J, Kjellman C, Winstedt L, Bengtsson M, Zhang X, Eich T, Toyoda M, Eriksson BM, Ge S, Peng A, Järnum S, Wood KJ, Lundgren T, Wennberg L, Bäckman L, Larsson E, Villicana R, Kahwaji J, Louie S, Kang A, Haas M, Nast C, Vo A, Tufveson G. IgG Endopeptidase in Highly Sensitized Patients Undergoing Transplantation. N Engl J Med 2017; 377: 442-453 [PMID: 28767349 DOI: 10.1056/NEJMoa1612567]
- Ge S, Chu M, Choi J, Louie S, Vo A, Jordan SC, Toyoda M. Imlifidase Inhibits HLA Antibody-mediated NK Cell Activation and Antibody-dependent Cell-mediated Cytotoxicity (ADCC) In Vitro. Transplantation 2020; 104: 1574-1579 [PMID: 32732834 DOI: 10.1097/TP.0000000000003023]
- Jordan SC, Legendre C, Desai NM, Lorant T, Bengtsson M, Lonze BE, Vo AA, Runström A, Laxmyr L, Sjöholm K, Schiött Å, Sonesson E, Wood K, Winstedt L, Kjellman C, Montgomery RA. Imlifidase Desensitization in Crossmatchpositive, Highly Sensitized Kidney Transplant Recipients: Results of an International Phase 2 Trial (Highdes). Transplantation 2021; 105: 1808-1817 [PMID: 33093408 DOI: 10.1097/TP.0000000000003496]
- Kjellman C, Maldonado AQ, Sjöholm K, Lonze BE, Montgomery RA, Runström A, Lorant T, Desai NM, Legendre C, Lundgren T, von Zur Mühlen B, Vo AA, Olsson H, Jordan SC. Outcomes at 3 years posttransplant in imlifidasedesensitized kidney transplant patients. Am J Transplant 2021; 21: 3907-3918 [PMID: 34236770 DOI: 10.1111/ajt.16754]





Published by Baishideng Publishing Group Inc

7041 Koll Center Parkway, Suite 160, Pleasanton, CA 94566, USA

Telephone: +1-925-3991568

E-mail: bpgoffice@wjgnet.com

Help Desk: https://www.f6publishing.com/helpdesk

https://www.wjgnet.com

