

Review

# Hair follicle-targeting drug delivery strategies for the management of hair follicle-associated disorders



# Yueting Gu<sup>a,#</sup>, Qiong Bian<sup>a,#</sup>, Yanjun Zhou<sup>b</sup>, Qiaoling Huang<sup>c,\*</sup>, Jianqing Gao<sup>a,d,\*</sup>

<sup>a</sup> Institute of Pharmaceutics, College of Pharmaceutical Sciences, Zhejiang University, Hangzhou 310058, China <sup>b</sup> Zhejiang Huanling Pharmaceutical Technology Company, Jinhua 321000, China

<sup>c</sup> The Third People's Hospital of Hangzhou, Hangzhou 310009, China

<sup>d</sup> Jiangsu Engineering Research Center for New-type External and Transdermal Preparations, Changzhou 213149, China

#### ARTICLE INFO

Article history: Received 6 January 2022 Revised 30 March 2022 Accepted 14 April 2022 Available online 5 May 2022

Keywords: Follicular drug delivery Androgenetic alopecia Acne vulgaris Nanotechnology Microneedles

#### ABSTRACT

The hair follicle is not only a critical penetration route in percutaneous absorption but also has been recognized to be a target for hair follicle-associated disorders, such as androgenetic alopecia (AGA) and acne vulgaris. Hair follicle-targeting drug delivery systems allow for controlled drug release and enhance therapeutic efficacy with minimal side effects, exerting a promising method for the management of hair follicle-associated dysfunctions. Therefore, they have obtained much attention in several fields of research in recent years. This review gives an overview of potential follicle-targeting drug delivery formulations currently applied based on the particularities of the hair follicles, including a comprehensive assessment of their preclinical and clinical performance.

© 2022 Shenyang Pharmaceutical University. Published by Elsevier B.V. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/)

# 1. Introduction

The stratum corneum (SC) and hair follicle delivery routes are the most crucial pathways allowing transdermal drugs to penetrate the skin [1]. Improving the bioavailability of transdermal medicine has been an urgent pharmaceutical challenge, as non-permeable corneocytes occupy large areas of skin [2,3]. Under physiological conditions, the intact SC significantly prevents large hydrophilic substances and particulate structures from skin penetration. In contrast, the presence of hair follicles provides a way to pass through this barrier. Below the skin surface, hair follicles located deep into the dermis offer a large surface area for the prospective absorption of drugs or substances. The hair follicles function as reservoirs and portals for dermally applied substances [4,5]. Over the past several years, hair follicle-targeting drug delivery has become a very appealing strategy for treating hair follicle-related disorders, including androgen-associated diseases, hair loss [6,7], and acne vulgaris [8]. The importance of follicular delivery systems reflects in promoting the topical treatment efficiency of such diseases and reducing

Corresponding authors.
E-mail addresses: hz3ygcp@163.com (Q.L. Huang), gaojianqing@zju.edu.cn (J.Q. Gao).

<sup>#</sup> These authors contributed equally to this work.

Peer review under responsibility of Shenyang Pharmaceutical University.

https://doi.org/10.1016/j.ajps.2022.04.003

<sup>1818-0876/© 2022</sup> Shenyang Pharmaceutical University. Published by Elsevier B.V. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/)

systemic side effects. Several studies have demonstrated the advantages of the transfollicular route on porcine ear skin [9,10]. Additionally, particles-containing formulations could be detected within the hair follicles even after 10 days, while nanoparticles in the SC reservoir stored for only one day, indicating that hair follicles can function as a promising tool for sustained drug release [11].

Various hair follicular diseases probably result from the follicles' complex cycle and structure. Hair loss and acne vulgaris are probably the most common disorders closely related to hair follicles, as hair follicles on the face and scalp occupy as much as 10% of entire hair follicles [12]. In such cases, hair follicle-targeting drug delivery is becoming an essential element of different therapies. Androgenetic alopecia (AGA), as a common form of alopecia, is a multifactorial and age-related chronic disease and seriously affects patients' psychology as well as the quality of life [13]. Nearly 80% and 50% of Caucasian men and women are reported to experience some degree of AGA by the age of 70 years [14]. Conventional medical treatment consists of topical minoxidil and oral finasteride (FIN) formulations to stimulate hair regrowth. However, the existing treatment regimens for AGA are not able to adequately deliver drugs directly to the hair follicles, resulting in the emergence of systemic side effects [15,16] and a lack of efficacy [6]. Acne vulgaris is a common skin dysfunction of pilosebaceous units (PSU) [17] and is related to excessive sebum production, follicular keratinization, immunological changes, and bacterial proliferation in the hair follicles. Topical retinoids, including adapalene, tazarotene, and tretinoin, have been approved by the US Food and Drug Administration to manage acne vulgaris [18]. However, common adverse effects such as skin irritation and discomfort may affect the tolerability of topical therapies [19]. Furthermore, hair follicles are partly or wholly occluded by excessive keratin and dead cells in the early stage of acne vulgaris [20], further aggravating the development of the disease. Thus, it is critical to create an effective and safe follicular drug delivery system for the topical treatment of AGA and acne vulgaris.

The preferential deposition of drugs in hair follicles is determined by the physicochemical properties of the drug itself and the type of carrier applied in the formulations. Many drug delivery systems, including polymeric nanoparticles (NPs), lipid nanoparticles, and microneedles, have been broadly investigated to enhance the specific delivery of active pharmaceutical ingredients to hair follicles. This review's major goal is to critically introduce and analyze hair follicletargeting delivery systems, thereby providing a reference for more advanced and safer formulations to cure follicular disorders.

# 2. Features of the hair follicle

## 2.1. The structure and barrier of the hair follicle

Previous studies have proven that hair follicles play a vital role in topical drug delivery as a trans-appendage pathway and trans-epidermal route [21]. The complex three-dimensional structure of the hair follicle and cyclical activity must be considered when designing follicular drug delivery systems. As Fig. 1I shows, the permanent part of the hair follicle includes the infundibulum, isthmus, and bulge region from top to bottom [22]. The infundibulum at the upper part of the hair follicle is covered by an impermeable and intact SC. In contrast, the permeability dramatically increases in the lower infundibulum as the epidermis differentiates into a trichilemmal pattern, which is readily accessible for transdermal compounds [23]. The inner root sheath surrounding the hair follicle canal is related to the shape of the hair. Furthermore, the outer root sheath plays a regulatory role through interactions with melanocytes. The representative character of the hair follicle is its cyclical growth (Fig. 1II), which undergoes the anagen, catagen, and telogen phases and then re-enters the anagen phase [24]. The bulge containing the hair follicle stem cells (HFSCs) in the outer root sheath plays a crucial role in regulating the hair cycle [25]. Furthermore, the upper hair follicles are surrounded by plenty of immune cells, such as dendritic cells and Langerhans cells, indicating that hair follicle targeting might be a meaningful tool for transcutaneous immunization [26]. Notably, the sebaceous glands provide extra target sites that have drawn therapeutic interest for conditions like AGA, acne vulgaris, and other sebaceous gland disorders [27].

# 2.2. The mechanism of hair follicle-targeting drug delivery

The penetration through the hair follicle route is a complex process resulting from the hair follicle's complex structure, and the exact mechanism has not been clearly understood [23]. Generally, the transepidermal transport reduces once the drug is encapsulated in particulate carriers because particles tend to penetrate and preferentially deposit in the orifices of hair follicles (Fig. 2I; [29]). The process of hair follicle delivery is affected by many factors involving the physicochemical characteristics of drug molecules, such as the oil-water partition coefficient, molecular size, molecular weight, etc., and carriers, such as particle size, lipophilicity, surface charge, etc. [30–32].

As shown in Fig. 2II, previous studies have underscored the strong effect of "particle size" on the depth and extent of follicular delivery systems [33,34]. Thus, appropriately sized carriers play a critical role in the penetration of different skin layers and target sites within the hair follicle. Furthermore, keratin cells can function as a pumping system, and they can transmit carriers mechanically into the hair follicles along with hair movement [35]. The hair movement potentiates the follicular penetration of nanoparticles with the mean diameter between the thickness of keratin cells and the diameter of hair follicles. The optimal size of particles was found to be in the range of 400 - 700 nm to penetrate deeply into the hair follicle. In contrast, smaller particles mainly reached the deeper skin layers, and larger particles primarily remained on the skin's surface [36]. Earlier studies have demonstrated that smaller nanoparticles (20 nm) had a quicker accumulation in hair follicles during a time-dependent process (Fig. 2III), while these smaller nanoparticles are more likely to reach the lower layer of skin in the end. In summary, smaller substances exhibit a tendency



Fig. 1 – Schematic overview of the (I) hair follicular structures and (II) hair cycle related to the follicular penetration pathway (Red box: target site for acne vulgaris and AGA; Blue box: target site for hair growth control). [28]



Fig. 2 – Mechanism and factors of the follicular penetration process. I. Illustration for the penetration route of transdermal drug delivery system. Penetration pathways of transdermal drug: (1) Trans-epidermal penetration pathway around the corneocytes; (2) Trans-epidermal penetration pathway across the corneocytes; (3) Follicular penetration pathway. Particles tend to penetrate and preferentially deposit in the orifices of hair follicles. II. The size effects of PLGA nanoparticles on penetration depths associated with the target sites in terminal hair follicles (THF) and vellus hair follicles (VHF). [40] III. x-y images showing the nanoparticle accumulation in hair follicles after applying nanoparticles (20 nm, 200 nm) for (a, d) 30 min, (b, e) 1 h, and (c, f) 2 h. The circles in white represent hair follicles. [41]

to penetrate the skin through the transfollicular pathway into the living tissues surrounding the hair follicles, but the particles with optimal size tend to accumulate preferentially within the hair follicles [37]. It is crucial to perform customized designs by controlling the particle size according to the medication positioning requirements. Moreover, dissolving microneedles could penetrate the skin and release their loaded therapeutics directly into dermal layers to enrich hair follicles [29], thereby making them a potential choice for targeting the hair follicles.

Furthermore, the influence of NPs' surface charge on hair follicle targeting was extensively studied but remains unclear. Interestingly, there are conflicting findings among different studies. A recent study proved that hair follicles could abundantly accumulate NPs with negative surface charge and lipophilic surface features. Negatively charged



Fig 3 – Scheme of current strategies for hair follicle targeting. I. Nanotechnology-based particles are utilized to deliver therapeutic substances into the hair follicle. II. Microneedles are utilized to deliver the therapeutic substances directly to the sites around the hair follicle. III. Exogenous stimulations lead to an active release of the drugs from the particles equipped with specific release mechanisms.

poly (D, L-lactide-co-glycolide) (PLGA) nanoparticles and PLGA nanoparticles coated with phospholipids facilitated follicular uptake on a pig ear skin model [38]. However, another study [39] indicated nanostructured lipid carriers (NLCs) containing cationic surfactants exhibited higher follicle accumulation.

# 3. Strategies for hair follicle targeting

Hair follicles can efficiently bypass complex intercellular permeation pathways and enter deeper skin layers, which makes the hair follicles become an exciting target site. Given the complexity of follicular penetration, three different strategies taking advantage of the hair follicle targeting have been applied and schematically illustrated in Fig. 3. Nanotechnology-based particles are utilized to deliver therapeutic substances into the pre-determined target sites of the hair follicle. Moreover, microneedles can break the skin barrier and deliver the therapeutic substances directly to the sites around the hair follicle, achieving accurate and fast mapping. Or nanoparticles equipped with a specific release mechanism are applied to deeply deliver medicinal substances into the hair follicle, where exogenous stimulations accelerate drugs' active release.

# 3.1. Nanotechnology-based formulations for follicular drug delivery

Current strategies for follicular drug delivery mainly focus on using nanostructured drug delivery systems, such as polymeric nanoparticles, nanoemulsions, lipid nanoparticles, and metallic nanoparticles. An overview of NPs related to hair follicle delivery is shown in Table 1. Various types of NPs possess individual characteristic behaviors and different physicochemical characteristics regarding skin and follicular penetration. Though the exact mechanism of hair follicle targeting by nanoformulations is unclear, particle size is a determinant for targeting delivery and retention in hair follicles. Besides, the occlusive effect is also an important factor in the process of nanoparticles entering into hair follicles [42]. Nanosized particles can guarantee close contact with superficial junctions of the SC. After the evaporation of water, the hydrated SC will enlarge the space between the keratinocytes, thereby enhancing the drug transport to hair follicles [43]. The general merits of nanoparticle local drug delivery systems can be summarized as follows: enhanced physical and chemical stability, increased drug accumulation, better solubility, and sustained release of drugs [44,45]. Though hair follicles account for less than 0.1% of the total skin area [46], the penetration of follicular drugs obtained through nanotechnology-based formulations improves by creating particles of small size to enlarge the surfaces with the SC. Particularly, drug-loaded nanoparticles access and accumulate preferentially in the orifices of hair follicles, which function as drug reservoirs for controlled drug diffusion that can reduce the doses and frequencies to overcome side effects of therapy.

### 3.1.1. Polymeric nanoparticles

Polymeric nanoparticles are solid particles in the nanometer range, prepared using biodegradable and biocompatible polymers or monomers, such as chitosan, cellulose, polystyrene, PLA, PLGA, polyvinylalcohol, and polyethyleneimine [87]. NPs with a diameter between 1nm and 1000 nm are more likely to reach their target organs or tissues through various barriers in the body. A previous study [88] traced the fate of NPs in the skin by incorporating Nile red into the NPs and demonstrating efficient drug delivery to hair follicles (Fig. 4I). Although NPs themselves can't pass through the skin barrier completely, they can be entrapped deeply within the hair follicle. Additionally, the high surface area to volume ratio enhances the contact between particles and target cells. NPs systems penetrate preferentially into the follicle in a time-dependent manner; meanwhile optimal size NPs exhibit higher follicular accumulation [41]. Thus, polymer-based synthetic nanotechnology-based formulations have gained growing interest since the last decade as a follicular drug delivery treatment.

Nanotechnology- based formulations	Encapsulated therapeutics	Composition	Size (nm)	Zeta potential (mV)	Application	Advantages	Ref.
Polymeric nanoparticles	Minoxidil	Methylcellulose	90–300	NA	In vivo AGA-induced C57BL/6 mice model	Enhance drug aggregation and expressions of hair-growth factors in hair bulbs No skin stimulation	[47]
	Minoxidil	Chitosan	$235.5\pm99.9$	$+38.6 \pm 6.0$	In vitro porcine ears skin permeation test	Sustained drug release Increased drug permeation into hair follicles	[48]
	FIN	PLGA	316.5±14.4	NA	In vitro polydimethylsiloxane membrane permeation test	Encapsulation efficiency 79.49% $\pm$ 0.47% Prolonged drug release Improved residence into hair follicles	[49]
	Minoxidil	Hyaluronic acid (HA)-PLGA	$243\pm44.5$	NA	In vitro rat skin permeation test	Higher skin permeability Uptake by hair follicle dermal papillary cells	[50]
	Latanoprost	Poly- <i>ɛ</i> -caprolactone	97.8±1.2	$-30.1 \pm 1.8$	In vitro porcine ears skin permeation test	Stable storage for 90 days Improved drug accumulation into hair follicles	[51]
	Rosuvastatin	Methyl-β-cyclodextrin 10% Polyvinyl Pyrrolidone K30	218	NA	In vivo hair loss Albino rat model	Sustained drug release Activation of epithelial stem cells of hair follicle	[52]
	Minoxidil	Poly-(q-caprolactone)-block- poly(ethyleneglycol)	40 - 130	NA	In vivo skin retention test	Penetrated mainly via hair follicles routes	[53]
	Quercetin	Dipalmotyl (DPPC)-PLGA	339±1.6	$-32.6\pm0.51$	In-vivo alopecia-induced rat models	Entrapment efficiency 78% $\pm$ 5.5% Uptake by hair follicles Inhibit hair follicle cells apoptosis in vivo	[54]
	$\alpha$ -Mangostin	Ethyl cellulose	436.0±11.5	NA	Therapeutic effect study in 10 acne patients	Sustained release in human synthetic sebum Excellent hair follicle entrapment	[55]
	Adapalene	PEG5K-b-oligo (DTO-SA)-b-PEG5K	64.7 - 81.6	NA	In vitro human cadaver and porcine ear skin permeation test	Increased drug accumulation in hair follicles	[56]
	Adapalene	Poly(amidoamine)	$256\pm12$	$19.0 \pm 3.1$ (0.05%) $27.6 \pm 2.3$ (0.07%)	In vitro abdominal porcine skin permeation test	Increased drug accumulation in hair follicles and skin.	[57]

(continued on next page)

# Table 1 (continued)

Nanotechnology- based formulations	Encapsulated therapeutics	Composition	Size (nm)	Zeta potential (mV)	Application	Advantages	Ref.
	Dexamethasone	Eudragit® L100	303.1±5.5	NA	In vitro porcine skin permeation test	pH-sensitive Significant transfollicular penetration	[58]
	Adapalene	poly- $\varepsilon$ -caprolactone	107.5±8.19	-13.1	In vitro full-thickness human skin permeation and distribution test	Preferential targeting to PSU	[59]
	Clindamycin	Chitosan HA	$\begin{array}{c} 362\pm19\\ 417\pm9 \end{array}$	$27.7 \pm 0.9 \\ -30.2 \pm 2.7$	In vitro skin penetration test using intact skin porcine, skin with the PSU artificially blocked and schescous skin	Enhanced targeted delivery to pilosebaceous structures	[60]
	Isotretinoin	Delonix polymer	$230\pm10$	$-67\pm3$	In vitro pig ear skin permeation test	Significant follicular targeting Function as follicular drug reservoir	[61]
	Cyclosporin A	Polylactic acid (PLA)	$152.2 \pm 5$	$-16\pm0.2$	In vitro porcine skin permeation test	Increased skin permeation/hair follicles accumulation	[62]
	Dutasteride	Poly-(ε-caprolactone)-lipid	$199.0\pm0.5$	$-13.6 \pm 0.6$	In vitro porcine's ear skin	5-fold increase of hair follicles	[63]
Polymeric micellar	Adapalene	D-α-tocopheryl polyethylene glycol succinate diblock copolymer	4 - 12	NA	In vitro full-thickness porcine and human skin permeation test	Preferential accumulation in the follicular orifice	[64]
	Benzoyl peroxide	Pluronic® F127	24.8 - 25.9	-2 to -13	In vitro porcine skin permeation test	Drug deposition in the follicular pathway	[65]
Nanoemulsions	Minoxidil	Clove oil Kolliphor® P188	10	NA	In vitro follicular drug penetration test	Controlled drug release 26-fold drug penetrated into hair follicles	[66]
	Minoxidil	Eucalyptol Oleic Acid	$29.6 \pm 3.1 \\ 19.5 \pm 1.3 \\ 8.0 \pm 0.5 \\ 12.4 \pm 0.1$	NA	In vitro full-thickness excised human skin permeation test	Promoted drug retention in deeper skin layers Greater hair follicle penetration	[67]
	FIN	Soya lecithin Polyethylene glycol 600	195.2±9.43	-7.61±1.35	In vivo AGA-induced Swiss albino mice model	Increased hair diameter and length Restored the follicle station Be safe and stable for more than 90 days	[68]
	Luteolin	Poly (ethylene oxide)-block- poly(ε-caprolactone) Lecithin	290	NA	In vivo alopecia-induced C57BL/6 mice model	Stability for long-term storage Hair growth-promotion activity	[69]
	Cedrol	Medium chain oil Span 80	$14.26 \pm 0.16$	NA	In vivo alopecia-induced C57BL/6 mice model	Improved drug solubility Increased growth rate of hair follicles	[70]

(continued on next page)

# Table 1 (continued)

Nanotechnology- based formulations	Encapsulated therapeutics	Composition	Size (nm)	Zeta potential (mV)	Application	Advantages	Ref.
Nanostructured lipid carriers	Minoxidil	Stearic acid Oleic acid	281.4±7.4	$-32.9 \pm 1.23$	In vitro rat skin permeation test	Drug entrapment efficiency 92.48% ± 0.31% Promoted hair follicles retention	[71]
	Minoxidil, Tretinoin Minoxidil	Phospholipid Cholesterol Squalene Precirol®	$\begin{array}{c} 149.33 \pm 1.4 \\ 236.0 \pm 3.3 \end{array}$	$7.74 \pm 0.22 \\ -43.8 \pm 0.9$	In vitro rat skin permeation test In vivo skin permeation test	Promoted hair layers retention Ameliorated follicular uptake Promoted proliferation of	[72] [73]
		Anti-platelet-derived growth factor	$194.5\pm4.7$	$-45.5\pm0.6$		dermal papilla cells Up regulation of hair regeneration related factor	
	Diphencyprone	Squalene	$236.3 \pm 3.2$	$-52.8 \pm 4.7$	In vivo nude mouse dorsal skin permeation test	Improved drug targeting to follicles	[39]
	Spironolactone	Olive oil Transcutol® Tween 80	$215.6 \pm 20.4$	$-18.7 \pm 0.92$	In vitro skin permeation test	Entrapment efficiency $87.36\% \pm 3.34\%$ Deliver the NLCs within the follicles	[74]
	Dutasteride	Stearic acid	$187.6\pm7.0$	$-18 \pm 0.9$ (uncoated) 25.8 $\pm$ 1.1 (coated)	In vitro porcine skin permeation test	Entrapment efficiency 97.8% $\pm$ 0.68% Promoted penetration in the hair follicular region	[75]
	Dutasteride	Lauric acid Chitosan	184.2±2.9	$-18 \pm 2.3$ (uncoated) $24.8 \pm 2.1$ (coated)	In vitro porcine skin permeation test	Physically stable for 180 days Enhanced cell proliferation of human dermal papilla cells	[76]
	Cyproterone acetate	Stearic acid Cholesterol Triolein	300	$-35\pm0.5$	In vivo hamsters skin permeation test	Enhanced accumulation in hair follicles Increased drug accumulation in dermis and epidermis	[77]
	Melatonin	Palmitostearate Evening primrose Olive Soybean Bitter almond	$\begin{array}{c} 683 \pm 27.08 \\ 307 \pm 18.31 \\ 307 \pm 3.68 \\ 303 \pm 16.24 \end{array}$	$\begin{array}{c} -17.2\pm0.53\\ -15.1\pm0.22\\ -6.6\pm0.14\\ -14.6\pm0.78\end{array}$	Therapeutic effect study in 40 male AGA patients	Increased hair density and thickness	[78]
	Arginine	Precirol® Oleic acid	87.34	-24.6	In vivo hamsters skin permeation test	Increased accumulation in the hair follicles Accelerate new hair follicle growth	[79]

(continued on next page)

# Table 1 (continued)

Nanotechnology- based formulations	Encapsulated therapeutics	Composition	Size (nm)	Zeta potential (mV)	Application	Advantages	Ref.
	17-α-estradiol	Buriti oil Ceramides	96±15	$-17\pm 6$	In vivo human skin permeation test	Encapsulation efficiency 99.6 $\% \pm 0.3\%$ Physical stability for 42 d Accumulation in the hair follicle	[80]
	Adapalene	Glyceryl distearate Glyceryl monostearate Tween 80 or Span 65	$300.3\pm1.45$	$-21.3 \pm 0.07$	Clinical study in 15 acne vulgaris patients	Sustained drug release Improvement in pilosebaceous follicles	[81]
	Clindamycin phosphate	Stearic acid Oeic acid	$400\pm14$	$-48.9 \pm 0.7$	In vivo skin permeation on porcine skin	Increased accumulation into hair follicles openings	[82]
Solid lipid nanoparticles	Flutamide	Precirol ATO-5® Span 80	$192\pm13$	NA	In vivo skin permeation and hair growth test	Good stability for 2 months Higher accumulation in the hair follicles	[33]
	Roxithromycin (ROX)	Compritol ATO 888 Poloxamer 188	172±2	$-31.68 \pm 3.10$	In vivo skin permeation on a male volunteer	Long term stability up to 26 weeks Increased penetration to human PSU	[12]
Transferosome	FIN	Phospholipon 90 G Span 65	$299.6 \pm 45.6$ $171.0 \pm 5.6$ $197.4 \pm 29.1$	NA	In vivo rat skin permeation test	Enhanced drug permeation in skin layer	[83]
	Minoxidil Caffeine	Polysorbate 20 Polysorbate 80	NA	NA	In vivo AGA-induced rat model	Enhanced hair length	[84]
Ethosome	Cryptotanshinone	Soybean phosphatidycholine Ethanol	69.1±1.9	NA	In vivo anti-acne effect in rabbit model	Increased anti-acne effect	[85]
Liquid crystal nanocarrier	Minoxidil	Monoglycerides Phospholipids Poloxamer 407	82±1	$-57\pm3$	In vivo hair regrowth efficacy test on rats	Selective delivery to pilosebaceous follicle	[86]



Fig. 4 – Application of polymer nanoparticles in hair follicle-targeting drug release system and for the treatment of hair loss. I. Nile red/ ROX-loaded nanoparticles for hair follicle targeting after 1 h of incubation time. [88] II. Fluorescence microphotographs of rat skin incubating with RhB labeled HA-PLGA NP for 4, 6, 8, and 12 h. Scale bar =  $100 \,\mu$ m. [50] III. Cyanoacrylate surface biopsies of (A) Nile red labeled TyroSpheres and (B) Nile red solution treated porcine ear skin. [56] IV. Confocal laser scanning microscope (CLSM) images of abdominal porcine skin deposited (a) FITC-labeled Polyamidoamine dendrimers and (b) free FITC solution for 8 h. [57] V (A) The x-y CLSM images of porcine skin treated with (a) Nile Red solution, (b) Nile Red-loaded polymeric micelles, and (c) x-z image of porcine skin treated with Nile Red loaded polymeric micelles. Scale bar =  $50 \,\mu$ m. (B) The CLSM images at depths of (a)  $14 \,\mu$ m, (b)  $150 \,\mu$ m, and (c)  $400 \,\mu$ m of Nile Red loaded polymeric micelles deposited porcine skin. Scale bar =  $100 \,\mu$ m. [65]

Though topical minoxidil is generally well tolerated, the efficacy of drug treatments remains low [89]. The low effectiveness is likely due to low absorption: about 1.4% of topically applied minoxidil can be absorbed through a normal intact scalp [90]. Minoxidil-encapsulated HA-PLGA NPs [50] have reportedly delivered drugs directly into hair follicles with time-dependent increase (Fig. 4II). At 4h, most of the fluorescence was measured in the SC and epidermis, which meant that Rhodamine B (RhB)-HA-PLGA NPs were more likely to remain only on the skin surface. After 6h, the total fluorescence of follicle cells increased while the SC decreased. Adapalene, the first-line treatment for acne vulgaris, belongs to a third-generation retinoid with the effects of prostration, keratinization, and antilipid secretion. Unfortunately, adapalene's physicochemical properties (logP = 8.04, pKa = 4.23) limit its topical application in the skin and hair follicles. "Retinoid reactions" are the most common adverse effects of topical adapalene therapy, including erythema, burning feeling, and scaling in dose-dependent. To overcome these limitations, Ramezanli et al. [56] developed biocompatible self-assembled nanoparticles (TyroSpheres) for follicular drug delivery of adapalene. *In vivo* cyanoacrylate follicular biopsies test exhibited the effective release of fluorescent dye-labeled TyroSpheres within the hair follicle (Fig. 4III). The accumulation of adapalene was  $51.5 \pm 10.8$  and  $33.0 \pm 11.8 \,\mu\text{g/mm}^2$  respectively in hair follicles as a result of TyroSpheres and Differin®, indicating that the TyroSphere could significantly increase the drug deposition to

the PSU. Besides, the partition coefficient values of the drug in sebum/water (Ksebum) and SC/water (Ksc) were calculated to be  $39.5\pm7.1$  and  $18.6\pm1.5$ , respectively. These results suggest that TyroSpheres have been transported through PSU. Furthermore, the location of drug carriers is vital in hair follicle-associated disorders treatment. Studies demonstrated that nanoparticles accumulated in the follicles rather than penetrated into the deeper layers of the skin (Fig. 4IV). For example, Pluronic® F127 micelles in nanoscale could be transmitted at the maximum 400  $\mu$ m of hair follicles (Fig. 4V), indicating they did not pass through the intact SC but selectively localized in skin structures.

#### 3.1.2. Lipid nanoparticles

Lipid nanoparticles include solid lipid nanoparticles (SLNs) and NLCs, which are considered as suitable drug delivery systems due to their biocompatibility, degradability, and low toxicity. SLNs with a size of 40–1000 nm are lipid-based nanoparticles formed only from solid lipids. NLCs were designed to overcome some potential shortcomings of SLNs and exhibit a higher loading capacity and long-term stability [91]. Hair follicles have been confirmed to be a potential pathway for NLCs penetration resulting from the lipophilic property of the follicular ducts (Fig. 51). Moreover, a previous study has revealed that free fatty acids used to prepare SLNs and NLCs can function as efficient  $5\alpha$ -reductase inhibitors and stimulate hair growth [75].

The application of SLNs and NLCs to AGA therapy has been extensively investigated because they are able to pass through the skin through the hair follicle route. Lipid nanoparticles with smaller size and occlusive properties can attach themselves to the skin surface, promoting adhesiveness, increasing hydration, and maintaining sustained release stably in the following phase. Moreover, the nanoparticle size appears to be a vital factor affecting the penetration depth and speed of substances through the skin. The previous study has found that SLNs below 100 nm can aggregate into deep layers of skin mainly via hair follicle pathways [92]. Wang et al. [71] prepared minoxidil-encapsulated NLCs with oleic acid as the liquid lipid and stearic acid as the solid lipid. Compared to the minoxidil-loaded SLNs, minoxidil-NLCs showed better storage stability and 10.7-folds higher drug retention in the skin. NLCs broadly promoted the drug deposition in the follicles, which may be due to the fusion with sebum or the opening of inactive follicles.

Not only NLCs alone, but the combinations of NLCs and functional substances also represent a novel delivery strategy and aim to have synergistic effects in promoting hair growth. Aljuffali et al. [73] used anti-platelet-derived growth factor (PDGF)-conjugated NLCs to achieve active targeting of specific cells, which could precisely deliver the drug to follicles and dermal papilla cells (DPCs). The encapsulated minoxidil was largely deposited in the follicles (Fig. 5II) and penetrated into the DPCs, thereby accelerating cell proliferation and upregulating the expression of angiogenesis-related factors. *In vivo* skin permeation analysis demonstrated that anti-PDGF-NLCs could increase the accumulation of minoxidil in follicles by three-fold compared to the solution group. Furthermore, chitosan is considered a highly favorable material and possesses excellent biocompatibility and nontoxicity properties. Positively charged chitosan may be beneficial for skin and hair delivery, as it would be attracted to negatively charged skin and hair. And it has been proved that NPs with chitosan facilitated the hair follicle targeting delivery compared to nanoparticles without chitosan. Besides, Lauric acid itself has been proven to have a remarkable inhibitory effect on both type I and type II  $5\alpha$ reductase activity compared to other fatty acids, possessing significant antiandrogen activity. A recent report from Noor et al. [75] prepared lauric acid-chitosan oligomer-coated dutasteride NLCs, and found that the formulations could promote cell proliferation especially in DPCs.

### 3.1.3. Metallic nanoparticles

Drug molecules can be encapsulated either into the particle core or adsorbed on the particle surface of metallic nanoparticles, including metals (such as iron, gold, or silver) or metal oxides (such as TiO<sub>2</sub>, ZnO [93], and iron oxide [94]). The previous study [95] showed that metallic nanoparticles with a diameter of less than 10 nm were able to pass passively through the hair follicle pores through the skin and reach the deep region of the hair follicle. Gold nanoparticles (GNPs) possess varied unique features, such as intense visible light absorption, and high control over size and shape, making it an attractive metallic nanoparticle for hair follicle-targeting delivery. Recently, gold microparticles (GMPs) with inner silica cores have been introduced to treat acne vulgaris [96]. GMPs applied to the skin surface accumulated abundantly into the PSU through the follicular pathway and combining diode laser pulses, which enabled the direct intercept to the pathophysiology by targeting the sebaceous glands.

Moreover, studies also showed that the size, shape, and surface ligands of metallic nanoparticles could affect the penetration performance of particles into hair follicles (Fig.6I A). For instance, Friedman et al. [97] explored the effects of size as well as the shape on GNPs regarding its possibility for hair follicle permeation. They found that particles with a diameter of 40 nm could penetrate a depth of 210 nm, and 250 nm particles penetrated up to 252 nm, while particles of 530 nm could not penetrate at all (Fig. 6I B). Studies on the morphology of GNP have shown that particles in spherical have lower penetration relative to nanorods. Furthermore, nanostars could abundantly accumulate in the PSU (Fig. 6II), indicating that the anisotropic shapes of nanoparticles play a significant role in particle accumulation. In addition to size and shape, functionalized surface ligands (such as neutral, anionic, cationic, and hydrophobic ligand) on gold nanorods (GNRs) have also been proven to influence the drug deposition into hair follicles in human skin sheets (Fig. 6III). Mahmoud et al. [98] indicated that hydrophobic polystyrene (PS)-GNRs might be more advantageous for hair follicle-targeting than PEGylated GNRs because of the lipophilic property of the follicular compartments (Fig. 6IV). Consequently, a brannew composite formulation may enable greater follicular penetration of metallic nanoparticles.

## 3.2. Microneedles

As a new mode of administration, microneedles (MNs) can easily break the SC barrier and release their loaded drug into



Fig. 5 – Lipid nanoparticles for hair follicle-targeting drug system. I. Confocal microscopy images of human skin incubating with the fluorescence labeled NLC formulation: (a) rhodamine labeled NLCs (red); (b) Alexa Fluor 488 phalloidin labeled actin (green); (c) TO-PRO-3 labeled nucleus (blue); (d) bright field; (e) merge of (a), (b) and (c). [80] Scale bar =20 µm. II. (A) H&E staining and fluorescence microphotographs of the skin treated with Nile red labeled minoxidil formulations after 4 h. Scale bar =150 µm. (B) Confocal micrographs showing the distribution of Nile red on the skin surface. Scale bar =150 µm. [73]

the epidermal and dermal layers, thereby delivering drugs into hair follicles directly. MNs with enough mechanical strength are able to puncture into the skin to form microchannels, possessing the merits of self-administration, negligible pain, and relatively inexpensive cost [99–102]. Remarkably, applying MNs alone could recruit growth factors through microwounds, at the same time, activate HFSCs and hair growth signaling pathways [103,104]. Recently, MNs have been applied to disorders associated with hair follicles to increase the efficiency of percutaneous absorption and reduce dose frequency, which can act synergistically to enhance follicular drug delivery (Table 2).



Fig. 6 – The effect of size, shape, and surface ligands on the capacity to follicular drug delivery of metallic nanoparticles. I. (A) Schematic illustration of size, shape, and surface ligand effect of GNPs on follicular penetration. (B) Fluorescence microscopy images of full human skin samples treated fluorescence-labeled Polystyrene nanoparticles with different diameters (Red: fluorescence-labeled nanoparticles; blue: DAPI). [97] II. The shape control of GNPs: (A) The transmission electron microscopy (TEM) and scanning electron microscopy (SEM) images of (A1) synthesized spheres, (A2) rods, and (A3) stars GNPs; (B) Follicular penetration of fluorescence-labeled gold (B1) nanospheres, (B2) nanorods, and (B3) nanostars. Scale bar = 200 µm. [97] III. The effect of surface ligands on the GNR deposition into the hair follicles in human skin (PS: hydrophobic polystyrene). [98] IV. (A) Microscopic and (B) LA-ICP-MS scanning image exhibiting the distribution of gold in SC, epidermis, dermis, and a hair follicle of human skin sample treated with PEG-GNR for 24h. Scale bar = 1000 µm. [98]

Recent advances by Yuan et al. developed ceria nanozyme (CeNZ)-encapsulated microneedles (Ce-MNs) patch, achieving faster hair regeneration with high quality. The CeNZs were efficiently delivered into skin (200-300 nm deep) through MNs, which could relieve oxidative stress and stimulate angiogenesis to remodel the hair follicle's microenvironment [105]. Besides, Fang et al. [106] fabricated a polyvinyl alcohol (PVA)-dissolvable microneedle patch loading mesoporous iron oxide (MIO) and minoxidil (MX-MIO@MNs) to facilitate hair regrowth. MX-MIO@MNs could be triggered by the external magnetic field, promoting local heating and controlled drug release. The releasing MIOs further captured hydrophobic pharmaceuticals, and the hydrophobic substances are the vital substances supporting hair regrowth in the human body. MX-MIO@MNs with appropriate mechanical property could successfully deliver the minoxidil-loaded MIOs to the skin and achieve more than 8-fold improvements in hair growth with no irritation. Others including Cao et al. [107] developed squalene-containing microneedles mediated with FIN NLCs to enhance the permeability to hair follicles and reduce the systemic adverse effects resulting from oral administration. The lipidic composition of NLCs mimicking natural components of hair follicles markedly improved the coumarin 6 (C6)-NLCs uptake in human dermal papilla cells. In vitro and in vivo permeation tests both revealed that the combination of NLCs with MN had significantly higher skin retention compared that in a FIN-NLCs and FIN suspension (Fig. 7I). Detailed permeation analysis showed that C6-NLCs-MN presented a better delivery of follicular targeting than C6-NLCs or C6-suspension groups. Moreover, FIN-NLCs-MNs exhibited higher therapeutic efficacy of hair regeneration and telogen-to-anagen conversion than the FIN-NLCs or the commercial minoxidil groups on AGA-induced mice.



Fig. 7 – Microneedles and exogenous stimulation-mediated formulations for follicular drug delivery. I. Visualization pathway of C6 suspension, C6-NLCs, and C6-NLCs-MN using CLSM after 24 h of permeation in rat skin. [107] II. Laser scanning microscopy (LSM) image of a follicular cross-section in the (a1) transmission and (a2) fluorescence mode from the AuNP-doped BSA particles treated group; LSM image of a follicular cross-section with a visible sebaceous gland in the (b1) transmission and (b2) fluorescence mode from the AuNP-doped BSA particles treated group. Scale bar = 250 µm. [110] III (A) Schematic illustration of UVA-responsive nanocapsules (NCs). (B) CLSM images of porcine ear skin after 2 min of UVA irradiation and incubation with fluorescence-labeled NCs for 30 min. Scale bar = 200 µm. (C) Three-dimensional plot images of porcine ear skin treated with fluorescence-labeled NCs after (a) 0 min, (b) 2 min, and (c) 8 min of irradiation with UVA (power density: 11.96 mW/cm<sup>2</sup>, peak wavelength = 365 nm). [117]

Therapeutics	Microneedle types	Microneedle matrix	Characterizations	Advantages	Application	Ref.
UK5099 Exosome	Dissolving MNs	Keratin HA	Height: 600 μm Base diameter:300 μm 15 × 15 array Depth in 200 μm	Accelerate HFSCs activation Reduced dosage Accelerated pigmentation and hair regrowth within 6 days	AGA	[16]
FIN	Dissolving MNs	НА	Mechanical strength:0.6 N/needle Depth into 400 µm	Punctured into the deeper layers of the HF site Promoted hair regeneration Accelerated the hair cycle transition	AGA	[107]
Valproic acid	Dissolving MNs	Carboxymethyl cellulose (CMC)	Height: $600 \pm 22.32 \mu\text{m}$ Tip diameter: $20 \pm 8 \mu\text{m}$ $7 \times 7$ micro-cavities	Higher accuracy than topical application Increased expression of hair regrowth related pathway	Hair regrowth	[108
MIO Minoxidil	Dissolving MNs	PVA	Height: 600 µm 11 × 11 array Mechanical strength >0.17 N/needle	Increased blood flow as well as vasodilation in skin Magneto-thermal drug release achieved great improvements of hair regeneration Low toxicity	AGA	[106
FIN powder	Dissolving MNs	СМС	Height: $801.5 \pm 6.2 \mu\text{m}$ Base diameter: $450.7 \pm 6.8 \mu\text{m}$ $5 \times 5 \text{array}$	Release drug into dermal layers Sustained release for 3 days after one application Promoted hair growth	AGA	[102
Ceria Nanozyme	Dissolving MNs	HA Polyvinylpyrrolidone K90	Height: $550 \mu\text{m}$ Base diameter: $200 \mu\text{m}$ $15 \times 15 array$ Mechanical strength: $4.13 \pm 0.83 \text{N}$ $200-300 \mu\text{m}$ deep in the skin	Remodeled the microvasculature in the hair loss area Lower administration frequency Speed up hair regrowth in the AGA mice model	AGA	[105
Clindamycin	Responsive MNs	PVA	Height: 600 µm Base diameter: 300 µm Mechanical strength: 2.2 N/needle	ROS-responsive MN patch Enhanced efficacy toward dermis lesions	Acne vulgaris	[109

# 3.3. Exogenous stimulation-mediated formulations for follicular drug delivery

Furthermore, nanoparticles should be equipped with an appropriate triggered release mechanism to achieve continuous drug release exogenously. In this regard, nanoparticles play an important role in transporting active substances to hair follicles. Different exogenous trigger mechanisms, including infrared light [110], pH [58], and ultrasound (US), ultraviolet (UV)-responsive, are investigated to stimulate a controlled drug release and enable translocation into hair follicles once released. Short periods of heat application may facilitate preferential drug delivery to hair follicles because the viscosity of human sebum reduces as the temperature increases [111]. Previous research by Jung et al. showed an increased follicular penetration depth at 37 °C compared to that at 21 °C and 32 °C [112], suggesting that formulations can achieve enhanced follicle deposition above their cloud point temperature. Lademann et al. demonstrated that water-filtered infrared A radiation could facilitate the release of model drugs targeting the follicular structures [110]. The plasmonic heat caused by radiation made stronger fluorescent signals for model drugs on the SC layers in addition to the sebaceous glands and follicular ducts in 95.2% of hair follicles (Fig 7II). Furthermore, the utilization of the pH gradient is very promising because it has been proven that pH values range from 6.5 on the pig ear skin surface to 7.4 in lower HF regions [113], providing a clear direction for pH-triggered intrafollicular drug delivery. More recently, Dong et al. demonstrated the possibility of a



Fig. 8 – Clinical application of hair follicle-targeting delivery in the treatment of AGA and acne vulgaris. I. (A) Two male patients with AGA (a, c) before therapy and (b, d) after 3-month treatment (top row: melatonin solution; bottom row: melatonin nanovesicles). (B) The hair shaft diameter of male patients with AGA (a, c) before therapy and (b, d) after 3-month treatment (top row: melatonin solution; bottom row: melatonin nanovesicles) [118] II. (A) The mean percent of inflammatory lesions in the gold microparticles-treated group and control group after 3 months (N: the number of patients at different time point; bar: standard error of the mean). (B) Images showing the lesions of inflammatory acne vulgaris on the cheeks (top row: before therapy; bottom row: after 6-month treatment of gold microparticles). [119]

pH-responsive burst drug release from Eudragit® L100 NPs. Electron paramagnetic resonance and CLSM investigations showed transfollicular penetration and increased aggregation into hair follicles [58]. What's more, Liao et al. developed an innovative integrated drug delivery platform by combing US energy with multifunctional microbubbles (MBs). The authors explored the effects of minoxidil-coated lysozyme-shelled MBs and albumin-shelled MBs with US. The results showed that the delivery platform could enhance the deposition of therapeutic drugs to the target structure of interest in the hair follicle, thereby offering new therapeutic opportunities both in hair loss and acne vulgaris [114-116]. Similar results were observed by Busch et al., who developed a follicular targeting ultraviolet A (UVA)-triggered drug system from nanocapsules ([117]; Fig. 7 III A and B). The fluorescence brightness of the model drug decreased with the increase of UVA irradiation time (Fig. 7 III C), possibly because the drug was insoluble in the intrafollicular environment. These results indicated that the drug preferentially accumulated in hair follicles.

## 4. Clinical application

Whereas hair follicle-targeting drug delivery strategies are still limited by regulatory approval in clinical trials, breakthrough therapies are currently underway in small groups of patients. Roque et al. prepared FINloaded PLGA nanoparticles using a modified method for emulsification/solvent diffusion [49]. In clinical safety testing, all formulation excipients did not cause any erythema or other untoward effects. Hatem et al. encapsulated melatonin in vitamin C-based nanovesicles to enhance its therapeutic efficacy and skin retention [118]. Further studies showed that melatonin nanovesicles clinically increased hair density and diameter and reduced hair loss compared to melatonin solution (Fig. 8I). Additionally, two independent prospective randomized controlled clinical trials have investigated the topical application of gold-coated silica microparticles in managing moderate to severe acne vulgaris [119]. Microparticles with a 15-nm-thick gold shell and 120-nm-diameter silica core, providing deep penetration into the sebaceous glands, appeared statistically effective treatment in each trial. In Trial 1, the lesions of inflammatory acne vulgaris were significantly cut down and reached -61% compared to the baseline at 28 weeks (Fig. 8II).

The current work demonstrates that hair follicle-targeting drug delivery could reduce side effects and generate more effective treatments. To date, there is a variety of researches concerning an increased follicular penetration when applied with strategies like nanocarrier systems. Still, only very few products based on the conception of hair follicle-targeting are available on the market. Most studies focused on the follicular permeation in vitro or in vivo, and the investigation about their clinical efficacy is rare at present. The relative translation still remains a considerable challenge. On one side, obtaining the same clinical outcomes from penetration experiments in the laboratory is extremely difficult. We often choose porcine, rat, mouse, or hamster skin as the substitute for human skin in the lab, and they invariably produce quite different permeation results. On the other hand, there are still concerns about nanoparticles' potential toxicity, requiring further explorations on the possible dangers and long-term fate of nanomaterials in hair follicles.

# 5. Conclusions and future perspectives

Hair follicle-targeting drug delivery formulations have been found to obtain overall performance enhancements, which improve drug efficacy by enhancing the skin and follicle drug delivery and decreasing side effects compared to conventional formulations. Based on the particularities of the hair follicles, the drugs trapped in hair follicles enable sustained and controlled release, allowing better patient compliance and treatment efficiency. During the past years, the potential of the hair follicle as a penetration pathway for the treatment of hair follicle-associated disorders like hair loss and acne vulgaris has been recognized.

So far and overall, hair follicle-targeting drug delivery represents a promising concept, and there are additional challenges (Fig. 9). Hair follicles are more than a complex structure. For example, most human hair follicles will remain for 2–6 years in the anagen phase. They go through extensive changes with the transformation of the hair follicle cycle. Different cycles status certainly affect the follicular penetration and must be taken into consideration while designing formulations. Furthermore, the sebaceous content also hampers nanoparticle accumulation into the hair follicle. Topically applied substances can't enter the 'closed' hair follicles, as the dry sebum and cell debris would block the hair follicles. Increasing the amounts of drugs that can enter the hair follicle with the presence of the SC should be the focus of future studies. Last but not least, every disease has its own characteristics, and it is difficult to design a



Fig. 9 – Research to clinical: the challenges in future prospect and clinical application for hair follicle-targeting drug delivery. IVIVC: in vivo and in vitro consistence.

compatible delivery system for a new bioactive molecule under a particular skin condition and purpose. It has been reported that the movement of the hair plays a crucial role in the hair follicle targeting process of nanoparticles [23]. In disease states like hair loss, the interactions between particles and hair follicles and also the influence of hair should be considered in further studies. Thus, as a future research direction, the selection of corresponding nanocarrier types and more reliable skin models that can mimic the diseased state is absolutely vital for permeation experiments in follicletargeting drug delivery.

## **CRediT** authorship contribution statement

Yueting Gu: Investigation, Writing-original draft. Qiong Bian: Writing-review& editing. Yanjun Zhou: Participation in discussion. Qiaoling Huang: Conceptualization. Jianqing Gao: Conceptualization, Supervision.

# **Conflicts of interest**

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

#### REFERENCES

[1] Roberts MS, Cheruvu HS, Mangion SE, Alinaghi A, Benson HAE, Mohammed Y, et al. Topical drug delivery: history, percutaneous absorption, and product development. Adv Drug Deliv Rev 2021;177:113929–71.

- [2] Jamaledin R, Yiu CKY, Zare EN, Niu LN, Vecchione R, Chen G, et al. Advances in antimicrobial microneedle patches for combating infections. Adv Mater 2020;32(33):1–29.
- [3] Sheng T, Luo B, Zhang W, Ge X, Yu J, Zhang Y, et al. Microneedle-mediated vaccination: innovation and translation. Adv Drug Deliv Rev 2021;179:113919–41.
- [4] Svenskaya YI, Genina EA, Parakhonskiy BV, Lengert EV, Talnikova EE, Terentyuk GS, et al. A simple non-invasive approach toward efficient transdermal drug delivery based on biodegradable particulate system. ACS Appl Mater Interfaces 2019;11(19):17270–82.
- [5] Elmahdy A, Cao Y, Hui X, Maibach H. Follicular pathway role in chemical warfare simulants percutaneous penetration. J Appl Toxicol 2021;41(6):964–71.
- [6] Randolph M, Tosti A . Oral minoxidil treatment for hair loss: a review of efficacy and safety. J Am Acad Dermatol 2021;84(3):737–46.
- [7] Yang G, Chen G, Gu Z. Transdermal drug delivery for hair regrowth. Mol Pharm 2021;18(2):483–90.
- [8] Wen T, Lin Z, Zhao Y, Zhou Y, Niu B, Shi C, et al. Bioresponsive nanoarchitectonics-integrated microneedles for amplified chemo-photodynamic therapy against acne vulgaris. ACS Appl Mater Interfaces 2021;13(41):48433–48.
- [9] Klein AL, Lubda M, Skov PS, Vogt A, Keck CM, Lademann J, et al. Investigation of transfollicular caffeine penetration using microdialysis on ex vivo porcine ear skin. Eur J Pharm Biopharm 2020;157:1–8.
- [10] Subongkot T, Sirirak T. Development and skin penetration pathway evaluation of microemulsions for enhancing the dermal delivery of celecoxib. Colloids Surfaces B Biointerfaces 2020;193:111103–9.
- [11] Lademann J, Richter H, Schaefer UF, Blume-Peytavi U, Teichmann A, Otberg N, et al. Hair follicle - a long-term reservoir for drug delivery. Skin Pharmacol Physiol 2006;19(4):232–6.
- [12] Wosicka-Frąckowiak H, Cal K, Stefanowska J, Główka E, Nowacka M, Struck-Lewicka W, et al. Roxithromycin-loaded lipid nanoparticles for follicular targeting. Int J Pharm 2015;495(2):807–15.
- [13] Yap CX, Sidorenko J, Wu Y, Kemper KE, Yang J, Wray NR, et al. Dissection of genetic variation and evidence for pleiotropy in male pattern baldness. Nat Commun 2018;9(1):1–12.
- [14] Yuan AR, Bian Q, Gao JQ. Current advances in stem cell-based therapies for hair regeneration. Eur J Pharmacol 2020;881:173197–208.
- [15] Gentile P, Garcovich S. Advances in regenerative stem cell therapy in androgenic alopecia and hair loss: Wnt Pathway, growth-factor, and mesenchymal stem cell signaling impact analysis on cell growth and hair follicle development. Cells 2019;8(5):466–86.
- [16] Yang G, Chen Q, Wen D, Chen Z, Wang J, Chen G, et al. A therapeutic microneedle patch made from hair-derived keratin for promoting hair regrowth. ACS Nano 2019;13(4):4354–60.
- [17] Mancuso A, Cristiano MC, Fresta M, Paolino D. The challenge of nanovesicles for selective topical delivery for acne treatment: enhancing absorption whilst avoiding toxicity. Int J Nanomedicine 2020;15:9197–210.
- [18] Kolli SS, Pecone D, Pona A, Cline A, Feldman SR. Topical retinoids in acne vulgaris: a systematic review. Am J Clin Dermatol 2019;20(3):345–65.
- [19] Habeshian KA, Cohen BA . Current issues in the treatment of acne vulgaris. Pediatrics 2020;145(2):225–32.
- [20] Oulès B, Philippeos C, Segal J, Tihy M, Vietri Rudan M, Cujba AM, et al. Contribution of GATA6 to homeostasis of

the human upper pilosebaceous unit and acne pathogenesis. Nat Commun 2020;11(1):5067–83.

- [21] Fuchs CSK, Ortner VK, Hansen FS, Philipsen PA, Haedersdal M . Subclinical effects of adapalene-benzoyl peroxide: a prospective in vivo imaging study on acne micromorphology and transfollicular delivery. J Eur Acad Dermatology Venereol 2021;35(6):1377–85.
- [22] Patzelt A, Lademann J. Drug delivery to hair follicles. Expert Opin Drug Deliv 2013;10(6):787–97.
- [23] Patzelt A, Lademann J. Recent advances in follicular drug delivery of nanoparticles. Expert Opin Drug Deliv 2020;17(1):49–60.
- [24] Flores A, Schell J, Krall AS, Jelinek D, Miranda M, Grigorian M, et al. Lactate dehydrogenase activity drives hair follicle stem cell activation. Nat Cell Biol 2017;19(9):1017–26.
- [25] Koester J, Miroshnikova YA, Ghatak S, Chacón-Martínez CA, Morgner J, Li X, et al. Niche stiffening compromises hair follicle stem cell potential during ageing by reducing bivalent promoter accessibility. Nat Cell Biol 2021;23(7):771–81.
- [26] Sakamoto K, Jin S-P, Goel S, Jo J-H, Voisin B, Kim D, et al. Disruption of the endopeptidase ADAM10-Notch signaling axis leads to skin dysbiosis and innate lymphoid cell-mediated hair follicle destruction. Immunity 2021;54(10):1–17.
- [27] Shamloul G, Khachemoune A . An updated review of the sebaceous gland and its role in health and diseases part 2: pathophysiological clinical disorders of sebaceous glands. Dermatol Ther 2021;34(2):1–8.
- [28] Chen CL, Huang WY, Wang EHC, Tai KY, Lin SJ. Functional complexity of hair follicle stem cell niche and therapeutic targeting of niche dysfunction for hair regeneration. J Biomed Sci 2020;27(1):1–11.
- [29] Vora LK, Donnelly RF, Larrañeta E, González-Vázquez P, Thakur RRS, Vavia PR. Novel bilayer dissolving microneedle arrays with concentrated PLGA nano-microparticles for targeted intradermal delivery: proof of concept. J Control Release 2017;265:93–101.
- [30] Wang Z, Xue Y, Chen T, Du Q, Zhu Z, Wang Y, et al. Glycyrrhiza acid micelles loaded with licochalcone A for topical delivery: co-penetration and anti-melanogenic effect. Eur J Pharm Sci 2021;167:106029–55.
- [31] Al Mahrooqi JH, Khutoryanskiy VV, Williams AC. Thiolated and PEGylated silica nanoparticle delivery to hair follicles. Int J Pharm 2021;593:120130–7.
- [32] Pelikh O, Eckert RW, Pinnapireddy SR, Keck CM . Hair follicle targeting with curcumin nanocrystals: influence of the formulation properties on the penetration efficacy. J Control Release 2021;329:598–613.
- [33] Hamishehkar H, Ghanbarzadeh S, Sepehran S, Javadzadeh Y, Adib ZM, Kouhsoltani M . Histological assessment of follicular delivery of flutamide by solid lipid nanoparticles: potential tool for the treatment of androgenic alopecia. Drug Dev Ind Pharm 2016;42(6):846–53.
- [34] Radtke M, Patzelt A, Knorr F, Lademann J, Netz RR. Ratchet effect for nanoparticle transport in hair follicles. Eur J Pharm Biopharm 2017;116:125–30.
- [35] Lademann J, Patzelt A, Richter H, Antoniou C, Sterry W, Knorr F. Determination of the cuticula thickness of human and porcine hairs and their potential influence on the penetration of nanoparticles into the hair follicles. J Biomed Opt 2009;14(2):210141–4.
- [36] Dragicevic N., Maibach H.I. Percutaneous penetration enhancers chemical methods in penetration enhancement: nanocarriers. 2016.
- [37] Lademann J, Richter H, Teichmann A, Otberg N, Blume-peytavi U, Luengo J, et al. Nanoparticles – an

efficient carrier for drug delivery into the hair follicles. Eur J Pharm Biopharm 2007;66:159–64.

- [38] Raber AS, Mittal A, Schäfer J, Bakowsky U, Reichrath J, Vogt T, et al. Quantification of nanoparticle uptake into hair follicles in pig ear and human forearm. J Control Release 2014;179:25–32.
- [39] Lin YK, Al-Suwayeh SA, Leu YL, Shen FM, Fang JY . Squalene-containing nanostructured lipid carriers promote percutaneous absorption and hair follicle targeting of diphencyprone for treating alopecia areata. Pharm Res 2013;30(2):435–46.
- [40] Patzelt A, Richter H, Knorr F, Schäfer U, Lehr CM, Dähne L, et al. Selective follicular targeting by modification of the particle sizes. J Control Release 2011;150(1):45–8.
- [41] Alvarez-Román R, Naik A, Kalia YN, Guy RH, Fessi H. Skin penetration and distribution of polymeric nanoparticles. J Control Release 2004;99(1):53–62.
- [42] Jain AK, Jain A, Garg NK, Agarwal A, Jain A, Agrawal GP. Biointerfaces Adapalene loaded solid lipid nanoparticles gel: an effective approach for acne treatment. Colloids Surfaces B Biointerfaces 2014;121:222–9.
- [43] Agrahari V, Mitra AK. Delivery and targeting of nanoparticles into hair follicles. Ther Deliv 2016;7(2):117–38.
- [44] Ghasemiyeh P, Mohammadi-Samani S . Potential of nanoparticles as permeation enhancers and targeted delivery options for skin: advantages and disadvantages. Drug Des Devel Ther 2020;14:3271–89.
- [45] Verma S, Utreja P. Vesicular nanocarrier based treatment of skin fungal infections: potential and emerging trends in nanoscale pharmacotherapy. Asian J Pharm Sci 2019;14(2):117–29.
- [46] Meidan VM, Bonner MC, Michniak BB. Transfollicular drug delivery - is it a reality? Int J Pharm 2005;306:1–14.
- [47] Nagai N, Iwai Y, Sakamoto A, Otake H, Oaku Y, Abe A, et al. Drug delivery system based on minoxidil nanoparticles promotes hair growth in C57BL/6 mice. Int J Nanomedicine 2019;14:7921–31.
- [48] Matos BN, Reis TA, Gratieri T, Gelfuso GM. Chitosan nanoparticles for targeting and sustaining minoxidil sulphate delivery to hair follicles. Int J Biol Macromol 2015;75:225–9.
- [49] Roque LV, Dias IS, Cruz N, Rebelo A, Roberto A, Rijo P, et al. Design of finasteride-loaded nanoparticles for potential treatment of alopecia. Skin Pharmacol Physiol 2017;30(4):197–204.
- [50] Jeong WY, Kim S, Lee SY, Lee H, Han DW, Yang SY, et al. Transdermal delivery of minoxidil using HA-PLGA nanoparticles for the treatment in alopecia. Biomater Res 2019;23(1):1–10.
- [51] Oliveira ACS, Oliveira PM, Cunha-Filho M, Gratieri T, Gelfuso GM. Latanoprost loaded in polymeric nanocapsules for effective topical treatment of alopecia. AAPS PharmSciTech 2020;21(8):1–7.
- [52] Maged A, Mahmoud AA, Salah S, Abd-elmonsif NM, Ghorab MM . Spray-dried rosuvastatin nanoparticles for promoting hair growth. AAPS PharmSciTech 2020;21(6):1–12.
- [53] Shim J, Kang HS, Park WS, Han SH, Kim J, Chang IS. Transdermal delivery of mixnoxidil with block copolymer nanoparticles. J Control Release 2004;97(3):477–84.
- [54] Das L, Kaurav M, Pandey RS. Phospholipid-polymer hybrid nanoparticle-mediated transfollicular delivery of quercetin: prospective implement for the treatment of androgenic alopecia. Drug Dev Ind Pharm 2019;45(10):1654–63.
- [55] Pan-In P, Wongsomboon A, Kokpol C, Chaichanawongsaroj N, Wanichwecharungruang S. Depositing α-mangostin nanoparticles to sebaceous gland area for acne treatment. J Pharmacol Sci 2015;129(4):226–32.
- [56] Ramezanli T, Zhang Z, Michniak-Kohn BB . Development

and characterization of polymeric nanoparticle-based formulation of adapalene for topical acne therapy. Nanomedicine Nanotechnol, Biol Med 2017;13(1):143–52.

- [57] Gökçe BB, Boran T, Emlik Çalık F, Özhan G, Sanyal R, Güngör S. Dermal delivery and follicular targeting of adapalene using PAMAM dendrimers. Drug Deliv Transl Res 2021;11(2):626–46.
- [58] Dong P, Sahle FF, Lohan SB, Saeidpour S, Albrecht S, Teutloff C, et al. pH-sensitive Eudragit® L 100 nanoparticles promote cutaneous penetration and drug release on the skin. J Control Release 2019;295:214–22.
- [59] Sallam MA, Marín Boscá MT . Mechanistic analysis of human skin distribution and follicular targeting of adapalene-loaded biodegradable nanospheres with an insight into hydrogel matrix influence, in vitro skin irritation, and in vivo tolerability. J Pharm Sci 2017;106(10):3140–9.
- [60] Tolentino S, Pereira MN, Cunha-Filho M, Gratieri T, Gelfuso GM . Targeted clindamycin delivery to pilosebaceous units by chitosan or hyaluronic acid nanoparticles for improved topical treatment of acne vulgaris. Carbohydr Polym 2021;253:117295–303.
- [61] Tolulope A, Chahud F, Lopez RF V. Isotretinoin-Delonix polymeric nanoparticles: potentials for skin follicular targeting in acne treatment. Int J Pharm 2021;610:121217–28.
- [62] Gomes AC, Cavaco-paulo A . Cyclosporin A-loaded poly(D, L-lactide) nanoparticles: a promising tool for treating alopecia. Nanomedicine 2020;15(2).
- [63] Ushirobira CY, Luana AFA, Pereira MN, Cunha-filho M, Gelfuso GM, Gratieri T . Dutasteride nanocapsules for hair follicle targeting: effect of chitosan-coating and physical stimulus. Int J Biol Macromol 2020;151:56–61.
- [64] Kandekar SG, Del Río-Sancho S, Lapteva M, Kalia YN . Selective delivery of adapalene to the human hair follicle under finite dose conditions using polymeric micelle nanocarriers. Nanoscale 2018;10(3):1099–110.
- [65] Kahraman E, Özhan G, Özsoy Y, Güngör S. Polymeric micellar nanocarriers of benzoyl peroxide as potential follicular targeting approach for acne treatment. Colloids Surfaces B Biointerfaces 2016;146:692–9.
- [66] Cardoso SA, Barradas TN . Developing formulations for drug follicular targeting: nanoemulsions loaded with minoxidil and clove oil. J Drug Deliv Sci Technol 2020;59:101908–17.
- [67] Abd E, Benson HAE, Roberts MS, Grice JE. Minoxidil skin delivery from nanoemulsion formulations containing eucalyptol or oleic acid: enhanced diffusivity and follicular targeting. Pharmaceutics 2018;10(1):1–12.
- [68] Upadhyay DK, Sharma A, Kaur N, Das Gupta G, Narang RK, Rai VK . Nanoemulgel for efficient topical delivery of finasteride against androgenic alopecia. J Pharm Innov 2020;3:1–12.
- [69] Shin K, Choi H, Song SK, Yu JW, Lee JY, Choi EJ, et al. Nanoemulsion vehicles as carriers for follicular delivery of luteolin. ACS Biomater Sci Eng 2018;4(5):1723–9.
- [70] Deng Y, Huang F, Wang J, Zhang Y, Zhang Y, Su G, et al. Hair growth promoting activity of cedrol nanoemulsion in C57BL/6 mice and its bioavailability. Molecules 2021;26(6):1–15.
- [71] Wang W, Chen L, Huang X, Shao A . Preparation and characterization of minoxidil loaded nanostructured lipid carriers. AAPS PharmSciTech 2017;18(2):509–16.
- [72] Kochar P, Nayak K, Thakkar S, Polaka S, Khunt D, Misra M. Exploring the potential of minoxidil tretinoin liposomal based hydrogel for topical delivery in the treatment of androgenic alopecia. Cutan Ocul Toxicol 2020;39(1):43–53.
- [73] Aljuffali IA, Pan TL, Sung CT, Chang SH, Fang JY . Anti-PDGF receptor  $\beta$  antibody-conjugated squarticles loaded with minoxidil for alopecia treatment by targeting hair follicles

and dermal papilla cells. Nanomedicine Nanotechnol, Biol Med 2015;11(6):1321–30.

- [74] Shamma RN, Aburahma MH. Follicular delivery of spironolactone via nanostructured lipid carriers for management of alopecia. Int J Nanomedicine 2014;9(1):5449–60.
- [75] Noor NM, Sheikh K, Somavarapu S, Taylor KMG . Preparation and characterization of dutasteride-loaded nanostructured lipid carriers coated with stearic acid-chitosan oligomer for topical delivery. Eur J Pharm Biopharm 2017;117:372–84.
- [76] Noor NM, Abdul-Aziz A, Sheikh K, Somavarapu S, Taylor KMG. In vitro performance of dutasteride-nanostructured lipid carriers coated with lauric acid-chitosan oligomer for dermal delivery. Pharmaceutics 2020;12(10):1–20.
- [77] Ghasemiyeh P, Azadi A, Daneshamouz S, Heidari R, Azarpira N, Mohammadi-Samani S . Cyproterone acetate-loaded nanostructured lipid carriers: effect of particle size on skin penetration and follicular targeting. Pharm Dev Technol 2019;24(7):812–23.
- [78] Hatem S, Nasr M, Moftah NH, Ragai MH, Geneidi AS, Elkheshen SA . Clinical cosmeceutical repurposing of melatonin in androgenic alopecia using nanostructured lipid carriers prepared with antioxidant oils. Expert Opin Drug Deliv 2018;15(10):927–35.
- [79] Yazdani-Arazi SN, Ghanbarzadeh S, Adibkia K, Kouhsoltani M, Hamishehkar H. Histological evaluation of follicular delivery of arginine via nanostructured lipid carriers: a novel potential approach for the treatment of alopecia. Artif Cells, Nanomedicine Biotechnol 2017;45(7):1379–87.
- [80] Caverzan J, de Jesus MB, Durán N. Nanostructured lipid carriers loaded with 17-α-estradiol accumulate into hair follicles. J Braz Chem Soc 2020;31(7):1345–51.
- [81] Ahmad Nasrollahi S, Koohestani F, Naeimifar A, Samadi A, Vatanara A, Firooz A . Preparation and evaluation of adapalene nanostructured lipid carriers for targeted drug delivery in acne. Dermatol Ther 2021;34(2):1–10.
- [82] Pereira MN, Tolentino S, Pires FQ, Anjos JLV, Alonso A, Gratieri T, et al. Nanostructured lipid carriers for hair follicle-targeted delivery of clindamycin and rifampicin to hidradenitis suppurativa treatment. Colloids Surfaces B Biointerfaces 2021;197:111448–58.
- [83] Ahmed OAA, Rizq WY . Finasteride nano-transferosomal gel formula for management of androgenetic alopecia: ex vivo investigational approach. Drug Des Devel Ther 2018;12:2259–65.
- [84] Ramezani V, Honarvar M, Seyedabadi M, Karimollah A, Ranjbar AM, Hashemi M . Formulation and optimization of transfersome containing minoxidil and caffeine. J Drug Deliv Sci Technol 2018;44:129–35.
- [85] Yu Z, Lv H, Han G, Ma K. Ethosomes loaded with cryptotanshinone for acne treatment through topical gel formulation. PLoS ONE 2016;11(7):1–11.
- [86] Fresta M, Mancuso A, Cristiano MC, Urbanek K, Cilurzo F, Cosco D, et al. Targeting of the pilosebaceous follicle by liquid crystal nanocarriers: in vitro and in vivo effects of the entrapped minoxidil. Pharmaceutics 2020;12(11):1–22.
- [87] Tahir MA, Ali ME, Lamprecht A. Nanoparticle formulations as recrystallization inhibitors in transdermal patches. Int J Pharm 2020;575:118886–95.
- [88] Główka E, Wosicka-Frąckowiak H, Hyla K, Stefanowska J, Jastrzębska K, Klapiszewski Ł, et al. Polymeric nanoparticles-embedded organogel for roxithromycin delivery to hair follicles. Eur J Pharm Biopharm 2014;88(1):75–84.
- [89] Goren A, Shapiro J, Roberts J, McCoy J, Desai N, Zarrab Z, et al. Clinical utility and validity of minoxidil response

testing in androgenetic alopecia. Dermatol Ther 2015;28(1):13–6.

- [90] Gupta AK, Talukder M, Venkataraman M, Bamimore MA. Minoxidil: a comprehensive review. J Dermatolog Treat 2021;32:1–11.
- [91] Souto EB, Baldim I, Oliveira WP, Rao R, Yadav N, Gama FM, et al. SLN and NLC for topical, dermal, and transdermal drug delivery. Expert Opin Drug Deliv 2020;17(3):357–77.
- [92] Adib ZM, Ghanbarzadeh S, Kouhsoltani M, Khosroshahi AY, Hamishehkar H. The effect of particle size on the deposition of solid lipid nanoparticles in different skin layers: a histological study. Adv Pharm Bull 2016;6(1):31–6.
- [93] Khorasani MT, Joorabloo A, Adeli H, Milan PB, Amoupour M . Enhanced antimicrobial and full-thickness wound healing efficiency of hydrogels loaded with heparinized ZnO nanoparticles: in vitro and in vivo evaluation. Int J Biol Macromol 2021;166:200–12.
- [94] Vazquez-Prada KX, Lam J, Kamato D, Xu ZP, Little PJ, Ta HT . Targeted molecular imaging of cardiovascular diseases by iron oxide nanoparticles. Arterioscler Thromb Vasc Biol 2021;41:601–13.
- [95] Baroli B, Ennas MG, Loffredo F, Isola M, Pinna R, López-Quintela MA . Penetration of metallic nanoparticles in human full-thickness skin. J Invest Dermatol 2007;127(7):1701–12.
- [96] Fuchs CSK, Ortner VK, Mogensen M, Philipsen PA, Haedersdal M. Transfollicular delivery of gold microparticles in healthy skin and acne vulgaris, assessed by in vivo reflectance confocal microscopy and optical coherence tomography. Lasers Surg Med 2019;51(5):430–8.
- [97] Friedman N, Dagan A, Elia J, Merims S, Benny O. Physical properties of gold nanoparticles affect skin penetration via hair follicles. Nanomedicine Nanotechnol, Biol Med 2021;36:102414–23.
- [98] Mahmoud NN, Alkilany AM, Dietrich D, Karst U, Al-Bakri AG, Khalil EA . Preferential accumulation of gold nanorods into human skin hair follicles: effect of nanoparticle surface chemistry. J Colloid Interface Sci 2017;503:95–102.
- [99] Seong KY, Seo MS, Hwang DY, O'Cearbhaill ED, Sreenan S, Karp JM, et al. A self-adherent, bullet-shaped microneedle patch for controlled transdermal delivery of insulin. J Control Release 2017;265:48–56.
- [100] Yang D, Chen M, Sun Y, Jin Y, Lu C, Pan X, et al. Microneedle-mediated transdermal drug delivery for treating diverse skin diseases. Acta Biomater 2021;121:119–33.
- [101] Zhou Z, Zhang S, Yang G, Gao Y. Enhanced delivery efficiency and sustained release of biopharmaceuticals by complexation-based gel encapsulated coated microneedles: rhIFNα-1b example. Asian J Pharm Sci 2021;16(5):612–22.
- [102] Kim S, Eum J, Yang H, Jung H. Transdermal finasteride delivery via powder-carrying microneedles with a diffusion enhancer to treat androgenetic alopecia. J Control Release 2019;316:1–11.
- [103] Hu C, Chu C, Liu L, Wang C, Jin S, Yang R, et al. Dissecting the microenvironment around biosynthetic scaffolds in murine skin wound healing. Sci Adv 2021;7(22):1–17.
- [104] Ocampo-Garza SS, Fabbrocini G, Ocampo-Candiani J, Cinelli E, Villani A . Micro needling: a novel therapeutic approach for androgenetic alopecia, a review of literature. Dermatol Ther 2020;33(6):1–7.
- [105] Yuan A, Xia F, Bian Q, Wu H, Gu Y, Wang T, et al. Ceria nanozyme-integrated microneedles reshape the perifollicular microenvironment for androgenetic alopecia treatment. ACS Nano 2021;15(8):13759–69.
- [106] Fang JH, Liu CH, Hsu RS, Chen YY, Chiang WH, Wang HMD, et al. Transdermal composite microneedle composed of mesoporous iron oxide nanoraspberry and PVA for

androgenetic alopecia treatment. Ploymers 2020;12(6):1392–405.

- [107] Cao S, Wang Y, Wang M, Yang X, Tang Y, Pang M, et al. Microneedles mediated bioinspired lipid nanocarriers for targeted treatment of alopecia. J Control Release 2021;329:1–15.
- [108] Fakhraei Lahiji S, Seo SH, Kim S, Dangol M, Shim J, Li CG, et al. Transcutaneous implantation of valproic acid-encapsulated dissolving microneedles induces hair regrowth. Biomaterials 2018;167:69–79.
- [109] Zhang Y, Feng P, Yu J, Yang J, Zhao J, Wang J, et al. ROS-responsive microneedle patch for acne vulgaris treatment. Adv Ther 2018;1(3):1–6.
- [110] Lademann J, Richter H, Knorr F, Patzelt A, Darvin ME, Rühl E, et al. Triggered release of model drug from AuNP-doped BSA nanocarriers in hair follicles using IRA radiation. Acta Biomater 2016;30:388–96.
- [111] Caserta F, Brown MB, McAuley WJ. The use of heat and chemical penetration enhancers to increase the follicular delivery of erythromycin to the skin. Eur J Pharm Sci 2019;132:55–62.
- [112] Jung S, Nagel G, Giulbudagian M, Calderón M, Patzelt A, Knorr F, et al. Temperature-enhanced follicular penetration of thermoresponsive nanogels. Zeitschrift Fur Phys Chemie 2018;232(5–6):805–17.
- [113] Dimde M, Sahle FF, Wycisk V, Steinhilber D, Camacho LC, Licha K, et al. Synthesis and validation of functional nanogels as pH-sensors in the hair follicle. Macromol Biosci 2017;17(10):1–8.

- [114] Liao AH, Lu YJ, Lin YC, Chen HK, Sytwu HK, Wang CH. Effectiveness of a layer-by-layer microbubbles-based delivery system for applying minoxidil to enhance hair growth. Theranostics 2016;6(6):817–27.
- [115] Liao A, Huang Y, Chuang H, Wang C, Shih C. Minoxidil-coated lysozyme-shelled microbubbes combined with ultrasound for the enhancement of hair follicle growth : efficacy in vitro and in vivo. Front Pharmacol 2021;12:1–13.
- [116] Liao A, Hung C, Lin C, Lin Y, Chen H. Treatment effects of lysozyme- shelled microbubbles and ultrasound in inflammatory skin disease. Sci Rep 2017:41325–33.
- [117] Busch L, Avlasevich Y, Zwicker P, Thiede G, Landfester K, Keck CM, et al. Release of the model drug SR101 from polyurethane nanocapsules in porcine hair follicles triggered by LED-derived low dose UVA light. Int J Pharm 2021;597:120339–50.
- [118] Hatem S, Nasr M, Moftah NH, Ragai MH, Geneidi AS, Elkheshen SA . Melatonin vitamin C-based nanovesicles for treatment of androgenic alopecia: design, characterization and clinical appraisal. Eur J Pharm Sci 2018;122:246–53.
- [119] Paithankar DY, Sakamoto FH, Farinelli WA, Kositratna G, Blomgren RD, Meyer TJ, et al. Acne treatment based on selective photothermolysis of sebaceous follicles with topically delivered light-absorbing gold microparticles. J Invest Dermatol 2015;135(7):1727–34.