No association between *ECSIT* germline mutations and hemophagocytic lymphohistiocytosis in natural killer/T-cell lymphoma

Hemophagocytic lymphohistiocytosis (HLH) is a clinical syndrome of excessive immune activation with fever, cytopenia, and organ infiltration by activated macrophages. Secondary HLH associated with natural killer (NK)/T-cell lymphoma (NKTCL) has extremely poor prognosis,¹ and biomarkers that may predict patients who are more likely to develop HLH are lacking. Wen et al.² recently showed an association between a somatic gene mutation in the evolutionarily conserved signaling intermediate in Toll pathway (ECSIT) gene (c.T419C; p.V140A) and HLH in NKTCL. The variant ECSIT protein triggered NF-kB signaling pathway more potently, leading to aberrant secretion of proinflammatory cytokines by ECSIT-T419C-transfected NKTCL cell lines. They found that the ECSIT-T419 mutation was significantly enriched in individuals with NKTCL-associated HLH, which developed in nine of 17 patients with and five of 36 patients without the mutation, respectively. Patients with ECSIT-T419 had elevated expression of proinflammatory cytokines and poorer prognosis. While intriguing, the prevalence of ECSIT-T419 and relation with HLH has not been assessed in independent cohorts. We therefore sought to examine whether the ECSIT-T419 mutation predisposes to HLH in multiple cohorts of patients with NKTCL and correlates its presence with clinical outcomes.

First, we studied the mutational profile of ECSIT in 25 subjects with sporadic NKTCL from China with available whole exome sequencing of paired tumour-blood samples.³ Samples were sequenced with Illumina HiSeq X and NextSeq 6000, and variant-calling was performed by Strelka2 using default single-sample settings.⁴ We found the ECSIT-T419 mutation in five of 25 subjects, but they were all germline mutations; heterozygously mutated in both matching tumour (variant allele frequency [VAF], mean, 43.8%, 95% Confidence Interval [CI]: 38.8-48.9) and blood (VAF mean, 53.8%, 95%CI: 51.5-56.2) samples from these five subjects (Figure 1A). The reported prevalence of somatic ECSIT-T419 mutation in Wen et al.'s study was 19.3% (17 of 88), similar to the mutation frequency of Jiang et al.'s cohort, but were all germline mutations.

In order to further verify whether ECSIT-T419 is a germline or somatic mutation, we studied 67 patients with NKTCL who provided written informed consent under respective institutions' Institutional Review Boards (IRB) from Singapore local hospitals and Sun Yat-Sen University Cancer Center in Guangzhou, China. We Sanger sequenced matched tumor- buccal swab (representative Sanger sequence in Figure 1B) or peripheral blood (representative Sanger sequence in Figure 1C) samples from NKTCL patients and ECSIT-T419 was validated in 7.5% (five of 67) of both the tumor and matching non-tumor samples. Targeted resequencing using nextgeneration sequencing method revealed the mean VAF of ECSIT-T419 to be 52.2%, 95%CI: 42.8-61.6 in the five tumors, 52.2%, 95%CI: 48.5-56.0 in four matched blood samples, and 53% in a matched buccal swab sample (P=0.90, 2-tailed Wilcoxon Rank-sum test, VAF of ECSIT-T419 between tumors and non-tumoral samples). The 50% VAF, the presence near and of ECSIT-T419 mutation observed in all matching tumor, blood and buccal swab DNA indicate that this is a germline heterozygous single-nucleotide polymorphism

(SNP), with a report SNP ID of rs145036301. Among the five patients with *ECSIT*-T419 mutation, HLH information was available for three patients and none developed HLH, as defined by the HLH 2004 criteria.⁵

Given the discrepant findings, we re-analyzed the initial discovery cohort of paired tumor-normal exome data (n=5) from Wen *et al.*² In the sample where *ECSIT*-T419 mutation was reported as a somatic mutation, VAF was 52% in the tumor (150 of 288; alternate allele depth/reference allele depth) and 10% (5 of 51) in the matched normal sample (Figure 1D). Furthermore, the VAF exceeded the thresholds of 30% in tumor and 5% in matched normal sample as specified by Wen *et al.*² Thus, this variant should not be considered as a somatic mutation as based on the authors' analysis criteria.

Notwithstanding the false somatic call, we wanted to examine whether the germline ECSIT-T419 mutation is associated with HLH in two independent cohorts of patients with NKTCL in Singapore and Taiwan. In Singapore, the cases were identified using local databases from two teaching hospitals and all samples and clinical information were collected after IRB approval. Cases were reviewed by a central pathologist and HLH was defined according to the HLH 2004 criteria. Sixty-four cases of NKTCL were identified between 2007-2017, and ECSIT-T419 mutations were found in 15.4% (two of 13) and 5.9% (two of 51) patients with and without HLH respectively. Out of the 13 patients with HLH, four were women. Median age was 43 (range, 18-60 years). At time of the last follow-up in December 2018, all patients had died. Seven of 12 patients received polychemotherapy, while one was treated with the HLH-2004 protocol (with dexamethasone, etoposide, cyclosporin), and two received steroids. Median survival was only 33 days (range, 1-389 days). Causes of death were lymphoma (n=6), HLH (n=6), and infection (n=1). The two individuals with ECSIT mutation succumbed at day 1 and day 89. Within these NKTCL patients with HLH in our Singapore cohort, there was no significant association of the ECSIT mutation with them (P=0.18, Fisher's exact test, Online Supplementary Table S1).

In the Taiwanese cohort of 85 NKTCL cases with clinical and sequencing data from the Chang Gung Memorial Hospital, ECSIT-T419 mutation frequency was observed at 11.8% (ten of 85). Nine cases developed HLH, and none of these samples harboured the ECSIT-T419C mutation. When both Singapore and Taiwan cohorts were combined for analysis, we did not find any statistical association between ECSIT-T419C mutation and HLH (OR=1.48, 95%CI: 0.38-5.76, P≈1.0, Fisher's exact test, Online Supplementary Table S2). There were also no significant associations between the ECSIT-T419C mutation with clinical characteristics such as sex, stage, Eastern Cooperative Oncology Group (ECOG) performance status and international prognostic index (Figure 1E; Online Supplementary Table S3). Overall survival (Online Supplementary Figure S1A) and progression-free survival (Online Supplementary Figure S1B) were also not significantly associated with ECSIT-T419 mutation.

Given the rarity and fulminant nature of malignancyassociated HLH hindering the collection of biopsy specimens, we combined data from multiple cohorts to examine associations between *ECSIT*-T419 and HLH in NKTCL in the largest study to date. Strict diagnostic inclusion criteria were used for both HLH and NKTCL. Some possible explanations for the discordant results between Wen *et al.* and our study need consideration. Patients in Singapore and Taiwan developed HLH at around the time of diagnosis or relapse, as opposed to Wen *et al.*'s cohort which developed HLH 3 to 6 months after diagnosis of NKTCL, during or after treatment. The onset of HLH might be triggered by the initiation of chemotherapy that leads to loss of immune homeostasis and further aggravates T-cell dysfunction which may further lower the threshold for triggering HLH in lymphoma

patients.⁶ It is possible that in the absence of chemotherapy in our patients, the activating effect of the *ECSIT*-T419 mutation on the NF-KB pathway is not strong enough to drive HLH. However, there were four *ECSIT* wild-type patients from Singapore who developed HLH again after chemotherapy initiation.



Figure 1. ECSIT-T419C is a germline mutation not associated with hemophagocytic lymphohistiocytosis in natural killer/T-cell lymphoma patients. (A) Sanger sequencing electropherogram profile for tumor-normal paired samples with heterozygous ECSIT-V140A mutation, identified as L12, L14, L20, L21, and L24 in Jiang et al.³ (B and C) Representative Sanger sequencing electropherogram profile for two tumor- peripheral blood (B) and buccal swab (C) samples for the ECSIT-V140A mutation from Singapore local hospitals and the Sun Yat-Sen University Cancer Center in Guangzhou, China. (D) Integrative Genomics Viewer (IGV) snap-shot centered around heterozygous germline ECSIT-T419C mutation of the paired tumor-normal exome sequencing data of sample NKT1 from Wen et al.² Variant allele frequencies (VAF) were calculated from the number of variant-supporting/total read-counts at ECSIT-T419C. Aligned reads were colored pink according to the read-strand that they were aligned with onto the human reference genome. (E) No association between ECSIT mutation and clinical characteristics of natural killer/T-cell lymphoma patients in Singapore and Taiwan. ECSIT: evolutionarily conserved signaling intermediate in Toll pathway; IPI: international prognostic index, ECOG: Eastern Cooperative Oncology Group, HLH: hemophagocytic lymphohisticcytosis, Mut: mutant; WT: wild+ype.

Differences in other patient characteristics may also explain the discordance (e.g., patients with HLH in Singapore had stage III or IV disease, while most patients with HLH in Wen *et al.*'s cohort had early stage disease).

In summary, our data from multiple cohorts do not support the risk effect of ECSIT-T419 mutation (SNP rs145036301) on HLH in NKTCL. Furthermore, this is a germline rather than somatic mutation that appears in SNP database (dbSNP v153) and has now been flagged as a common polymorphic variant by Catalogue of Somatic Mutations in Cancer (COSMIC v90) databases.⁷ Additionally, there were no differences in clinical characteristics or prognosis between NKTCL patients with and without ECSIT-T419 mutation. One limitation of our study is not being able to examine whether germline variants in genes associated with familial HLH are enriched in patients with NKTCL-associated HLH. However, recent studies have not shown an association of biallelic pathogenic variants in HLH-associated genes with adult HLH, albeit in cohorts that comprise a mixture of lymphoma and non-lymphoma subtypes.9,10 Ultimately, additional efforts to define disruptive variants in a larger number of genes, in expanded cohorts of adults with lymphoma associated HLH, may further refine our understanding and treatment of this devastating condition.

Shin Yeu Ong,^{1*} Jing Quan Lim,^{2,3*} Nicholas Grigoropoulos,¹ Yurike Laurensia,² Dachuan Huang,^{2,3} Burton Kuan Hui Chia,² Daryl Cheah Ming Zhe,² Sahil Ajit Saraf,⁴ Chee Leong Cheng,⁴ Wen-Yu Chuang,⁵ Ming-Chung Kuo,⁶ Yi-Jiun Su,⁶ Colin Phipps,¹ Chandramouli Nagarajan,¹ Yuh Shan Lee,¹ Daryl Tan Chen Lung,¹ Lee-Yung Shih,⁶ Yeow Tee Goh,¹ Soon Thye Lim^{2,3#} and Choon Kiat Ong^{2,3,7#}

¹Department of Haematology, Singapore General Hospital, Singapore; ²National Cancer Center, Singapore; ³Duke-NUS Medical School, Singapore; ⁴Department of Pathology, Singapore General Hospital, Singapore; ⁵Department of Pathology, Chang Gung Memorial Hospital at Linkou and Chang Gung University, Taoyuan, Taiwan; ⁶Division of Hematology-Oncology, Chang Gung Memorial Hospital at Linkou, and Chang Gung University, Taoyuan, Taiwan and ⁷Genome Institute of Singapore, A*STAR, Singapore.

*SYO and JQL contributed equally as co-first authors.

**STL and CKO contributed equally as co-senior authors.*

Correspondence:

CHOON KIAT ONG - cmrock@nccs.com.sg

SOON THYE LIM - lim.soon.thye@singhealth.com.sg doi:10.3324/haematol.2020.269209

Received: August 12, 2020. Accepted: September 30, 2020. Pre-published: October 13, 2020. Disclosures: no conflicts of interest to disclose.

Contributions: CKO conceived the project and designed the study; SYO drafted the initial manuscript; CCL, SAS, and WYC performed pathological studies; YL performed sequencing studies; JQL and BKHC performed the bioinformatics analysis; SYO, JQL, DCH, DCMZ, CP, CN, YSL, DTCL, STL, MCK, YJS, and LYS recruited participants, managed subject information and tissue samples, and contributed to data analysis; CKO, JQL, NG and YTG participated in critical revision of the manuscript.

Funding: the study was supported by grants from the Singapore Ministry of Health's National Medical Research Council (NMRC-OFLCG-18May0028 and NMRC-ORIRG16nov090), Tanoto Foundation Professorship in Medical Oncology, New Century International Pte Ltd, Ling Foundation, and Chang Gung Memorial Hospital (OMRPG3C0021), Taiwan.

References

- Jin Z, Wang Y, Wang J, et al. Multivariate analysis of prognosis for patients with natural killer/T cell lymphoma-associated hemophagocytic lymphohistiocytosis. Hematology. 2018;23(4):228-234.
- 2. Wen H, Ma H, Cai Q, et al. Recurrent ECSIT mutation encoding V140A triggers hyperinflammation and promotes hemophagocytic syndrome in extranodal NK/T cell lymphoma. Nat Med. 2018; 24(2):154-164.
- Jiang L, Gu ZH, Yan ZX, et al. Exome sequencing identifies somatic mutations of DDX3X in natural killer/T-cell lymphoma. Nat Genet. 2015;47(9):1061-1066.
- Kim S, Scheffler K, Halpern AL, et al. Strelka2: fast and accurate calling of germline and somatic variants. Nat Methods. 2018;15(8):591-594.
- 5. Henter JI, Horne A, Arico M, et al. HLH-2004: diagnostic and therapeutic guidelines for hemophagocytic lymphohistiocytosis. Pediatr Blood Cancer. 2007;48(2):124-131.
- Daver N, McClain K, Allen CE, et al. A consensus review on malignancy-associated hemophagocytic lymphohistiocytosis in adults. Cancer. 2017;123(17):3229-3240.
- Smigielski EM, Sirotkin K, Ward M, et al. dbSNP: a database of single nucleotide polymorphisms. Nucleic Acids Res. 2000;28(1):352-355.
- Tate JG, Bamford S, Jubb HC, et al. COSMIC: the Catalogue of Somatic Mutations In Cancer. Nucleic Acids Res. 2019; 47(D1):D941-D947.
- 9. Miller PG, Niroula A, Ceremsak JJ, et al. Identification of germline variants in adults with hemophagocytic lymphohistiocytosis. Blood Adv. 2020;4(5):925-929.
- 10. Carvelli J, Piperoglou C, Farnarier C, et al. Functional and genetic testing in adults with HLH reveals an inflammatory profile rather than a cytotoxicity defect. Blood. 2020;136(5):542-552.