SAGE-Hindawi Access to Research International Journal of Hepatology Volume 2011, Article ID 841407, 10 pages doi:10.4061/2011/841407

## Review Article

## **Management of Hepatic Encephalopathy**

## G. Wright, 1 A. Chattree, 2 and R. Jalan 1

<sup>1</sup> University College London Institute of Hepatology, The Royal Free Hospital, Pond Street, London NW3 2PF, UK

Correspondence should be addressed to G. Wright, gavin.wright@ucl.ac.uk

Received 28 April 2011; Accepted 8 June 2011

Academic Editor: Deepak Amarapurkar

Copyright © 2011 G. Wright et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Hepatic encephalopathy (HE), the neuropsychiatric presentation of liver disease, is associated with high morbidity and mortality. Reduction of plasma ammonia remains the central therapeutic strategy, but there is a need for newer novel therapies. We discuss current evidence supporting the use of interventions for both the general management of chronic HE and that necessary for more acute and advanced disease.

### 1. Introduction

There are a plethora of therapeutic approaches to targeting varying severities of hepatic encephalopathy (HE), the neuropsychiatric presentations of liver disease. There is a need for newer therapies for patients with advanced HE and worsening acute liver injury. Reduction of plasma ammonia remains the central strategy although novel strategies may be beneficial. We discuss current evidence supporting the use of therapeutic interventions for both the general management of chronic HE and that necessary for more acute and advanced disease.

# 2. General Management of Chronic Encephalopathy (Table 1)

#### 2.1. Ammonia-Lowering Strategies

2.1.1. Dietary Protein Supplementation. Patients with cirrhosis often have a poor nutritional reserve due to anorexia, poor diet, malabsorption, and altered metabolic state. Hospitalized patients are often hypermetabolic and hypercatabolic, worsened by complications such as gastrointestinal bleeding, continued anorexia, and fasting for tests. Yet dietary protein has the potential to drive further ammoniagenesis, and so previously dietary protein restriction was common practice. However, protein restriction is no longer advocated as does not improve HE and may be harmful [1]. In fact high-protein diets are well tolerated in cirrhotic patients [2], with consensus supporting the need for normal or high dietary

protein (1–1.5 g/kg protein and 25–40 kcal/kg per day) [2, 3]. Rare exceptions arise occasionally with inborn errors of metabolism or acute liver failure (ALF) patients intubated for grade 3-4 HE associated with high circulating ammonia when protein restriction with maintained calorie intake (e.g., dextrose infusion) is necessary.

2.1.2. Branched-Chain Amino Acids (BCAAs). BCAAs are chiefly derived from dairy products and vegetables and account for 25% of total dietary protein. They are a good substrate for protein synthesis, both conserving and restoring muscle mass in advanced liver disease. In cirrhosis, poor dietary intake leads to a deficiency of BCAA and resultant accumulation of aromatic amino acids, both worsening protein-energy deficits and glutaminergic neurotransmission (increased false neurotransmitter precursors). In "high protein diet" intolerant and severely malnourished patients, BCAA supplements may be useful to provide the necessary nitrogen intake without a decline in mental state, with vegetable proteins likely to be better tolerated due to their higher BCAA content. As BCAAs are under the influence of circulating insulin, the insulin resistance state of cirrhosis may limit there nutritional benefit unless systemic insulin replacement is implemented. However, a number of metaanalyses have failed to find consensus on the use of BCAAs in cirrhosis from a wealth of conflicting data [4, 5].

In most cirrhotic patients, a modified eating pattern, based on several meals and a late evening snack, is adequate [4, 6].

<sup>&</sup>lt;sup>2</sup> Department of Gastroenterology, King Georges Hospital, Barley Lane, Goodmayes, Ilford, Essex IG3 8YB, UK

TABLE 1: Treatment stratagems used in HE.

#### HE grade: I-II

#### General management

Hyperammonemia

Dietary protein supplementation

Purgatives

- (i) Nonabsorbable disaccharides
- (ii) Enemas

Non-absorbable antibiotics

Modulation of interorgan ammonia

- (i) L-ornithine, L-aspartate (LOLA)
- (ii) Sodium benzoate
- (iii) Phenylacetate

#### Others

(i) Flumazenil "Bromocriptine" acarbose

Emerging therapies

(i) Probiotics

HE grade: III-IV

Cerebral edema & elevated ICP

#### General

- (i) Ventilate
- (ii) Sedate (e.g., Propofol)

#### Specific

- (i) Antimicrobials
- (ii) Hypertonic saline
- (iii) Mannitol
- (iv) Dexamethasone
- (v) Induced hypothermia
- (vi) Thiopentone
- (vii) Indomethacin
- (viii) Antiepileptic drugs (AED'S)
- (ix) N-acetylcysteine (NAC)

#### Transplantation

Orthotopic liver transplant (OLT)

Partial hepatectomy

Liver assist devices

- 2.1.3. Glycaemic Control. Disturbed glycaemic and lipid control is common in progressive liver disease and only worsened by the stress response in critically unwell patients. Therefore, once feeding has commenced, tight glycaemic control using insulin may be necessary to reduce oxidative stress (which triggers insulin resistance), limit mitochondrial liver damage, and improve endothelial activation (e.g., NO production), which will improve blood flow, limiting tissue injury, and improve outcome [7, 8].
- 2.1.4. Vitamins and Nutrients. Cirrhosis also leads to deficiencies of lipid-soluble vitamins, minerals, and micronutrients. For example, Zinc is a cofactor in the urea cycle [9] and also found in vesicles of predominately glutamatergic presynaptic terminals thereby having a role in neurotransmission [10]. Zinc supplementation (600 mg/day) has been studied without obvious benefit though replacement should be

considered if the patient is deficient [11]. Autopsy specimens from patients with hepatic coma and pallidal MR images of patients with HE suggest that manganese deposition in the basal ganglia may be a factor [12, 13]. However, as with earlier studies evaluating the role of gut bacterial products like mercaptans, phenols and medium- and short-chain fatty acids [14], there has been little cumulative evidence to support targeted treatment strategies.

#### 3. Probiotics

Most of the ammonia produced by the gut is from the deamination of dietary amino acids by bacteria, with a small contribution from the urea produced by urease-positive bacteria. In the critically ill and malnourished patient, levels of the predominant defensive bacteria strains (Bifidobacterium and Lactobacillus) decline. Antibiotics may further lead to ammonia-producing bacteria ameliorating hyperammonaemia. Probiotics are living nonpathogenic microorganisms utilized as food ingredients that may have a role in the treatment of HE. Probiotics are thought to exert an effect in HE by reducing intestinal ammonia production by enterocyte glutaminase and reduce bacterial translocation, modulate proinflammatory responses, and modulate gut permeability [15]. Furthermore, probiotics bypass the small bowel and get fermented by colonic bacteria to form lactic, acetic, and butyric acids, and gas (mainly hydrogen); any resultant intestinal hurry may increase the expulsion of ammoniagenic bacteria. In randomized placebo controlled trials [16], probiotics have been shown to reduce gut ammonia production and inflammation [16, 17]. It is worth noting that fermentable fibres alone were also beneficial in that study. This is not unexpected as the common effect of probiotics, aside from a decline of substrate for other bacteria [18] and reduced translocation, is the fermentation of nonabsorbed sugars (e.g., mono-, di- and oligosaccharides). This fermentation of sugars leads to the production of differential amounts of lactic acid, ethanol, and CO2 to modulate intestinal acidity and gas production.

#### 4. Purgatives

A purgative is an agent which cleanses the bowel by increasing the evacuation of luminal contents. This is beneficial in HE as it allows for reduced intestinal ammonia production and despite limited evidence from randomized controlled trials remain the most widely used therapy for HE.

4.1. Nonabsorbable Disaccharides. It is unclear how non-absorbable disaccharides exert a beneficial effect. There have been many proposed mechanisms (1) enhanced growth of nonurease-producing bacteria [19], (2) catharsis secondary to bowel acidification reducing ammonia absorption [20, 21], (3) proliferation of healthy bacteria by providing additional carbohydrate and thus nitrogen (even as ammonia) into protein, and/or (4) providing carbon and energy and so spare bacterial ammonia metabolism [22]. More specifically, lactulose (a sugar) passes through the small bowel completely undigested (unlike glucose, sucrose, and lactose, which are

easily fermented in the small bowel). Once in the colon, lactulose is fermented by anaerobic bacteria, especially Bacteroides spp. Fermentation of lactulose by colonic bacteria yields important weak acids (lactic, acetic & butyric) and gases (e.g., hydrogen). This leads to the acidification of ammonia into ammonium which is poorly absorbed. However, physiologically a total daily dose of 10-20 g is small compared to 500–1000 g faeces/day, such that the impact on acidity/reduced faecal pH on the faecal flora is likely to be limited. This is supported by the failure of mannitol and sorbitol, which both cause low pH, to improve HE [23]. The production of colonic hydrogen may be more important as only 7 g of lactulose produces 1 Litre of hydrogen that could induce intestinal hurry and shift massive amounts of colonic bacteria [24]. However, it may be the provision of energy in preference to ammonia that accounts for the benefit of nonabsorbable disaccharides.

A comprehensive meta-analysis of non-absorbable disaccharides has suggested that current data from randomised clinical trials do not support its routine use in clinical practice [25] though newer clinical studies suggest benefit with lactulose conferring improved neuropsychometric and quality of life scores [26], which lends weight to the overwhelming amount of anecdotal evidence that disaccharides are beneficial. It is likely that the impact of other therapies initiated at the same time often confounds any benefit on HE severity by the established ammonia-lowering effect of non-absorbable disaccharides.

Compliance, adverse effects, clinical safely, and cost effectiveness are necessary concerns. It is often overlooked that aggressive use of lactulose causes significant gaseous distension, discomfort, and diarrhoea which may lead to poor compliance. Furthermore, frank dehydration, prerenal uraemia, hyponatraemia, or aspiration of lactulose can occur. Therefore, although non-absorbable disaccharides are relatively cheap, their cost effectiveness should be balanced against clinical outcomes.

4.2. Other Purgatives. Enemas are beneficial as a means of expelling ammonia-producing gut flora by both cleansing and colonic acidification [27] but are no better than oral purgatives like lactulose. Therefore, if bowel motions can be maintained at  $\geq 2/\text{day}$ , then enemas may not offer any additional benefit.

#### 5. Nonabsorbable Antibiotics

The contribution of intestinal urease-positive bacteria to gut ammonia production is mainly in the colon rather than gastric mucosa (e.g., Helicobacter pylori), due to their number and more alkaline colonic pH which favours enhanced ammonia diffusion, such that Helicobacter pylori eradication has no therapeutic benefit [28]. Oral, non-absorbable, synthetic antibacterial agents such as Neomycin and Rifaximin have been used to inhibit the growth or kill susceptible ammoniagenic bacterial species, showing comparable efficacy to lactulose [29]. Rifaximin is a synthetic antibiotic related to rifamycin, with wide antibacterial activity against both aerobic and anaerobic gram-negative and gram-positive

bacteria. In random controlled studies Rifaximin is proven efficacious (maintaining remission and reducing hospitalization with HE even in patients already on lactulose), and a superior safety profile and thus preferred to neomycin [30, 31], Although beneficial, non-absorbable antibiotics are often reserved for patients who fail to respond to non-absorbable disaccharides.

## 6. Modulators of Interorgan Ammonia Metabolism (Figure 1)

The concept of manipulating endogenous biosynthetic pathways to eliminate nonurea waste nitrogen as a substitute for defective urea synthesis is well established [32]. Despite abnormal urea-cycle functioning, reducing total body nitrogen by promoting the synthesis of non-urea nitrogen-containing metabolites with high excretion rates appears to be of benefit.

6.1. Arginine Supplementation. L-arginine is an important dietary substrate for the urea cycle which allows for ammonia detoxification to urea (via arginase). L-arginine is a semiessential amino acid, as although metabolically produced, in some disease states may require dietary supplementation. In cases of the childhood urea cycle disorders (e.g., deficiency of argininosuccinate synthetase (AS) and argininosuccinase (AL)), dietary restriction of L-arginine triggers the rapid development (15-68 hours) of symptomatic hyperammonaemia (e.g., vomiting, lethargy, or irritability) [33]. In these disorders, there is a significant reduction in urea production, with nitrogen instead accumulating as mainly glutamine, ammonium, and to a limited extent alanine and glutamate. In AS and AL deficiency, the provision of additional dietary L-arginine promotes the synthesis of citrulline and argininosuccinate, allowing for the urinary excretion of nitrogen.

In ALF, systemic hypotension and cerebral oedema may be associated with increased plasma nitric oxide (NO) levels. L-arginine is the rate-limiting substrate for NO production but is deficient in ALF due possibly to increased arginase activity in the liver which converts it to urea and ornithine. There have been no clinical studies evaluating a role for L-arginine supplementation in HE, though animal studies suggest that correcting L-arginine deficiency may alter portal hypertension and cerebral oedema via arginase-dependent reduction in hyperammonaemia and/or NO-dependent mechanism(s).

6.2. Phenylbutyrate. Phenylbutyrate (converted to phenylacetate in vivo) is an established therapy for hyperammonaemia associated with urea cycle disorders [34], which are characterized by elevated glutamine levels. This excess can be mopped up by phenylacetate, which covalently combines with circulating glutamine to form renally excreted phenylacetylglutamine, removing glutamine as a substrate for ammoniagenesis. So far phenylbutyrate has proved ineffective in the treatment of HE associated with liver failure, probably because a high glutamate state, a prerequisite for phenylacetate to work, is absent in liver failure.

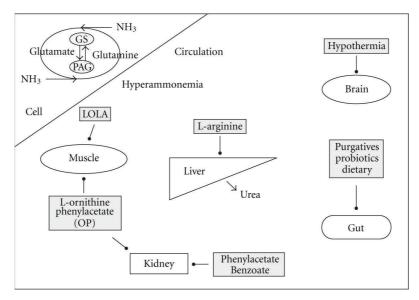


FIGURE 1: Therapies manipulating interorgan ammonia and amino acid metabolism. In liver failure, the relative activities of cellular glutamine syntheses (GS) and phosphate-activated glutaminase (PAG) in different organs influence interorgan ammonia and amino acid metabolism. With a loss of hepatic urea cycle capacity, hyperammonaemia is predominately due to worsening intestinal and renal ammonia efflux, with skeletal muscle having the potential to increase its ability to detoxify ammonia. Though the brain also detoxifies ammonia, this is counterproductive as resultant astrocyte glutamine accumulation induces brain swelling. This schematic highlights not only current standard therapies for hyperammonaemia which principally act on individual organs (e.g., purgatives targeting intestinal ammonia production), but also newer interventions targeting multiple organs (e.g., LOLA and OP).

6.3. Sodium Benzoate. Similarly sodium benzoate increases the renal excretion of ammonia but as hippuric acid (hippurate), the glycine conjugate of benzoic acid [32]. Sodium benzoate also improves the encephalopathy with inborn errors of metabolism [35] and is as effective as lactulose in the treatment of acute portosystemic HE [36].

6.4. Combined Intravenous Sodium Phenylbutyrate and Benzoate (Ammonul, Ucyclyd Pharma). In urea-cycle disorders, combination therapy results in a 79% reduction in plasma ammonia, and 84–98% improved survival with late onset disease, though poor in neonates and high peak ammonia values [37]. If untreated, only 16% of neonates survive, compared to 72% with late onset disease [38]. However, as the N-acyltransferases that conjugate glutamine to phenylacetate and glycine to benzoate are located in the liver and kidney, the severe hepatotoxicity of ALF may eventually lead to response failure, especially with the saturation of enzyme capacity (e.g., phenylacetate to PAG) [39, 40].

6.5. L-Ornithine L-Aspartate (LOLA). LOLA provides L-ornithine and L-aspartate as substrates for glutamate production in muscle leading to a reduction in circulating ammonia and in models of liver failure further suggest that LOLA reduces brain oedema of advanced HE [41]. In a double-blind randomized control study of cirrhotics with mild HE, one week of LOLA reduced ammonia and improved mental function [42]. A cross-over study showed that 20–40 g/day of infused LOLA ameliorated postprandial increases in ammonia following oral protein loading [43]. However, at higher doses, this study increased plasma glutamate, unchanged glutamine, and increased urea production [43] contradicting

the muscle ammonia detoxification hypothesis. Furthermore, 40g dosing induced hyperglycaemia and hyperinsulinaemia [43]. As yet, there are no studies in patients with ALF, and its use in ALF is currently not recommended. Critically, there are concerns that the ammonia-lowering effects of LOLA may only be transient, due to rebound hyperammonaemia on stopping LOLA [44], as a significant rise in glutamine levels eventually becomes a source for ammoniagenesis by the kidney and intestines (through glutaminase) [45]. Additionally, aspartate is unlikely to offer added benefit as in animal models it failed to reduce ammonia [46].

6.6. L-Ornithine Phenylacetate (OP). OP is a novel therapy targeting interorgan ammonia and amino acid metabolism [44]. OP reduces toxic levels of ammonia by ornithine acting as a substrate for glutamine synthesis from ammonia in skeletal muscle. This combination unlike other therapies targeting interorgan ammonia metabolism (e.g., LOLA), by stopping the recycling of ammonia (trapped as ornithine-glutamine) via phenylacetate excreting the ornithine-related glutamine as phenylacetylglutamine in the kidneys. It has been shown to correct the hyperammonemic state in animal models of cirrhosis [47] and ALF [48], limiting brain oedema and rises in ICP. Clinical studies are currently underway.

#### 7. Others

7.1. Acarbose. The hypoglycaemic agent acarbose which stimulates gut motility, through the inhibition of intestinal glucose absorption by promoting intestinal saccharolytic

TABLE 2: Precipitating factors in hepatic encephalopathy.

Precipitating factors in HE

Constipation

Dehydration

Gastrointestinal bleeding

Infection

Excessive dietary protein

Hypokalaemia

Hypoglycaemia

Hypothyroidism

Hypoxia

Metabolic alkalosis

Anaemia

Azotaemia/uraemia

Medications (narcotics, sedatives, etc.)

Hepatoma

TIPS, surgical shunt

Vascular occlusion

bacterial flora in preference to proteolytic flora, thereby reducing substrate for ammonia production. In a cross-over randomized trial of cirrhotic patients with low-grade HE and type-2 diabetes mellitus, 8 weeks of acarbose (100 mgs TDS) significantly decreased ammonia blood levels, and intellectual function, aside from decreasing fasting and postprandial glucose, and reducing glycosylated haemoglobin levels [49]. However, acarbose is unlikely to be an option except in those with coexistent type-2 diabetes mellitus.

7.2. Bromocriptine. Bromocriptine, a dopamine agonist, has been used with limited success for disturbances in dopaminergic neurotransmission associated with chronic intractable HE [50, 51], but such studies failed to show a clear benefit over standard therapy [49]. Furthermore, in cirrhotic patients with ascites, it can induce hyponatraemia [52]. However, there is anecdotal evidence to suggest a benefit in a small number of cirrhotic patients with low-grade encephalopathy and basal ganglia injury with associated dopamine deficiency.

# 8. Correction of Precipitating Factors (Table 2)

Worsening encephalopathy is often precipitated by a number of factors which can be anticipated and promptly corrected. Though HE may be triggered by uncommon events, it is important to outline the management of the more common precipitants.

8.1. Constipation. Enemas are beneficial as a mean of expelling ammonia producing gut flora either due by cleansing or colonic acidification [27]. However, there is only limited evidence to show a benefit over the use of oral purgatives like lactulose. Therefore, if bowel motions can be maintained at  $\geq 2/\text{day}$ , enemas are only used as an adjunct to the primarily used non-absorbable disaccharides.

- 8.2. Infections. bacterial infections predispose to variceal bleeding in cirrhotic patients. A meta-analysis of antibiotic use in variceal bleeding reported a 30% decrease in rate of infection and 9% improvement in short-term survival [53, 54]. Septic encephalopathy may also confound or mimic HE.
- 8.3. Gastrointestinal Bleeding. Due to the high-protein content of blood and thus nitrogenous load, there is increased intestinal ammonia production. This ammoniagenic blood meal and precipitation of HE are potentially related to an absence of the branched-chain amino acid isoleucine which protects the inhibitory effect of ammonia on the TCA cycle in neuronal cells.
- 8.4. Portosystemic Shunts. Persistent shunts may account for worsening HE poorly responsive to standard oral therapies and may be best treated by shunt closure.
- 8.5. TIPSS Insertion. The creation of a portosystemic shunt (used to stabilize patients with uncontrolled variceal bleeding or intractable ascites) may induce HE (especially within the first few months). Prophylaxis against encephalopathy with Lactitol (60 g/day) or Rifaximin (1200 mg/day) is not proven to be effective during the first month after TIPSS [55]. Therefore, careful selection of patients for a TIPSS or surgical shunt is necessary.

## 9. Acute Severe HE: Intracranial Hypertension and Cerebral Oedema (Table 1)

ALF is characterised by rapid onset HE with cerebral oedema and intracranial hypertension and progression to coma stages, independently associated with a 30% mortality [56]. Early ventilation, intensive care unit admission and judicious use of available therapies have led to a significant decline in deaths as a result of cerebral oedema. Aiding liver recovery by prompt and specific treatment of the cause of acute liver injury, treating precipitating factors such as dehydration, electrolyte and acid-base imbalance, [57], infection [58], and ameliorating hyperammonaemia remain at the forefront of therapy. The following therapeutic strategies are utilized in the management of severe HE requiring ventilation.

#### 10. General

Early airway maintenance is necessary to protect the airway and prevent high carbon dioxide tension and hypoxia which can result in cerebral hyperaemia [59]. Sedation and mechanical ventilation is also essential to safely manage agitation. Once intubated, the head should be elevated by  $10-20^{\circ}$  with minimal intervention and care when moving patients and optimize intracranial pressure (ICP) without compromising the cerebral perfusion pressure [60, 61]. Airway protection will also reduce the likelihood of aspiration, pneumonia, defective gas exchange, and infection. Sedative requirements (e.g., fentanyl, midazolam, or propofol) are low with worsening severity of HE but are likely to increase with recovery. Propofol is a useful sedative because it will

reduce ICP, and because of its nonhepatic metabolism will not accumulate. It may, however, induce hypotension [62].

10.1. Circulatory Support and Fluid Management. ALF is a hyperdynamic state with high cardiac output, low mean arterial pressure, and low systemic vascular resistance [63]. Generalized vasodilatation, which produces profound activation of the neurohormonal system, culminates in vasoconstriction of regional vascular beds [64]. Mean arterial pressure should be maintained at a level to keep the cerebral perfusion pressure between 50 and 65 mmHg [65]. The onset of multiorgan failure often necessitates the use of inotropes. Circulatory failure often becomes refractory to inotropes and up to 70% of patients die [66]. A routine short synacthen test on admission to guide the use of steroids is important as adrenal insufficiency is a common complication of ALF [67].

10.2. Renal Support. Renal dysfunction is common due to either prerenal, hepatorenal, or nephrotoxic (e.g., acetaminophen) injury [68]. This frequently requires renal replacement [66] with continuous (compared to intermittent) haemofiltration [69]. This avoids rapid water shifts seen with intermittent therapy [70], providing greater haemodynamic stability and improved cerebral perfusion pressure [69, 71]. Furthermore, due to impaired hepatic lactate metabolism, lactate-free dialysates are preferred [72].

10.3. Electrolyte Imbalance. Electrolyte imbalance should be corrected aggressively. Hyponatraemia ≤125 mmol/L may precipitate cerebral oedema and is a contraindication for orthotopic liver transplant (OLT) [73, 74]. Induced hypernatraemia has been shown to improve ICP and reduce inotropic requirements in traumatic brain injury and ALF [75].

10.4. Antimicrobial Agents. The incidence of sepsis in ALF is a significant factor in mortality and a contraindication to transplantation. Around 75% develop bacterial and 30% fungal infections [76, 77]. The administration of broadspectrum antibiotics/antifungal therapy should be initiated at the first sign of infection, with focused treatment once the organism is identified. Despite the absence of randomized control trials of prophylactic systemic antimicrobials in ALF, their use is widespread [78, 79].

10.5. Glycaemic Control. Both hyper- and hypoglycaemia need rapid correction as they may worsen brain oedema. The role of tight glycaemic control in ALF has not been ascertained but must be instituted with caution because of the tendency for the development of hypoglycaemia.

#### 11. Specific

11.1. Mannitol. Mannitol (an osmotic diuretic) increases brain capillary osmolality, drawing water from the brain tissue into the capillaries, and has been shown to significantly reduce the extent of cerebral oedema and improve survival [80, 81]. Bolus doses of 20% mannitol at 1 g/kg are preferred. Plasma osmolality should be kept <320 Osm/L, as mannitol is less effective with increasing osmolality. If patient is

oliguric, mannitol may accumulate and can only be used with concomitant haemofiltration.

11.2. Dexamethasone. In ALF, reducing inflammation (whether systemic or local) by utilizing the anti-inflammatory effects of steroids may improve cerebral haemodynamics and prevent/treat intracranial hypertension [79, 82, 83]. However, trials using dexamethasone in advanced ALF have shown little effect on the frequency of cerebral oedema or survival [80].

11.3. Mild Hypothermia. In models of ALF, induced hypothermia significantly reduces brain water, duration of encephalopathy, and improved outcome [84–86]. Using cooling blankets to induce moderate hypothermia (target core temp. 32–33°C) can lead to a reduction in ICP, even in patients unresponsive to mannitol and/or ultrafiltration [87, 88]. Hypothermia also significantly improves cardiovascular haemodynamics with reduced noradrenaline requirements [88], likely related to a reduction in arterial ammonia and also brain ammonia extraction and flux [87, 89]. As yet, there is no data from randomized control trials on the use of hypothermia in ALF but is worth considering in patients with uncontrolled intracranial hypertension.

11.4. Thiopental Sodium. By inducing cerebral vasoconstriction through the inhibition of nitric oxide synthetase, intermittent bolus injections of thiopental (1.5–3.5 mg/kg) reduce elevations of ICP [90]. However, its use is limited to intractable increases in ICP unresponsive to other therapies. Because of profound negative effects on systemic haemodynamics, its use is limited.

11.5. Indomethacin. Nonsteroidal anti-inflammatory (NSAIDS) may modulate brain function [91] (with possible effects on cognitive function via modulation of the glutamate-nitric oxide-cyclic GMP pathway [92]). Indomethacin (0.5 mg/kg), a nonselective cyclooxygenase inhibitor [93], can reduce ICP and cerebral oedema independent of a change in cerebral blood flow [94]. However, its use is limited by nephrotoxicity, platelet dysfunction, and risk of gastrointestinal bleeding. Poor brain penetration of NSAIDs at therapeutic levels requires high doses which increases the risk of toxicity [92, 95].

11.6. Antiepileptic Drugs (AED's). In some ALF patients with grade 3-4 HE, subclinical seizures occur, and the use of phenytoin was shown to significantly reduced seizure frequency and the development of increased ICP [96].

11.7. N-Acetylcysteine (NAC). In a case of acetaminophen overdose, NAC must be continued irrespective of the time between the overdose and presentation and acetaminophen level as it can prevent the progression of ALF and reduces mortality especially in those who progress to grade III–IV HE [97]. There is less convincing evidence for NAC in nonacetaminophen overdose [98, 99]. In nonacetaminophen ALF, NAC may improve survival by its effects on cardiac output, oxygen extraction and consumption, and due to its

antioxidant effects that ameliorate the significant oxidative stresses that occur with liver failure.

11.8. Flumazenil. In a large placebo controlled trial focusing on intensive care patients with advanced HE (grade III-IV), the short-acting benzodiazepine-receptor antagonist flumazenil was shown to rapidly improve the neurological score in 15% and electroencephalogram (EEG) findings in 30% of patients within minutes of its administration [100]. However, flumazenil does not lead to any lasting effect or correct HE, unless coadministered with a long-acting therapy [101], and as such is not recommended.

### 12. Liver Support and Transplantation

12.1. Transplantation. Transplantation offers definitive intervention for liver failure with a swift return to a normal mental state though minimal HE may persist in a few due to some as yet unknown irreversible cerebral changes [102]. Disparity between donor organs and recipients has led to a plethora of extracorporeal liver assist devices [103, 104] and even partial hepatectomy [83, 105] to aid or supplant the failing liver.

12.2. Extracorporeal Liver Assist Devices. Such devices may be either "biological" (using either immortalised cultured hepatocytes or whole animal livers), or "nonbiological" (using extracorporeal blood purification to dialyse albumin-bound hydrophobic substances), ultimately mimicking endogenous excretory and synthetic liver function. The extracorporeal devices under clinical evaluation include the following.

Molecular Adsorbent Recirculating System (MARS). It provides counter-current haemodialysis against albumin and bicarbonate circuits.

Single-Pass Albumin Dialysis (SPAD). It provides countercurrent albumin dialysis against high-flow blood in a fibre haemodiafilter, which unlike MARS is discarded after passing the filter. As it uses a standard renal dialysis device, continuous venovenous haemodiafiltration is possible.

*Prometheus system.* It provides direct albumin adsorption with high-flux haemodialysis after selective filtration of the albumin fraction through a specific polysulfone filter.

All devices successfully remove protein-bound toxins but have variable effects on systemic (versus portal) haemodynamics, and the potential to worsen coagulopathy. The clinical benefit of such devices is unclear but may at least offer a bridge to either transplantation or liver recovery.

#### 13. Conclusion

Ammonia-lowering therapy remains at the cornerstone of standard medical care for HE, along with measures to treat precipitating factors and specific interventions for the cerebral sequelae of advanced disease. Understanding interorgan ammonia metabolism and the pathophysiological basis of HE are most likely to lead to the development of new therapeutic approaches [45]. However, there is a lack of conclusive evidence from clinical studies even for current best practice [25, 106] and, therefore, a requirement for robust

randomized controlled trials to drive a more evidence-based approach.

#### References

- [1] J. Cordoba, J. Lopez-Hellin, M. Planas et al., "Normal protein diet for episodic hepatic encephalopathy: results of a randomized study," *Journal of Hepatology*, vol. 41, pp. 38–43, 2004.
- [2] M. Plauth, M. Merli, J. Kondrup, A. Weimann, P. Ferenci, and M. J. Muller, "ESPEN guidelines for nutrition in liver disease and transplantation," *Clinical Nutrition*, vol. 16, no. 2, pp. 43–55, 1997.
- [3] H. Lochs and M. Plauth, "Liver cirrhosis: rationale and modalities for nutritional support—the European Society of Parenteral and Enteral Nutrition consensus and beyond," *Current Opinion in Clinical Nutrition and Metabolic Care*, vol. 2, no. 4, pp. 345–349, 1999.
- [4] G. Marchesini, R. Marzocchi, M. Noia, and G. Bianchi, "Branched-chain amino acid supplementation in patients with liver diseases," *Journal of Nutrition*, vol. 135, no. 6, pp. 1596S–1601S, 2005.
- [5] B. Als-Nielsen, R. L. Koretz, L. L. Kjaergard, and C. Gluud, Branched-Chain Amino Acids for Hepatic Encephalopathy (Cochrane Review), Cochrane Library, John Wiley & Sons, Chichester, UK, 2004.
- [6] G. Marchesini, G. Bianchi, B. Rossi, M. Brizi, and N. Melchionda, "Nutritional treatment with branched-chain amino acids in advanced liver cirrhosis," *Journal of Gastroenterology*, vol. 35, 12, pp. 7–12, 2000.
- [7] L. Langouche, I. Vanhorebeek, D. Vlasselaers et al., "Intensive insulin therapy protects the endothelium of critically ill patients," *Journal of Clinical Investigation*, vol. 115, no. 8, pp. 2277–2286, 2005.
- [8] N. Houstis, E. D. Rosen, and E. S. Lander, "Reactive oxygen species have a causal role in multiple forms of insulin resistance," *Nature*, vol. 440, no. 7086, pp. 944–948, 2006.
- [9] G. Marchesini, A. Fabbri, G. Bianchi, M. Brizi, and M. Zoli, "Zinc supplementation and amino acid-nitrogen metabolism in patients with advanced cirrhosis," *Hepatology*, vol. 23, no. 5, pp. 1084–1092, 1996.
- [10] Z. S. Agus, I. D. Dukes, and M. Morad, "Divalent cations modulate the transient outward current in rat ventricular myocytes," *American Journal of Physiology*, vol. 261, no. 2, pp. C310–C318, 1991.
- [11] P. Reding, J. Duchateau, and C. Bataille, "Oral zinc supplementation improves hepatic encephalopathy. Results of a randomised controlled trial," *The Lancet*, vol. 2, no. 8401, pp. 493–495, 1984.
- [12] C. Rose, R. F. Butterworth, J. Zayed et al., "Manganese deposition in basal ganglia structures results from both portal-systemic shunting and liver dysfunction," *Gastroenterology*, vol. 117, no. 3, pp. 640–644, 1999.
- [13] G. P. Layrargues, C. Rose, L. Spahr, J. Zayed, L. Normandin, and R. F. Butterworth, "Role of manganese in the pathogenesis of portal-systemic encephalopathy," *Metabolic Brain Disease*, vol. 13, no. 4, pp. 311–317, 1998.
- [14] L. Zieve, W. M. Doizaki, and F. J. Zieve, "Synergism between mercaptans and ammonia or fatty acids in the production of coma: a possible role for mercaptans in the pathogenesis of hepatic coma," *Journal of Laboratory and Clinical Medicine*, vol. 83, no. 1, pp. 16–28, 1974.

- [15] G. W. Elmer and L. V. McFarland, "Biotherapeutic agents in the treatment of infectious diarrhea," *Gastroenterology Clinics* of North America, vol. 30, no. 3, pp. 837–854, 2001.
- [16] Q. Liu, Z. P. Duan, D. K. Ha, S. Bengmark, J. Kurtovic, and S. M. Riordan, "Synbiotic modulation of gut flora: effect on minimal hepatic encephalopathy in patients with cirrhosis," *Hepatology*, vol. 39, no. 5, pp. 1441–1449, 2004.
- [17] S. F. Solga, "Probiotics can treat hepatic encephalopathy," *Medical Hypotheses*, vol. 61, no. 2, pp. 307–313, 2003.
- [18] G. Bongaerts, R. Severijnen, and H. Timmerman, "Effect of antibiotics, prebiotics and probiotics in treatment for hepatic encephalopathy," *Medical Hypotheses*, vol. 64, no. 1, pp. 64– 68, 2005.
- [19] J. Bircher, J. Muller, P. Guggenheim, and U. P. Haemmerli, "Treatment of chronic portal-systemic encephalopathy with lactulose," *The Lancet*, vol. 1, no. 7443, pp. 890–892, 1966.
- [20] A. Bennett and K. G. Eley, "Intestinal pH and propulsion: an explanation of diarrhoea in lactase deficiency and laxation by lactulose," *Journal of Pharmacy and Pharmacology*, vol. 28, no. 3, pp. 192–195, 1976.
- [21] A. Bennett and K. G. Eley, "Proceedings: stimulation of intestinal propulsion by lowered intraluminal pH," *British Journal of Surgery*, vol. 63, no. 2, pp. 160–161, 1976.
- [22] A. Vince, M. Killingley, and O. M. Wrong, "Effect of lactulose on ammonia production in a fecal incubation system," *Gastroenterology*, vol. 74, no. 3, pp. 544–549, 1978.
- [23] L. Agostini, P. F. Down, J. Murison, and O. M. Wrong, "Faecal ammonia and pH during lactulose administration in man: comparison with other cathartics," *Gut*, vol. 13, no. 11, pp. 859–866, 1972.
- [24] C. Cherbut, "Motor effects of short-chain fatty acids and lactate in the gastrointestinal tract," *Proceedings of the Nutrition Society*, vol. 62, no. 1, pp. 95–99, 2003.
- [25] B. Als-Nielsen, L. L. Gluud, and C. Gluud, "Non-absorbable disaccharides for hepatic encephalopathy: systematic review of randomised trials.," *British Medical Journal*, vol. 328, no. 7447, pp. 1046–1050, 2004.
- [26] S. Prasad, R. K. Dhiman, A. Duseja, Y. K. Chawla, A. Sharma, and R. Agarwal, "Lactulose improves cognitive functions and health-related quality of life in patients with cirrhosis who have minimal hepatic encephalopathy," *Hepatology*, vol. 45, no. 3, pp. 549–559, 2007.
- [27] M. Uribe, O. Campollo, F. Vargas et al., "Acidifying enemas (lactitol and lactose) vs. nonacidifying enemas (tap water) to treat acute portal-systemic encephalopathy: a double-blind, randomized clinical trial," *Hepatology*, vol. 7, pp. 639–643, 1987.
- [28] J. Miquel, R. Barcena, D. Boixeda et al., "Role of Helicobacter pylori infection and its eradication in patients with subclinical hepatic encephalopathy," *European Journal of Gastroenterology & Hepatology*, vol. 13, pp. 1067–1072, 2001.
- [29] H. O. Conn, C. M. Leevy, Z. R. Vlahcevic et al., "Comparison of lactulose and neomycin in the treatment of chronic portalsystemic encephalopathy. A double blind controlled trial," *Gastroenterology*, vol. 72, pp. 573–583, 1977.
- [30] F. Miglio, D. Valpiani, S. R. Rossellini, A. Ferrieri, and N. Canova, "Rifaximin, a non-absorbable rifamycin, for the treatment of hepatic encephalopathy. A double-blind, randomised trial," *Current Medical Research and Opinion*, vol. 13, no. 10, pp. 593–601, 1997.
- [31] S. Di Piazza, M. G. Filippazzo, L. M. Valenza et al., "Rifaximine versus neomycin in the treatment of portosystemic encephalopathy," *Italian Journal of Gastroenterology*, vol. 23, no. 7, pp. 403–407, 1991.

- [32] S. W. Brusilow, D. L. Valle, and M. L. Batshaw, "New pathways of nitrogen excretion in inborn errors of urea synthesis," *The Lancet*, vol. 2, no. 8140, pp. 452–454, 1979.
- [33] S. W. Brusilow, "Arginine, an indispensable amino acid for patients with inborn errors of urea synthesis," *Journal of Clinical Investigation*, vol. 74, no. 6, pp. 2144–2148, 1984.
- [34] I. Smith, "The treatment of inborn errors of the urea cycle," *Nature*, vol. 291, no. 5814, pp. 378–380, 1981.
- [35] M. L. Batshaw, S. Brusilow, and L. Waber, "Treatment of inborn errors of urea synthesis: activation of alternative pathways of waste nitrogen synthesis and excretion," *New England Journal of Medicine*, vol. 306, no. 23, pp. 1387–1392, 1982.
- [36] S. Sushma, S. Dasarathy, R. K. Tandon, S. Jain, S. Gupta, and M. S. Bhist, "Sodium benzoate in the treatment of acute hepatic encephalopathy: a double-blind randomized trial," *Hepatology*, vol. 16, no. 1, pp. 138–144, 1992.
- [37] G. M. Enns, S. A. Berry, G. T. Berry, W. J. Rhead, S. W. Brusilow, and A. Hamosh, "Survival after treatment with phenylacetate and benzoate for urea-cycle disorders," *The New England Journal of Medicine*, vol. 356, no. 22, pp. 2282–2292, 2007.
- [38] M. C. Nassogne, B. Heron, G. Touati, D. Rabier, and J. M. Saudubray, "Urea cycle defects: management and outcome," *Journal of Inherited Metabolic Disease*, vol. 28, no. 3, pp. 407–414, 2005.
- [39] A. Thibault, M. R. Cooper, W. D. Figg et al., "A phase I and pharmacokinetic study of intravenous phenylacetate in patients with cancer," *Cancer Research*, vol. 54, no. 7, pp. 1690–1694, 1994.
- [40] V. Praphanphoj, S. A. Boyadjiev, L. J. Waber, S. W. Brusilow, and M. T. Geraghty, "Three cases of intravenous sodium benzoate and sodium phenylacetate toxicity occurring in the treatment of acute hyperammonaemia," *Journal of Inherited Metabolic Disease*, vol. 23, no. 2, pp. 129–136, 2000.
- [41] C. Rose, A. Michalak, K. V. R. Rao, G. Quack, G. Kircheis, and R. F. Butterworth, "L-ornithine-L-aspartate lowers plasma and cerebrospinal fluid ammonia and prevents brain edema in rats with acute liver failure," *Hepatology*, vol. 30, no. 3, pp. 636–640, 1999.
- [42] G. Kircheis, R. Nilius, C. Held et al., "Therapeutic efficacy of L-ornithine-L-aspartate infusions in patients with cirrhosis and hepatic encephalopathy: results of a placebo-controlled, double-blind study," *Hepatology*, vol. 25, no. 6, pp. 1351– 1360, 1997.
- [43] U. Staedt, H. Leweling, R. Gladisch, C. Kortsik, E. Hagmuller, and E. Holm, "Effects of ornithine aspartate on plasma ammonia and plasma amino acids in patients with cirrhosis. A double-blind, randomized study using a four-fold cross-over design," *Journal of Hepatology*, vol. 19, no. 3, pp. 424–430, 1993.
- [44] R. Jalan, G. Wright, N. A. Davies, and S. J. Hodges, "L-ornithine phenylacetate (OP): a novel treatment for hyperammonemia and hepatic encephalopathy," *Medical Hypotheses*, vol. 69, no. 5, pp. 1064–1069, 2007.
- [45] S. W. Olde Damink, N. E. P. Deutz, C. H. C. Dejong, P. B. Soeters, and R. Jalan, "Interorgan ammonia metabolism in liver failure," *Neurochemistry International*, vol. 41, no. 2-3, pp. 177–188, 2002.
- [46] L. Zieve, C. Lyftogt, and D. Raphael, "Ammonia toxicity: comparative protective effect of various arginine and ornithine derivatives, aspartate, benzoate, and carbamyl glutamate," *Metabolic Brain Disease*, vol. 1, no. 1, pp. 25–35, 1986.

- [47] N. A. Davies, G. Wright, L. M. Ytrebo et al., "L-ornithine and phenylacetate synergistically produce sustained reduction in ammonia and brain water in cirrhotic rats," *Hepatology*, vol. 50, no. 1, pp. 155–164, 2009.
- [48] L. M. Ytrebø, R. G. Kristiansen, H. Maehre et al., "L-ornithine phenylacetate attenuates increased arterial and extracellular brain ammonia and prevents intracranial hypertension in pigs with acute liver failure," *Hepatology*, vol. 50, pp. 165–174, 2009.
- [49] S. Gentile, G. Guarino, M. Romano et al., "A randomized controlled trial of acarbose in hepatic encephalopathy," *Clinical Gastroenterology and Hepatology*, vol. 3, no. 2, pp. 184–191, 2005.
- [50] M. Y. Morgan, A. Jakobovits, and A. Elithorn, "Successful use of bromocriptine in the treatment of a patient with chronic portasystemic encephalopathy," *The New England Journal of Medicine*, vol. 296, no. 14, pp. 793–794, 1977.
- [51] M. Y. Morgan, A. W. Jakobovits, I. M. James, and S. Sherlock, "Successful use of bromocriptine in the treatment of chronic hepatic encephalopathy," *Gastroenterology*, vol. 78, no. 4, pp. 663–670, 1980.
- [52] M. Uribe, A. Farca, and M. A. Marquez, "Treatment of chronic portal systemic encephalopathy with bromocriptine: a double-blind controlled trial," *Gastroenterology*, vol. 76, no. 6, pp. 1347–1351, 1979.
- [53] B. Bernard, J. D. Grange, E. N. Khac, X. Amiot, P. Opolon, and T. Poynard, "Antibiotic prophylaxis for the prevention of bacterial infections in cirrhotic patients with gastrointestinal bleeding: a meta-analysis," *Hepatology*, vol. 29, no. 6, pp. 1655–1661, 1999.
- [54] M. C. Hou, H. C. Lin, T. T. Liu et al., "Antibiotic prophylaxis after endoscopic therapy prevents rebleeding in acute variceal hemorrhage: a randomized trial," *Hepatology*, vol. 39, no. 3, pp. 746–753, 2004.
- [55] O. Riggio, A. Masini, C. Efrati et al., "Pharmacological prophylaxis of hepatic encephalopathy after transjugular intrahepatic portosystemic shunt: a randomized controlled study," *Journal of Hepatology*, vol. 42, no. 5, pp. 674–679, 2005.
- [56] A. J. Makin, J. Wendon, and R. Williams, "A 7-year experience of severe acetaminophen-induced hepatotoxicity (1987– 1993)," *Gastroenterology*, vol. 109, no. 6, pp. 1907–1916, 1995.
- [57] R. Jalan and D. Kapoor, "Reversal of diuretic-induced hepatic encephalopathy with infusion of albumin but not colloid," *Clinical Science*, vol. 106, no. 5, pp. 467–474, 2004.
- [58] D. L. Shawcross, N. A. Davies, R. Williams, and R. Jalan, "Systemic inflammatory response exacerbates the neuropsychological effects of induced hyperammonemia in cirrhosis," *Journal of Hepatology*, vol. 40, no. 2, pp. 247–254, 2004.
- [59] R. J. Ede, A. E. S. Gimson, D. Bihari, and R. Williams, "Controlled hyperventilation in the prevention of cerebral oedema in fulminant hepatic failure," *Journal of Hepatology*, vol. 2, no. 1, pp. 43–51, 1986.
- [60] A. Davenport, E. J. Will, and A. M. Davison, "Effect of posture on intracranial pressure and cerebral perfusion pressure in patients with fulminant hepatic and renal failure after acetaminophen self-poisoning," *Critical Care Medicine*, vol. 18, no. 3, pp. 286–289, 1990.
- [61] G. Strauss, B. A. Hansen, G. M. Knudsen, and F. S. Larsen, "Hyperventilation restores cerebral blood flow autoregulation in patients with acute liver failure," *Journal of Hepatology*, vol. 28, no. 2, pp. 199–203, 1998.

- [62] E. F. M. Wijdicks and S. L. Nyberg, "Propofol to control intracranial pressure in fulminant hepatic failure," *Transplantation Proceedings*, vol. 34, no. 4, pp. 1220–1222, 2002.
- [63] A. Ellis and J. Wendon, "Circulatory, respiratory, cerebral, and renal derangements in acute liver failure: pathophysiology and management," *Seminars in Liver Disease*, vol. 16, no. 4, pp. 379–387, 1996.
- [64] R. Jalan, "Acute liver failure: current management and future prospects," *Journal of Hepatology*, vol. 42, no. 1, pp. S115– S123, 2005.
- [65] M. H. Davies, D. Mutimer, J. Lowes, E. Elias, and J. Neuberger, "Recovery despite impaired cerebral perfusion in fulminant hepatic failure," *The Lancet*, vol. 343, no. 8909, pp. 1329–1330, 1994.
- [66] R. Jalan, "Intracranial hypertension in acute liver failure: pathophysiological basis of rational management," *Seminars in Liver Disease*, vol. 23, no. 3, pp. 271–282, 2003.
- [67] R. Harry, G. Auzinger, and J. Wendon, "The clinical importance of adrenal insufficiency in acute hepatic dysfunction," Hepatology, vol. 36, no. 2, pp. 395–402, 2002.
- [68] K. Moore, "Renal failure in acute liver failure," European Journal of Gastroenterology & Hepatology, vol. 11, no. 9, pp. 967–975, 1999.
- [69] A. Davenport, E. J. Will, A. M. Davison et al., "Changes in intracranial pressure during haemofiltration in oliguric patients with grade IV hepatic encephalopathy," *Nephron*, vol. 53, no. 2, pp. 142–146, 1989.
- [70] R. J. Winney, D. M. Kean, J. J. K. Best, and M. A. Smith, "Changes in brain water with haemodialysis," *The Lancet*, vol. 2, no. 8515, pp. 1107–1108, 1986.
- [71] A. Davenport, E. J. Will, and A. M. Davidson, "Improved cardiovascular stability during continuous modes of renal replacement therapy in critically ill patients with acute hepatic and renal failure," *Critical Care Medicine*, vol. 21, no. 3, pp. 328–338, 1993.
- [72] N. D. Murphy, S. K. Kodakat, J. A. Wendon et al., "Liver and intestinal lactate metabolism in patients with acute hepatic failure undergoing liver transplantation," *Critical Care Medicine*, vol. 29, no. 11, pp. 2111–2118, 2001.
- [73] R. Jalan, K. Dabos, D. N. Redhead, A. Lee, and P. C. Hayes, "Elevation of intracranial pressure following transjugular intrahepatic portosystemic stent-shunt for variceal haemorrhage," *Journal of Hepatology*, vol. 27, no. 5, pp. 928–933, 1997.
- [74] J. Cordoba, J. Gottstein, and A. T. Blei, "Chronic hyponatremia exacerbates ammonia-induced brain edema in rats after portacaval anastomosis," *Journal of Hepatology*, vol. 29, no. 4, pp. 589–594, 1998.
- [75] N. Murphy, G. Auzinger, W. Bernel, and J. Wendon, "The effect of hypertonic sodium chloride on intracranial pressure in patients with acute liver failure," *Hepatology*, vol. 39, no. 2, pp. 464–470, 2004.
- [76] N. Rolando, J. Philpott-Howard, and R. Williams, "Bacterial and fungal infection in acute liver failure," *Seminars in Liver Disease*, vol. 16, no. 4, pp. 389–402, 1996.
- [77] N. Rolando, J. J. Wade, A. Stangou et al., "Prospective study comparing the efficacy of prophylactic parenteral antimicrobials, with or without enteral decontamination, in patients with acute liver failure," *Liver Transplantation and Surgery*, vol. 2, no. 1, pp. 8–13, 1996.
- [78] N. Rolando, F. Harvey, J. Brahm et al., "Prospective study of bacterial infection in acute liver failure: an analysis of fifty patients," *Hepatology*, vol. 11, no. 1, pp. 49–53, 1990.

- [79] N. Rolando, J. Wade, M. Davalos, J. Wendon, J. Philpott-Howard, and R. Williams, "The systemic inflammatory response syndrome in acute liver failure," *Hepatology*, vol. 32, no. 4 I, pp. 734–739, 2000.
- [80] J. Canalese, A. E. S. Gimson, and C. Davis, "Controlled trial of dexamethasone and mannitol for the cerebral oedema of fulminant hepatic failure," *Gut*, vol. 23, no. 7, pp. 625–629, 1982.
- [81] M. A. Hanid, M. Davies, P. J. Mellon et al., "Clinical monitoring of intracranial pressure in fulminant hepatic failure," *Gut*, vol. 21, pp. 866–869, 1980.
- [82] R. Jalan and R. Williams, "The inflammatory basis of intracranial hypertension in acute liver failure," *Journal of Hepatology*, vol. 34, no. 6, pp. 940–942, 2001.
- [83] R. Jalan, A. Pollok, S. H. A. Shah, K. K. Madhavan, and K. J. Simpson, "Liver derived pro-inflammatory cytokines may be important in producing intracranial hypertension in acute liver failure," *Journal of Hepatology*, vol. 37, no. 4, pp. 536–538, 2002.
- [84] M. Peignoux, J. Bernuau, and J. P. Benhamou, "Total hepatectomy and hepatic vascular exclusion in the rat: a comparison, with special reference to the influence of body temperature," *Clinical Science*, vol. 62, no. 3, pp. 273–277, 1982.
- [85] P. Traber, M. DalCanto, D. Ganger, and A. T. Blei, "Effect of body temperature on brain edema and encephalopathy in the rat after hepatic devascularization," *Gastroenterology*, vol. 96, no. 3, pp. 885–891, 1989.
- [86] S. Eguchi, A. Kamlot, J. Ljubimova et al., "Fulminant hepatic failure in rats: Survival and effect on blood chemistry and liver regeneration," *Hepatology*, vol. 24, no. 6, pp. 1452–1459, 1996
- [87] R. Jalan, S. W. M. O. Damink, N. E. P. Deutz, A. Lee, and P. C. Hayes, "Moderate hypothermia for uncontrolled intracranial hypertension in acute liver failure," *The Lancet*, vol. 354, no. 9185, pp. 1164–1168, 1999.
- [88] R. Jalan, S. W. M. Olde Damink, N. E. P. Deutz, P. C. Hayes, and A. Lee, "Moderate hypothermia in patients with acute liver failure and uncontrolled intracranial hypertension," *Gastroenterology*, vol. 127, no. 5, pp. 1338–1346, 2004.
- [89] G. I. Strauss, G. M. Knudsen, J. Kondrup, K. Moller, and F. S. Larsen, "Cerebral metabolism of ammonia and amino acids in patients with fulminant hepatic failure," *Gastroenterology*, vol. 121, no. 5, pp. 1109–1119, 2001.
- [90] A. Forbes, G. J. M. Alexander, J. G. O'Grady et al., "Thiopental infusion in the treatment of intracranial hypertension complicating fulminant hepatic failure," *Hepatology*, vol. 10, no. 3, pp. 306–310, 1989.
- [91] G. R. Bernard, A. P. Wheeler, J. A. Russell et al., "The effects of ibuprofen on the physiology and survival of patients with sepsis. The Ibuprofen in sepsis study group," *The New England Journal of Medicine*, vol. 336, no. 13, pp. 912–918, 1997.
- [92] O. R. Cauli, R. Piedrafita, B. Boix, and J. B. Felipo, "Inflammation and hepatic encephalopathy: ibuprofen restores learning ability in rats with chronic liver failure," *Hepatology*, vol. 46, no. 2, pp. 514–519, 2007.
- [93] K. Jensen, J. Ohrstrom, G. E. Cold, and J. Astrup, "Indomethacin (Confortid) in severe head injury and elevated intracranial pressure (ICP)," Acta Neurochirurgica Supplementum, vol. 55, pp. 47–48, 1992.
- [94] F. Tofteng and F. S. Larsen, "The effect of indomethacin on intracranial pressure, cerebral perfusion and extracellular lactate and glutamate concentrations in patients with ful-

- minant hepatic failure," Journal of Cerebral Blood Flow and Metabolism, vol. 24, no. 7, pp. 798–804, 2004.
- [95] M. S. Abdel-Halim, B. Sjoquist, and E. Anggard, "Inhibition of prostaglandin synthesis in rat brain," *Acta Pharmacol Toxicol*, vol. 43, pp. 266–272, 1978.
- [96] A. J. Ellis, J. A. Wendon, and R. Williams, "Subclinical seizure activity and prophylactic phenytoin infusion in acute liver failure: a controlled clinical trial," *Hepatology*, vol. 32, no. 3, pp. 536–541, 2000.
- [97] P. M. Harrison, R. Keays, G. P. Bray, G. J. M. Alexander, and R. Williams, "Improved outcome of paracetamol-induced fulminant hepatic failure by late administration of acetylcysteine," *The Lancet*, vol. 335, no. 8705, pp. 1572–1573, 1990.
- [98] R. Keays, P. M. Harrison, J. A. Wendon et al., "Intravenous acetylcysteine in paracetamol induced fulminant hepatic failure: a prospective controlled trial," *British Medical Journal*, vol. 303, no. 6809, pp. 1026–1029, 1991.
- [99] Z. Ben-Ari, H. Vaknin, and R. Tur-Kaspa, "N-acetylcysteine in acute hepatic failure (non-paracetamol-induced)," *Hepa-to-Gastroenterology*, vol. 47, no. 33, pp. 786–789, 2000.
- [100] G. Barbaro, G. Di Lorenzo, M. Soldini et al., "Flumazenil for hepatic encephalopathy grade III and IVa in patients with cirrhosis: an Italian multicenter double-blind, placebo-controlled, cross-over study," *Hepatology*, vol. 28, no. 2, pp. 374– 378, 1998.
- [101] K. D. Mullen, P. Amodio, and M. Y. Morgan, "Therapeutic studies in hepatic encephalopathy," *Metabolic Brain Disease*, vol. 22, no. 3-4, pp. 407–423, 2007.
- [102] S. Mechtcheriakov, I. W. Graziadei, M. Mattedi et al., "Incomplete improvement of visuo-motor deficits in patients with minimal hepatic encephalopathy after liver transplantation," *Liver Transplantation*, vol. 10, no. 1, pp. 77–83, 2004.
- [103] S. Sen, R. Jalan, and R. Williams, "Liver failure: basis of benefit of therapy with the molecular adsorbents recirculating system," *International Journal of Biochemistry and Cell Biology*, vol. 35, no. 9, pp. 1306–1311, 2003.
- [104] V. Stadlbauer, N. A. Davies, S. Sen, and R. Jalan, "Artificial liver support systems in the management of complications of cirrhosis," *Seminars in Liver Disease*, vol. 28, no. 1, pp. 96– 109, 2008.
- [105] B. Ringe, N. Lubbe, E. Kuse, U. Frei, and R. Pichlmayr, "Total hepatectomy and liver transplantation as two-stage procedure," *Annals of Surgery*, vol. 218, no. 1, pp. 3–9, 1993.
- [106] D. Shawcross and R. Jalan, "Dispelling myths in the treatment of hepatic encephalopathy," *The Lancet*, vol. 365, no. 9457, pp. 431–433, 2005.