

## ***In vitro* activity of econazole in comparison with three common antifungal agents against clinical *Candida* strains isolated from superficial infections**

Abastabar M<sup>1</sup>, Shokohi T<sup>1\*</sup>, Rouhi Kord R<sup>2</sup>, Badali H<sup>1</sup>, Hashemi SJ<sup>3</sup>, Ghasemi Z<sup>3</sup>, Ghoghghi A<sup>4</sup>, Baghi N<sup>2</sup>, Abdollahi M<sup>2,5</sup>, Hosseinpour S<sup>2</sup>, Rahimi N<sup>2</sup>, Seifi Z<sup>2</sup>, Gholami S<sup>1</sup>, Haghani I<sup>1</sup>, Jabari MR<sup>2</sup>, Pagheh A<sup>2,4</sup>

<sup>1</sup> Invasive Fungi Research Center (IFRC), Department of Medical Mycology and Parasitology, School of Medicine, Mazandaran University of Medical Sciences, Sari, Iran

<sup>2</sup> Student Research Committee, Mazandaran University of Medical Sciences, Sari, Iran

<sup>3</sup> Department of Medical Parasitology and Mycology, School of Medicine, Tehran University of Medical Sciences, Tehran, Iran

<sup>4</sup> Toxoplasmosis Research Center, Mazandaran University of Medical Sciences, Sari, Iran

<sup>5</sup> Department of Microbiology and Immunology, School of Medicine, Mazandaran University of Medical Sciences, Sari, Iran

\*Corresponding author: **Tahereh Shokohi**, Invasive Fungi Research Center (IFRC)/ Department of Medical Mycology and Parasitology, School of Medicine, Km 18 Khazarabad Road, Iran. Email: shokohi.tahereh@gmail.com

(Received: 23 November 2015; Revised: 22 December 2015; Accepted: 26 December 2015)

### **Abstract**

**Background and Purpose:** *Candida* species are the most common organisms involved in superficial fungal infections, worldwide. Although econazole is among the most frequently used topical formulations for the treatment of candidiasis, no information is available regarding the susceptibility profiles of *Candida* species in Iran.

**Materials and Methods:** *In vitro* susceptibility of 100 clinical *Candida* isolates belonging to 6 species from superficial candidiasis of Iran towards to econazole was compared with three other common antifungal agents including itraconazole, fluconazole, and miconazole. Minimum inhibitory concentrations (MICs) values were analyzed according to the Clinical and Laboratory Standards Institute (CLSI) M38-A3 document. All isolates were previously identified to the species level, using polymerase chain reaction-restriction fragment length polymorphism (PCR-RFLP) on ITS region.

**Results:** The MIC of econazole, itraconazole, miconazole, and fluconazole were within the range of 0.016-16, 0.032-16, 0.016-16, and 0.25-64 µg/ml, respectively. In general, econazole and miconazole were more active against *Candida* isolates, compared to the other two agents.

**Conclusion:** The present study demonstrated that for *Candida albicans* isolates, miconazole and econazole had the best effect, but in non-*albicans* *Candida* species, itraconazole and miconazole displayed more activity than other antifungal agents.

**Keywords:** Antifungal, Econazole, Candidiasis

#### ➤ How to cite this paper:

Abastabar M, Shokohi T, Rouhi Kord R, Badali H, Hashemi SJ, Ghasemi Z, Ghoghghi A, Baghi N, Abdollahi M, Hosseinpour S, Rahimi N, Seifi Z, Gholami S, Haghani I, Jabari MR, Pagheh A. *In vitro* activity of econazole in comparison with three common antifungal agents against clinical *Candida* strains isolated from superficial infections. *Curr Med Mycol*. 2015; 1(4): 7-12. DOI: [10.18869/acadpub.cmm.1.4.7](https://doi.org/10.18869/acadpub.cmm.1.4.7)

### **Introduction**

Superficial mycoses are among the most prevalent fungal infections, worldwide. These infections are caused by various fungi including *Candida* species, dermatophytes, and rarely other pathogens [1]. *Candida albicans* is currently the main causative agent in the majority of superficial candidiasis, followed by non-*Candida albicans* species, *C. tropicalis*, *C. glabrata*, *C. parapsilosis*, and *C. krusei* [1].

Over the past decades, there has been a remarkable increase in the number of infections caused by *Candida* species mainly due to the rising number of immunocompromised hosts

such as transplant recipients, diabetic patients, and HIV-infected individuals [1, 2]. Oral and/or topical formulations of fluconazole, itraconazole, miconazole, clotrimazole, amphotericin B, and nystatin are usually the treatment of choice for different types of superficial candidiasis.

Azole-based therapy is the preferred treatment option, despite recent reports on the resistance of *Candida* species to these agents as the most prevalent type of antifungal resistance [3, 4]. In fact, widespread use of these antifungal agents for prophylaxis and treatment of *Candida* infections has resulted in the

emergence of resistant *Candida* species [5].

Acquired resistance to fluconazole has been reported in *C. albicans* isolates from patients with advanced AIDS, receiving prolonged azole treatment [6, 7]. As evidences from different recent studies suggest, there has been a shift towards fluconazole resistance in other *Candida* species, particularly *C. glabrata* [8-10]. Generally, the rate of azole resistance among *Candida* species is vague and variable in Iran, with the reported rates ranging from 10% to 85% for fluconazole and 12% to 62% for itraconazole [11-16].

Although a large number of azole-based antifungal agents have been used by this time for the treatment of candidiasis, further researches are required regarding the increasing resistance to these agents in Iran. Econazole is among the most frequently used topical formulations for the treatment of candidiasis [17]. However, econazole has not been used in Iran for the treatment of patients and the susceptibility profiles of different fungi against these agents have not been yet identified.

In view of the aforementioned background, in this study, we aimed to compare the *in vitro* activity of econazole with three common antifungal agents including fluconazole (FLC), itraconazole (ITC), and miconazole (MIC) against 100 clinical *Candida* strains belonging to six different species (i.e., *C. albicans*, *C. glabrata*, *C. parapsilosis*, *C. tropicalis*, *C. krusei*, and *C. guilliermondii*) isolated from patients with superficial candidiasis in Iran.

## Material and Methods

### *Fungal isolates*

Clinical strains were isolated from different specimens obtained from patients with superficial candidiasis, referred to the medical mycology laboratory of Razi Hospital in Tehran, Iran during 2014-2015. All clinical isolates were previously identified, using phenotypic criteria and polymerase chain reaction-restriction fragment length polymorphism (PCR-RFLP) of internal transcribed spacer (ITS) region of rDNA [18].

In total, 100 *Candida* isolates belonging to six species including *C. albicans* (n=67), *C. glabrata* (n=4), *C. parapsilosis* (n=15), *C.*

*tropicalis* (n=10), *C. krusei* (n=2), and *C. guilliermondii* (n=2) were identified. To ensure purity and viability, the isolates were cultured on Sabouraud dextrose agar (Difco Laboratories, Detroit, MI, USA) and incubated at 35°C until use.

### *In vitro susceptibility testing*

The MIC of antifungal agents was determined for *Candida* isolates, using the reference procedure described by the Clinical and Laboratory Standards Institute (CLSI) in accordance with guideline M38-A3 [19]. *C. krusei* (ATCC 6258) and *C. parapsilosis* (ATCC 22019) strains were used as the controls.

The reference powders of fluconazole (Pfizer), itraconazole, econazole, and miconazole (Sigma, St. Louis, MO, USA) were obtained from respective manufacturers. Fluconazole was dissolved in sterile distilled water, while other agents were prepared in 100% dimethyl sulfoxide. All agents were diluted in RPMI 1640 medium (Sigma Chemical Co.), supplemented with L-glutamine, without sodium bicarbonate buffered at pH 7.0 with 0.165 M morpholinepropanesulfonic acid (MOPS) (Sigma, Aldrich Chemie) to provide the following concentrations: 0.016–16 mg/ml for econazole, itraconazole, and miconazole and 0.064–64 for fluconazole.

### *Inoculum preparation*

The inoculum of all *Candida* species was prepared from 24 h cultures, grown on Sabouraud dextrose agar (SDA, Difco Laboratories, Detroit, MI, USA) at 35°C. At first, several colonies were picked and suspended in 5 mL of sterile saline. The obtained suspension was vortexed for few seconds by a mixer. The cell densities were measured by a spectrophotometer at a wavelength of 530 nm, and the transmission was adjusted to 75–77%.

The suspension was diluted in RPMI 1640 medium to yield a final inoculum concentration ranged from 0.5-2.5 ×10<sup>3</sup> CFU/ml. MIC was determined after incubation of the 96-well microplates at 35 °C for 24-48 hours as the 80% or more of reduction of growth for all agents compared to the growth rate of the drug-free control well.

### Data analysis

MIC values were measured for antifungal agents and presented as geometric mean (GM), MIC range, MIC<sub>50</sub>, and MIC<sub>90</sub>. All the tests were performed in duplicate.

### Results

Table 1 summarizes the *in vitro* susceptibility of 100 isolates from six *Candida* species to fluconazole, itraconazole, econazole, and miconazole. In the present study, MIC<sub>50</sub>, MIC<sub>90</sub>, GM, and MIC range of all *C. albicans* isolates were determined. Since the number of *C. tropicalis*, *C. parapsilosis*, *C. glabrata*, *C. krusei*, and *C. guilliermondii* isolates was insufficient, MIC<sub>50</sub> and MIC<sub>90</sub> values were not measured for these species.

The MICs of econazole, itraconazole, miconazole, and fluconazole were within the range of 0.016-16, 0.032-16, 0.016-16, and 0.25-64 µg/ml, respectively. The rates of resistance to fluconazole (MIC<sub>≥</sub> 8 µg/ml), itraconazole (MIC<sub>≥</sub> 1 µg/ml), and miconazole (MIC<sub>≥</sub>1 µg/ml) were 57% (n=57), 69% (n=69), and 54% (n=54), respectively. However, the rate of resistance to econazole could not be calculated, given the absence of published interpretive criteria for this agent.

Overall, among *C. albicans* isolates, miconazole and econazole were more active than the two other used agents. As presented in Table 1, MIC<sub>50</sub> values of miconazole and econazole against *C. albicans* isolates were 4 and 8 µg/ml, while MIC<sub>50</sub> values of itraconazole and fluconazole were 16 and 64 µg/ml, respectively. Also, GM values of miconazole and econazole against *C. albicans* isolates were 0.75 and 1.16 µg/ml, while GM values for itraconazole and fluconazole were 5.34 and 13.45 µg/ml, respectively.

Based on the findings, miconazole, followed by econazole showed the best activity against *C. albicans* isolates (n=67). However, for all non-*Candida albicans* species (n=33), itraconazole and miconazole displayed better activity, compared to econazole and fluconazole. In addition, itraconazole was more active against *C. tropicalis* and *C. parapsilosis* isolates, compared to the other three agents.

Considering the MIC values of all agents, *C. albicans* isolates were more resistant than other species (Table 1). Among all isolates, *C. krusei* species was fully resistant to all used azoles, while *C. tropicalis* was the most sensitive species.

### Discussion

Despite the fact that more than 100 fungal species have been identified as important clinical pathogens causing superficial to life-threatening mycoses, infections due to *Aspergillus* and *Candida* species are the most common. Among these infections, superficial candidiasis is generally community-acquired and is considered responsible for remarkable morbidity. These infections are frequently caused by *Candida albicans* and non-*Candida albicans* species such as *C. tropicalis*, *C. glabrata*, *C. parapsilosis*, and *C. krusei* [1, 2].

The incidence of infections caused by azole-resistant *Candida* and non-*Candida albicans* species has increased over the past decades owing to the excessive use of azoles, especially triazoles such as fluconazole [20-22]. Considering the scarcity of available data on the susceptibility profiles of *Candida* species to econazole in Iran, in the present study, *in vitro* activity of 100 clinical *Candida* isolates from six species from patients with superficial candidiasis towards econazole was compared with three common antifungal agents including fluconazole, itraconazole, and miconazole.

In a previous study by Salehei et al. in Iran, the susceptibility patterns of vaginal *Candida* isolates to eight antifungal drugs including clotrimazole, miconazole, itraconazole, fluconazole, ketoconazole, econazole, nystatin, and terbinafine were determined, using disk diffusion method. They found that the highest sensitivity of *C. albicans* to antifungal drugs was observed against miconazole, whereas 43 (81%) isolates were resistant to fluconazole and econazole antifungals [16]. Similarly, Al-Mamari [23] using disk diffusion reported that the highest sensitivity of *C. albicans* was seen against miconazole (95%) whereas 73 isolates (78%) were resistant to fluconazole and econazole antifungals.

**Table 1.** *In vitro* antifungal susceptibilities of 100 clinical *Candida* isolates against four antifungal agents

Isolate	Antifungal Agent	MIC ( $\mu\text{g} / \text{ml}$ )												MIC range	MIC 50	MIC 90	G mean	Resistant (Number)	
		0.016	0.032	0.0625	0.125	0.25	0.5	1	2	4	8	16	32						64
<i>Candida albicans</i> (n=67)	ECO	0	1	1	1	2	3	10	10	1	15	23			0.016-16	8	16	1.16	-
	ITR	0	0	0	3	3	2	5	2	3	3	46			0.125-16	16	16	5.34	57
	MIC	2	0	0	4	1	3	5	6	5	1	30			0.016-16	4	16	0.75	43
	FLU			0	0	3	2	4	5	9	1	0	5	38	0.25-64	64	64	13.45	46
<i>Candida parapsilosis</i> (n=15)	ECO	0	0	0	1	1	1	2	5	2	1	2			0.0625-16	2		1.56	-
	ITR	0	1	2	0	4	3	1	2	0	0	2			0.032-16	0.5		0.45	3
	MIC	1	0	0	1	0	5	3	2	1	1	2			0.016-16	1		0.7	2
	FLU			0	0	1	2	5	2	2	0	1	1	2	0.25-64	1		1.81	2
<i>Candida tropicalis</i> (n=10)	ECO	0	0	0	0	0	2	5	0	1	0	2			0.5-16	1		1.7	-
	ITR	0	0	0	0	1	2	1	1	0	0	1			0.125-16	0.5		0.46	3
	MIC	0	0	0	0	0	2	4	1	4	0	1			0.5-16	1		1.62	3
	FLU			0	0	0	0	5	1	1	2	0	0	1	1-64	1		2.82	3
<i>Candida glabrata</i> (n=4)	ECO	0	0	0	0	0	0	0	2	0	1	1			2-16	-		4.75	-
	ITR	0	0	0	0	1	0	1	0	0	1	1			0.25-16	-		2.37	3
	MIC	0	0	0	0	0	0	1	0	2	0	1			1-16	-		4	3
	FLU			0	0	0	0	0	0	1	0	0	1	2	4-64	-		26.90	2
<i>C. guilliermondi</i> (n=2)	ECO	0	0	0	0	0	0	0	1	0	0	1			2-16	-		5.65	-
	ITR	0	0	0	0	0	0	0	1	0	0	1			0.32-16	-		5.65	2
	MIC	0	0	0	0	0	0	1	0	0	0	1			0.016-16	-		4	1
	FLU			0	0	0	0	0	0	0	0	0	1	1	0.25-64	-		45.25	2
<i>C. krusei</i> (n=2)	ECO	0	0	0	0	0	0	0	0	0	0	2			16	-		16	-
	ITR	0	0	0	0	0	0	0	1	1	0	0			2-4	-		2.82	2
	MIC	0	0	0	0	0	0	0	0	0	0	2			16	-		16	2
	FLU			0	0	0	0	0	0	0	0	0	1	1	32-64	-		45.25	2
<i>All Candidia spp.</i> (n=100)	ECO	0	1	1	2	3	6	17	18	4	17	31			0.016-16	-		3.11	-
	ITR	0	1	2	3	9	7	8	7	4	4	55			0.032-16	-		2.7	69
	MIC	1	0	0	5	1	10	14	9	12	12	36			0.016-16	-		2.4	54
	FLU	0	0	0	0	4	4	14	8	13	3	1	9	44	0.25-64	-		10.43	57

In the current study, based on the microdilution method, miconazole showed the lowest MIC, while fluconazole exhibited the highest MIC value. Previous studies in Iran, comparing the efficacy of fluconazole and other agents against *Candida* species of superficial infections, have reported inconsistent results [24-26]. In studies by Katirae et al. [24], Pakshir et al. [25], Badiie et al. [26], and Shokohi et al. [27], resistance of *C. albicans* isolates to fluconazole was estimated at 25.7%, 26%, 10%, and 2.6%, respectively.

On the other hand, in a study by Gross et al. [28], 96.5% and 98% of *C. albicans* isolates were susceptible to fluconazole and itraconazole, respectively; however, we

observed a higher rate of fluconazole resistance (46%). In the current study, all *C. krusei* isolates were resistant to the evaluated azoles, whereas in the study by Güzel et al., which evaluated vaginitis isolates [29], resistance rates of 57.1% and 32.1% against fluconazole and itraconazole were reported, respectively; also, all isolates were sensitive to miconazole.

Based on studies by Badiie et al. [26] and El Feky et al. [30], 17.4% and 40% of *C. krusei* isolates were resistant to fluconazole, respectively; conversely, these isolates were susceptible to other azoles. Similar to a study by Shokohi et al. on oropharyngeal lesions isolates [27], we reported *Candida* isolates including *C. albicans* (n=36), *C. glabrata* (n=2), *C. krusei*

(n=2), *C. guilliermondii* (n=1), *C. tropicalis* (n=1), and *C. parapsilosis* (n=1), which were fully resistant to four antifungals.

In agreement with studies by Badiee et al. [26] and El Feky et al., *C. tropicalis* was the most susceptible species to all antifungal agents in the present study. In total, azole resistance was more prevalent in non-*C. albicans* species, particularly *C. glabrata*, *C. krusei*, and *C. guilliermondii* as compared to *C. albicans*.

In contrast with a study by Salehei et al. in Iran [16], in the present study, two azoles, i.e., econazole and miconazole, typically showed better activity against all the isolates, compared to the other azoles. In fact, 42% of the isolates were susceptible to miconazole, while susceptibility to fluconazole and itraconazole was estimated at 40% and 28%, respectively.

Although econazole is among the most commonly used topical formulations for the treatment of candidiasis and dermatophytosis, this agent has not been routinely used in Iran. Based on our findings, econazole, similar to miconazole, showed suitable activity with a MIC range of 0.016-16 µg/ml. In summary, this study revealed that econazole is more potent than fluconazole and itraconazole against all *Candida* species.

## Conclusion

Based on the findings, it can be concluded that econazole is a suitable alternative choice for treatment of Iranian isolates of candida isolated from superficial infections.

## Acknowledgments

This study was funded by the Invasive Fungi Research Center (IFRC) at Faculty of Medicine, Mazandaran University of Medical Sciences, Sari, Iran (No.: 760). This research was approved by the ethics committee of the university (ethical code: IR.MAZUMS.REC.94).

## Authors' Contributions

M.A. and T.S. designed and supervised the research, M.A., T.S., H.B. and S.J.H. edited the final manuscript and R.R., Z.G., I.H., S.P., A.G., N.B., M.A., S.H., N.R., M.R.J., S.G. and Z.S. performed the tests.

## Conflicts of Interest

The authors report no conflicts of interest. The authors alone are responsible for the content and writing of the paper.

## Financial Disclosure

The authors declare no financial interests related to the materials of the study.

## References

1. Dias MF, Quaresma-Santos MV, Bernardes-Filho F, Amorim AG, Schechtman RC, Azulay DR. Update on therapy for superficial mycoses: review article part I. *An Bras Dermatol*. 2013; 88(5):764–74.
2. Zaoutis TE, Argon J, Chu J, Berlin JA, Walsh TJ, Feudtner C. The epidemiology and attributable outcomes of candidemia in adults and children hospitalized in the United States: a propensity analysis. *Clin Infect Dis*. 2005; 41(9):1232–9.
3. Kanafani ZA, Perfect JR. Antimicrobial resistance: resistance to antifungal agents: mechanisms and clinical impact. *Clin Infect Dis*. 2008; 46(1):120-8.
4. Goldman M, Cloud GA, Smedema M, LeMonte A, Connolly P, McKinsey DS, et al. Does long-term itraconazole prophylaxis result in in vitro azole resistance in mucosal *Candida albicans* isolates from persons with advanced human immunodeficiency virus infection? The National Institute of Allergy and Infectious Diseases Mycoses study group. *Antimicrob Agents Chemother*. 2000; 44(6):1585–7.
5. Sheikh N, Jahagirdar V, Kothadia S, Nagoba B. Antifungal drug resistance in *Candida* species. *Eur J Gen Med*. 2013; 10(4):254-8.
6. Mulu A, Kassu A, Anagaw B, Moges B, Gelaw A, Alemayehu M, et al. Frequent detection of 'azole' resistant *Candida* species among late presenting AIDS patients in northwest Ethiopia. *BMC Infect Dis*. 2013; 13:82.
7. Rosana Y, Yasmon A, Lestari DC. Overexpression and mutation as a genetic mechanism of fluconazole resistance in *Candida albicans* isolated from human immunodeficiency virus patients in Indonesia. *J Med Microbiol*. 2015; 64(9):1046-52.
8. Pfaller MA, Diekema DJ, Sheehan DJ. Interpretive breakpoints for fluconazole and *Candida* revisited: a blueprint for the future of antifungal susceptibility testing. *Clin Microbiol Rev*. 2006; 19(2):435–47.
9. Lee I, Fishman NO, Zaoutis TE, Morales KH, Weiner MG, Synnestvedt M, et al. Risk factors for fluconazole-resistant *Candida glabrata* bloodstream infections. *Arch Intern Med*. 2009; 169(4):379-83.
10. Farmakiotis D, Tarrand JJ, Kontoyiannis DP. Drug-resistant *Candida glabrata* infection in cancer patients. *Emerg Infect Dis*. 2014; 20(11):1833-40.
11. Shekari Ebrahim Abad H, Zaini F, Kordbacheh P, Mahmoudi M, Safara M, Mortezaee V. In vitro activity of caspofungin against fluconazole-resistant *Candida* species isolated from clinical samples in

- Iran. Jundishapur J Microbiol. 2015; 8(6):e18353.
12. Mohamadi J, Motaghi M, Panahi J, Havasian MR, Delpisheh A, Azizian M, et al. Anti-fungal resistance in candida isolated from oral and diaper rash candidiasis in neonates. *Bioinformation*. 2014; 10(11):667-70.
  13. Haddadi P, Zareifar S, Badiie P, Alborzi A, Mokhtari M, Zomorodian K, et al. Yeast colonization and drug susceptibility pattern in the pediatric patients with neutropenia. *Jundishapur J Microbiol*. 2014; 7(9):e11858.
  14. Badiie P, Alborzi A, Davarpanah MA, Shakiba E. Distributions and antifungal susceptibility of *Candida* species from mucosal sites in HIV positive patients. *Arch Iran Med*. 2010; 13(4):282-7.
  15. Jafari-nodoushan AA, Kazemi A, Mirzaii F, Dehghani M. Fluconazole susceptibility profile of *Candida* isolates recovered from patients specimens admitted to Yazd central laboratory. *Iran J Pharm Res*. 2010; 7(1):69-75.
  16. Salehei Z, Seifi Z, Mahmoudabadi A. Sensitivity of vaginal isolates of *Candida* to eight antifungal drugs isolated from Ahvaz, Iran. *Jundishapur J Microbiol*. 2012; 5(4):574-7.
  17. Dias MF, Bernardes-Filho F, Quaresma-Santos MV, Amorim AG, Schechtman RC, Azulay DR. Treatment of superficial mycoses: review. Part II. *An Bras Dermatol*. 2013; 88(6):937-44.
  18. Mohammadi R, Badiie P, Badali H, Abastabar M, Safa AH, Hadipour M, et al. Use of restriction fragment length polymorphism to identify *Candida* species, related to onychomycosis. *Adv Biomed Res*. 2015; 4(1):95.
  19. Clinical and Laboratory Standards Institute. Reference method for broth dilution antifungal susceptibility testing of yeasts; approved standard. 3<sup>rd</sup> ed. Wayne, PA: Clinical and Laboratory Standards Institute; 2008.
  20. Bulik CC, Sobel JD, Nailor MD. Susceptibility profile of vaginal isolates of *Candida albicans* prior to and following fluconazole introduction - impact of two decades. *Mycoses*. 2011; 54(1):34-8.
  21. Chakrabarti A, Chatterjee SS, Rao KL, Zameer MM, Shivaprakash MR, Singhi S, et al. Recent experience with fungaemia: change in species distribution and azole resistance. *Scand J Infect Dis*. 41(4):275–84.
  22. Trick WE, Fridkin SK, Edwards JR, Hajjeh RA, Gaynes RP. Secular trend of hospital-acquired candidemia among intensive care unit patients in the United States during 1989–1999. *Clin Infect Dis*. 2002; 35(5):627–30.
  23. Al-mamari A, Al-buryhi M, Al-heggami MA, Al-hag S. Identify and sensitivity to antifungal drugs of *Candida* species causing vaginitis isolated from vulvovaginal infected patients in Sana'a city. *Der Pharma Chemica*. 2014; 6(1):336-42.
  24. Katirae F, Khosravi AR, Khalaj V, Hajiabdolbaghi M, Khaksar AA, Rasoulinejad M. In vitro antifungal susceptibility of oral candida species from Iranian HIV infected patients. *Tehran Univ Med J*. 2012; 70(2):96-103.
  25. Pakshir K, Akbarzadeh MA, Bonyadpour B, Mohagheghzadeh AA. In vitro activity and comparison of clotrimazol, fluconazol and nystatin against *Candida* vaginitis isolates in Shiraz, 2008. *J Rafsanjan Univ Med Sci*. 2010; 9(3):210-20.
  26. Badiie P, Alborzi A, Davarpanah MA, Shakiba E. Distributions and antifungal susceptibility of *Candida* species from mucosal sites in HIV positive patients. *Arch Iran Med*. 2010; 13(4):282-7.
  27. Shokohi S, Bandalizadeh Z, Hedayati MT, Mayahi S. In vitro antifungal susceptibility of *Candida* species isolated from oropharyngeal lesions of patients with cancer to some antifungal agents. *Jundishapur J Microbiol*. 2011; 4(2):S19-26.
  28. Gross NT, Arias ML, Moraga M, Baddasarow Y, Jarstrand C. Species distribution and susceptibility to azoles of vaginal yeasts isolated prostitutes. *Infect Dis Obstet Gynecol*. 2007; 2007:82412.
  29. Güzel AB, Aydın M, Meral M, Kalkanci A, Ilkit M. Clinical characteristics of Turkish women with *Candida krusei* vaginitis and antifungal susceptibility of the *C. krusei* isolates. *Infect Dis Obstet Gynecol*. 2013; 2013:698736.
  30. ElFeky DS, Gohar NM, El-Seidi EA, Ezzat MM, AboElew SH. Species identification and antifungal susceptibility pattern of *Candida* isolates in cases of vulvovaginal candidiasis. *Alex J Med*. 2015; In Press.