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Novel *CAD-ALK* gene rearrangement is drugable by entrectinib in colorectal cancer

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Background: Activated anaplastic lymphoma kinase (*ALK*) gene fusions are recurrent events in a small fraction of colorectal cancers (CRCs), although these events have not yet been exploited as in other malignancies.

Methods: We detected *ALK* protein expression by immunohistochemistry and gene rearrangements by fluorescence *in situ* hybridisation in the ALKA-372-001 phase I study of the pan-Trk, ROS1, and *ALK* inhibitor entrectinib. One out of 487 CRCs showed *ALK* positivity with a peculiar pattern that prompted further characterisation by targeted sequencing using anchored multiplex PCR.

Results: A novel *ALK* fusion with the carbamoyl-phosphate synthetase 2, aspartate transcarbamylase, and dihydroorotase (*CAD*) gene (*CAD-ALK* fusion gene) was identified. It resulted from inversion within chromosome 2 and the fusion of exons 1–35 of *CAD* with exons 20–29 of *ALK*. After failure of previous standard therapies, treatment of this patient with the *ALK* inhibitor entrectinib resulted in a durable objective tumour response.

Conclusions: We describe the novel *CAD-ALK* rearrangement as an oncogene and provide the first evidence of its drugability as a new molecular target in CRC.

Activated anaplastic lymphoma kinase (*ALK*) gene fusions, found in haematological and solid malignancies, have been successfully exploited as therapeutic targets in lung and inflammatory myofibroblastic tumours using the *ALK* kinase inhibitors crizotinib and ceritinib (Grande *et al*, 2011; Awad and Shaw, 2014). In colorectal cancer (CRC), *ALK* rearrangements are recurrent events found in 0.4–3% of samples analysed and involve as partner genes *EML4*, *C2orf44*, or *PRKARIA* (reviewed by Aisner *et al* (2014)). Although expression of the resulting chimaeric transcripts was shown in some patients by PCR with reverse transcription (RT-PCR) analysis, the final evidence of *ALK* fusion protein expression

in these tumours has been lacking, hampering the exploitation of these observations in the therapeutic setting of CRC.

Entrectinib is a selective pan-TRK, ROS1, and *ALK* kinase inhibitor with strong preclinical activity in multiple cancer models where these targets are constitutively activated (Ardini *et al*, 2013; De Braud *et al*, 2014; Sartore-Bianchi *et al*, 2015). Entrectinib is currently being developed in phases I–II clinical studies in patients with tumours harbouring *NTRK*, *ROS1*, or *ALK* gene aberrations (De Braud *et al*, 2014).

Screening of 487 CRC formalin-fixed, paraffin-embedded (FFPE) samples for *ALK* expression by immunohistochemistry

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(IHC) within the ALKA-372-001 phase I, first-in-human study of entrectinib identified a sample with high levels of ALK protein expression. This finding was extremely atypical and highly suggestive of a rearrangement event placing the *ALK* gene, not normally expressed in the colon, under ectopic control of a fusion partner promoter. We then confirmed by fluorescence *in situ* hybridisation (FISH) that an *ALK* gene rearrangement was in fact present (Medico *et al*, 2015). Here we report the molecular and clinical study of this CRC patient, which led to the discovery of a novel *ALK* rearrangement that was present in the primary tumour, in a thoracic lymph node, and in liver metastases. We also describe the objective response of this patient to entrectinib treatment, providing the first evidence of an *ALK* gene rearrangement as a clinically relevant therapeutic target in CRC.

MATERIALS AND METHODS

We detected ALK protein expression by IHC using the ALK mouse monoclonal antibody 5A4 (N-Histofine ALK Detection Kit; Nichirei Biosciences Inc., Histo-Line Laboratories s.r.l, Milan, Italy) and gene rearrangements by FISH using the LSI ALK Break Apart FISH Probe Kit (Vysis, Abbott Molecular, Abbott Park, IL, USA) as previously described (Medico *et al*, 2015).

To identify the *ALK* rearrangement partner, we subjected RNA from the primary tumour FFPE clinical samples to next-generation sequencing (NGS) analysis using targeted sequencing technology based on anchored multiplex PCR (Archer AMP; ARCHERDX, Boulder, CO, USA), customised for the detection of rearrangements of selected tyrosine kinases, including ALK, and allowing simultaneous identification of the fusion partner through an unbiased approach that did not require *a priori* knowledge of the

gene fusion event. PCR/Sanger sequencing of the chimaeric transcript using primers spanning the fusion junction region confirmed the rearrangement. Mutations in *KRAS* and *NRAS* exons 2, 3, and 4 and *BRAF* exon 15; *HER2* amplification; and *ROS1* and *NTRK* rearrangements were assessed as previously described (Sartore-Bianchi *et al*, 2015; Siravegna *et al*, 2015; Valtorta *et al*, 2015).

The patient was enrolled in the ALKA-372-001 phase I study (EudraCT Number: 2012-000148-88) and received treatment with entrectinib 400 mg m⁻² po qd, which was recently determined to be the recommended phase II dose (RP2D). Objective tumour response was measured by computed tomography (CT) using the Response Evaluation Criteria in Solid Tumours (RECIST version 1.1) (Eisenhauer *et al*, 2009).

RESULTS

The patient was a 53-year-old woman with metastatic CRC (brain, thoracic lymph nodes and liver) who was in disease progression after standard therapies, including surgery (right hemicolectomy), external beam radiation therapy to the CNS and thoracic lymph nodes, and two lines of chemotherapy, both based on oxaliplatin, 5-fluorouracil/leucovorin, and bevacizumab, administered before and after the radiation therapy. The primary tumour resected in March 2012 was a grade 3 adenocarcinoma of the right colon metastatic to the supraclavicular lymph node biopsied in April 2012. At progression in March 2015, the patient gave written informed consent to a biopsy of liver metastases. All samples showed histology of primary CRC and CRC metastases (Figure 1A–C1). High levels of ALK protein were observed by IHC in the primary tumour, thoracic lymph node and liver

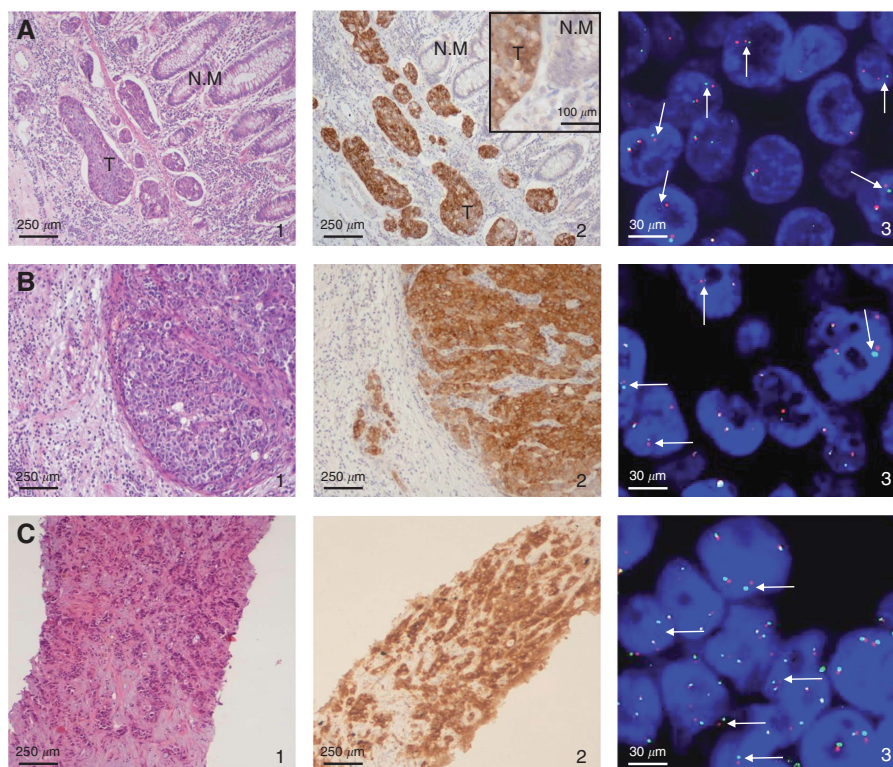


Figure 1. Histological, immunohistochemical, and fluorescence *in situ* hybridisation analyses of the primary right colonic tumour, lymph node, and liver metastasis of the case presented. Haematoxylin & eosin, immunohistochemical and FISH images of the primary colon tumour (A 1–3; N.M: normal mucosa, T: tumour), lymph node (B 1–3), and liver metastasis (C 1–3) of the patient presented in this report, showing malignant tumour (A1, B1, C1), ALK protein overexpression (A2, B2, C2) and ALK gene rearrangements (A3, B3, C3, white arrows). Original magnification of images: $\times 100$ for haematoxylin & eosin and immunohistochemical staining, Insert $\times 400$; $\times 630$ for FISH analysis.

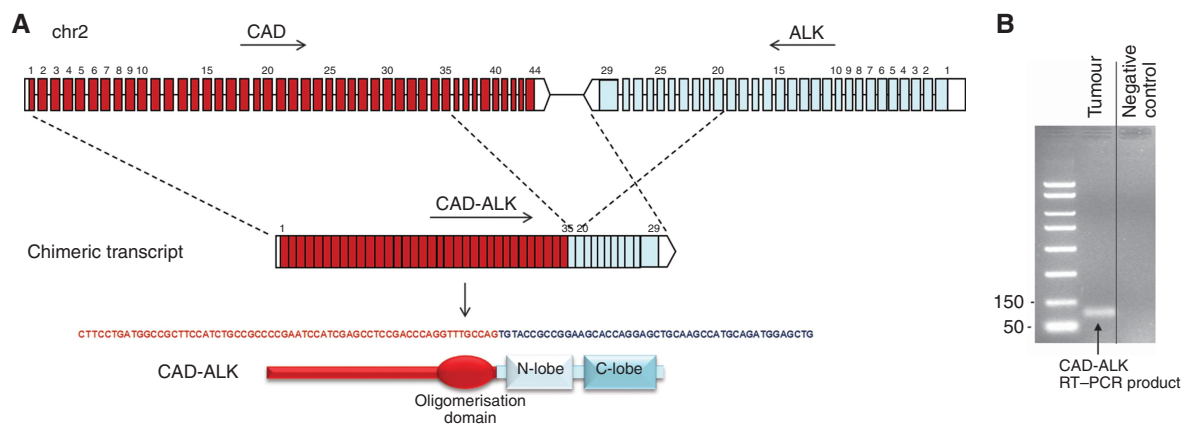


Figure 2. Identification of the *CAD-ALK* gene rearrangement. **(A)** The upper section shows a schematic representation of the *CAD-ALK* genomic DNA rearrangement and the resulting transcript. The sequence spanning the rearrangement junction is also shown. Exons are represented by coloured boxes, and introns are represented by lines: *CAD* in red and *ALK* in light blue. The lower section shows the functional domains conserved in the chimeric *CAD-ALK* protein. **(B)** Characterisation of the *CAD-ALK* transcript by PCR. Agarose gel showing amplification with primers for the rearranged *CAD-ALK* chimaeric transcript, spanning *CAD* exon 35 to *ALK* exon 20. The tumour sample was compared with a negative control sample (U138-MG cell line, expressing *ALK* full length).

metastasis (Figure 1A–C2), and the underlying *ALK* abnormality consisted of an *ALK* rearrangement as demonstrated by FISH (Figure 1A–C3). Further molecular characterisation showed wild-type *KRAS* and *NRAS* exons 2, 3, and 4 and *BRAF* exon 15, no amplification of *HER2*, and no *ROS1* or *NTRK* rearrangements. Moreover, we found no amplification of the *ALK* gene. Further investigation by NGS identified an *ALK* rearrangement resulting from an inversion within chromosome 2, fusing exons 1–35 of the carbamoyl-phosphatase 2, aspartate transcarbamylase, and dihydroorotase (*CAD*) gene with exons 20–29 of *ALK* (C35-A20), and confirmed by PCR/Sanger sequencing (Figure 2A and B).

These findings prompted us to enrol the patient in the ALKA-372-001 phase I study at the RP2D of 400 mg m⁻² po qd of entrectinib, starting in March 2015. The patient presented at baseline with ECOG performance status 0, stable and asymptomatic CNS disease, and progressive liver metastases. The first-response assessment via CT, which was performed in April 2015, 4 weeks after the beginning of treatment, showed a partial response per RECIST v1.1 with a decrease in the sum of the target lesions by 38% (Figure 3). Computed tomography performed 4 weeks later in May 2015 confirmed this response. CNS metastases (brain and cerebellum) were stable. No drug-related adverse events were recorded, and the patient was responding and still under treatment with entrectinib as of July 2015.

DISCUSSION

In CRC, recurrent genetic lesions conferring oncogene addiction are only recently emerging as experimental therapeutic targets. These lesions are presently confined to gene amplifications and mutations, such as *HER2* (Siena *et al*, 2015b), *MET* (Bardelli *et al*, 2013), and *BRAF* (Yaeger *et al*, 2015). Activating gene rearrangements representing new potential therapeutic opportunities have only recently started to be exploited in the clinic (Ardini *et al*, 2014). We recently reported the existence of *NTRK1* rearrangements as recurrent events in CRC, and we discovered TRKA as a target in CRC by identifying an *LMNA-NTRK1* rearrangement, leading to sensitivity to treatment with entrectinib in a patient with CRC refractory to standard therapies (Ardini *et al*, 2014; Sartore-Bianchi *et al*, 2015). *ALK* rearrangements have also been reported in CRC (Aisner *et al*, 2014), but no data are available

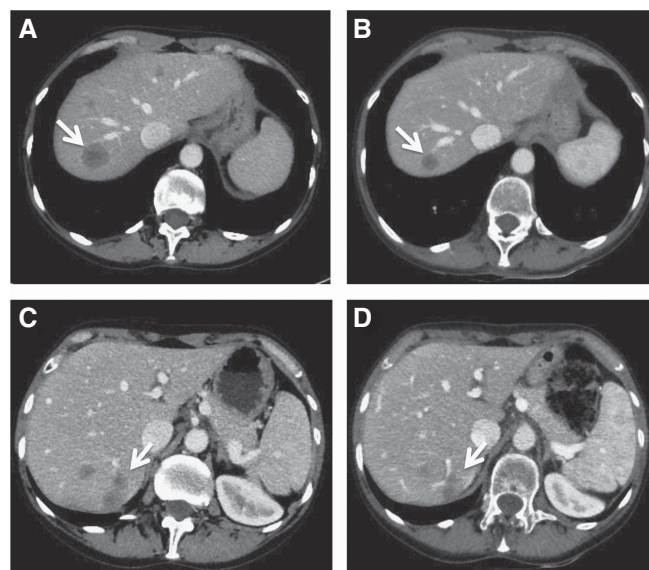


Figure 3. Computed tomography (CT) scans showing the objective tumour response to entrectinib. The baseline abdominal CT scan of March 2015 demonstrated liver involvement with the two largest lesions both in hepatic segment VII, measuring 27 and 33 mm in longest diameter, respectively (**A**, **C** arrows). At the first-response assessment, in April 2015, 4 weeks after the initiation of treatment, CT showed a RECIST partial response with an overall decrease in the sum of the target lesions of 38%, and lesions in segment VII displaying longest diameters of 15 and 22 mm (**B**, **D** arrows).

regarding the effectiveness of specific *ALK* therapeutic inhibition in this clinical setting.

We recently applied IHC as a screening strategy to detect *ALK* rearrangements in CRC, leading to the identification of a positive sample harbouring an *ALK* rearrangement, as confirmed by FISH (Medico *et al*, 2015). Here we report the molecular characterisation of this metastatic CRC, revealing a novel *CAD-ALK* gene rearrangement, and the successful treatment of this patient with entrectinib.

The *CAD* gene encodes a trifunctional protein that is associated with the enzymatic activities of the first three enzymes in the

six-step pathway of pyrimidine biosynthesis: carbamoylphosphate synthetase (CPS II), aspartate transcarbamoylase, and dihydroorotase (Grande-García *et al*, 2013). *CAD* is expressed in normal colonic tissue, and its expression is increased in inflammatory bowel disease (Richmond *et al*, 2012). *CAD* is also detected at medium-high levels in CRC by IHC (Uhlén *et al*, 2015). The *CAD-ALK* chimaeric gene encompassed *CAD* exons 1–35 fused to the canonical exon 20 recombination site that was previously reported for *ALK* gene fusions (Grande *et al*, 2011; Awad and Shaw, 2014). The chimaeric protein comprises the carbamoyl phosphate synthetase large-chain oligomerisation domain fused to the entire *ALK* kinase domain, potentially resulting in constitutive dimerisation causing *ALK* kinase activation. The chimaeric protein is expected to be sensitive to ATP competitive inhibitors such as entrectinib because it encompasses a drug-binding region identical to the corresponding wild-type *ALK* kinase. A recent IHC screening effort to detect *ALK* expression in 172 Korean CRC cases resulted in the identification of a strongly positive rectal adenocarcinoma sample, which was found to harbour a *CAD-ALK* (C35-A20) rearrangement (Lee *et al*, 2015). In addition, one of the 50 CRC patients enrolled in a pathway-directed therapeutic trial (NEXT-1) was also found to harbour an *EML4-ALK* (E21, A20) rearrangement (Lee *et al*, 2015). These data confirm the existence of *ALK* rearrangements as rare recurrent events in CRC.

Therapeutic inhibition of *EML4-ALK* with entrectinib in patients with non-small cell lung cancer has resulted in remarkable objective responses and clinical benefit (Siena *et al*, 2015a). We provide the first evidence that treatment with an *ALK* inhibitor is effective in metastatic CRC involving an *ALK* gene rearrangement. Entrectinib at RP2D rapidly induced tumour shrinkage, achieving an objective response within 4 weeks of commencing treatment, which lasted >4 months and was maintained at the time of the present report (July 2015). Although activated *ALK* rearrangements are rare events in CRC (Aisner *et al*, 2014; Medico *et al*, 2015), a screening strategy based on simple *ALK* assessment by IHC followed by targeted NGS based on anchored multiplex PCR (Zheng *et al*, 2014) represents a feasible strategy, which will enhance the identification of patients who can benefit from entrectinib treatment in CRC and other histologies (Siena *et al*, 2015b). The reason for reporting on a single case is the rarity of gene fusions in CRC; a large series for clinical studies would be possible only if pharmaceutical interest and rationale are triggered by the knowledge of successful uncommon cases such as this and other recently reported translocations (Sartore-Bianchi *et al*, 2015).

The innovation of the present report resides in the discovery of the novel *CAD-ALK* rearranged gene in CRC, whose pharmaceutical blockade with a single agent, entrectinib, led to a clinically meaningful anti-tumour effect. Thus for the first time, we provide the proof of concept that *ALK* alterations can act as drivers in CRC, building a new step towards personalised therapy in this clinical setting.

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CONFLICT OF INTEREST

ASB is a consultant/advisory member for Amgen, Bayer, and Merck-Serono. SS is a consultant/advisory member for Amgen, Bayer, Eli-Lilly, Merck-Serono, Merus, Novartis, Roche, Sanofi, and Ignyta. DL, ZH, PM, DM, and RS are employees of Ignyta. AS, RB, EA, AG and AI are employees of Nerviano Medical Sciences. The remaining authors declare no conflict of interest.

REFERENCES

- Aisner DL, Nguyen TT, Paskulin DD, Le AT, Haney J, Schulte N, Chionh F, Hardingham J, Mariadason J, Tebbutt N, Doebele RC, Weickhardt AJ, Varella-García M (2014) *ROS1* and *ALK* fusions in colorectal cancer, with evidence of intratumoral heterogeneity for molecular drivers. *Mol Cancer Res* **12**: 111–118.
- Ardini E, Bosotti R, Borgia AL, De Ponti C, Somaschini A, Cammarota R, Amboldi N, Radrizzani L, Milani A, Magnaghi P, Ballinari D, Casero D, Gasparri F, Banfi P, Avanzi N, Saccardo MB, Alzani R, Bandiera T, Felder E, Donati D, Pesenti E, Sartore-Bianchi A, Gambacorta M, Pierotti MA, Siena S, Veronese S, Galvani A, Isacchi A (2014) The *TPM3-NTRK1* rearrangement is a recurring event in colorectal carcinoma and is associated with tumor sensitivity to *TRKA* kinase inhibition. *Mol Oncol* **8**: 1495–1507.
- Ardini E, Meninchichi M, Banfi P, Casero D, Giorgini ML, Saccardo MB, Amboldi N, Avanzi N, Orsini P, Isacchi A, Presenti E, Galvani A (2013) The *ALK* inhibitor NMS-E628 also potently inhibits *ROS1* and induces tumor regression in ROS-driven models. Proceedings of the 104th Annual Meeting of the American Association for Cancer Research; 6–10 April 2013; Washington, DC. *Cancer Res* **73**(8 Suppl): Abstract no. 2092.
- Awad MM, Shaw AT (2014) *ALK* inhibitors in non-small cell lung cancer: crizotinib and beyond. *Clin Adv Hematol Oncol* **12**: 429–439.
- Bardelli A, Corso S, Bertotti A, Hobor S, Valtorta E, Siravegna G, Sartore-Bianchi A, Scala E, Cassingena A, Zecchin D, Apicella M, Migliardi G, Galimi F, Lauricella C, Zanon C, Perera T, Veronese S, Corti G, Amatu A, Gambacorta M, Diaz LA, Sausen M, Velculescu VE, Comoglio P, Trusolino L, Di Nicolantonio F, Giordano S, Siena S (2013) Amplification of the *MET* receptor drives resistance to anti-EGFR therapies in colorectal cancer. *Cancer Discov* **3**: 658–673.
- De Braud F, Pilla L, Niger M, Damian S, Bardazza B, Martinetti A, Pelosi G, Marrapese G, Palmeri L, Cera G, Valtorta E, Veronese S, Sartore-Bianchi A, Ardini E, Martignoni M, Isacchi A, Luo D, Freddo L, Siena S (2014) *RDX-101*, an oral Pan-Trk, *ROS1*, and *ALK* inhibitor, in patients with advanced solid tumors with relevant molecular alterations. *Ann Oncol* **25**: iv146–iv164.
- Eisenhauer EA, Therasse P, Bogaerts J, Schwartz LH, Sargent D, Ford R, Dancy J, Arbuck S, Gwyther S, Mooney M, Rubinstein L, Shankar L, Dodd L, Kaplan R, Lacombe D, Verweij J (2009) New response evaluation criteria in solid tumours: revised RECIST guideline (version 1.1). *Eur J Cancer* **45**: 228–247.
- Grande E, Bolós M-VV, Arriola E (2011) Targeting oncogenic *ALK*: a promising strategy for cancer treatment. *Mol Cancer Ther* **10**: 569–579.
- Grande-García A, Lallous N, Díaz-Tejada C, Ramón-Maiques S (2013) Structure, functional characterization, and evolution of the dihydroorotase domain of human *CAD*. *Structure* **22**: 185–198.
- Lee J, Kim H, Hong J, Wang K, Kim S, Jang J, Kim S, Park J, Lim H, Kang W, Park Y, Lee J, Lee W, Park Y, Huh J, Yun S, Do I-G, Kim S, Balasubramanian S, Stephens P, Ross J, Li G, Hornby Z, Ali S, Miller V, Kim K-M, Ou S-H (2015) Detection of novel and potentially actionable anaplastic lymphoma kinase (*ALK*) rearrangement in colorectal adenocarcinoma by immunohistochemistry screening. *Oncotarget* **6**(27): 24320–24332.
- Medico E, Russo M, Picco G, Cancelliere C, Valtorta E, Corti G, Buscarino M, Isella C, Lamba S, Martinoglio B, Veronese S, Siena S, Sartore-Bianchi A, Beccuti M, Mottolise M, Linnebacher M, Cordero F, Di Nicolantonio F, Bardelli A (2015) The molecular landscape of colorectal cancer cell lines unveils clinically actionable kinase targets. *Nat Commun* **6**: 7002.

- Richmond AL, Kabi A, Homer CR, Marina-García N, Nickerson KP, Nesvizhskii AI, Sreekumar A, Chinnaiyan AM, Nuñez G, McDonald C (2012) The nucleotide synthesis enzyme CAD inhibits NOD2 antibacterial function in human intestinal epithelial cells. *Gastroenterology* **142**: 1483–92.e6.
- Sartore-Bianchi A, Ardini E, Bosotti R, Amatu A, Valtorta E, Somaschini A (2015) A novel LMNA-NTRK1 gene fusion confers sensitivity to entrectinib in metastatic colorectal cancer. *J Natl Cancer Inst* **108**(1).
- Siena S, Drillon A, Ou S-H, Farago A, Patel M, Bauer T, Hong D, Liu S, Lee J, Patel R, Schechet L, Luo D, Chow Maneval E, Multani P, De Braud F (2015a) Entrectinib (RXDX-101), an oral pan-Trk, ROS1, and ALK inhibitor in patients with advanced solid tumors harboring gene rearrangements. *Eur J Cancer* **51**(Suppl 3): S724–S725.
- Siena S, Sartore-Bianchi A, Lonardi S, Trusolino L, Martino C, Bencardino K, Leone F, Zagonel V, Torri V, Siravegna G, Amatu A (2015b) Trastuzumab and lapatinib in HER2-amplified metastatic colorectal cancer patients (mCRC): The HERACLES trial. *J Clin Oncol* **33**: 3508.
- Siravegna G, Mussolin B, Buscarino M, Corti G, Cassingena A, Crisafulli G, Ponzetti A, Cremolini C, Amatu A, Lauricella C, Lamba S, Hobor S, Avallone A, Valtorta E, Rospo G, Medico E, Motta V, Antoniotti C, Tatangelo F, Bellosillo B, Veronese S, Budillon A, Montagut C, Racca P, Marsoni S, Falcone A, Corcoran RB, Di Nicolantonio F, Loupakis F, Siena S, Sartore-Bianchi A, Bardelli A (2015) Clonal evolution and resistance to EGFR blockade in the blood of colorectal cancer patients. *Nat Med* **21**: 795–801.
- Uhlén M, Fagerberg L, Hallström BMM, Lindskog C, Oksvold P, Mardinoglu A, Sivertsson Å, Kampf C, Sjöstedt E, Asplund A, Olsson I, Edlund K, Lundberg E, Navani S, Szigarty CA, Odeberg J, Djureinovic D, Takanen JO, Hober S, Alm T, Edqvist P-HH, Berling H, Tegel H, Mulder J, Rockberg J, Nilsson P, Schwenk JM, Hamsten M, von Feilitzen K, Forsberg M, Persson L, Johansson F, Zwahlen M, von Heijne G, Nielsen J, Pontén F (2015) Proteomics. Tissue-based map of the human proteome. *Science* **347**: 1260419.
- Valtorta E, Martino C, Sartore-Bianchi A, Penault-Llorca F, Viale G, Risio M, Rugge M, Grigioni W, Lonardi S, Zagonel V, Leone F, Noe J, Ciardiello F, Pinto C, Labianca R, Mosconi S, Graiff C, Aprile G, Frau B, Garufi C, Loupakis F, Racca P, Tonini G, Lauricella C, Veronese S, Truini M, Siena S, Marsoni S, Gambacorta M (2015) Assessment of a HER2 scoring system for colorectal cancer: results from a validation study. *Mod Pathol* **28**(11): 1481–1491.
- Yaeger R, Cercek A, O'Reilly EM, Reidy DL, Kemeny N, Wolinsky T, Capanu M, Gollub MJ, Rosen N, Berger MF, Lacouture ME, Vakiani E, Saltz LB (2015) Pilot trial of combined BRAF and EGFR inhibition in BRAF-mutant metastatic colorectal cancer patients. *Clin Cancer Res* **21**: 1313–1320.
- Zheng Z, Liebers M, Zhelyazkova B, Cao Y, Panditi D, Lynch KD, Chen J, Robinson HE, Shim HS, Chmielecki J, Pao W, Engelman JA, Iafrate AJ, Le LP (2014) Anchored multiplex PCR for targeted next-generation sequencing. *Nat Med* **20**: 1479–1484.



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