### PROTOCOL

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# Exosomal microRNAs as potential circulating biomarkers in gastrointestinal tract cancers: a systematic review protocol

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#### Abstract

**Background:** Metastasis is the most frequent type of recurrence in gastrointestinal (GI) cancers, and there is an emerging potential for new diagnostic and therapeutic approaches, especially in the cases of metastatic GI carcinomas. The expression profiles of circulating exosomal microRNAs are of particular interest as novel non-invasive diagnostic and prognostic biomarkers for improved detection of GI cancers in body fluids, especially in the serum of patients with recurrent cancers. The aim of this study is to systematically review primary studies and identify the miRNA profiles of serum exosomes of GI cancers.

**Methods and design:** This systematic review will be reported in line with the Preferred Reporting Items for Systematic Reviews and Meta-analyses (PRISMA) guidance. Relevant studies will be identified through a comprehensive search of the following main electronic databases: PubMed, Web of Science, Embase, Scopus, and Google Scholar, with no language restrictions (up to July 2017). Full copies of articles will be identified by a defined search strategy and will be considered for inclusion against pre-defined criteria. The quality assessment of the included studies will be performed by the Newcastle-Ottawa Scale (NOS). Data will be analyzed using Stata software V.12. Publication bias will be assessed by funnel plots, Beggs' and Eggers' tests. The levels of evidence for primary outcomes will be evaluated using the GRADE criteria.

**Discussion:** The analysis of circulating exosomal miRNA profiles provides attractive screening and non-invasive diagnostic tools for the majority of solid tumors including GI cancers. There is limited information regarding the relationship between serum exosomal miRNA profiles and the pathological condition of patients with different GI cancers. Since there is no specific biomarker for GI cancers, we aim to suggest a number of circulating exosomal miRNA candidates as potential multifaceted GI cancer biomarkers for clinical utility.

Systematic review registration: PROSPERO CRD42017057129

Keywords: Circulating, Exosomal microRNAs, Gastrointestinal tract, Cancer

#### Background

Gastrointestinal tract (GI) cancers represent a major challenge to healthcare systems given that they constitute 25% of cancer-related mortalities [1]. The incidence rate of GI cancers varies between developed and

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developing countries [2]. Among the 15 most frequent malignancies worldwide, one out of four new cancer cases and one out of three annual cancer-related deaths are attributed to GI cancers [3].

miRNAs are small non-coding RNA molecules consisting of approximately 22 nucleotides that control gene expression through targeting specific mRNAs bearing partially complementary target sequences for degradation and/or translational repression. miRNAs have been shown to be involved in regulating several cellular processes including cell differentiation, proliferation, and



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apoptosis in both normal and pathological conditions [4, 5]. miRNAs can act as either oncogenes or tumor suppressor genes in the networks specifically altered during cancer development and progression [4, 6, 7]. They are present as mobile genetic components for cell to cell communication or through the systemic circulation system within the body [4, 8].

Exosomes are nano-sized membrane vesicles, ranging from 30 to 100 nm in size, that are secreted by several cell types, including cancer cells, by exocytosis and can be isolated from different body fluids and malignant ascites [1, 4, 9]. Exosome content is composed of unique miRNAs, mRNA, DNA, lipids, and proteins that indicate the genetic information of their parental cells [10]. Exosomes play an important role in transferring bioactive oncogenic cargo to non-transformed recipient cells which cause tumor invasion, drug resistance, metastasis, and modulation of cancer metabolism [1, 11]. Findings have shown that the discovery of specific exosomal content and markers may represent a novel diagnostic tool [12, 13]. Since the majority of serum miRNAs are encapsulated in exosomes in order to be more stable, the exosomal miRNAs could be applied as promising non-invasive biomarkers and potential targetable factors in cancer diagnosis and treatment [9, 14, 15]. Recent studies have focused on "tumor-specific" or "tumor-enriched" miRNAs which are specifically packaged into exosomes [16-19].

Exosomal miRNA profiling of serum from different cancer patients versus healthy controls has revealed significant differences in relation to cancer development and metastasis, indicating a possible use of these miRNAs as prognostic biomarkers [20].The role of specific exosomal miRNAs as biomarkers in cancer diagnosis, prognosis, and screening have been studied in several types of cancers, including GI cancers [15–17, 21–32].

Some of the serum exosomal miRNAs can be used as biomarkers to predict the recurrence of GI carcinomas such as hepatocellular cancer and esophageal squamous cell carcinoma [33, 34]. The diagnostic and prognostic roles of exosomal miRNA profiles of colorectal cancer (CRC), as the most common malignancy of the GI system, have been evaluated in several studies [15, 31, 35–40].

Previous studies had been developed to investigate the preclinical and clinical characteristics of GI cancers and have led to improved overall and progression-free survival rates. However, there is an emerging potential for screening, diagnosis, or surveillance of cancer and the designing of new therapeutic approaches, especially in the cases of metastatic GI carcinomas. The identification and validation of biomarkers that can be measured routinely in easily accessible samples, including plasma, and that are able to diagnose cancer and predict treatment efficacy and the risk of progression or relapse is a major challenge in cancer research. To date, many researchers have published their data on the clinical relevance of micoRNAs and exosomal miRNA expression and circulating exosomal miRNAs in different clinical and nonclinical samples including tissue, stool, serum, and cell lines of GI cancers [15, 31, 35, 40–44].

Recently, a published review showed that the exosomal miRNAs could be detected and isolated from body fluids such as saliva. This review primarily provided an overview on exosomal miRNAs as diagnostic markers of GI cancers with a focus on the origin and trafficking of exosomes between cells and techniques to isolate exosomal miRNAs, micoRNAs, and exosomal miRNAs expression [44]. However, there is currently no systematic review on circulating exosomal miRNAs as promising non-invasive biomarkers for GI cancers. In the current systematic review, we focus on studies that have evaluated circulating exosomal miR-NAs as non-invasive biomarkers in serum of patients with primary GI tumors. We also aim to carry out a systematic review of published exosomal miRNAs, profiling studies that were performed by microarray and real-time PCR methods and then compare the exosomal miRNA expression profiles between primary and metastatic tumors in the serum or plasma of GI cancer patients. Additionally, we intend to identify the most consistently downregulated and upregulated circulating exosomal microRNAs in GI cancers and introduce some of them as diagnostic and prognostic biomarkers for non-invasive clinical applications. Although identification of exosomal miRNAs in other clinical specimen may be significant, in practice, it is difficult to apply invasive methods on patients. On the other hand, it would be challenging to apply findings about exosomal miRNAs expression in non-clinical samples into clinical contexts. Therefore, this systematic review aims to evaluate the circulating exosomal miRNAs as promising non-invasive biomarkers of GI cancers, for the first time.

#### Methods/Design

#### Aim of the study

The primary objective of this review is to provide a systematic review of primary studies to estimate the frequency of exosome-encapsulated miRNAs in the circulating blood of patients with GI cancers. The secondary objectives are to identify the expression pattern (up- and downregulation) of exosomal miRNAs in patients with different gastrointestinal tract cancers and to identify the specific circulating exosomal miRNAs involved in the diagnosis and prognosis of GI cancers. This review will complement the findings of an existing review published in 2016 [44].

#### **Review questions**

This review of studies should address the following points:

- 1. To establish the frequency of circulating exosomal miRNAs in GI cancers.
- 2. To establish the up- and downregulated circulating exosomal miRNAs in GI cancers.
- 3. To identify the specific circulating exosomal miRNAs involved in the diagnosis and prognosis of GI cancers.

#### Study design

This review protocol has been published in the PROS-PERO International prospective register of systematic reviews (http://www.crd.york.ac.uk/PROSPERO), with the registration number CRD 42017057129. In addition, the completed review will be reported in line with the Preferred Reporting Items for Systematic Reviews and Meta-Analyses (PRISMA) guidance [45] (Additional file 1).

## Criteria for considering studies for the review *Inclusion criteria*

Observational studies (cross-sectional, case-control, and cohort studies) investigating circulating exosomal miR-NAs in diagnosis and prognosis of GI malignancies, including upper and lower GI, and hepatopancreato-biliary will be included.

#### Participants

The participants are patients with GI cancers.

#### Types of studies to be excluded

Studies assessing in vitro and in vivo models, reviews, letters, editorials, case reports, and case series studies will be excluded.

## Search methods for identification of studies *Electronic searches*

The search strategy for identification of relevant studies will comprise the following main electronic databases: PubMed, Web of Science, Embase, and Scopus, as well as Google Scholar search engines with no language restrictions (up to July 2017). An example of the PubMed

Table 1 Search strategy	for	PubMed
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Quary	Search syntax
	Related terms/synonyms
#1	(mesentery[Mesh] OR stomach[Mesh] OR Pancreas[Mesh] OR rectum[Mesh] OR duodenum[Mesh] OR jejunum[Mesh] OR ileum[Mesh] OR cecum[Mesh] OR colon[Mesh] OR mouth[Mesh] OR esophagus[Mesh] OR Gastrointestinal Tracts[Mesh] OR "Gastrointestinal Tracts"[tiab] OR "GI Tract"[tiab] OR "Gastrointestinal tract"[tiab] OR "Gastrointestinal Tracts[Mesh] OR "Digestive Tract"[tiab] OR "Digestive Tract"[tiab] OR "Lower GI Tract"[tiab] OR GI[tiab] OR oral[tiab] OR mouth[tiab] OR esophagus[tiab] OR gullet[tiab] OR gullet[tiab] OR gastrointestinal OR duodenum[tiab] OR jejunum[tiab] OR ileum[tiab] OR cecum[tiab] OR colon[tiab] OR colorectal[tiab] OR sigmoid[tiab] OR rectum[tiab] OR anus[tiab] OR mesentery[tiab] OR hepatic[tiab] OR liver[tiab] OR hepatocellular[tiab] OR stomach[tiab] OR Pancrea*[tiab])

#2 (Neoplasms [Mesh] OR Cancer\*[tiab] OR Neoplasm\*[tiab] OR Carcinoma[tiab] OR Tumo\*[tiab])

#3 (extracellular vesicles[Mesh] OR exosomes[Mesh] OR exosome\*[tiab] OR "extracellular vesicle"[tiab] OR "extracellular vesicles"[tiab] OR microvesicle[tiab] OR "Shedding Microvesicles"[tiab])

The asterisk (\*) is used to signify truncation

search strategy is shown in Table 1. The search syntax will be modified in other databases accordingly.

#### Other resources

Reference lists of relevant primary studies, reviews, and key journals will be searched for additional studies.

#### Screening of the studies

Duplicate studies will be removed. All databases will be searched, and the titles and abstracts will be extracted (EGh). All the retrieved titles and abstracts from the electronic search will be screened, and the relevant data will be independently extracted (EGh and AR) according to the previously described inclusion criteria. The extracted data will then be cross-checked to rule out any discrepancies. Unresolved discrepancies will be referred to a third author to solve (LJ). All the reasons for exclusion of ineligible studies will be recorded.

#### Data extraction (selection and coding)

Data for each of the included studies will be extracted as follows:

- 1. Study characteristics (first author's surname, publication year, publication language, country, study design, setting, locations, criteria for sample selection and sample size, diagnostic criteria, outcomes measured, patient enrolment strategies, exosome isolation and identification methods, miRNA profiling, real-time PCR results of serum samples)
- 2. Participants' characteristics (age, gender)
- 3. Frequency estimates of expression of circulating exosomal miRNAs

Discrepancies between two reviewers will be resolved by consensus. An independent investigator will be consulted through discussion to reach consensus where there is uncertainty or disagreement between reviewers.

<sup>#4 #1</sup> AND #2 AND #3

Corresponding authors will be contacted if further information is needed. If no response is received after sending a reminder, studies will be excluded.

#### Risk of bias (quality) assessment

Quality assessment of observational studies is challenging due to their methodological complexities and the subjective nature of guality evaluations [46]. The Cochrane Collaboration endorsed the use of the Newcastle-Ottawa Scale (NOS) to assess the quality of observational studies in its 2011 handbook [47, 48]. The final risk of bias of included studies will be performed by the above mentioned tool. Two reviewers will independently assess the methodological quality of primary studies by the NOS. This tool is used for assessing the quality of non-randomized studies included in a systematic review and/or metaanalyses. The method was developed as a collaboration between the University of Newcastle, Australia, and the University of Ottawa, Canada, using a Delphi process to define variables for data extraction. Using the tool, each study is judged on eight items, categorized into three groups: the selection of the study groups, the comparability of the groups, and the ascertainment of either the exposure or outcome of interest for case-control or cohort studies respectively [49]. Separate tools were developed for cohort and case-control studies. It has also been adapted for cross-sectional studies [50-52]. For more details, see three appendix scales for evaluating the quality assessment of the above mentioned observational studies (Additional files 2, 3, and 4).

#### Strategy for data synthesis

Given the considerable variability among cancers and the difficulty in detecting common exosomal miRNAs and characteristics of the studies, the extracted outcomes will be summarized and reported using descriptive statistics without conducting any meta-analyses. Two separate tables will be used in all included studies. The first table will provide details on study quality according to the mentioned tool, and the other one will include study design, participants, and frequency of exosomal miRNAs in patients with GI cancers. The publication bias will be assessed by funnel plots (i.e., plots of study results against precision) and Beggs' and Eggers' tests. We will assess the overall quality of the evidence for each main outcome based on the GRADE (Grading of Recommendations Assessment, Development and Evaluation) system [53, 54]. GRADE provides explicit criteria that address study design, risk of bias, imprecision, inconsistency, indirectness, and magnitude of the effect to rate the quality of evidence across studies. This method rates the quality of the evidence from high (very confident that the true effect lies close to that of the estimated effect) to very low (very little confidence in the effect estimate).

#### Sensitivity analysis

Given the diversity of cancer types and different reported exosomal miRNAs across the studies, we anticipate that we cannot perform a meta-analysis; therefore, no sensitivity analysis will be applicable to this study.

#### Analysis of subgroups or subsets

If the data is determined to be heterogeneous, we will conduct a narrative synthesis of the findings from the included studies; therefore, a subgroup analysis will not be applicable to this study.

#### Discussion

Emerging evidence supports the hypothesis that exosomal microRNAs can be used as circulating diagnostic biomarkers of several cancers. However, there is currently no collective view of which circulating exosomal microRNAs are suitable candidates. The distinctive properties of miR-NAs within exosomes can be manipulated to render them suitable for the development of effective diagnostic and targetable tools for non-invasive screening and targeting of tumor cells in cancer patients. Reviews showing the serum exosomal microRNAs in different cancers have already emerged [1, 39, 44], but to date, this will be the first systematic review to evaluate the circulating exosomal miRNAs in the serum of patients with GI cancers. The findings of this review may be used in clinical settings and in designing and developing of the novel non-invasive biomarkers for early diagnosis and pathological condition of patients with GI cancers.

#### **Additional files**

Additional file 1: PRISMA-P (Preferred Reporting Items for Systematic Review and Meta-Analysis Protocols) 2015 checklist [45]. (DOC 85 kb) Additional file 2: Newcastle-Ottawa quality assessment scale for crosssectional studies. (PDF 189 kb) Additional file 3: Newcastle-Ottawa quality assessment scale for case-

control studies. (PDF 17 kb)

Additional file 4: Newcastle-Ottawa quality assessment scale for cohort studies. (PDF 93 kb)

#### Abbreviations

GI: Gastrointestinal; microRNAs: miRNAs, miRs; PRISMA: Preferred Reporting Items for Systematic Reviews and Meta-Analyses

#### Acknowledgements

Not applicable.

#### Funding None.

#### Availability of data and materials

All recorded data from the data extraction process will be available on request to the extent that they are not included in the systematic review article.

#### Authors' contributions

ZM, LJ, EGh, AR, and RGh contributed to the concept and study design. EGh and AR developed the search strategies. EGh and LJ contributed to the implementation and analysis of the studies and will also screen potential studies and extract data and assess quality, while ZM, RGH, AR, FA, SB, and MhAL will review the work. EGh and AR contributed to the initial drafting and critical revision the manuscript, and all authors gave feedback to the final draft of the protocol. EGh approved the manuscript for submission. All authors read and approved the final manuscript.

#### Ethics approval and consent to participate

As this systematic review will only be based on published data already in the public domain, an ethical approval is not required. The findings of the current study will be disseminated through a publication in a peer-reviewed journal and conference presentations. To the best of our knowledge, there are no systematic reviews that have specifically looked at the frequency of exosomeencapsulated miRNAs in the circulating blood of patients with GI cancers. We will submit the findings of this systematic review for publication and presentation at national and international conferences to update information on the circulating exosomal miRNAs as candidates of GI cancers. This review will ultimately help inform and guide future studies in the field.

#### Consent for publication

Not applicable.

#### **Competing interests**

The authors declare that they have no competing interests.

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#### Received: 18 June 2017 Accepted: 8 November 2017 Published online: 17 November 2017

#### References

- Rahbari M, Rahbari N, Reissfelder C, Weitz J, Kahlert C. Exosomes: novel implications in diagnosis and treatment of gastrointestinal cancer. Langenbeck's Arch Surg. 2016;401(8):1097–110.
- Sahin IH, Hassan MM, Garrett CR. Impact of non-steroidal anti-inflammatory drugs on gastrointestinal cancers: current state-of-the science. Cancer Lett. 2014;345(2):249–57.
- Neuzillet C, Rousseau B, Kocher H, Bourget P, Tournigand C. Unravelling the pharmacologic opportunities and future directions for targeted therapies in gastro-intestinal cancers part 1: GI carcinomas. Pharmacol Ther. 2017;174:145–72.
- Hayashi T, Lombaert IM, Hauser BR, Patel VN, Hoffman MP. Exosomal microRNA transport from salivary mesenchyme regulates epithelial progenitor expansion during organogenesis. Dev Cell. 2017;40(1):95–103.
- Cho YE, Kim SH, Lee BH, Baek MC. Circulating plasma and exosomal microRNAs as indicators of drug-induced organ injury in rodent models. Biomol Ther. 2017;25(4):367–73.
- Schwarzenbach H. The clinical relevance of circulating, exosomal miRNAs as biomarkers for cancer. Expert Rev Mol Diagn. 2015;15(9):1159–69.
- Babashah S, Soleimani M. The oncogenic and tumour suppressive roles of microRNAs in cancer and apoptosis. Eur J Cancer. 2011;47(8):1127–37.

- Tokuhisa M, Ichikawa Y, Kosaka N, Ochiya T, Yashiro M, Hirakawa K, et al. Exosomal miRNAs from peritoneum lavage fluid as potential prognostic biomarkers of peritoneal metastasis in gastric cancer. PLoS One. 2015;10(7): e0130472.
- Liu Q, Yu Z, Yuan S, Xie W, Li C, Hu Z, et al. Circulating exosomal microRNAs as prognostic biomarkers for non-small-cell lung cancer. Oncotarget. 2017; 8(8):13048–58.
- Wang H, Hou L, Li A, Duan Y, Gao H, Song X. Expression of serum exosomal microRNA-21 in human hepatocellular carcinoma. Biomed Res Int. 2014; 2014;864894.
- Charoenviriyakul C, Takahashi Y, Morishita M, Matsumoto A, Nishikawa M, Takakura Y. Cell type-specific and common characteristics of exosomes derived from mouse cell lines: yield, physicochemical properties, and pharmacokinetics. Eur J Pharm Sci. 2017;96:316–22.
- Yoshioka Y, Uehara K, Ebata T, Yokoyama Y, Mitsuma A, Ando Y, et al. Postoperative complications following neoadjuvant bevacizumab treatment for advanced colorectal cancer. Surg Today. 2014;44(7):1300–6.
- van der Meel R, Fens MH, Vader P, van Solinge WW, Eniola-Adefeso O, Schiffelers RM. Extracellular vesicles as drug delivery systems: lessons from the liposome field. J Control Release. 2014;195:72–85.
- Uratani R, Toiyama Y, Kitajima T, Kawamura M, Hiro J, Kobayashi M, et al. Diagnostic potential of cell-free and exosomal microRNAs in the identification of patients with high-risk colorectal adenomas. PLoS One. 2016;11(10):e0160722.
- Ogata-Kawata H, Izumiya M, Kurioka D, Honma Y, Yamada Y, Furuta K, et al. Circulating exosomal microRNAs as biomarkers of colon cancer. PLoS One. 2014;9(4):e92921.
- Tomasetti M, Lee W, Santarelli L, Neuzil J. Exosome-derived microRNAs in cancer metabolism: possible implications in cancer diagnostics and therapy. Exp Mol Med. 2017;49(1):e285.
- 17. Lindner K, Haier J, Wang Z, Watson DI, Hussey DJ, Hummel R. Circulating microRNAs: emerging biomarkers for diagnosis and prognosis in patients with gastrointestinal cancers. Clin Sci. 2015;128(1):1–15.
- Lai X, Wang M, McElyea SD, Sherman S, House M, Korc M. A microRNA signature in circulating exosomes is superior to exosomal glypican-1 levels for diagnosing pancreatic cancer. Cancer Lett. 2017;393:86–93.
- Donnarumma E, Fiore D, Nappa M, Roscigno G, Adamo A, laboni M, et al. Cancer-associated fibroblasts release exosomal microRNAs that dictate an aggressive phenotype in breast cancer. Oncotarget. 2017;8(12):19592–608.
- Ye S-b, Li Z-L, Luo D-h, Huang B-j, Chen Y-S, Zhang X-s, et al. Tumor-derived exosomes promote tumor progression and T-cell dysfunction through the regulation of enriched exosomal microRNAs in human nasopharyngeal carcinoma. Oncotarget. 2014;5(14):5439–52.
- Alegre E, Sanmamed MF, Rodriguez C, Carranza O, Martín-Algarra S, González Á. Study of circulating microRNA-125b levels in serum exosomes in advanced melanoma. Arch Pathol Lab Med. 2014;138(6):828–32.
- Cazzoli R, Buttitta F, Di Nicola M, Malatesta S, Marchetti A, Rom WN, et al. microRNAs derived from circulating exosomes as noninvasive biomarkers for screening and diagnosing lung cancer. J Thorac Oncol. 2013;8(9):1156–62.
- Giallombardo M, Borrás JC, Castiglia M, Van Der Steen N, Mertens I, Pauwels P, et al. Exosomal miRNA analysis in non-small cell lung cancer (NSCLC) patients' plasma through qpcr: a feasible liquid biopsy tool. JoVE (Journal of Visualized Experiments). 2016;(111):e53900:1–6. https://www.jove.com/video/ 53900/exosomal-mirna-analysis-non-small-cell-lung-cancer-nsclc-patients.
- 24. Huang X, Yuan T, Liang M, Du M, Xia S, Dittmar R, et al. Exosomal miR-1290 and miR-375 as prognostic markers in castration-resistant prostate cancer. Eur Urol. 2015;67(1):33–41.
- Liu J, Sun H, Wang X, Yu Q, Li S, Yu X, et al. Increased exosomal microRNA-21 and microRNA-146a levels in the cervicovaginal lavage specimens of patients with cervical cancer. Int J Mol Sci. 2014;15(1):758–73.
- Tanaka Y, Kamohara H, Kinoshita K, Kurashige J, Ishimoto T, Iwatsuki M, et al. Clinical impact of serum exosomal microRNA-21 as a clinical biomarker in human esophageal squamous cell carcinoma. Cancer. 2013;119(6):1159–67.
- Taverna S, Giallombardo M, Gil-Bazo I, Carreca AP, Castiglia M, Chacártegui J, et al. Exosomes isolation and characterization in serum is feasible in non-small cell lung cancer patients: critical analysis of evidence and potential role in clinical practice. Oncotarget. 2016;7(19):28748.
- Taylor DD, Gercel-Taylor C. MicroRNA signatures of tumor-derived exosomes as diagnostic biomarkers of ovarian cancer. Gynecol Oncol. 2008;110(1):13–21.
- 29. Vaksman O, Tropé C, Davidson B, Reich R. Exosome-derived miRNAs and ovarian carcinoma progression. Carcinogenesis. 2014;35(9):2113–20.

- Wang J, Zhou Y, Lu J, Sun Y, Xiao H, Liu M, et al. Combined detection of serum exosomal miR-21 and HOTAIR as diagnostic and prognostic biomarkers for laryngeal squamous cell carcinoma. Med Oncol. 2014;31(9):1–8.
- Kogure T, Lin WL, Yan IK, Braconi C, Patel T. Intercellular nanovesiclemediated microRNA transfer: a mechanism of environmental modulation of hepatocellular cancer cell growth. Hepatology. 2011;54(4):1237–48.
- Manterola L, Guruceaga E, Gallego Perez-Larraya J, Gonzalez-Huarriz M, Jauregui P, Tejada S, et al. A small noncoding RNA signature found in exosomes of GBM patient serum as a diagnostic tool. Neuro-Oncology. 2014;16(4):520–7.
- Sugimachi K, Matsumura T, Hirata H, Uchi R, Ueda M, Ueo H, et al. Identification of a bona fide microRNA biomarker in serum exosomes that predicts hepatocellular carcinoma recurrence after liver transplantation. Br J Cancer. 2015;112(3):532–8.
- Takeshita N, Hoshino I, Mori M, Akutsu Y, Hanari N, Yoneyama Y, et al. Serum microRNA expression profile: miR-1246 as a novel diagnostic and prognostic biomarker for oesophageal squamous cell carcinoma. Br J Cancer. 2013;108(3):644–52.
- Tauro BJ, Greening DW, Mathias RA, Mathivanan S, Ji H, Simpson RJ. Two distinct populations of exosomes are released from LIM1863 colon carcinoma cell-derived organoids. Mol Cell Proteomics. 2013;12(3):587–98.
- 36. Ji H, Chen M, Greening DW, He W, Rai A, Zhang W, et al. Deep sequencing of RNA from three different extracellular vesicle (EV) subtypes released from the human LIM1863 colon cancer cell line uncovers distinct miRNAenrichment signatures. PLoS One. 2014;9(10):e110314.
- Tovar-Camargo OA, Toden S, Goel A. Exosomal microRNA biomarkers: emerging frontiers in colorectal and other human cancers. Expert Rev Mol Diagn. 2016;16(5):553–67.
- Wang J, Yan F, Zhao Q, Zhan F, Wang R, Wang L, et al. Circulating exosomal miR-125a-3p as a novel biomarker for early-stage colon cancer. Sci Rep. 2017;7(1):1–9.
- Matsumura T, Sugimachi K, linuma H, Takahashi Y, Kurashige J, Sawada G, et al. Exosomal microRNA in serum is a novel biomarker of recurrence in human colorectal cancer. Br J Cancer. 2015;113(2):275–81.
- Yamada N, Nakagawa Y, Tsujimura N, Kumazaki M, Noguchi S, Mori T, et al. Role of intracellular and extracellular microRNA-92a in colorectal cancer. Transl Oncol. 2013;6(4):482–92.
- Chiba M, Kimura M, Asari S. Exosomes secreted from human colorectal cancer cell lines contain mRNAs, microRNAs and natural antisense RNAs, that can transfer into the human hepatoma HepG2 and lung cancer A549 cell lines. Oncol Rep. 2012;28(5):1551–8.
- Senfter D, Holzner S, Kalipciyan M, Staribacher A, Walzl A, Huttary N, et al. Loss of miR-200 family in 5-fluorouracil resistant colon cancer drives lymphendothelial invasiveness in vitro. Hum Mol Genet. 2015;24(13):3689–98.
- Ohshima K, Inoue K, Fujiwara A, Hatakeyama K, Kanto K, Watanabe Y, et al. Let-7 microRNA family is selectively secreted into the extracellular environment via exosomes in a metastatic gastric cancer cell line. PLoS One. 2010;5(10):e13247.
- Nedaeinia R, Manian M, Jazayeri M, Ranjbar M, Salehi R, Sharifi M, et al. Circulating exosomes and exosomal microRNAs as biomarkers in gastrointestinal cancer. Cancer Gene Ther. 2017;24(2):48–56.
- Moher D, Liberati A, Tetzlaff J, Altman DG, Group P. Preferred reporting items for systematic reviews and meta-analyses: the PRISMA statement. PLoS Med. 2009;6(7):e1000097.
- Neyarapally GA, Hammad TA, Pinheiro SP, Iyasu S. Review of quality assessment tools for the evaluation of pharmacoepidemiological safety studies. BMJ Open. 2012;2(5):e001362.
- Higgins JP, Green S. Cochrane handbook for systematic reviews of interventions: John Wiley & Sons; 2011. books.google.com. https://scholar. google.com/scholar?hl=en&as\_sdt=0%2C5&q=Higgins+JPT%2C+Green+S +%28editors%29.+Cochrane+Handbook+for+Systematic+Reviews+of +Interventions+Version+5.1.0+%5Bupdated+March+2011%5D.+The +Cochrane+Collaboration%2C+2011.+Available+from +http%3A%2F%2Fhandbook.cochrane.org.&btnG=.
- Wels GASB, O'Connell D, et al. The Newcastle-Ottawa Scale (NOS) for assessing the quality of nonrandomised studies in meta-analyses [webpage on the Internet]. Ottawa: Ottawa Hospital Research Institute; 2011. [Accessed 5 Feb 2013]
- Deeks J, Dinnes J, D'amico R, Sowden A, Sakarovitch C, Song F, et al. Evaluating non-randomised intervention studies. Health Technology Assessment (Winchester, England). 2003;7(27):iii-x, 1-173.

- Rotenstein LS, Ramos MA, Torre M, Segal JB, Peluso MJ, Guille C, et al. Prevalence of depression, depressive symptoms, and suicidal ideation among medical students: a systematic review and meta-analysis. JAMA. 2016;316(21):2214–36.
- Mata DA, Ramos MA, Bansal N, Khan R, Guille C, Di Angelantonio E, et al. Prevalence of depression and depressive symptoms among resident physicians: a systematic review and meta-analysis. JAMA. 2015;314(22):2373–83.
- Zeng X, Zhang Y, Kwong JS, Zhang C, Li S, Sun F, et al. The methodological quality assessment tools for preclinical and clinical studies, systematic review and meta-analysis, and clinical practice guideline: a systematic review. Journal of evidence-Based Medicine. 2015;8(1):2–10.
- Atkins D, Best D, Briss PA, Eccles M, Falck-Ytter Y, Flottorp S, et al. Grading quality of evidence and strength of recommendations. BMJ (Clinical research ed). 2004;328(7454):1490.
- Balshem H, Helfand M, Schünemann HJ, Oxman AD, Kunz R, Brozek J, et al. GRADE guidelines: 3. Rating the quality of evidence. J Clin Epidemiol. 2011; 64(4):401–6.

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