LincSNP 3.0: an updated database for linking functional variants to human long non-coding RNAs, circular RNAs and their regulatory elements

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ABSTRACT

We describe an updated comprehensive database, LincSNP 3.0 (http://bioinfo.hrbmu.edu.cn/LincSNP), which aims to document and annotate disease or phenotype-associated variants in human long noncoding RNAs (IncRNAs) and circular RNAs (circR-NAs) or their regulatory elements. LincSNP 3.0 has updated with several novel features, including (i) more types of variants including single nucleotide polymorphisms (SNPs), linkage disequilibrium SNPs (LD SNPs), somatic mutations and RNA editing sites have been expanded; (ii) more regulatory elements including transcription factor binding sites (TFBSs), enhancers, DNase I hypersensitive sites (DHSs), topologically associated domains (TADs), footprintss, methylations and open chromatin regions have been added; (iii) the associations among circRNAs, regulatory elements and variants have been identified; (iv) more experimentally supported variant-IncRNA/circRNA-disease/phenotype associations have been manually collected; (v) the sources of IncRNAs, circRNAs, SNPs, somatic mutations and RNA editing sites have been updated. Moreover, four flexible online tools including Genome Browser, Variant Mapper, Circos Plotter and Functional Annotation have been developed to retrieve, visualize and analyze the data. Collectively, LincSNP 3.0 provides associations among functional variants, regulatory elements, IncRNAs and circRNAs in diseases. It will serve as an important and continually updated resource for investigating functions and mechanisms of IncRNAs and circRNAs in diseases.

INTRODUCTION

Emerging evidences have shown the importance of long non-coding RNAs (lncRNAs) as novel regulators of many physical or pathological processes (1). LncRNAs are a highly versatile class of transcripts that have sparked new lines of research in nearly all fields of life sciences (2). More recently, accumulating evidence has indicated that lncRNAs are major players in many types of disease processes. Circular RNAs (circRNAs) are considered as a kind of special lncRNAs and represent a recent research hotspot in the field of RNA. Unlike linear RNA molecules, circRNAs are closed circular molecules with a covalently closed loop structure that lack 5'-3' polarity or a polyadenylated tail (3). CircRNAs play a critical role in biological processes and are reported to participate in multiple disease processes, and thus, these molecules offer new potential opportunities for therapeutic intervention (4).

With the in-depth research for lncRNA, many researchers continue focusing on the influences of genetic variants on lncRNA function. In order to facilitate the study of lncRNA-related genetic variants, we reported the first and second versions of the LincSNP database (Linc-SNP 1.0 and 2.0) that allows users to search all known disease-associated single nucleotide polymorphisms (SNPs) to human lncRNAs and their transcription factor binding sites (TFBSs). In fact, there are many other kinds of functional variants besides SNPs. For example, somatic mutations, considered as genomic variation phenomena, can directly or indirectly alter lncRNA expression, protein activities, and signaling pathways (5). RNA editing is discovered as a process of variant at RNA level and represents a widespread and prominent post-transcriptional mechanism (6). Its deregulation has been implicated in the pathogenesis of many functions and diseases (7).

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Encyclopedia of DNA elements (ENCODE) project announces >1.2 million candidate functional regulatory elements in human and mouse. This project shows that focusing on variants in regulatory elements within the noncoding regions can help to discover the real pathogenic variations (8). The ENCODE project has put forward a lot of novel insights on the regulated ways at the genome level, and identified many new types of regulatory elements such as TFBSs, enhancers, DNase I hypersensitive sites (DHSs), topologically associated domains (TADs), footprints and open chromatin regions (9). These functional elements could directly or indirectly influence the lncRNAs and circRNAs (10–12). In addition, SNPs can contribute to disease progression by influencing DNA methylation. DNA methylation quantitative trait loci (meOTL) have been identified in physiological and pathological contexts (13). Exploring the functional variants in these regulatory elements of lncRNAs and circRNAs could provide novel insights for investigating the functions and mechanisms of lncRNAs and circRNAs in human diseases.

To bridge this gap, we have updated LincSNP (14,15) to version 3.0 (LincSNP 3.0) (Figure 1 and Table 1). The database aims specifically to store and annotate disease or phenotype-associated variants including SNPs, linkage disequilibrium SNPs (LD SNPs), somatic mutations and RNA editing in human lncRNAs and circRNAs or their regulatory elements including TFBS, enhancer, DHS, TAD, footprint and open chromatin region. In addition, the effects of SNPs on methylation for lncRNAs and circRNAs are also included. Specially, four useful web-based tools are also provided to facilitate data analysis, extraction and visualization. LincSNP 3.0 bridges the gap between functional variants and human lncRNAs or circRNAs to enhance our understanding of lncRNA and circRNA function, particularly their potential roles in human disease.

MATERIALS AND METHODS

Data collection and processing

Experimentally verified functional variants in lncRNAs, circRNAs and their regulatory elements. All experimentally supported variant-lncRNA/circRNA-disease/trait associations were manually collected through several steps, as previously described (16): (i) We downloaded all published literatures through the PubMed database (17) with a list of keywords (before March 2020), such as 'lncRNA/circRNA SNP disease,' 'lncRNA/circRNA mutation disease,' 'lncRNA/circRNA variant disease,' 'lncRNA/circRNA polymorphism disease' and 'lncRNA/circRNA variant trait.' (ii) Based on above published papers, experimentally supported variant-lncRNA/circRNA-disease/trait associations were manually curated by at least two researchers. We retrieved the lncRNAs, circRNAs, variants, phenotype and disease names, experimental samples and methods, PubMed IDs, paper titles and a brief description from the original studies. Only high-quality associations with multiple lines of strong experimental evidence were collected, including those confirmed by genotyping, western blot, qRT-PCR or luciferase reporter assays. (iii) All selected studies were rechecked for the lncRNAs, circRNAs, variants and disease names. In addition, some names were

replaced with their official or recommended name respectively, which acquired from Ensembl(18), circBase(19) and dbGAP(20) databases.

Functional variants. (i) SNP, the functional SNP information was obtained from nine high-quality databases including dbGaP (20), GAD (21), GWAS Central (22), Johnson and O'Donnell (23), the NHGRI GWAS Catalog (24). PharmGKb (25), GWASdb (Version 2) (26), GRASP (Version 2) (27) and LnCeVar (28). As the integrated strategy in LincSNP 2.0, functional SNPs were selected from original publications with moderate thresholds (*P*-values < 1.0 \times 10⁻³) and only the most significant SNP was selected in cases where the same SNP could be obtained from different publications. In total, 1 489 332 unique functional SNPs were collected. (ii) LD SNP, the SNPs that had linkage disequilibrium (LD-SNP, $r^2 \ge 0.8$) relationships with functional SNPs from the 1000 Genomes Project (Phase I, version 3) were extracted. After LD analysis by VCFtools (29), 4 315 056 LD-SNPs were collected in LincSNP 3.0. (iii) So*matic mutation*, the somatic mutations were collected from COSMIC (Catalogue Of Somatic Mutations In Cancer) database (30). Furthermore, the FATHMM scores in COS-MIC were used to screen functional somatic mutations. Following the suggestion of COSMIC, score >0.7 was considered as 'Pathogenic'. Thus, we extracted 2 492 320 functional somatic mutations. (iv) RNA editing site, RNA editing sites were collected and integrated from DARNED (31) and RADAR (32). The tissue information of RNA editing was obtained from REDIportal (33). Lastly, 3 881 664 RNA editing sites were extracted.

Regulatory elements. (i) TFBS, the TFBS information was downloaded from GTRD (34) and rSNPBase 3.0 (35). In total, we identified 222 740 257 and 108 488 967 TFBSs in the defined promoter regions of human lncRNAs and circRNAs. (ii) Enhancer, first, the enhancer annotation information was obtained from SELER (36) and SEdb (37). Second, we obtained chromatin interaction region data detected by Hi-C (High-throughput/resolution chromosome conformation capture) seq from ENCODE based on HiC-Pro (38). Then, genome intersection between enhancer and chromatin interaction region was considered as a specific enhancer for this cell line. (iii) DHS, DHSs were identified by DNase-seq which can be downloaded from UCSC genome browser (39). Only those identified in more than three samples were extracted. Totally, 1 511 655 DHSs were contained for following analysis. (iv) Footprint, similar to DHS, footprints were also screened by DNase-seq in more than three samples. 1 791 645 footprints were obtained based on above steps. (v) TAD, TADs of lncRNAs and circRNAs were download from database rSNPBase 3.0 (35). 10 223 498 and 5 617 695 TADs of lncRNAs and circRNAs were obtained. (vi) Open chromatin region, the open chromatin regions were obtained from a previous study which detected by ATAC-seq (Assay for Transposase-Accessible Chromatin with high throughput sequencing) of 410 tumor samples spanning 23 cancer types (40). Only the open chromatin regions identified in multiple kinds of cancer types were extracted. 562 709 open chromatin regions were ob-



Figure 1. Database content and user interface of LincSNP 3.0. The left area contains the database content, which includes collection of confirmed functional variants from literatures, identification of functional variants, regulatory elements and RNAs and construction of web tools. The methods of analyzing raw data and constructing tools are also provided. The right area contain the user interface of LincSNP 3.0. A panel of basic functions and tools have been developed to infer associations among functional variants, regulatory elements, lncRNAs and circRNAs.

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Features	LincSNP 2.0	LincSNP 3.0	Fold increase
Functional variant			
SNP	809 451	1 489 332	1.84
LD SNP	$\sim \! 1 \ 160 \ 000$	4 315 056	~ 3.72
Somatic mutation	_	2 492 320	New
RNA editing site	_	3 881 664	New
Regulatory element			
TFBS for lncRNA/circRNA	593 492/-	222 740 257/108 488 967	375.30/New
Enhancer for lncRNA/circRNA	_/	209 968/173 438	New/New
DHS for lncRNA/circRNA	_/_	5 560 190/2 868 132	New/New
Footprint for lncRNA/circRNA	_/_	9 084 051/3 258 258	New/New
TAD for lncRNA/circRNA	_/_	10 223 498/5 617 695	New/New
Open chromatin region for lncRNA/circRNA	_/ _	1 806 843/882 305	New/New
Meth-QTL for lncRNA/circRNA	_/ _	974 853/408 967	New/New
RNA			
LncRNA	244 545	287 313	1.17
CircRNA	-	173 207	New

tained. (*vii*) *Meth-QTL*, we obtained DNA methylation quantitative trait loci from database Pancan-meQTL (41).

LncRNA and circRNA. (i) lncRNA, five databases including Ensembl (18), LncRBase (42), NONCODE (43), LNCipedia (44) and GENCODE (45) were integrated for getting lncRNA information. To provide a universal lncRNA annotation for users, lncRNA transcripts downloaded from different sources were considered to be the same transcripts if they have the same positions. Lastly, 287 313 human lncRNAs were extracted. (ii) CircRNA, to construct comprehensive and systematic circRNA annotations, we collected predicted circRNAs supported by back-spliced junction sites from the circBase database (19) and another five studies (46-50). Combining all these studies, a total of 173 207 unique circRNAs were included for subsequent analyses. All the annotations of functional variants, regulatory elements and RNAs were transformed to GRCh38 (The Genome Reference Consortium Human Genome Build 38).

Linking functional variants to human lncRNAs, circRNAs and their regulatory elements

In order to construct associations among functional variants, lncRNAs, circRNAs and their regulatory elements, BEDTools (51) was applied to match genomic locations. The associations between regulatory elements and lncR-NAs or circRNAs were based on following two sections: (i) *trans*-regulatory elements were obtained from current highquality databases; (ii) *cis*-regulatory elements were located in the promoter regions of human lncRNAs or circRNAs (5 kb upstream to 1 kb downstream region of the transcription start site for each lncRNA or circRNA). The functional variants must be located at regions of lncRNAs, circRNAs or their regulatory elements. After the above steps, functional variants to lncRNAs, circRNAs and their regulatory elements were linked.

Database construction and improved user interface

All data in LincSNP 3.0 were stored and managed using MySQL (version 5.5.58). The web interfaces were upgraded by applying Linux, Apache, MySQL and JSP (pre hypertext processor) (LAMP) technologies. LincSNP 3.0 is freely available to the research community at http:// bioinfo.hrbmu.edu.cn/LincSNP. The LincSNP 3.0 online web server was developed using Java Server Pages within Tomcat software (v6). Result tables and visualization of data were implemented by using jQuery (v1.11.3), data table (1.10.10) and Highcharts (V4.0) plugin software. All statistical analyses were performed on R framework (V3.6.3). The LincSNP 3.0 website is well supported by several popular web browsers, such as Microsoft Edge, Google Chrome, Firefox and Safari. In addition, for the convenience of users who have used LincSNP 1.0 and 2.0, the old versions are still in service. Researchers can enter them by clicking on the gateways in the LincSNP 3.0 homepage.

RESULTS

Newly designed and more humanized user-friendly interface

LincSNP 3.0 provides a user-friendly web interface that enables users to search, browse, visualize and download data in a few easy steps (Figure 2) including (*i*) Variant Confirm (Figure 2A, B), a retrieval module for experimentally verified lncRNA/circRNA-variants associations. (*ii*) Functional Variant-Centric (Figure 2C), a retrieval module for searching diverse types of functional variants located on lncRNAs, circRNAs or their regulatory elements. (*iii*) RNA-centric (Figure 2D, E), a retrieval module for searching diverse types of RNAs associated with their regulatory elements and different kinds of functional variants. (*iv*) Download, a module for downloading the associations among variants and lncRNAs, circRNAs and (*v*) Help, a module with detailed documentation of user tutorials.

Web-based online analysis tools of LincSNP 3.0

In order to allow users to explore the features and functions of variants on lncRNAs and circRNAs, four newly designed web-based online tools were developed. These tools could provide assistant for users conveniently to browse, retrieve, visualize and analyze the data. *(i) Genome Browser* (Figure 2F), provides a user-friendly interface for navigating the transcript structures and visualizing the variant sites of the lncRNA or circRNA transcripts in specific diseases.



Figure 2. Workflow and examples of using LincSNP 3.0. (A) The variant confirmed search interface of LincSNP 3.0 by querying an example of lung cancer. (B) The query results and detailed information of variants, RNAs, disease, biomarkers and literatures in LincSNP 3.0. (C) The functional variant search interface of LincSNP 3.0 by querying an example of lung cancer and blood. (D) The RNA-centric search interface of LincSNP 3.0 by querying an example of lung cancer and blood. (D) The RNA-centric search interface of LincSNP 3.0. (F) The query results and detailed information of variants, RNAs and regulatory elements in LincSNP 3.0. (F) The variants and regulatory elements landscape of PCAT19 in LincSNP 3.0 Genome Browser. (G) Interested variants map to lncRNAs and circRNAs in LincSNP 3.0 Variant Mapper. (H) The circos plot diagram including variants, regulatory and lncRNAs for lung cancer in LincSNP 3.0 Circos Plotter. (I) The functional enrichment analysis for variants in LincSNP 3.0 Functional Annotation.

Users could browse the loci of the lncRNAs, circRNAs and the variants by submitting the genomic interval. Tracks for types of variants and regulatory elements could be selected and added into the 'Search' page. (ii) Variant Mapper (Figure 2G), allows users to give interested genomic locations of variants. This tool could implement an integrated pipeline to match the variants on lncRNAs, circRNAs and their regulatory elements. (iii) Circos Plotter (Figure 2H), is a data visualizer which could provide a genome circos plot for variants. lncRNAs, cirRNAs and their regulatory elements in a specific disease or trait. It visualizes data in a circular layout for exploring relationships between variants, lncRNAs, circRNAs or their regulatory elements. (iv) *Functional Annotation* (Figure 2I), provides functional annotations from Gene ontology (GO) and Kyoto Encyclopedia of Genes and Genomes (KEGG) pathways based on GREAT (52).

DISCUSSION AND FUTURE EXTENSIONS

With the completion of the latest research phase of EN-CODE project, the researchers discover that 80% of the genome contains elements link to biochemical functions. It also shows that many variants previously correlated with certain diseases lie within or very near the non-coding functional DNA elements, providing new leads for linking genetic variations and diseases (53). The functions and mechanisms of lncRNAs, including circRNAs, need to be further explored and analyzed. Understanding and exploring the variants, regulatory elements, lncRNA and circRNA associations in disease or phenotype will greatly help to reveal the mystery of biological functions. Thus, we construct an updated database, LincSNP 3.0, which aims to integrate functional variants, lncRNAs, circRNAs and their regulatory elements. We believe that LincSNP 3.0 will be a useful resource for functional variants in lncRNAs and circRNAs.

More types of variants including SNPs, LD SNPs, somatic mutations and RNA editing were identified and annotated. We annotate RNA editing sites on DNA regulatory elements to show genome distribution for them. In future work, we will focus on if there are something transcribed from these regulatory regions and annotate the RNA editing sites on RNA level to improve our database. Many kinds of regulatory elements containing TFBSs, enhancers, DHSs, TADs, footprints and open chromatin regions were included. CircRNA information was also added and improved. To improve the functions of data processing and database access, four web-based tools, Genome Browser, Variant Mapper, Circos Plotter and Functional annotation, were developed. We expect that more disease and phenotype-associated variants are mapped to lncR-NAs, circRNAs and their regulatory elements and to collect the newly identified regulatory elements for lncRNAs and circRNAs. We will continually maintain and update the LincSNP database and integrate more data sets. We expect that through continuous improvements, LincSNP can become an effective tool to analyze the functional variants of IncRNAs and circRNAs and even contribute to disease diagnosis and treatment.

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