Potential roads for reaching the summit: an overview on target therapies for high-grade gliomas

Alice Giotta Lucifero¹, Sabino Luzzi^{1,2}, Ilaria Brambilla³, Lucia Schena.³, Mario Mosconi⁴, Thomas Foiadelli³, Salvatore Savasta³

¹ Neurosurgery Unit, Department of Clinical-Surgical, Diagnostic and Pediatric Sciences, University of Pavia, Pavia, Italy; ² Neurosurgery Unit, Department of Surgical Sciences, Fondazione IRCCS Policlinico San Matteo, Pavia, Italy; ³ Pediatric Clinic, Department of Pediatrics, Fondazione IRCCS Policlinico San Matteo, University of Pavia, Pavia, Italy; ⁴ Orthopaedic and Traumatology Unit, Department of Clinical-Surgical, Diagnostic and Pediatric Sciences, University of Pavia, Pavia, Italy

Abstract. Background: The tailored targeting of specific oncogenes represents a new frontier in the treatment of high-grade glioma in the pursuit of innovative and personalized approaches. The present study consists in a wide-ranging overview of the target therapies and related translational challenges in neuro-oncology. Methods: A review of the literature on PubMed/MEDLINE on recent advances concerning the target therapies for treatment of central nervous system malignancies was carried out. In the Medical Subject Headings, the terms "Target Therapy", "Target drug" and "Tailored Therapy" were combined with the terms "Highgrade gliomas", "Malignant brain tumor" and "Glioblastoma". Articles published in the last five years were further sorted, based on the best match and relevance. The ClinicalTrials.gov website was used as a source of the main trials, where the search terms were "Central Nervous System Tumor", "Malignant Brain Tumor", "Brain Cancer", "Brain Neoplasms" and "High-grade gliomas". Results: A total of 137 relevant articles and 79 trials were selected. Target therapies entailed inhibitors of tyrosine kinases, PI3K/AKT/mTOR pathway, farnesyl transferase enzymes, p53 and pRB proteins, isocitrate dehydrogenases, histone deacetylases, integrins and proteasome complexes. The clinical trials mostly involved combined approaches. They were phase I, II, I/II and III in 33%, 42%, 16%, and 9% of the cases, respectively. Conclusion: Tyrosine kinase and angiogenesis inhibitors, in combination with standard of care, have shown most evidence of the effectiveness in glioblastoma. Resistance remains an issue. A deeper understanding of the molecular pathways involved in gliomagenesis is the key aspect on which the translational research is focusing, in order to optimize the target therapies of newly diagnosed and recurrent brain gliomas. (www.actabiomedica.it)

Key words: Glioblastoma; Malignant Brain Tumors; Neuro-Oncology; Target Therapy; Tyrosine Kinase Inhibitors.

Background

High-grade gliomas, with glioblastoma (GBM) being the progenitor, are the most lethal primary brain tumors of all because of the certainty of recurrence and mortality.¹⁻⁴ As a matter of fact, the median overall survival is no longer than 15 months, despite current multimodality treatment including surgery, radiotherapy and chemotherapy.^{5,6}

The significant resistance of GBM to therapy is related to the heterogeneous genetic landscape of the tumor. High-grade gliomas harbor recurrent molecular abnormalities which are involved in the maintenance of the cell's cycle and growth, the tumor microenvironment, pathological angiogenesis, DNA repair and apoptosis.⁷⁻¹⁰

Advances in genetics and the studies of epigenetics in many pathologies affecting the central nervous system (CNS) have allowed the molecular characterization, as well as the identification of the anomalies in the cellular signaling pathways¹¹⁻¹⁴. The same insights have been of utmost importance also in neuro-oncological field, GBM first, where they led to a better understanding of tumor progression and cancer drug escape.¹⁵⁻²⁰ A deeper understanding of the malignant GBM phenotype has recently improved the knowledge about the biology of cancer, which is the starting point for identifying specific biomarkers and for developing new agents for targeting specific steps in the transduction pathways of glioma cells.²¹ Novel tailored therapies include drugs aimed at counteracting the effects of the neoplastic genetic deregulation, pathological angiogenesis and growth factor receptors; the latter with their downstream signaling pathways.

An overview of the target therapeutic strategies and challenges in developing effective agents is reported as follows.

Methods

The search of the literature was performed on the PubMed/MEDLINE (https://pubmed.ncbi.nlm. nih.gov) search engine, with combinations of Medical Subject Headings (MeSH) terms and text words, and on the ClinicalTrials.gov website (https://clinicaltrials.gov). The MeSH terms "Target Therapy", "Target drug" and "Tailored Therapy" were combined with the MeSH terms "High-grade gliomas", "Malignant brain tumor" and "Glioblastoma". In addition to original articles, our research involved reviews and editorials. The sorting of articles was carried out focusing on the most relevant studies chosen according to titles and abstracts.

On the ClinicalTrials.gov database the texts words "Central Nervous System Tumor", "Malignant Brain Tumor", "Brain Cancer", "High-grade gliomas" and "Brain Tumor" were used for the field "condition/disease". Only trials regarding target therapies, without restrictions for localization, study phase and recruitment status were selected. Filtering included articles published in the last five years, in English or translated into English. A descriptive analysis was provided.

Results

1. Volume of the Literature

The search retrieved a total of 178 articles and 148 clinical trials. After the implementation of the exclusion criteria and removal of duplicates, 137 articles and 79 randomized and non-randomized clinical trials were collected.

About the clinical trials, 33% were phase I, 42% phase II, 16% phase I/II and 9% phase III (Graph 1). Table 1 summarizes the most relevant clinical trials on target therapies for high-grade gliomas (Table 1).

2. Classification of The Target Therapies

The target therapies are mostly categorized according to the targets, which, in their turn, include molecular alterations and oncogenic signaling. The



Graph 1. Pie graph showing the distribution of the selected clinical trials according to the study phase.

#	ClinicalTrials.gov Identifier	Conditions	# of Patients Enrollment	Interventions	Study Phase	Status	Locations
1	NCT00025675	Brain and Central Nervous System Tumors	105	Gefitinib	2	Completed	USA
2	NCT00016991	Brain and Central Nervous System Tumors	53	Gefitinib	2	Completed	USA
3	NCT00238797	Glioblastoma Multiforme	36	Gefitinib	2	Completed	SW
4	NCT00027625	Brain and Central Nervous System Tumors	n/a	Gefitinib, Temozolomide	1	Completed	USA
5	NCT00418327	Malignant Brain Tumor	48	Erlotinib	1	Completed	FR
6	NCT00201419	Glioblastoma Multiforme	11	E al a cha th	1.0	0 1.1	USA
6	NC100301418	Anaplastic Astrocytoma		Erlotinib	1, 2	Completed	
7	NCT00086879	Brain and Central Nervous System Tumors	110	Carmustine, Erlotinib, Temozolomide	2	Completed	BE, FR, IT, NL, UK
8	NCT01591577	Newly Diagnosed Glioblas- toma Multiforme	50	Lapatinib, Temozolomide, Radiotherapy	2	Completed	USA
9	NCT00099060	Brain and Central Nervous System Tumors	24	Lapatinib	1, 2	Completed	CN
10	NCT02423525	Brain Cancer	24	Afatinib	1	Completed	USA
11	NCT00977431	Glioblastoma Multiforme	36	Afatinib, Te- mozolomide, Radiotherapy	1	Completed	UK
10	NCT01 C20070	Glioblastoma Multiforme	40	D		Completed	EC
12	NC101520870	Brain Tumor, Recurrent	49	Dacomitinib	2	Completed	ES
13	NCT01112527	Glioblastoma Multiforme	58	Dacomitinib	2	Completed	USA
14	NCT00463073	Malignant Gliomas	32	Cetuximab, Bevacizumab, Irinotecan	2	Completed	DK
15	NCT01800695	Glioblastoma Multiforme	202	Depatuxizumab mafodotin (ABT- 414) , Temozolo- mide, Whole Brain Radiation	1	Completed	AU
16	NCT02573324	Glioblastoma Multiforme	691	Depatuxizumab mafodotin (ABT-414) , Temozolomide	3	Active, not recruiting	USA
17	NCT04083976	Advanced Solid Tumor	280	Erdafitinib	2	Recruiting	USA
18	NCT00049127	Recurrent Adult Brain Neo- plasm	64	Imatinib	1, 2	Completed	USA
19	NCT00613054	Glioblastoma Multiforme	27	Imatinib, Hydroxyurea	1	Completed	USA
20	NCT01331291	Glioblastoma Multiforme	36	Bosutinib	2	Completed	USA
21	NCT00601614	Glioblastoma Multiforme Gliosarcoma	119	Temozolomide, Vandetanib	1.2	Completed	USA

Table 1. Clinical Trials on Target Therapies for High-Grade Gliomas.

#	ClinicalTrials.gov Identifier	Conditions	# of Patients Enrollment	Interventions	Study Phase	Status	Locations
22	NCT00427440	Advanced Malignant Glioma	61	AMG 102	2	Completed	USA
23	NCT01632228	Glioblastoma Multiforme	135	Onartuzumab, Bevacizumab	2	Completed	CN, FR, DE, IT, ES, SW, UK , USA
24	NCT01113398	Glioblastoma Multiforme Gliosarcoma	36	AMG 102, Bevacizumab	2	Completed	USA
25	NCT01632228	Glioblastoma Multiforme	135	Bevacizumab, Onartuzumab	2	Completed	USA
26	NCT00606879	Advanced Cancer	46	SGX523	1	Terminated	USA
27	NCT00607399	Advanced Cancer	46	SGX523	1	Terminated	USA
28	NCT00784914	Brain and Central Nervous System Tumors	12	Temsirolimus	1	Completed	USA
29	NCT00016328	Adult Glioblastoma Multiforme Adult Gliosarcoma Recurrent Adult Brain Tumor	33	Temsirolimus	2	Completed	USA
30	NCT00047073	Brain and Central Nervous System Tumors	13	Sirolimus, Surgery	1,2	Completed	USA
31	NCT00672243	Glioblastoma Multiforme Gliosarcoma	32	Erlotinib, Sirolimus	2	Completed	USA
32	NCT00553150	Brain and Central Nervous System Tumors	122	Everolimus, Temozolomide, Radiotherapy	1.2	Completed	USA
33	NCT00085566	Brain and Central Nervous System Tumors Prostate Cancer	61	Everolimus, Gefitinib	1.2	Completed	USA
34	NCT01339052	Glioblastoma Multiforme	65	Buparlisib, Surgery	2	Completed	USA
35	NCT01473901	Glioblastoma Multiforme	38	Buparlisib, Temozolomide, Radiotherapy	1	Completed	USA
36	NCT01349660	Glioblastoma Multiforme	88	Buparlisib, Bevaci- zumab	1,2	Active, not recruiting	USA
37	NCT00590954	Malignant Gliomas Brain Cancer	32	Perifosine	2	Completed	USA
38	NCT00005859	Brain and Central Nervous System Tumors	136	Tipifarnib	1.2	Completed	USA
39	NCT00049387	Adult Giant Cell Glioblastoma Adult Glioblastoma Adult Gliosarcoma	19	Tipifarnib, Temozolomide, Radiotherapy	1	Completed	USA
40	NCT00015899	Brain and Central Nervous System Tumors	53	Lonafarnib	1	Completed	USA

#	ClinicalTrials.gov Identifier	Conditions	# of Patients Enrollment	Interventions	Study Phase	Status	Locations
41	NCT00038493	Glioblastoma Multiforme	23	Temozolomide, Lonafarnib	2	Completed	USA
42	NCT01748149	Pediatric BRAFV600E-mu- tant Gliomas	40	Vemurafenib	1	Active, not recruiting	USA
40	NOTION	Glioblastoma		Ribociclib		Active, not recruiting	USA
43	NC102345824	Glioma	3		1		
44	NCT02896335	Metastatic Malignant Brain Tumors	30	Palbociclib	2	Recruiting	USA
15	NCT02924740	Glioblastoma Multiforme	24	Ribociclib,	1	Descriting	USA
43	INC 103834740	Brain Gliomas	24	Everolimus	1	Recruiting	
		Astrocytoma, Grade III		Zotiraciclib,			SW
46	NCT03224104	Glioblastoma	81	Temozolomide, Radiotherapy	1	Recruiting	
		Brain Tumors					
47	NCT02942264	Astrocytoma, Astroglioma	152	Zotiraciclib,	12	Recruiting	USA
	1101912201	Glioblastoma	132	Temozolomide	1, 2	Recruiting	
		Gliosarcoma					
		Cholangiocarcinoma			1	Active, not recruiting	USA, FR
	NCT02073994	Chondrosarcoma	170	Ivosidenib			
48		Glioma					
		Other Advanced Solid Tumors					
49	NCT02481154	Glioma	150	Vorasidenib	1	Active, not recruiting	USA
	NCT00884741	Glioblastoma Multiforme	637	Bevacizumab, Temozolomide, Radiotherapy		Completed	USA
50		Gliosarcoma			3		
		Supratentorial Glioblastoma					
51	NCT00731731	Adult Glioblastoma	125	Temozolomide, Vorinostat	1,2	Active, not recruiting	USA
52	NCT00128700	Brain and Central Nervous System Tumors	20	Temozolomide, Vatalanib, Radio- therapy	1, 2	Completed	BE, DE, IT, NL, SW
53	NCT00108056	Glioma	26	Enzastaurin	1	Terminated	USA
54	NCT00190723	Malignant Glioma	120	Enzastaurin	2	Completed	USA
55	NCT00503724	Brain and Central Nervous System Tumors	32	Enzastaurin	1	Completed	USA
		Neuroblastoma				1	
56	NCT00006247	Brain and Central Nervous System Tumors	33	Semaxanib	1	Terminated	USA
57	NCT01229644	Glioma	10	Crenolanib	2	Terminated	USA
50	NCT01393912	Diffuse Intrinsic Pontine Glioma					
58		Progressive or Refractory High-Grade Glioma	55	Crenolanib	1	Completed	USA

#	ClinicalTrials.gov Identifier	Conditions	# of Patients Enrollment	Interventions	Study Phase	Status	Locations
		Adult Giant Cell Glioblas- toma					
59	NCT00305656	Adult Glioblastoma	31	Cediranib	2	Completed	USA
		Adult Gliosarcoma				1	
		Recurrent Adult Brain Tumor					
60	NCT00326664	Recurrent Glioblastoma	55	Cediranib	1	Completed	USA
61	NCT00503204	Brain Tumor	20	Cediranib, Lomustine	1	Completed	USA, UK
62	NCT00704288	Glioblastoma Multiforme	222	Cabozantinib	2	Completed	USA
		Glioblastoma Multiforme		Cabozantinib,			
63	NCT00960492	Gliosarcoma	26	Temozolomide, Radiotherapy	1	Completed	USA
64	NCT00337207	Brain and Central Nervous System Tumors	55	Bevacizumab	2	Completed	USA
		Malignant Glioma			2	Completed	USA
6	NCT01740250	Grade IV Malignant Glioma	(0	Bevacizumab,			
65	NC101740258	Glioblastoma	- 69	Radiotherapy			
		Gliosarcoma					
66	NCT00271609	Recurrent High-Grade Gliomas	88	Bevacizumab	2	Completed	USA
		Malignant Gliomas				1	
67	NCT01290939	Glioblastoma Multiforme	433			Unknown	
		Cognition Disorders		Bevacizumab,	3		USA
		Disability Evaluation		Lomustine			
68	NCT01860638	Glioblastoma Multiforme	296	Bevacizumab, Lomustine	2	Completed	AU
		Glioblastoma Multiforme		Bevacizumab,			
69	NCT00884741	GliosarcomaSupratentorial	637	Chemiotherapy, Radiotherapy	3	Completed	USA
70	NCT00943826	Glioblastoma Multiforme	921	Bevacizumab, Temozolomide, Radiotherapy	3	Completed	USA
71	NCT00895180	Adult Glioblastoma Multi- forme	80	Olaratumab, Ramucirumab	2	Completed	USA
	NCT00369590	Adult Anaplastic Astrocy- toma	58	Aflibercept	2	Completed	USA
		Adult Anaplastic Oligoden- droglioma					
72		Adult Giant Cell Glioblas- toma					
		Adult Gliosarcoma					
		Recurrent Adult Brain Tumor					
73	NCT00093964	Glioblastoma Multiforme	81	Cilengitide	2	Completed	USA

#	ClinicalTrials.gov Identifier	Conditions	# of Patients Enrollment	Interventions	Study Phase	Status	Locations
	NCT00085254	Adult Giant Cell Glioblas- toma	112	Cilengitide, Temozolomide, Radiotherapy	1,2	Completed	USA
74		Adult Glioblastoma					
		Adult Gliosarcoma		Radiotiferapy			
75	NCT00689221	Glioblastoma Multiforme	545	Cilengitide, Temozolomide, Radiotherapy	3	Completed	USA, DE
		Glioblastoma Multiforme			2	Completed	USA
76	NCT00165477	Gliosarcoma	23	Lenalidomide, Radiotherapy			
		Malignant Gliomas					
77	NCT03345095	Newly Diagnosed Glioblastoma	750	Marizomib, Temozolomide, Radiotherapy	3	Recruiting	AU, BE
	NCT00006773	Adult Anaplastic Astrocy- toma	_	Bortezomib	1	Terminated	USA
		Adult Anaplastic Oligoden- droglioma					
78		Adult Giant Cell Glioblastom	42				
		Adult Glioblastoma					
		Adult Gliosarcoma					
		Recurrent Adult Brain Tumor					
79	NCT00998010	Brain and Central Nervous System Tumors	25	Bortezomib, Temozolomide, Radiotherapy	2	Completed	USA

AU: Austria; BE: Belgium; CA: Canada; DE: Germany; DK: Denmark; ES: Spain; FR: France; IT: Italy; NL: Netherlands; SW: Switzerland; UK: United Kingdom; USA: United States of America

majority of approaches are directed against signaling pathways related to cell proliferation and glioma invasion, angiogenesis and inhibition of apoptosis.²²⁻²⁵ Table 2 reports the classification of the target therapies used for malignant brain tumors (Table 2).

2.1. Tyrosine Kinase Inhibitors

Tyrosine kinase receptors consist in an extracellular ligand-binding and a transmembrane tyrosine kinase domain containing sites for autophosphorylation. Upon the binding of its ligand, the receptors undergo dimerization and phosphorylation of specific tyrosines, those become binding sites, recruit proteins and activate downstream intracellular pathways, ultimately resulting in tumor maintenance and proliferation.²⁶⁻²⁸ The most widely studied tyrosine kinase receptors are the epidermal growth factor receptor (EGFR), the platelet-derived growth receptor (PDGFR), the fibroblast growth factor receptor (FGFR) and the hepatocyte growth factor receptor (HGFR). All of them are constantly overexpressed or mutated in GBMs. Tyrosine kinase inhibitors (TKIs) are molecules which bind the aforementioned receptors, blocking their downstream signals.

2.1.1 EGFR

The EGFR gene is amplified or overexpressed in 40% to 60% of the primary GBMs, whereas loss of exons 2 to 7 (EGFRvIII) is present in 40-50% of the cases.²⁹⁻³¹

Target Therapy						
Candidate Drugs	Target	Biological Role in GBM				
	EGFRvIII	Proliferation, migration, invasion,				
	PDGFR					
1 K15	FGFR	and resistance to				
	HGFR	apoptosis				
	PI3K	Growth, metabo-				
PI3K/AKT/ mTOR Is	AKT	lism, proliferation, migration				
	mTORC1					
	RAS/MAPK	Cell cycle				
FTIs	BRAF V600E	maintenance and proliferation				
p53Is	MDM2/	Cell cycle progres- sion and resistance to				
	MDM4					
pRBIs	CDK4/CDK6	apoptosis				
IDHIs	IDH1	Metabolism, pro- liferation, invasion, angiogenesis				
HDACIs	Histones	Dysregulation DNA transcription, expansion of gene mutations				
	VEGF-A	Blood vessel forma-				
AIs	VEGFR1	tion, proliferation, therapeutic resistance				
	РКС	Tumor microenviron- ment maintenance				
IIs	Integrins	Cell adhesion, migra- tion, metastasis				
PIs	Proteasome complex	Homeostasis, growth and resistance to apoptosis				

Table 2. Classification of Target Therapies for Malignant BrainTumors

AIs Angiogenesis Inhibitors, EGFR: Epidermal Growth Factor Receptor; FGFR: Fibroblast Growth Factor Receptor FTIs: Farnesyl Transferase Inhibitors; HDACIs: Histone Deacetylases Inhibitors; HGFR: Hepatocyte Growth Factor Receptor; IDH1: Isocitrate Dehydrogenase 1; IDHIs: Isocitrate Dehydrogenase Inhibitors; IIs: Integrin Inhibitors; mTOR: Mammalian Target of Rapamycin; mTORC1: Mammalian Target of Rapamycin Complex 1; PDGFR: Platelet- Derived Growth Receptor; PI3K: Phosphatidylinositol 4,5-Bisphosphate–3; PIs: Proteasome Inhibitors; VEGF-A: Vascular Endothelial Growth Factor A; VEGFR1: Vascular Endothelial Growth Factor Receptor 1 EGFRvIII mutation leads to a ligand-independent kinase activity and, accordingly, an EGFR-pathway overactivation, resulting in increased cell proliferation, invasiveness and resistance to chemotherapeutic agents.^{32, 33} Gefinitib (Iressa®) and erlotinib (Tarceva®) are approved TKIs directed against EGFRvI-II. Three phase II clinical trials (#NCT00025675, #NCT00238797, #NCT00016991) highlighted the efficacy of gefinitib, pointing out a progression-free survival at 6 months (PFS-6) of 13%.³⁴ Erlotinib lacked success as a monotherapy, but enhanced the efficacy of chemo-radiotherapy, especially if associated with temozolomide (TMZ) and carmustine at a dose of 150 or 300 mg/daily.^{35, 36} Similar results have been reported for lapatinib, afatinib and dacomitinib.³⁷

In addition, two monoclonal antibodies (MAbs) are under observation. Cetuximab, a chimeric murinehuman IgG1 Mab that binds the extracellular EGFR domain inducing tumor apoptosis.³⁸ As a monotherapy, it demonstrated a PFS-6 of 9.2% and an increased overall survival (OS) of 5 months. In combination with bevacizumab and irinotecan cetuximab, it showed a PFS-6 of 30% and a median OS of 7.2 months.³⁹ ABT-414, an EGFR-directed MAb conjugated to an anti-microtubulin agent, had a PFS-6 of 28.3% in monotherapy or when combined with standard temozolomide chemoradiotherapy (#NCT02573324).⁴⁰

2.1.2. PDGFR

PDGFR gene amplification is found in nearly 15% of GBMs, and the receptor's overexpression, which leads to tumor growth and angiogenesis, is frequently associated with transition from low- to highgrade glioma.³⁰ Imatinib is the most famous PDGFR inhibitor, used in many hematological tumors for its activity against the mast/stem cell growth factor receptor (c-KIT), and oncogene fusion protein BCR-ABL.

Many phase II clinical trials have proven that imatinib monotherapy failed to improve PFS-6 or OS in patients with GBM,⁴¹ but resulted in a good response in combination with hydroxyurea.⁴²

Sorafenib, vandetanib, dasatinib and bosutinib are other PDGFR inhibitors. However, many clinical trials have failed to demonstrate the efficacy of dasatinib, both as monotherapy and combined with radiotherapy, TMZ and lomustine. $^{\scriptscriptstyle 43,\,44}$

2.1.3. FGFR

Erdafitinib, a selective FGFR TKI, showed promising results in patients with GBM harboring oncogenic FGFR-TACC fusion.^{45,46}

2.1.4. HGFR/c-MET

HGFR, also known as c-Met, amplification/ mutation has a role in promoting gliomagenesis and drug resistance.^{47, 48} Crizotinib, specifically designed against c-Met, has given some results in combination with dasatinib.^{49, 50} Analogous results have been reported for SGX523^{51, 52} (#NCT00606879, #NCT00607399). Conversely, onartuzumab and rilotumumab (AMG102) basically demonstrated no clinical benefits.^{53, 54} Two phase II clinical trials have been completed, one with AMG102 as monotherapy (#NCT00427440), and the other with AMG102 plus bevacizumab (#NCT01113398), both for patients with recurrent high-grade gliomas.

2.2. PI3K/AKT/mTOR Inhibitors

The Cancer Genome Atlas analysis highlighted the presence of PI3K/AKT/ mTOR signaling pathway dysregulation in 50-60% of GBMs.^{55, 56} The activation of phosphatidylinositol 4.5-bisphosphate-3 (PI3K) regulates the activity of many kinase proteins, such as AKT. It transduces the signals to many downstream intracellular effectors, like the mammalian target of rapamycin (mTOR). A fundamental intracellular protein is mTOR, involved in cell growth signaling and tumorigenesis. It is composed of two subunits, mTORC1-2, with different roles, and mTORC1, particularly involved in the transition of the cell cycle from G1 to S. The Food and Drug Administration (FDA) approved three mTORC1 inhibitors: sirolimus (Rapamycin, Rapamune[®]), everolimus[®] and temsirolimus[®].

Temsirolimus has been evaluated in some significant clinical trials; one of these was a phase II study involving 65 patients with recurrent GBM. It demonstrated a radiographic improvement in 36% of the patients, a PFS-6 of 7.8% and median OS of 4.4 months. 57

Sirolimus has been tested in combination with surgery (#NCT00047073), gefitinib in 34 recurrent glioma patients, and erlotinib (#NCT00672243), demonstrating moderate effectiveness.⁵⁸

Everolimus was studied in combination with gefitinib (#NCT00085566), bevacizumab or chemioradiotherapy. A phase II clinical trial tested the combination of everolimus, TMZ and radiotherapy versus conventional standard of care (#NCT00553150).

However, mTOR inhibitors have not demonstrated significant clinical activity, if not in combination with other treatments. This is due to their selectivity for mTORC1 and not mTORC2, ensuring only a partial blocking of the mTOR function.

In fact, two novel ATP-competitive mTORC2 inhibitors (CC214-1 and CC214-2) are under investigation, in order to overcome the resistance of mTOR inhibitors.⁵⁹

Other promising strategies involve the selective PI3K inhibitor, buparlisib, which has an antitumor activity, especially when associated with bevacizumab in patients with recurrent GBM.⁵⁹

Perifosine is a novel selective AKT inhibitor, currently tested in some ongoing trials. A phase II study investigated perifosine as a monotherapy for recurrent malignant gliomas⁶⁰ (#NCT00590954).

2.3. Farnesyl Transferase Inhibitors

Following the activation of TK receptors, the intracellular RAS protein family undergoes posttranslational modifications and triggers multiple effector pathways, including the RAF and MAP kinases (MAPK) involved in cell proliferation, differentiation and survival.

However, translocation of RAS to the cell membrane requires a post-translational alteration catalyzed by the farnesyl transferase enzyme.^{30, 61}

Farnesylation is the limiting step in RAS activities and the specific farnesyl transferase inhibitors (FTIs) lock all its functions upstream, and consequently the intracellular RAS-RAF-MEK-MAPK pathway.⁶² Lonafarnib, an FTI, was tested in a phase I clinical trial in combination with TMZ and radiotherapy, with promising results⁶⁵ (#NCT00049387).

2.3.1. BRAF V600E

RAF kinases, also triggered by the RAS system, are involved in intracellular growth pathways and stimulation.

Several studies reported the presence of BRAF V600E mutation, especially in infant gliomas.⁶⁶ Vemurafenib, a BRAF inhibitor, is under investigation in a phase I ongoing trial, for children with recurrent BRAFV600E-Mutant gliomas⁶⁷ (#NCT01748149).

2.4. MDM2/MDM4/p53 inhibitors

The dysregulation of p53 signaling pathways is found in more than 80% of high-grade gliomas. The p53 is fundamental in cell-cycle arrest and apoptosis; mutation results in clonal expansion of tumor cells and genetic instability.^{68,69}

In 20% of the patients, the p53 inactivity is due to the MDM2 or MDM4 overexpression. MDM2/ MDM4 inactivates p53 and consequently leads to loss of cancer suppression.^{30,70}

Therefore, an effective strategy rationale is to restore the p53 activity, by molecules targeting MDM2 or MDM4. Preclinical studies demonstrated the successful suppression of GBM growth with several MDM2 inhibitors, including RG7112,⁷¹ RG7388 and AMG232 as well as many others in progress (#NCT03107780).

2.5. CDK4/CDK6/pRB inhibitors

The altered function of retinoblastoma protein (pRB) contributes to gliomagenesis in 78% of the cases and the overexpression of CDK4/CDK6 plays a fundamental role in the modulation of this pathway, involved in cell growth.⁷²⁻⁷⁴

Novel agents directed to CDK4 and CDK6 demonstrated strong antitumor efficacy in RB1-wild-type GBM, such as ribociclib and palbociclib.

Ribociclib was tested in a phase I trial for recurrent glioblastoma or anaplastic glioma⁷⁵ (#NCT02345824); palbociclib was employed as a monotherapy for brain metastases⁷⁶ (#NCT02896335).

Zotiraclib, a multi-CDK inhibitor, has been explored in clinical trials for newly diagnosed or recurrent gliomas (#NCT02942264, #NCT03224104).

2.6. Isocitrate dehydrogenase-1 inhibitors

Isocitrate dehydrogenase-1 (IDH1) mutation is one of the most frequent abnormalities found in highgrade gliomas, and according to the World Health Organization, is a new classification of brain tumors also having predictive value of treatment response. This mutation consists in the gain-of-function with the production of D-2-hydroxyglutarate, which interferes with cellular metabolism ^{77, 78}. Ivosidenib, an IDH1 inhibitor, is being evaluated in a phase I ongoing trial, as a monotherapy, for advanced solid tumors including IDH-mutated gliomas (#NCT02073994).

2.7. Histone deacetylases inhibitors

Histone deacetylases (HDAC) are enzymes involved in the regulation of histones, which are proteins that organize the DNA structure and regulate gene transcription.

HDAC inhibitors have an emerging role in the treatment of GBMs, potentially promoting the apoptosis of the cancer cells.⁷⁹

Vorinostat, an oral quinolone HDAC inhibitor, is being studied in phase I/II clinical trials, as a monotherapy in recurrent GBM,⁸⁰ and in combination with TMZ, showing good tolerance and giving promising results⁸¹ (#NCT00731731).

Panobinostat, Romidepsin and other HDAC inhibitors are still under evaluation.

2.8. Angiogenesis inhibitors

The tumor's microenvironment, together with pathological angiogenesis and neovascularization, play

a fundamental role in the development and progression of high-grade gliomas.

Acting as managers for the angiogenesis process, as well as for a wide range of CNS vascular pathologies, they are mainly vascular growth factors of all the vascular endothelial growth factor-A (VEGF-A) and its receptors, VEGFR1 and VEGFR2, found on the glioma's endothelial cells.⁸²⁻⁸⁵

Efforts to downregulate this pathway have been pursued through the development of agents directed to VEGF/VEGFR, which not only block neoangiogenesis, but also have an effect on the vascular phenotype.

The inhibition of VEGF signaling also changes the vessels' diameter, permeability and tortuosity, decreasing tumor hypoxia and consequently disrupting the survival mechanism in glioma cells as well as increasing chemotherapy delivery and radiosensitivity.⁸³⁻⁸⁵

2.8.1. VEGFR

Several studies evaluated VEGFR inhibitors for patients with newly diagnosed, as well as recurrent GBM.

Vatalanib has been tested in phase I/II studies in combination with TMZ and radiotherapy (#NCT00128700). Cediranib demonstrated no clinical benefits in a phase II clinical trial as a monotherapy (#NCT00305656), yet there was greater benefit together with lomustine in a randomized phase III study⁸⁶ (#NCT00503204).

Cabozantinib is a promising agent against VEG-FR and MET signaling, evaluated in two phase II studies involving newly diagnosed (#NCT00960492) and recurrent GBM (#NCT00704288). Ramucirumab and icrucumab are new MAbs under evaluation, directed to VEGFR-2 and VEGFR-1, respectively.⁸⁷

2.8.2. VEGF

The most relevant of the VEGF inhibitors is bevacizumab, a humanized IgG1 monoclonal antibody against VEGF-A, which in 2009 received FDA-approval for the treatment of recurrent GBM, after the high radiographic response rates (ranging from 28% to 59%) achieved in two clinical trials.^{88,89} The significant antitumor potential of bevacizumab has been proven in many studies, using it as a monotherapy or in combination with lomustine (#NCT01290939) and radiochemiotherapy.^{90,91}

Combinations of bevacizumab with the standard of care were examined in two phase III clinical trials, AVAglio⁹² (#NCT00943826) and RTOG- 0825⁹³ (#NCT00884741), and although both demonstrated encouraging results in PFS survival benefit, bevacizumab remains only an alternative treatment in the recurrent setting.

Another promising agent is aflibercept, known as VEGF-trap, a recombinant product fusion protein which has been studied in phase II trials with a PFS-6 of 7.7% and median OS of 3 months.^{94,95}

2.8.3. Protein kinase C

Protein kinase C (PKC) is implicated in activation of the angiogenesis process, cell proliferation and constitution of the microenvironment, therefore, it is a potentially attractive therapeutic target.

Enzastaurin, a potent PKC inhibitor, demonstrated in a phase I/II trial a 25% radiographic response and a PFS-6 of 7% in GBM.[%]

Tamoxifen, a modulator of the estrogen receptor, has been described as a PKC inhibitor and was tested in GBM therapy with a median OS of 9.7 months.^{97, 98}

2.9. Integrin inhibitors

The integrins are transmembrane proteins which bind multiple extracellular ligands and mediate cell adhesion and migration. They are expressed at a high level in malignant glioma cells and play a central role in the angiogenesis, development, invasion and metastasis of the tumor.^{99, 100} Integrin inhibitors are being investigated as a means of reducing this mechanism.

Cilengitide, which competitively inhibits integrin ligand binding,¹⁰¹ has been evaluated in a phase I/II study stand-alone;¹⁰² or in a phase III trial, associated to TMZ and radiotherapy, resulting in a good improvement of PFS-6¹⁰³ (#NCT00689221).

Thalidomide and lenalidomide, which interfere with the expression of integrin receptors and have an

antiangiogenic effect, are being studied for GBM therapy, with results that are still unsatisfactory.¹⁰⁴⁻¹⁰⁶

2.10. Proteasome inhibitors

Proteasomes are proteins with enzymatic activities involved in the regulation of homeostasis, cell growth and apoptosis.

Bortezomib (Velcade®), the most used proteasome inhibitor in the oncological field, has also been tested for GBM therapy in combination with chemioradiotherapy¹⁰⁷ (#NCT00006773).

The pan-proteasome inhibitor, Marizomib, is currently undergoing phase III evaluation in newly diagnosed GBMs¹⁰⁸ (#NCT03345095).

Discussion

The present literature review highlights the current role of a series of target therapies, especially tyrosine kinase and angiogenesis inhibitors, in the treatment of malignant CNS tumors.

Several steps forward have been done in the recent years toward a deep understanding of complex pathophysiologic pathways associated with a wide spectrum of neurological and neuro-oncological pathologies of adulthood and pediatric age. ¹⁰⁹⁻¹¹¹ Nevertheless, the lack of success of the standard of care and the still largely dismal prognosis of patients affected by high-grade gliomas dictate the urgent need of new and more effective therapeutic approaches.

In this scenario, the improved understanding of genome mutations underlying the GBM phenotype has led to greater insight into the biology of the tumor, at the same time providing the opportunity for designing novel and personalized treatment strategies.^{82, 112, 113}

Data from the Cancer Genome Atlas project ⁵⁵ revealed the complicated genetic profile of GBMs and recognized the core signaling and transduction pathways commonly involved in the growth, proliferation, angiogenesis and spreading of the tumor.¹¹⁴

A further tangible aspect of these advances is the latest World Health Organization's classification of brain tumors, which integrates data from traditional histological analysis with biomolecular connotation obtained by specific genetic analysis and characterizations.¹¹⁵ Accordingly, the target therapies developed on the basis of the above have detected molecular abnormalities, and have made use of pharmacological agents tailored to specific mutations, specific to tumor subtypes.

Typical genetic alterations of GBMs are the overexpression of the tyrosine kinase receptors, especially the EGFR, PDGFR, FGFR and HGFR, dysregulation of PI3K/AKT/mTOR and RAS/MAPK pathways, as well as p53 or pRB mutations.^{30, 116, 117}

TKIs have long been investigated in several clinical trials with disappointing results. Despite the extreme specificity of these agents, they were not efficacious as a monotherapy, thus the current approach consists in the combination of multiple molecular agents within the same targets or between separate pathways.^{33, 118, 119}

PI3K/AKT/mTOR pathway and farnesyltransferase inhibitors show low tolerability and safe profiles during clinical studies, but have a synergistic effect only in combination with standard of care.^{58, 120}

Likewise, agents directed at restoring p53 and pRB activity gave encouraging results in association with chemotherapy and whole brain radiotherapy.^{76, 121} The newly discovered alterations in metabolic pathways, including IDH1 and HDAC enzymes, seem to be up-and-coming targets. Currently, anti-angiogenetic drugs are among the most promising. They focused on the blocking of VEGF/VEGFR,^{122, 123} along with components of the tumor microenvironment, such as protein kinase C, integrins and proteasome complexes.^{89, 124, 125}

Despite the rationale of the target therapies, the vast intratumoral heterogeneity and GBM cell plasticity have caused a rapid shift toward resistant tumor phenotypes, the latter responsible for the failure of the therapy.¹²⁶⁻¹²⁸

Additionally, the route of drug administration still presents a limitation for the efficacy of these therapies. Recent progress has been made through the use of stereotactic or endoscopic techniques for the intrathecal administration of pharmacological agents directly into the tumor site, also benefiting from the minimal invasiveness of these approaches, well evident also for other neurosurgical pathologies.¹²⁹⁻¹³¹

Last but not least, the immunological tumor microenvironment, composed of glia cells and lymphocytes, consistently modulates tumor sensitivity to treatment.¹³²⁻¹³⁴

Conclusion

The improved knowledge of the biology of tumors has recently made it possible to transform the molecular alterations at the base of the high malignancy of GBM, into different treatment strategies.

Good results came from tyrosine kinase inhibitors, primarily erlotinib and gefinitinb. Similarly, PI3K/ AKT/mTOR inhibitors and p53 restoring agents proved their efficacy in several clinical trials. Bevacizumab, in association with TMZ and radiotherapy, has been approved for recurrent GBMs.

An in-depth identification of driver molecular alterations may make it possible to appropriately select those patients who are candidates for a target therapy.

The greatest challenge of the near future consists in overcoming the issue of escape of GBM that is present in all of these therapies.

Acknowledgements

We want to thank Giorgia Di Giusto, Engineer, for her invaluable technical support during data collection and analysis.

Conflict of interest: Each author declares that he or she has no commercial associations (e.g. consultancies, stock ownership, equity interest, patent/licensing arrangement etc.) that might pose a conflict of interest in connection with the submitted article.

Refernces

- Jiang T, Mao Y, Ma W, et al. CGCG clinical practice guidelines for the management of adult diffuse gliomas. Cancer Lett. 2016;375(2): 263–273. https://doi.org/10.1016/j.canlet.2016.01.024.
- Ricard D, Idbaih A, Ducray F, Lahutte M, Hoang-Xuan K, Delattre JY. Primary brain tumours in adults. Lancet. 2012;379(9830): 1984–1996. https://doi.org/10.1016/S0140-6736(11)61346-9.
- Cloughesy TF, Cavenee WK, Mischel PS. Glioblastoma: from molecular pathology to targeted treatment. Annu Rev Pathol. 2014;9: 1–25. https://doi.org/10.1146/annurevpathol-011110-130324.
- Ostrom QT, Gittleman H, Liao P, et al. CBTRUS statistical report: primary brain and central nervous system tumors diagnosed in the United States in 2007–2011. Neuro Oncol. 2014;16 Suppl 4: iv1–63. https://doi.org/10.1093/neuonc/ nou223.

- Stupp R, Mason WP, van den Bent MJ, et al. Radiotherapy plus concomitant and adjuvant temozolomide for glioblastoma. N Engl J Med. 2005;352(10): 987–996. https://doi. org/10.1056/NEJMoa043330.
- 6. Stupp R, Hegi ME, Mason WP, et al. Effects of radiotherapy with concomitant and adjuvant temozolomide versus radiotherapy alone on survival in glioblastoma in a randomised phase III study: 5-year analysis of the EORTC-NCIC trial. Lancet Oncol. 2009;10(5): 459–466. https:// doi.org/10.1016/S1470-2045(09)70025-7.
- Hardee ME, Zagzag D. Mechanisms of glioma-associated neovascularization. Am J Pathol. 2012;181(4): 1126–1141. https://doi.org/10.1016/j.ajpath.2012.06.030.
- Auffinger B, Spencer D, Pytel P, Ahmed AU, Lesniak MS. The role of glioma stem cells in chemotherapy resistance and glioblastoma multiforme recurrence. Expert Rev Neurother. 2015;15(7): 741–752. https://doi.org/10.1586/14737175.2 015.1051968.
- Roos A, Ding Z, Loftus JC, Tran NL. Molecular and Microenvironmental Determinants of Glioma Stem-Like Cell Survival and Invasion. Front Oncol. 2017;7: 120. https:// doi.org/10.3389/fonc.2017.00120.
- Prelaj A, Rebuzzi SE, Grassi M, et al. Multimodal treatment for local recurrent malignant gliomas: Resurgery and/or reirradiation followed by chemotherapy. Mol Clin Oncol. 2019;10(1): 49–57. https://doi.org/10.3892/ mco.2018.1745.
- Pascual-Castroviejo I, Lopez-Pereira P, Savasta S, Lopez-Gutierrez JC, Lago CM, Cisternino M. Neurofibromatosis type 1 with external genitalia involvement presentation of 4 patients. J Pediatr Surg. 2008;43(11): 1998–2003. https:// doi.org/10.1016/j.jpedsurg.2008.01.074.
- 12. Savasta S, Chiapedi S, Perrini S, Tognato E, Corsano L, Chiara A. Pai syndrome: a further report of a case with bifid nose, lipoma, and agenesis of the corpus callosum. Childs Nerv Syst. 2008;24(6): 773–776. https://doi.org/10.1007/ s00381-008-0613-9.
- Salpietro V, Mankad K, Kinali M, et al. Pediatric idiopathic intracranial hypertension and the underlying endocrine-metabolic dysfunction: a pilot study. J Pediatr Endocrinol Metab. 2014; 27(1-2): 107–115. https://doi.org/10.1515/jpem-2013-0156.
- Nosadini M, Granata T, Matricardi S, et al. Relapse risk factors in anti-N-methyl-D-aspartate receptor encephalitis. Dev Med Child Neurol. 2019;61(9): 1101–1107. https:// doi.org/10.1111/dmcn.14267.
- Cheng CY, Shetty R, Sekhar LN. Microsurgical Resection of a Large Intraventricular Trigonal Tumor: 3-Dimensional Operative Video. Oper Neurosurg (Hagerstown). 2018;15(6): E92-E93. https://doi.org/10.1093/ons/opy068.
- Palumbo P, Lombardi F, Siragusa G, et al. Involvement of NOS2 Activity on Human Glioma Cell Growth, Clonogenic Potential, and Neurosphere Generation. Int J Mol Sci. 2018;19(9). https://doi.org/10.3390/ijms19092801.
- Luzzi S, Crovace AM, Del Maestro M, et al. The cellbased approach in neurosurgery: ongoing trends and future perspectives. Heliyon. 2019;5(11): e02818. https://doi. org/10.1016/j.heliyon.2019.e02818.

- Luzzi S, Giotta Lucifero A, Del Maestro M, et al. Anterolateral Approach for Retrostyloid Superior Parapharyngeal Space Schwannomas Involving the Jugular Foramen Area: A 20-Year Experience. World Neurosurg. 2019;132: e40– e52. https://doi.org/10.1016/j.wneu.2019.09.006.
- Spena G, Roca E, Guerrini F, et al. Risk factors for intraoperative stimulation-related seizures during awake surgery: an analysis of 109 consecutive patients. J Neurooncol. 2019;145(2): 295–300. https://doi.org/10.1007/s11060-019-03295-9.
- Antonosante A, Brandolini L, d'Angelo M, et al. Autocrine CXCL8-dependent invasiveness triggers modulation of actin cytoskeletal network and cell dynamics. Aging (Albany NY). 2020;12(2): 1928–1951. https://doi.org/10.18632/ aging.102733.
- Pearson JRD, Regad T. Targeting cellular pathways in glioblastoma multiforme. Signal Transduct Target Ther. 2017;2: 17040. https://doi.org/10.1038/sigtrans.2017.40.
- Parsons DW, Jones S, Zhang X, et al. An integrated genomic analysis of human glioblastoma multiforme. Science. 2008;321(5897): 1807–1812. https://doi.org/10.1126/science.1164382.
- 23. Phillips HS, Kharbanda S, Chen R, et al. Molecular subclasses of high-grade glioma predict prognosis, delineate a pattern of disease progression, and resemble stages in neurogenesis. Cancer Cell. 2006;9(3): 157–173. https://doi. org/10.1016/j.ccr.2006.02.019.
- 24. Ceccarelli M, Barthel FP, Malta TM, et al. Molecular Profiling Reveals Biologically Discrete Subsets and Pathways of Progression in Diffuse Glioma. Cell. 2016;164(3): 550–563. https://doi.org/10.1016/j.cell.2015.12.028.
- Szopa W, Burley TA, Kramer-Marek G, Kaspera W. Diagnostic and Therapeutic Biomarkers in Glioblastoma: Current Status and Future Perspectives. Biomed Res Int. 2017;2017: 8013575. https://doi.org/10.1155/2017/8013575.
- Nakada M, Kita D, Teng L, et al. Receptor Tyrosine Kinases: Principles and Functions in Glioma Invasion. Adv Exp Med Biol. 2020;1202: 151–178. https://doi.org/10.1007/978-3-030-30651-9_8.
- Carrasco-Garcia E, Saceda M, Martinez-Lacaci I. Role of receptor tyrosine kinases and their ligands in glioblastoma. Cells. 2014;3(2): 199–235. https://doi.org/10.3390/ cells3020199.
- Wang K, Huang R, Wu C, et al. Receptor tyrosine kinase expression in high-grade gliomas before and after chemoradiotherapy. Oncol Lett. 2019;18(6): 6509–6515. https:// doi.org/10.3892/ol.2019.11017.
- Pelloski CE, Ballman KV, Furth AF, et al. Epidermal growth factor receptor variant III status defines clinically distinct subtypes of glioblastoma. J Clin Oncol. 2007;25(16): 2288– 2294. https://doi.org/10.1200/JCO.2006.08.0705.
- Brennan CW, Verhaak RG, McKenna A, et al. The somatic genomic landscape of glioblastoma. Cell. 2013;155(2): 462– 477. https://doi.org/10.1016/j.cell.2013.09.034.
- Gan HK, Kaye AH, Luwor RB. The EGFRvIII variant in glioblastoma multiforme. J Clin Neurosci. 2009;16(6): 748– 754. https://doi.org/10.1016/j.jocn.2008.12.005.

- 32. An Z, Aksoy O, Zheng T, Fan QW, Weiss WA. Epidermal growth factor receptor and EGFRvIII in glioblastoma: signaling pathways and targeted therapies. Oncogene. 2018; 37(12): 1561–1575. https://doi.org/10.1038/s41388-017-0045-7.
- 33. Felsberg J, Hentschel B, Kaulich K, et al. Epidermal Growth Factor Receptor Variant III (EGFRvIII) Positivity in EGFR-Amplified Glioblastomas: Prognostic Role and Comparison between Primary and Recurrent Tumors. Clin Cancer Res. 2017;23(22): 6846–6855. https://doi. org/10.1158/1078-0432.CCR-17-0890.
- Rich JN, Reardon DA, Peery T, et al. Phase II trial of gefitinib in recurrent glioblastoma. J Clin Oncol. 2004;22(1): 133–142. https://doi.org/10.1200/JCO.2004.08.110.
- 35. Raizer JJ, Abrey LE, Lassman AB, et al. A phase II trial of erlotinib in patients with recurrent malignant gliomas and nonprogressive glioblastoma multiforme postradiation therapy. Neuro Oncol. 2010;12(1): 95–103. https://doi. org/10.1093/neuonc/nop015.
- 36. van den Bent MJ, Brandes AA, Rampling R, et al. Randomized phase II trial of erlotinib versus temozolomide or carmustine in recurrent glioblastoma: EORTC brain tumor group study 26034. J Clin Oncol. 2009;27(8): 1268–1274. https://doi.org/10.1200/JCO.2008.17.5984.
- Reardon DA, Nabors LB, Mason WP, et al. Phase I/randomized phase II study of afatinib, an irreversible ErbB family blocker, with or without protracted temozolomide in adults with recurrent glioblastoma. Neuro Oncol. 2015;17(3): 430–439. https://doi.org/10.1093/neuonc/nou160.
- Fukai J, Nishio K, Itakura T, Koizumi F. Antitumor activity of cetuximab against malignant glioma cells overexpressing EGFR deletion mutant variant III. Cancer Sci. 2008;99(10): 2062–2069. https://doi.org/10.1111/j.1349-7006.2008.00945.x.
- 39. Hasselbalch B, Eriksen JG, Broholm H, et al. Prospective evaluation of angiogenic, hypoxic and EGFR-related biomarkers in recurrent glioblastoma multiforme treated with cetuximab, bevacizumab and irinotecan. APMIS. 2010;118(8): 585–594. https://doi.org/10.1111/j.1600-0463.2010.02631.x.
- 40. Phillips AC, Boghaert ER, Vaidya KS, et al. ABT-414, an Antibody-Drug Conjugate Targeting a Tumor-Selective EGFR Epitope. Mol Cancer Ther. 2016;15(4): 661–669. https://doi.org/10.1158/1535-7163.MCT-15-0901.
- 41. Wen PY, Yung WK, Lamborn KR, et al. Phase I/II study of imatinib mesylate for recurrent malignant gliomas: North American Brain Tumor Consortium Study 99–08. Clin Cancer Res. 2006;12(16): 4899–4907. https://doi. org/10.1158/1078-0432.CCR-06-0773.
- Reardon DA, Egorin MJ, Quinn JA, et al. Phase II study of imatinib mesylate plus hydroxyurea in adults with recurrent glioblastoma multiforme. J Clin Oncol. 2005;23(36): 9359–9368. https://doi.org/10.1200/JCO.2005.03.2185.
- Lassman AB, Pugh SL, Gilbert MR, et al. Phase 2 trial of dasatinib in target-selected patients with recurrent glioblastoma (RTOG 0627). Neuro Oncol. 2015;17(7): 992–998. https://doi.org/10.1093/neuonc/nov011.

- 44. Galanis E, Anderson SK, Twohy EL, et al. A phase 1 and randomized, placebo-controlled phase 2 trial of bevacizumab plus dasatinib in patients with recurrent glioblastoma: Alliance/North Central Cancer Treatment Group N0872. Cancer. 2019;125(21): 3790–3800. https://doi.org/10.1002/ cncr.32340.
- 45. Di Stefano AL, Fucci A, Frattini V, et al. Detection, Characterization, and Inhibition of FGFR-TACC Fusions in IDH Wild-type Glioma. Clin Cancer Res. 2015;21(14): 3307–3317. https://doi.org/10.1158/1078-0432.CCR-14-2199.
- Singh D, Chan JM, Zoppoli P, et al. Transforming fusions of FGFR and TACC genes in human glioblastoma. Science. 2012;337(6099): 1231–1235. https://doi.org/10.1126/science.1220834.
- Sierra JR, Tsao MS. c-MET as a potential therapeutic target and biomarker in cancer. Ther Adv Med Oncol. 2011;3(1 Suppl): S21–35. https://doi.org/10.1177/1758834011422557.
- 48. Xie Q, Bradley R, Kang L, et al. Hepatocyte growth factor (HGF) autocrine activation predicts sensitivity to MET inhibition in glioblastoma. Proc Natl Acad Sci U S A. 2012;109(2): 570–575. https://doi.org/10.1073/pnas. 1119059109.
- 49. Broniscer A, Jia S, Mandrell B, et al. Phase 1 trial, pharmacokinetics, and pharmacodynamics of dasatinib combined with crizotinib in children with recurrent or progressive high-grade and diffuse intrinsic pontine glioma. Pediatr Blood Cancer. 2018;65(7): e27035. https://doi.org/10.1002/ pbc.27035.
- 50. Chi AS, Batchelor TT, Kwak EL, et al. Rapid radiographic and clinical improvement after treatment of a MET-amplified recurrent glioblastoma with a mesenchymal-epithelial transition inhibitor. J Clin Oncol. 2012;30(3): e30–33. https://doi.org/10.1200/JCO.2011.38.4586.
- 51. Buchanan SG, Hendle J, Lee PS, et al. SGX523 is an exquisitely selective, ATP-competitive inhibitor of the MET receptor tyrosine kinase with antitumor activity in vivo. Mol Cancer Ther. 2009;8(12): 3181–3190. https://doi. org/10.1158/1535-7163.MCT-09-0477.
- 52. Guessous F, Zhang Y, diPierro C, et al. An orally bioavailable c-Met kinase inhibitor potently inhibits brain tumor malignancy and growth. Anticancer Agents Med Chem. 2010;10(1): 28–35. https://doi.org/10.2174/187152061100 9010028.
- 53. Martens T, Schmidt NO, Eckerich C, et al. A novel onearmed anti-c-Met antibody inhibits glioblastoma growth in vivo. Clin Cancer Res. 2006;12(20 Pt 1): 6144–6152. https://doi.org/10.1158/1078-0432.CCR-05-1418.
- 54. Buchanan IM, Scott T, Tandle AT, et al. Radiosensitization of glioma cells by modulation of Met signalling with the hepatocyte growth factor neutralizing antibody, AMG102. J Cell Mol Med. 2011;15(9): 1999–2006. https://doi. org/10.1111/j.1582-4934.2010.01122.x.
- 55. Cancer Genome Atlas Research N. Comprehensive genomic characterization defines human glioblastoma genes and core pathways. Nature. 2008;455(7216): 1061–1068. https://doi. org/10.1038/nature07385.

- Polivka J, Jr., Janku F. Molecular targets for cancer therapy in the PI3K/AKT/mTOR pathway. Pharmacol Ther. 2014; 142(2):164–175.https://doi.org/10.1016/j.pharmthera.2013. 12.004.
- 57. Wick W, Gorlia T, Bady P, et al. Phase II Study of Radiotherapy and Temsirolimus versus Radiochemotherapy with Temozolomide in Patients with Newly Diagnosed Glioblastoma without MGMT Promoter Hypermethylation (EORTC 26082). Clin Cancer Res. 2016;22(19):4797–4806. https://doi.org/10.1158/1078-0432.CCR-15-3153.
- Chang SM, Wen P, Cloughesy T, et al. Phase II study of CCI-779 in patients with recurrent glioblastoma multiforme. Invest New Drugs. 2005;23(4): 357–361. https:// doi.org/10.1007/s10637-005-1444-0.
- 59. Gini B, Zanca C, Guo D, et al. The mTOR kinase inhibitors, CC214-1 and CC214-2, preferentially block the growth of EGFRvIII-activated glioblastomas. Clin Cancer Res. 2013;19(20): 5722–5732. https://doi.org/10.1158/1078-0432.CCR-13-0527.
- 60. Momota H, Nerio E, Holland EC. Perifosine inhibits multiple signaling pathways in glial progenitors and cooperates with temozolomide to arrest cell proliferation in gliomas in vivo. Cancer Res. 2005;65(16): 7429–7435. https://doi. org/10.1158/0008-5472.CAN-05-1042.
- Pandey V, Bhaskara VK, Babu PP. Implications of mitogenactivated protein kinase signaling in glioma. J Neurosci Res. 2016;94(2): 114–127. https://doi.org/10.1002/jnr.23687.
- 62. Sebti SM, Adjei AA. Farnesyltransferase inhibitors. Semin Oncol. 2004;31(1 Suppl 1): 28–39. https://doi. org/10.1053/j.seminoncol.2003.12.012.
- 63. Cloughesy TF, Wen PY, Robins HI, et al. Phase II trial of tipifarnib in patients with recurrent malignant glioma either receiving or not receiving enzyme-inducing antiepileptic drugs: a North American Brain Tumor Consortium Study. J Clin Oncol. 2006;24(22): 3651–3656. https://doi. org/10.1200/JCO.2006.06.2323.
- 64. Lustig R, Mikkelsen T, Lesser G, et al. Phase II preradiation R115777 (tipifarnib) in newly diagnosed GBM with residual enhancing disease. Neuro Oncol. 2008;10(6): 1004–1009. https://doi.org/10.1215/15228517-2008-070.
- Chaponis D, Barnes JW, Dellagatta JL, et al. Lonafarnib (SCH66336) improves the activity of temozolomide and radiation for orthotopic malignant gliomas. J Neurooncol. 2011; 104(1):179–189. https://doi.org/10.1007/s11060-010-0502-4.
- 66. Kleinschmidt-DeMasters BK, Aisner DL, Birks DK, Foreman NK. Epithelioid GBMs show a high percentage of BRAF V600E mutation. Am J Surg Pathol. 2013;37(5): 685–698. https://doi.org/10.1097/PAS.0b013e31827f9c5e.
- Hyman DM, Puzanov I, Subbiah V, et al. Vemurafenib in Multiple Nonmelanoma Cancers with BRAF V600 Mutations. N Engl J Med. 2015;373(8): 726–736. https://doi. org/10.1056/NEJMoa1502309.
- Vousden KH, Lane DP. p53 in health and disease. Nat Rev Mol Cell Biol. 2007;8(4): 275–283. https://doi. org/10.1038/nrm2147.
- 69. Duffy MJ, Synnott NC, McGowan PM, Crown J, O'Connor D, Gallagher WM. p53 as a target for the treatment of can-

cer. Cancer Treat Rev. 2014;40(10): 1153–1160. https://doi. org/10.1016/j.ctrv.2014.10.004.

- Reifenberger G, Liu L, Ichimura K, Schmidt EE, Collins VP. Amplification and overexpression of the MDM2 gene in a subset of human malignant gliomas without p53 mutations. Cancer Res. 1993;53(12): 2736-2739.
- Verreault M, Schmitt C, Goldwirt L, et al. Preclinical Efficacy of the MDM2 Inhibitor RG7112 in MDM2-Amplified and TP53 Wild-type Glioblastomas. Clin Cancer Res. 2016;22(5): 1185–1196. https://doi.org/10.1158/1078-0432.CCR-15-1015.
- 72. Ohgaki H, Kleihues P. Genetic alterations and signaling pathways in the evolution of gliomas. Cancer Sci. 2009;100(12): 2235–2241. https://doi.org/10.1111/j.1349-7006.2009.01308.x.
- Wiedemeyer WR, Dunn IF, Quayle SN, et al. Pattern of retinoblastoma pathway inactivation dictates response to CDK4/6 inhibition in GBM. Proc Natl Acad Sci U S A. 2010;107(25): 11501–11506. https://doi.org/10.1073/ pnas.1001613107.
- 74. Barton KL, Misuraca K, Cordero F, et al. PD-0332991, a CDK4/6 inhibitor, significantly prolongs survival in a genetically engineered mouse model of brainstem glioma. PLoS One. 2013;8(10): e77639. https://doi.org/10.1371/ journal.pone.0077639.
- 75. Tien AC, Li J, Bao X, et al. A Phase 0 Trial of Ribociclib in Recurrent Glioblastoma Patients Incorporating a Tumor Pharmacodynamic- and Pharmacokinetic-Guided Expansion Cohort. Clin Cancer Res. 2019;25(19): 5777–5786. https://doi.org/10.1158/1078-0432.CCR-19-0133.
- 76. Taylor JW, Parikh M, Phillips JJ, et al. Phase-2 trial of palbociclib in adult patients with recurrent RB1-positive glioblastoma. J Neurooncol. 2018;140(2): 477–483. https://doi. org/10.1007/s11060-018-2977-3.
- 77. Xia L, Wu B, Fu Z, et al. Prognostic role of IDH mutations in gliomas: a meta-analysis of 55 observational studies. Oncotarget. 2015;6(19): 17354–17365. https://doi. org/10.18632/oncotarget.4008.
- Polivka J, Polivka J, Jr., Rohan V, et al. Isocitrate dehydrogenase-1 mutations as prognostic biomarker in glioblastoma multiforme patients in West Bohemia. Biomed Res Int. 2014;2014: 735659. https://doi.org/10.1155/2014/735659.
- Alvarez AA, Field M, Bushnev S, Longo MS, Sugaya K. The effects of histone deacetylase inhibitors on glioblastoma-derived stem cells. J Mol Neurosci. 2015;55(1): 7–20. https://doi.org/10.1007/s12031-014-0329-0.
- Galanis E, Jaeckle KA, Maurer MJ, et al. Phase II trial of vorinostat in recurrent glioblastoma multiforme: a north central cancer treatment group study. J Clin Oncol. 2009;27(12): 2052-2058. https://doi.org/10.1200/ JCO.2008.19.0694.
- Lee EQ, Puduvalli VK, Reid JM, et al. Phase I study of vorinostat in combination with temozolomide in patients with high-grade gliomas: North American Brain Tumor Consortium Study 04-03. Clin Cancer Res. 2012;18(21):

6032-6039. https://doi.org/10.1158/1078-0432.CCR-12-1841.

- Kamran N, Calinescu A, Candolfi M, et al. Recent advances and future of immunotherapy for glioblastoma. Expert Opin Biol Ther. 2016;16(10): 1245–1264. https://doi.org/1 0.1080/14712598.2016.1212012.
- Bao S, Wu Q, Sathornsumetee S, et al. Stem cell-like glioma cells promote tumor angiogenesis through vascular endothelial growth factor. Cancer Res. 2006;66(16): 7843–7848. https://doi.org/10.1158/0008-5472.CAN-06-1010.
- Popescu AM, Purcaru SO, Alexandru O, Dricu A. New perspectives in glioblastoma antiangiogenic therapy. Contemp Oncol (Pozn). 2016;20(2): 109–118. https://doi. org/10.5114/wo.2015.56122.
- Jain RK, di Tomaso E, Duda DG, Loeffler JS, Sorensen AG, Batchelor TT. Angiogenesis in brain tumours. Nat Rev Neurosci. 2007;8(8): 610–622. https://doi.org/10.1038/ nrn2175.
- 86. Batchelor TT, Duda DG, di Tomaso E, et al. Phase II study of cediranib, an oral pan-vascular endothelial growth factor receptor tyrosine kinase inhibitor, in patients with recurrent glioblastoma. J Clin Oncol. 2010;28(17): 2817–2823. https://doi.org/10.1200/JCO.2009.26.3988.
- Hsu JY, Wakelee HA. Monoclonal antibodies targeting vascular endothelial growth factor: current status and future challenges in cancer therapy. BioDrugs. 2009;23(5): 289– 304. https://doi.org/10.2165/11317600-000000000-00000.
- Friedman HS, Prados MD, Wen PY, et al. Bevacizumab alone and in combination with irinotecan in recurrent glioblastoma. J Clin Oncol. 2009;27(28): 4733–4740. https:// doi.org/10.1200/JCO.2008.19.8721.
- Kreisl TN, Kim L, Moore K, et al. Phase II trial of singleagent bevacizumab followed by bevacizumab plus irinotecan at tumor progression in recurrent glioblastoma. J Clin Oncol. 2009;27(5): 740–745. https://doi.org/10.1200/ JCO.2008.16.3055.
- 90. Taal W, Oosterkamp HM, Walenkamp AM, et al. Singleagent bevacizumab or lomustine versus a combination of bevacizumab plus lomustine in patients with recurrent glioblastoma (BELOB trial): a randomised controlled phase 2 trial. Lancet Oncol. 2014;15(9): 943–953. https://doi. org/10.1016/S1470-2045(14)70314-6.
- 91. Wick W, Gorlia T, Bendszus M, et al. Lomustine and Bevacizumab in Progressive Glioblastoma. N Engl J Med. 2017;377(20): 1954–1963. https://doi.org/10.1056/NEJ-Moa1707358.
- 92. Chinot OL, de La Motte Rouge T, Moore N, et al. AVAglio: Phase 3 trial of bevacizumab plus temozolomide and radiotherapy in newly diagnosed glioblastoma multiforme. Adv Ther. 2011;28(4): 334–340. https://doi.org/10.1007/ s12325-011-0007-3.
- 93. Gilbert MR, Dignam JJ, Armstrong TS, et al. A randomized trial of bevacizumab for newly diagnosed glioblastoma. N Engl J Med. 2014;370(8): 699–708. https://doi. org/10.1056/NEJMoa1308573.

- 94. de Groot JF, Lamborn KR, Chang SM, et al. Phase II study of aflibercept in recurrent malignant glioma: a North American Brain Tumor Consortium study. J Clin Oncol. 2011;29(19): 2689–2695. https://doi.org/10.1200/ JCO.2010.34.1636.
- Gomez-Manzano C, Holash J, Fueyo J, et al. VEGF Trap induces antiglioma effect at different stages of disease. Neuro Oncol. 2008;10(6): 940–945. https://doi. org/10.1215/15228517-2008-061.
- Kreisl TN, Kotliarova S, Butman JA, et al. A phase I/II trial of enzastaurin in patients with recurrent high-grade gliomas. Neuro Oncol. 2010;12(2): 181–189. https://doi. org/10.1093/neuonc/nop042.
- Baltuch GH, Couldwell WT, Villemure JG, Yong VW. Protein kinase C inhibitors suppress cell growth in established and low-passage glioma cell lines. A comparison between staurosporine and tamoxifen. Neurosurgery. 1993;33(3): 495–501; discussion 501. https://doi.org/10. 1227/00006123-199309000-00021.
- Robins HI, Won M, Seiferheld WF, et al. Phase 2 trial of radiation plus high-dose tamoxifen for glioblastoma multiforme: RTOG protocol BR-0021. Neuro Oncol. 2006;8(1): 47–52. https://doi.org/10.1215/S1522851705000311.
- Tabatabai G, Tonn JC, Stupp R, Weller M. The role of integrins in glioma biology and anti-glioma therapies. Curr Pharm Des. 2011;17(23): 2402–2410. https://doi. org/10.2174/138161211797249189.
- 100. Corsini NS, Martin-Villalba A. Integrin alpha 6: anchors away for glioma stem cells. Cell Stem Cell. 2010;6(5): 403–404. https://doi.org/10.1016/j.stem.2010.04.003.
- 101. Reardon DA, Nabors LB, Stupp R, Mikkelsen T. Cilengitide: an integrin-targeting arginine-glycine-aspartic acid peptide with promising activity for glioblastoma multiforme. Expert Opin Investig Drugs. 2008;17(8): 1225–1235. https://doi.org/10.1517/13543784.17.8.1225.
- 102. Gilbert MR, Kuhn J, Lamborn KR, et al. Cilengitide in patients with recurrent glioblastoma: the results of NABTC 03-02, a phase II trial with measures of treatment delivery. J Neurooncol. 2012;106(1): 147–153. https://doi. org/10.1007/s11060-011-0650-1.
- 103. Nabors LB, Fink KL, Mikkelsen T, et al. Two cilengitide regimens in combination with standard treatment for patients with newly diagnosed glioblastoma and unmethylated MGMT gene promoter: results of the open-label, controlled, randomized phase II CORE study. Neuro Oncol. 2015; 17(5): 708–717. https://doi.org/10.1093/neuonc/nou356.
- 104. Fadul CE, Kingman LS, Meyer LP, et al. A phase II study of thalidomide and irinotecan for treatment of glioblastoma multiforme. J Neurooncol. 2008;90(2): 229–235. https://doi.org/10.1007/s11060-008-9655-9.
- 105. Alexander BM, Wang M, Yung WK, et al. A phase II study of conventional radiation therapy and thalidomide for supratentorial, newly-diagnosed glioblastoma (RTOG 9806). J Neurooncol. 2013;111(1): 33–39. https://doi. org/10.1007/s11060-012-0987-0.

- 106. Fine HA, Kim L, Albert PS, et al. A phase I trial of lenalidomide in patients with recurrent primary central nervous system tumors. Clin Cancer Res. 2007;13(23): 7101– 7106. https://doi.org/10.1158/1078-0432.CCR-07-1546.
- 107. Vlachostergios PJ, Hatzidaki E, Befani CD, Liakos P, Papandreou CN. Bortezomib overcomes MGMT-related resistance of glioblastoma cell lines to temozolomide in a schedule-dependent manner. Invest New Drugs. 2013;31(5): 1169–1181. https://doi.org/10.1007/s10637-013-9968-1.
- 108. Potts BC, Albitar MX, Anderson KC, et al. Marizomib, a proteasome inhibitor for all seasons: preclinical profile and a framework for clinical trials. Curr Cancer Drug Targets. 2011;11(3): 254–284. https://doi. org/10.2174/156800911794519716.
- 109. Parisi P, Vanacore N, Belcastro V, et al. Clinical guidelines in pediatric headache: evaluation of quality using the AGREE II instrument. J Headache Pain. 2014;15: 57. https://doi.org/10.1186/1129-2377-15-57.
- 110. Foiadelli T, Piccorossi A, Sacchi L, et al. Clinical characteristics of headache in Italian adolescents aged 11-16 years: a cross-sectional questionnaire school-based study. Ital J Pediatr. 2018;44(1): 44. https://doi.org/10.1186/ s13052-018-0486-9.
- 111. Garone G, Reale A, Vanacore N, et al. Acute ataxia in paediatric emergency departments: a multicentre Italian study. Arch Dis Child. 2019;104(8): 768–774. https://doi. org/10.1136/archdischild-2018-315487.
- 112. Polivka J, Jr., Polivka J, Rohan V, Topolcan O, Ferda J. New molecularly targeted therapies for glioblastoma multiforme. Anticancer Res. 2012;32(7): 2935-2946.
- 113. Chen R, Cohen AL, Colman H. Targeted Therapeutics in Patients With High-Grade Gliomas: Past, Present, and Future. Curr Treat Options Oncol. 2016;17(8): 42. https:// doi.org/10.1007/s11864-016-0418-0.
- 114. Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. Cell. 2011;144(5): 646–674. https://doi. org/10.1016/j.cell.2011.02.013.
- 115. Louis DN, Perry A, Reifenberger G, et al. The 2016 World Health Organization Classification of Tumors of the Central Nervous System: a summary. Acta Neuropathol. 2016;131(6): 803–820. https://doi.org/10.1007/s00401-016-1545-1.
- 116. Capper D, Jones DTW, Sill M, et al. DNA methylationbased classification of central nervous system tumours. Nature. 2018;555(7697): 469–474. https://doi.org/10.1038/ nature26000.
- 117. Le Rhun E, Preusser M, Roth P, et al. Molecular targeted therapy of glioblastoma. Cancer Treat Rev. 2019;80: 101896. https://doi.org/10.1016/j.ctrv.2019.101896.
- 118. Lassman AB, Rossi MR, Raizer JJ, et al. Molecular study of malignant gliomas treated with epidermal growth factor receptor inhibitors: tissue analysis from North American Brain Tumor Consortium Trials 01-03 and 00-01. Clin Cancer Res. 2005;11(21): 7841–7850. https://doi. org/10.1158/1078-0432.CCR-05-0421.

- 119. van den Bent MJ, Gao Y, Kerkhof M, et al. Changes in the EGFR amplification and EGFRvIII expression between paired primary and recurrent glioblastomas. Neuro Oncol. 2015;17(7): 935–941. https://doi.org/10.1093/neuonc/ nov013.
- 120. Ma DJ, Galanis E, Anderson SK, et al. A phase II trial of everolimus, temozolomide, and radiotherapy in patients with newly diagnosed glioblastoma: NCCTG N057K. Neuro Oncol. 2015;17(9): 1261–1269. https://doi. org/10.1093/neuonc/nou328.
- 121. Wick W, Dettmer S, Berberich A, et al. N2M2 (NOA-20) phase I/II trial of molecularly matched targeted therapies plus radiotherapy in patients with newly diagnosed non-MGMT hypermethylated glioblastoma. Neuro Oncol. 2019;21(1): 95–105. https://doi.org/10.1093/neuonc/ noy161.
- 122. Anthony C, Mladkova-Suchy N, Adamson DC. The evolving role of antiangiogenic therapies in glioblastoma multiforme: current clinical significance and future potential. Expert Opin Investig Drugs. 2019;28(9): 787–797. https://doi.org/10.1080/13543784.2019.1650019.
- 123. Mastrella G, Hou M, Li M, et al. Targeting APLN/ APLNR Improves Antiangiogenic Efficiency and Blunts Proinvasive Side Effects of VEGFA/VEGFR2 Blockade in Glioblastoma. Cancer Res. 2019;79(9): 2298–2313. https://doi.org/10.1158/0008-5472.CAN-18-0881.
- 124. Roth P, Silginer M, Goodman SL, et al. Integrin control of the transforming growth factor-beta pathway in glioblastoma. Brain. 2013;136(Pt 2): 564–576. https://doi. org/10.1093/brain/aws351.
- 125. Kong XT, Nguyen NT, Choi YJ, et al. Phase 2 Study of Bortezomib Combined With Temozolomide and Regional Radiation Therapy for Upfront Treatment of Patients With Newly Diagnosed Glioblastoma Multiforme: Safety and Efficacy Assessment. Int J Radiat Oncol Biol Phys. 2018;100(5): 1195–1203. https://doi.org/10.1016/j. ijrobp.2018.01.001.
- 126. Esteller M, Garcia-Foncillas J, Andion E, et al. Inactivation of the DNA-repair gene MGMT and the clinical response of gliomas to alkylating agents. N Engl J Med. 2000;343(19): 1350–1354. https://doi.org/10.1056/ NEJM200011093431901.
- 127. Osuka S, Van Meir EG. Overcoming therapeutic resistance in glioblastoma: the way forward. J Clin Invest. 2017;127(2): 415–426. https://doi.org/10.1172/JCI89587.

- 128. Noch EK, Ramakrishna R, Magge R. Challenges in the Treatment of Glioblastoma: Multisystem Mechanisms of Therapeutic Resistance. World Neurosurg. 2018;116: 505–517. https://doi.org/10.1016/j.wneu.2018.04.022.
- 129. Luzzi S, Zoia C, Rampini AD, et al. Lateral Transorbital Neuroendoscopic Approach for Intraconal Meningioma of the Orbital Apex: Technical Nuances and Literature Review. World Neurosurg. 2019;131: 10–17. https://doi. org/10.1016/j.wneu.2019.07.152.
- 130. Arnaout MM, Luzzi S, Galzio R, Aziz K. Supraorbital keyhole approach: Pure endoscopic and endoscope-assisted perspective. Clin Neurol Neurosurg. 2020;189: 105623. https://doi.org/10.1016/j.clineuro.2019.105623.
- 131. Millimaggi DF, Norcia VD, Luzzi S, Alfiero T, Galzio RJ, Ricci A. Minimally Invasive Transforaminal Lumbar Interbody Fusion with Percutaneous Bilateral Pedicle Screw Fixation for Lumbosacral Spine Degenerative Diseases. A Retrospective Database of 40 Consecutive Cases and Literature Review. Turk Neurosurg. 2018;28(3): 454–461. https://doi.org/10.5137/1019-5149.JTN.19479-16.0.
- 132. Da Ros M, De Gregorio V, Iorio AL, et al. Glioblastoma Chemoresistance: The Double Play by Microenvironment and Blood-Brain Barrier. Int J Mol Sci. 2018;19(10). https://doi.org/10.3390/ijms19102879.
- 133. Jia D, Li S, Li D, Xue H, Yang D, Liu Y. Mining TCGA database for genes of prognostic value in glioblastoma microenvironment. Aging (Albany NY). 2018;10(4): 592– 605. https://doi.org/10.18632/aging.101415.
- 134. Chen Z, Hambardzumyan D. Immune Microenvironment in Glioblastoma Subtypes. Front Immunol. 2018;9: 1004. https://doi.org/10.3389/fimmu.2018.01004.

Neurosurgery Unit, Department of Clinical-Surgical, Diagnostic and Pediatric Sciences, University of Pavia Polo Didattico "Cesare Brusotti", Viale Brambilla, 74

Received: 10 May 2020

Accepted: 1 June 2020

Correspondence:

Sabino Luzzi M.D., Ph.D.

^{27100 -} Pavia (Italy)

E-mail: sabino.luzzi@unipv.it