

Taxonogenomics description of *Bacillus dakarensis* sp. nov., *Bacillus sinesaloumensis* sp. nov. and *Bacillus massiliogabonensis* sp. nov., three new species isolated from human stools

M. Sarr^{1,4}, C. I. Lo^{2,3}, M. L. Tall^{1,2}, A. Fadlane^{1,2}, B. Senghor^{1,4}, C. Sokhna^{1,4}, D. Raoult^{1,2}, M. Million^{1,2} and F. Fenollar^{2,3}

1) Aix Marseille Université, IRD, AP-HM, MEΦI, 2) IHU-Méditerranée Infection, 3) Aix Marseille Université, IRD, AP-HM, SSA, VITROME, Marseille, France and 4) Campus Commun UCAD-IRD of Hann, Dakar, Senegal

Abstract

Using microbial culturomics, three *Bacillus* strains were isolated, identified and characterized following the taxonogenomics strategy. *Bacillus dakarensis* strain Marseille-P3515^T (=CSURP3515), *Bacillus sinesaloumensis* strain Marseille-P3516^T (=CSURP3516), and *Bacillus massiliogabonensis* strain Marseille-P2639^T (=CSURP2639) were isolated from human stool samples. The phylogenetic analysis, phenotypic characteristics and genotypic data presented here prove that these three bacteria are different from previously known bacterial species with standing in nomenclature and represent new *Bacillus* species.

© 2020 Published by Elsevier Ltd.

Keywords: Africa, *Bacillus* sp., culturomics, human stool, taxonogenomics

Original Submission: 10 January 2020; **Revised Submission:** 12 May 2020; **Accepted:** 17 June 2020

Article published online: 31 July 2020

Corresponding author: F. Fenollar, Institut Hospitalo-Universitaire Méditerranée-Infection, 19–21 Boulevard Jean Moulin, 13385, Marseille cedex 05, France.

E-mail: florence.fenollar@univ-amu.fr

Introduction

Genus *Bacillus* was created in 1872 by Ferdinand Julius Cohn [1]. To date, there are 379 species and seven subspecies with validly published names. Most *Bacillus* species are environmental bacteria found in food, soil, freshwater and the sea. Other *Bacillus* species may be saprophytic [2] or endophytic on plants [3]. Two species are important in public health, namely *Bacillus cereus* (associated with food poisoning) and *Bacillus anthracis* (responsible for anthrax) [4,5].

Bacteria involved in normal physiological functions and with a predisposition to human diseases should be studied for better understanding [6]. The culturomics method that isolates bacteria under different culture conditions is complemented in our laboratory by the systematic sequencing of the 16S rRNA gene,

which allows us to explore the microbial diversity of the human gut [7–9]. The new species, which we report here, were described using a combination of genotypic and phenotypic characteristics, following a previously described taxonogenomics strategy [10,11].

We present the details of the isolation and taxonogenomics characterization of strain Marseille-P3515^T, strain Marseille-P3516^T and strain Marseille-P2639^T as type strains of *Bacillus dakarensis* sp. nov., *Bacillus sinesaloumensis* sp. nov. and *Bacillus massiliogabonensis* sp. nov., respectively.

Isolation and growth conditions

In 2016, three stool samples were collected with the aim of studying halophilic bacteria involved in human gut functioning [12]. Marseille-P3515^T and Marseille-P3516^T were isolated from the stool samples of a 17-year-old boy and a 10-year-old girl, respectively, both living in Ndiop, a rural area in Senegal. Strain Marseille-P2639^T was isolated from the stool sample of a 16-year-old boy living in Gabon. After numerous attempts at identification by matrix-assisted laser desorption/ionization time-of-flight (MALDI-TOF) mass spectrometry, no reliable

recognition was obtained for the three strains. The screening was performed on a Microflex LT spectrometer (Bruker Daltonics, Bremen, Germany) as previously described [13]. The spectra obtained (Fig. 1) were imported into MALDI Biotyper 3.0 software and analysed against the Bruker database, which is permanently improved with the MEPHI database (<https://www.mediterranee-infection.com/urms-data-base>). The strains (Marseille-P3515^T, Marseille-P3516^T and Marseille-P2639^T) were first isolated after 1–2 days of pre-incubation of stool samples in aerobic conditions in blood-culture bottles enriched with 5% rumen fluid sterilized by filtration at 0.2 µm and seeded on 5% sheep-blood Columbia agar (bioMérieux, Marcy l'Étoile, France) under aerobic conditions at 37°C.

Strain identification

After missing identification using MALDI-TOF, the 16S rRNA genes for each strain were sequenced to classify these bacteria.

These genes were amplified using universal primer pairs fD1 and rP2 (Eurogentec, Angers, France) and sequenced with the Big Dye® Terminator v1.1 Cycle Sequencing Kit and 3500xL Genetic Analyzer capillary sequencer (ThermoFisher, Saint-Aubin, France), as previously described [14]. CODONCODE ALIGNER software was used for assembly and to correct the 16S rRNA nucleotide sequences (<http://www.codoncode.com>). The sequences of each strain were submitted for BLAST in the NCBI database to determine the phylogenetically closest species with standing in nomenclature. It is in this context that we found that strain Marseille-P3515^T exhibited a 97.96% sequence identity with *Bacillus circulans* strain NBRC 13626 (GenBank Accession no.: AY724690), strain Marseille-P3516^T showed a 98.51% sequence identity with *Bacillus humi* strain LMG 22167 (AJ627210), and strain Marseille-P2639^T displayed 98.41% sequence identity with *Bacillus ciccensis* strain 105-2 (KP965576). These values are below the threshold value recommended (<98.7% sequence similarity of the 16S rRNA gene) by authors to delineate new bacterial species within a genus without performing DNA–DNA hybridization [15,16]. Based

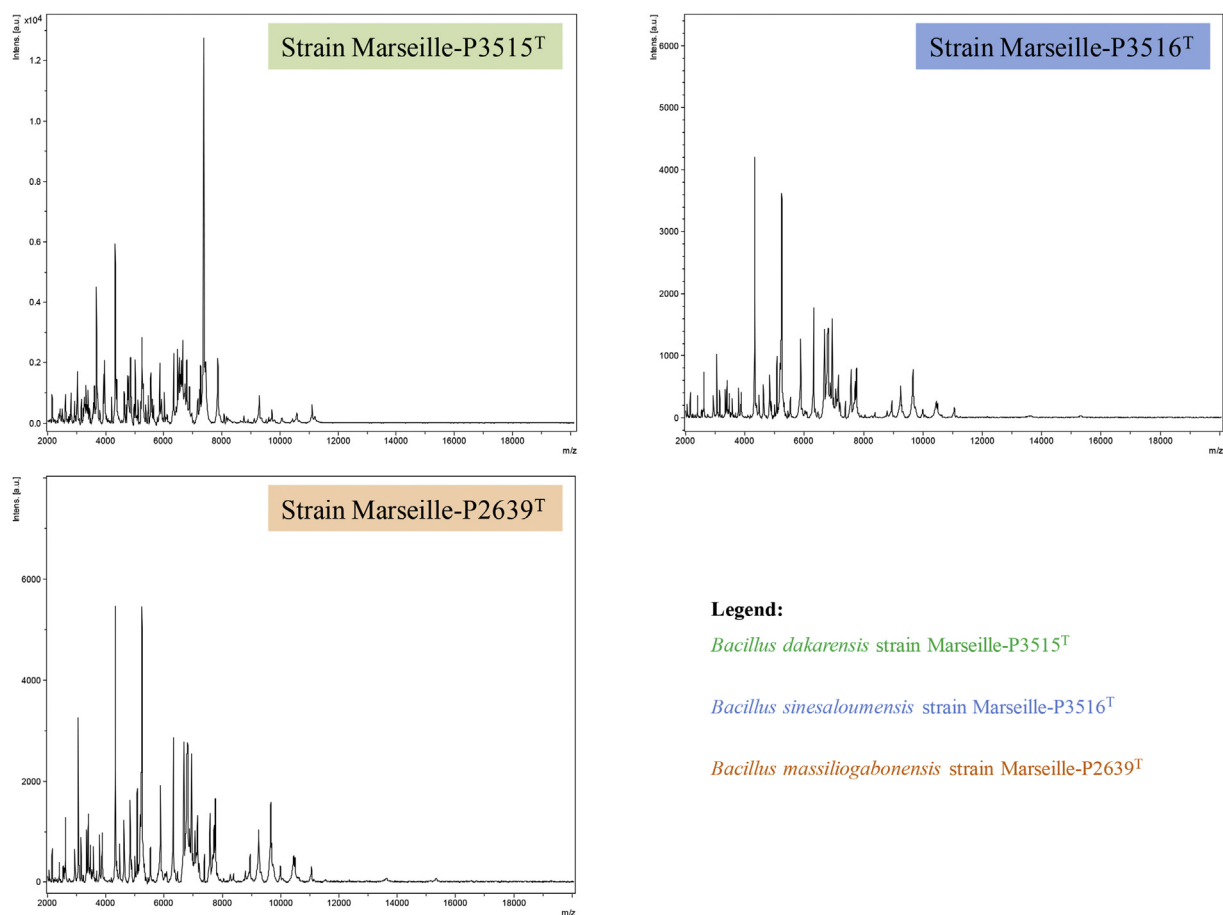


FIG. 1. MALDI-TOF MS reference spectrum of the three new species described above. The reference spectra were generated by comparison of spectra from 12 individual colonies for each species.

on this observation, we declare that these strains are new members of the genus *Bacillus* belonging to the family *Bacillaceae* within the phylum *Firmicutes* (Fig. 2).

Phenotypic characteristics

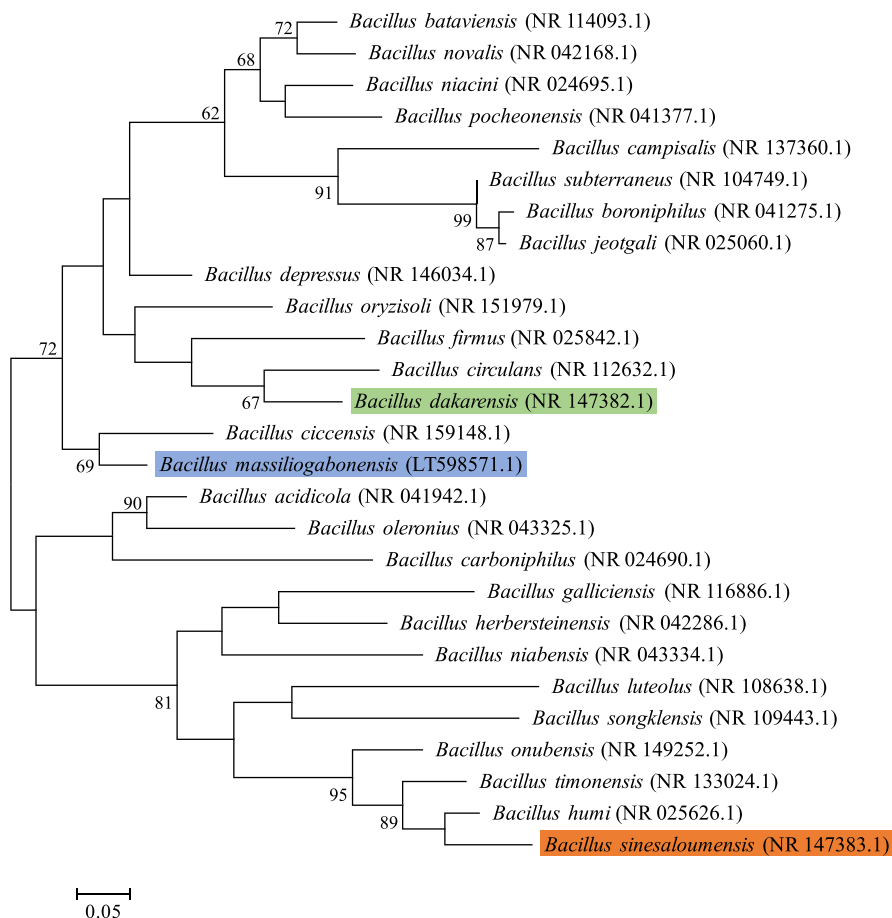
The strains from Senegal were easily grown in an aerobic atmosphere. Apparent colonies were obtained after 24 h of incubation at 37°C on 5% sheep's blood–Columbia agar medium (bioMérieux). Strains Marseille-P3515^T and Marseille-P3516^T were recovered from human stool sample from a Senegalese village named Ndiop. Their colonies appear rounded, beige and shiny with a mean diameter of 1.2 mm. Cells were Gram-positive bacteria, rod-shaped and catalase positive. In addition, the oxidase reaction test was positive for these two strains, which were mobile and spore forming. These strains were cultured on halophilic media with NaCl concentrations of 50, 75, 100 and 150 g/L of NaCl. In parallel, the growth of bacteria was tested on media at different pH (pH 6, 6.5, 7, 7.5 and 8).

Tests have shown that the two strains grow better in 48 h at pH 7.5, at 75 g/L of NaCl and 37°C. These data indicate that strains Marseille-P3515^T and Marseille-P3516^T are halophilic bacteria.

Strain Marseille-P2639^T was endospore forming and motile. It was a Gram-negative bacterium that exhibited catalase activity. Bacterial cells did not have an oxidase reaction. They measured 3.7 µm in length and 0.8 µm in diameter. Strain Marseille-P2639^T was an aerobic bacterium that grew between 23°C and 45°C in <1 day of incubation. Colonies of strain Marseille-P2639^T were white with a mean diameter of 3 mm on 5% sheep's blood-enriched Columbia agar. Strain Marseille-P2639^T is a bacterium that weakly tolerates salt concentrations >50 g/L of NaCl but is able to grow on media with a pH ranging from 6 to 10. The optimal growth temperature is 37°C under aerobic conditions.

The shape of these bacteria was highlighted with the Tecnai G20 transmission electron microscope (FEI Company) (Fig. 3). The biochemical characteristics of these strains were tested using the API ZYM and API 50 CH strips (bioMérieux) and are

FIG. 2. Phylogenetic tree highlighting the position of three new bacterial species relative to their most closely related and validly published type strains. GenBank accession numbers of 16S rRNA are indicated in parentheses. Sequences were aligned using MUSCLE with default parameters, phylogenetic inference were obtained using the maximum likelihood method and the MEGA 7 software. Numbers at the nodes are percentages of bootstrap values obtained by repeating the analysis 1000 times to generate a majority consensus tree. The scale bar indicates a 5% nucleotide sequence divergence.



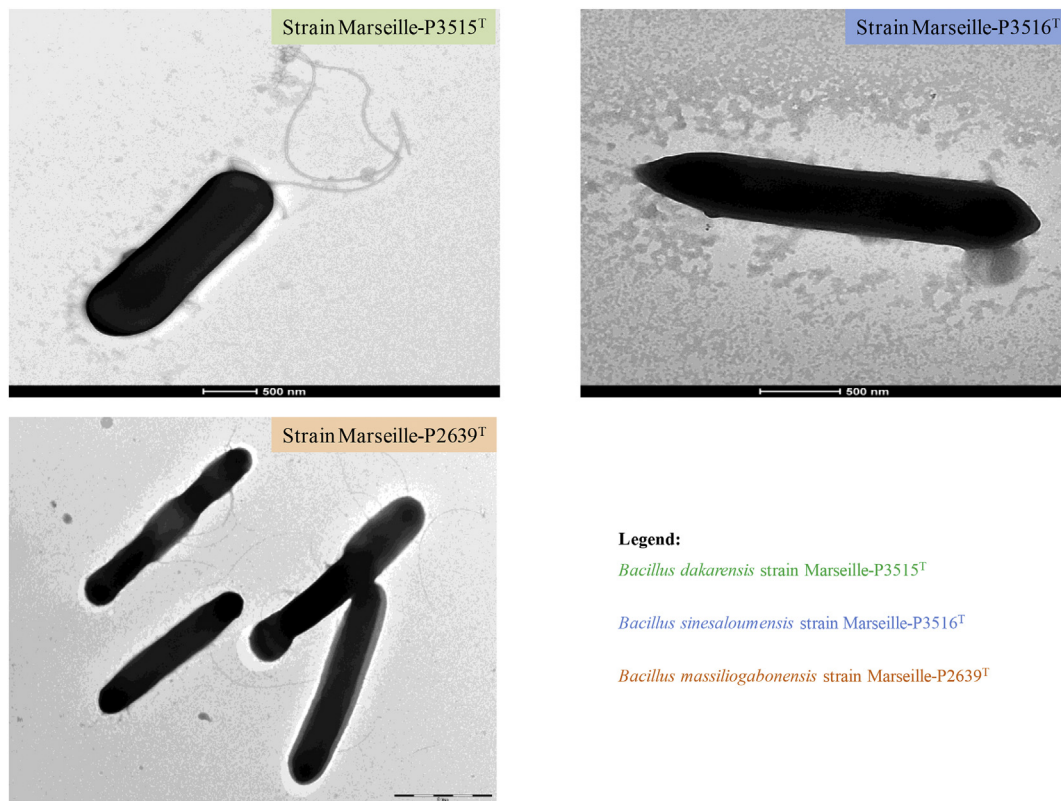
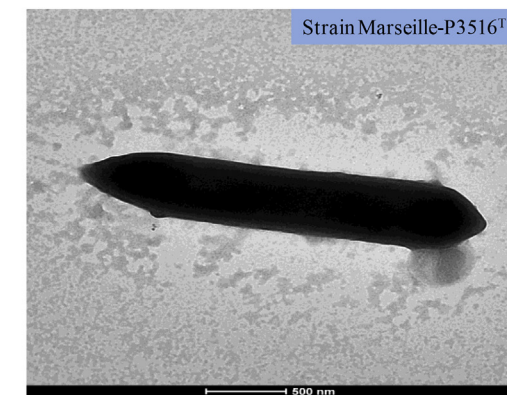


FIG. 3. Transmission electron microscopy of three new *Bacillus* species. Cells are observed on Tecnai G20 transmission electron microscope operated at 200 keV. Scales are displayed on the figures.

presented in Table 1. A comparative study of the differential characteristics of these strains with other closely related species is displayed in Table 2.

Genome sequencing

Genomic DNA was extracted using the EZ1 biorobot (Qiagen, Courtaboeuf, France) with the EZ1 DNA tissue Kit and then sequenced using MiSeq technology (Illumina, San Diego, CA, USA) with the Nextera Mate Pair sample prep kit and Nextera XT Paired end (Illumina), as previously described [17]. The assembly was performed with a pipeline incorporating different softwares (VELVET [18], SPADES [19] and SOAP DENOVO [20]), and trimmed data (MISEQ and TRIMMOMATIC [21] softwares) or untrimmed data (only MISEQ software). GAPCLOSER was used to reduce assembly gaps. Scaffolds <800 bp and scaffolds with a depth value < 25% of the mean depth were removed. The best assembly was selected using different criteria (number of scaffolds, N50, number of N). The degree of genomic similarity of these three strains with closely related species was estimated using ORTHOANI software [22]. ORTHOANI values among



Legend:

Bacillus dakarensis strain Marseille-P3515^T

Bacillus sinesaloumensis strain Marseille-P3516^T

Bacillus massiliogabonensis strain Marseille-P2639^T

Bacillus species (Fig. 4) ranged from 68.03% between *Bacillus acidicola* and *B. humi* to 88.91% between *B. ciccensis* and *B. massiliogabonensis*. These values were <95%, the threshold value suggested for delineating new bacterial species in an ORTHOANI comparison [23].

Genome properties

The genome of strain Marseille-P3515^T is 5 482 351 bp long with 38.6 mol% G + C content (Table 3). It is composed of 11 scaffolds (composed of 62 contigs). Of the 5282 predicted genes, 5040 are protein-coding genes and 242 are RNAs (15 genes are 5S rRNA, 7 genes are 16S rRNA, 9 genes are 23S rRNA, 211 genes are tRNA genes). A total of 3600 genes (71.43%) were assigned as putative function (by cogs or by NR blast); 206 genes were identified as ORFans (4.09%). The remaining genes were annotated as hypothetical proteins (1014 genes, ≥20.12%).

Strain Marseille-P3516^T has a genome size of 4 556 426 bp long with 37.9 mol% G + C content (Table 3). It is composed of three scaffolds with 26 contigs. Of the 4466 predicted genes, 4323 are protein-coding genes and 143 are RNAs. These RNA

TABLE 1. Phenotypic characterization of *Bacillus dakarensis* sp. nov., strain Marseille-P3515^T, *Bacillus sinesaloumensis* sp. nov., strain Marseille-P3516^T and *Bacillus massiliogabonensis* sp. nov., strain Marseille-P2639^T sp. nov., based on analytical profile index (API) tests

Tests	Number	Characteristics	Marseille-P3515 ^T	Marseille-P3516 ^T	Marseille-P2639 ^T
API ZYM	2	Alkaline phosphatase	-	-	+
	3	Esterase (C4)	+	+	+
	4	Esterase lipase (C8)	-	-	-
	5	Lipase (C14)	-	-	-
	6	Leucine arylamidase	-	-	-
	7	Valine arylamidase	-	-	-
	8	Cystine arylamidase	-	-	-
	9	Trypsin	-	-	-
	10	α-chymotrypsin	-	-	+
	11	Acid phosphatase	+	-	+
	12	Naphthol-AS-BI-phosphohydrolase	-	-	-
	13	α-galactosidase	-	+	-
	14	β-galactosidase	-	+	-
	15	β-glucuronidase	-	+	-
	16	α-glucosidase	-	+	-
	17	β-glucosidase	-	+	-
	18	N-acetyl-β-glucosaminidase	-	+	-
	19	α-mannosidase	-	-	-
	20	α-fucosidase	-	-	-
	API 50CH	1	Glycerol	-	-
2		Erythritol	-	-	-
3		D-arabinose	+	-	-
4		L-arabinose	+	-	-
5		D-ribose	-	-	-
6		D-xylose	+	-	-
7		L-xylose	-	-	-
8		D-Adonitol	+	-	-
9		Methyl βD-xylopyranoside	-	-	-
10		D-galactose	-	-	+
11		D-glucose	-	-	-
12		D-fructose	-	-	-
13		D-mannose	-	-	-
14		L-sorbose	-	-	-
15		L-rhamnose	+	-	-
16		Dulcitol	-	-	-
17		Inositol	+	-	-
18		D-mannitol	-	-	-
19		D-sorbitol	-	-	-
20		Methyl αD-mannopyranoside	-	-	-
21		Methyl αD-glucopyranoside	-	-	+
22		N-acetyl-glucosamine	-	-	+
23		Amygdalin	-	-	+
24		Arbutin	-	-	+
25		Esculin ferric citrate	+	+	+
26		Salicin	-	-	+
27		D-cellobiose	-	-	-
28		D-maltose	-	-	-
29		D-lactose	-	-	-
30	D-melibiose	+	-	-	
31	Sucrose	-	-	-	
32	D-trehalose	-	-	+	
33	Inulin	-	-	+	
34	D-melezitose	-	-	+	
35	D-raffinose	-	-	+	
36	Starch	-	-	+	
37	Glycogen	-	-	+	
38	Xylitol	+	-	-	
39	Gentiobiose	-	-	-	
40	D-turanose	-	-	-	
41	D-lyxose	+	-	-	
42	D-tagalose	-	-	-	
43	D-fucose	+	-	-	
44	L-fucose	-	-	-	
45	D-arabitol	-	-	-	
46	L-arabitol	-	-	+	
47	Potassium gluconate	-	-	-	
48	Potassium 2-ketogluconate	-	-	-	
49	Potassium 5-ketogluconate	-	-	-	

genes are divided into 12 5S rRNA genes, 13 16S rRNA genes, 13 23S rRNA genes and 105 tRNA genes. Strain Marseille-P3516^T possesses 3095 genes (71.59%) as putative function (by cogs or by NR blast) and 130 genes as ORFans (3.01%); 868 genes (20.08%) were annotated as hypothetical proteins.

The genome of strain Marseille-P2639^T is 5 224 786 bp in length with 37.9 mol% G + C content and is composed of nine scaffolds with 69 contigs (Table 3). Overall, in 5234 predicted genes, 5045 genes encode proteins and 189 genes are RNAs (12 genes are 5S rRNA, 18 genes are 16S rRNA, 16 genes are

TABLE 2. Differential characteristics of *Bacillus dakarensis* strain Marseille-P3515^T, *Bacillus sinesaloumensis* strain Marseille-P3516^T and *Bacillus massiliogabonensis* strain Marseille-P2639^T compared with *Bacillus ndiopicus* strain FF3^T and *Bacillus dielmoensis* strain FF4^T

Property	P3515 ^T	P3516 ^T	P2639 ^T	FF3 ^T	FF4 ^T
Cell diameter (µm)	0.5–1	1–1.9	0.7–1	0.8–1.6	0.5–0.8
Oxygen requirement	Aerobic	Aerobic	Aerobic	Aerobic	Aerobic
Gram stain	+	+	–	+	+
Motility	+	+	–	+	+
Endospore formation	+	+	+	+	–
Production of:					
Alkaline phosphatase	–	–	+	+	+
Acid phosphatase	+	–	+	–	+
Catalase	+	+	+	+	+
Oxidase	+	+	–	–	–
β-Galactosidase	–	+	–	–	+
α-Glucosidase	–	+	–	–	+
Esterase	+	+	+	+	+
Esterase lipase	–	–	–	+	+
Naphthol-AS-BI-phosphohydrolase	–	–	–	–	+
N-acetyl-β-glucosaminidase	–	+	–	–	–
Utilization of:					
Potassium 5-ketogluconate	–	–	–	–	–
D-Xylose	+	–	–	–	–
D-Fructose	–	–	–	–	–
D-Glucose	–	–	–	–	–
D-Mannose	–	–	–	–	–
Habitat	Human sample	Human sample	Stool sample	Human skin	Human skin

Note: +, positive result; –, negative result; NA, data not available.

23S rRNA, 143 genes are tRNA genes). A total of 3476 genes (68.9%) were assigned as putative function (by the cogs or by NR blast), 179 genes were recognized as ORFans (3.55%) and the remaining 1151 genes (22.81%) were annotated as hypothetical proteins.

The digital DNA–DNA hybridization between the genomes of three new *Bacillus* species and other available genomes of the phylogenetically closest species, was calculated using the Genome-to-Genome Distance Calculator online calculator with formula 2 (Table 4). A 70% threshold is fixed by Meier-

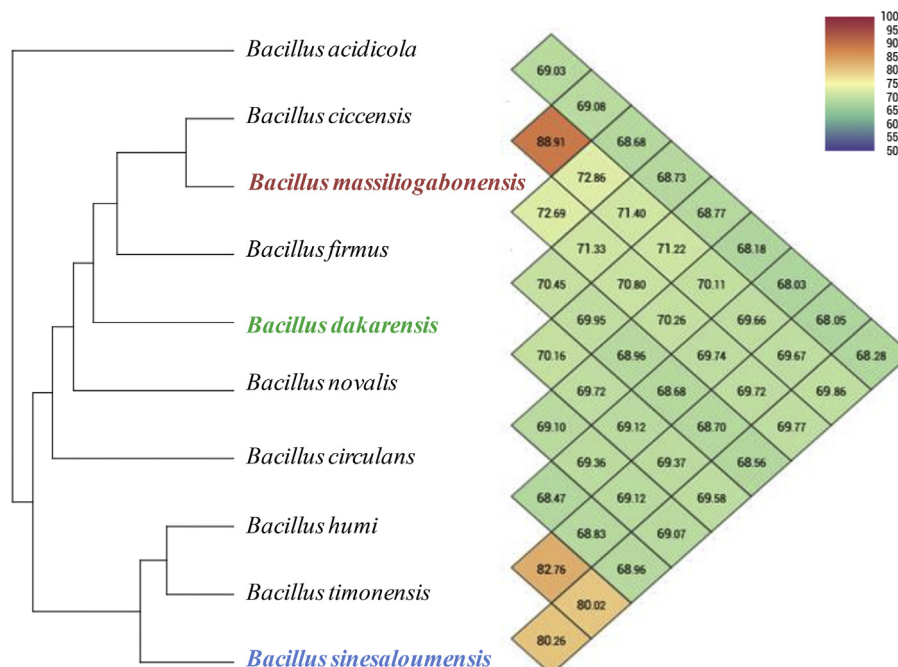
**FIG. 4.** Heatmap generated with ORTHOANI values calculated using the OAT software for *Bacillus dakarensis* strain Marseille-P3515^T, *Bacillus sinesaloumensis* strain Marseille-P3516^T and *Bacillus massiliogabonensis* strain Marseille-P2639^T among other related *Bacillus* species with standing in nomenclature.

TABLE 3. Nucleotide content and gene count levels of genomes of *Bacillus dakarensis* strain Marseille-P3515^T, *Bacillus sinesaloumensis* strain Marseille-P3516^T and *Bacillus massiliogabonensis* strain Marseille-P2639^T

Attribute	<i>B. dakarensis</i>		<i>B. sinesaloumensis</i>		<i>B. massiliogabonensis</i>	
	Value	% Of total ^a	Value	% Of total ^a	Value	% Of total ^a
Size (bp)	5 482 351	100	4 556 426	100	5 224 786	100
G + C content (bp)	2 057 335	38.6	1 711 281	37.9	1 944 649	37.9
Total of genes	5282	100	4466	100	5234	100
RNA genes	242	4.5	143	3.2	189	3.6
Coding sequence size (bp)	4 474 355	81.6	3 886 129	85.3	4 337 909	83.0
Protein-coding genes	504	100	4323	100	5045	100
Protein assigned to COGs	3246	64.4	2741	63.4	3063	60.7
Genes with peptide signals	595	11.8	478	11.0	561	11.1
Genes with transmembrane helices	1189	23.6	1183	27.3	1278	25.3
Genes associated with mobilome	2147	42.6	1714	39.6	2068	40.9
Genes associated with virulence	951	18.8	790	18.2	948	18.7

^aTotal is based on total number of protein-coding genes in annotated genome.

Kolthoff et al., to differentiate two distinct species [24]. DNA–DNA hybridization values ranged from 21.5% between *B. ciccensis* and *Bacillus firmus* to 34.9% between *Bacillus sinesaloumensis* and *Bacillus acidicola*, which supported the previous data indicating the classification of these as a new bacterial species (Table 4). Likewise, repartition of genes into the 25 general COG categories is illustrated in Table 5 and Fig. 5. The average percentage nucleotide identity calculated using the Average Genomic Identity of Orthologous Gene Sequences (AGIOS) in-house software [10] and number of orthologous genes of *B. dakarensis*, *B. sinesaloumensis* and *B. massiliogabonensis* shared with others species are displayed in Table 6.

Conclusion

Based on unique phenotypic characteristics, including API test strips, MALDI-TOF spectra, and phylogenetic and genomic analyses such as 16S rRNA sequence similarity <98.7% and ORTHOANI values < 95% with the phylogenetically closest

species with standing in nomenclature, we propose strains Marseille-P3515^T, Marseille-P3516^T and Marseille-P2639^T, respectively, as being the type strains of *Bacillus dakarensis* sp. nov., *Bacillus sinesaloumensis* sp. nov. and *Bacillus massiliogabonensis* sp. nov., which are new species in the genus *Bacillus*.

Description of *Bacillus dakarensis* sp. nov.

Bacillus dakarensis (da.ka.ren'sis, N.L. masc. adj. *dakarensis* of Dakar, the name of the capital of Senegal where the stool sample was collected). The colonies of the strain appear beige and circular on blood agar with a mean diameter of 1.2 mm. The cells are mobile and spore-forming. They are Gram-positive bacilli and present positive oxidase and positive catalase activities. The draft genome size of strain Marseille-P3515^T is about 5.33 Mb with a 38.6 mol% of G + C content. The 16S rRNA gene sequence and whole-genome shotgun sequence of *B. dakarensis* strain Marseille-P3515^T were deposited in GenBank under accession numbers LT671589 and FTOZ00000000, respectively. The type strain Marseille-P3515^T (=CSURP3515) was isolated from the stool sample of 17-year-old-boy living in Senegal.

TABLE 4. Genome comparison between three new bacterial strains and closely related species using GGDC and formula 2 (dDDH estimates based on identities over HSP length), upper right. The inherent uncertainty in assigning dDDH values from intergenomic distances is presented in the form of confidence intervals

	Bdak	Bmas	Bsin	Baci	Bcic	Bcir	Bfir	Bnov	Btim
Bdak	100%	24.3% ± 4.8	27.5% ± 4.9	28.4% ± 4.9	24.5% ± 4.8	28.5% ± 4.9	22.7% ± 4.7	24.3% ± 4.8	23.2% ± 4.8
Bmas		100%	31.5% ± 4.9	32.1% ± 4.9	37.6% ± 5	30.3% ± 4.9	22.0% ± 4.7	26.3% ± 4.8	25.9% ± 4.8
Bsin			100%	34.9% ± 4.9	27.8% ± 4.8	27.1% ± 4.8	26.4% ± 4.9	32.1% ± 5	23.8% ± 4.7
Baci				100%	32.7% ± 4.9	33.9% ± 4.9	27.3% ± 4.9	29.3% ± 4.9	29.3% ± 4.9
Bcic					100%	28.9% ± 4.8	21.5% ± 4.7	25.6% ± 4.8	25.0% ± 4.8
Bcir						100%	23.4% ± 4.8	32.6% ± 4.9	25.0% ± 4.8
Bfir							100%	22.2% ± 4.7	22.6% ± 4.7
Bnov								100%	26.2% ± 4.8
Btim									100%

dDDH, digital DNA–DNA hybridization; HSP, high scoring pair; GGDC, genome-to-genome distance calculator.

Bacillus dakarensis strain Marseille-P3515^T (Bdak), *Bacillus massiliogabonensis* strain Marseille-P2639^T (Bmas), *Bacillus sinesaloumensis* strain Marseille-P3516^T (Bsin), *Bacillus acidicola* strain DSM 14745^T (Baci), *Bacillus ciccensis* strain KCTC 33663^T (Bcic), *Bacillus circulans* strain NBRC 13626 (Bcir), *Bacillus firmus* strain NBRC 15306 (Bfir), *Bacillus novalis* strain NBRC 102450 (Bnov) and *Bacillus timonensis* strain Marseille-P162 (Btim).

TABLE 5. Number of genes associated with 25 general COG functional categories

Code	<i>Bacillus sinesaloumensis</i>		<i>Bacillus dakarensis</i>		<i>Bacillus massiliogabonensis</i>		Description
	Value	% Of total	Value	% Of total	Value	% Of total	
[J]	236	5.45	237	4.70	250	4.95	Translation
[A]	0	0	0	0	0	0	RNA processing and modification
[K]	219	5.06	217	4.30	255	5.05	Transcription
[L]	104	2.40	111	2.20	126	2.49	Replication, recombination and repair
[B]	1	0.02	1	0.01	1	0.01	Chromatin structure and dynamics
[D]	52	1.20	56	1.11	58	1.14	Cell cycle control, mitosis and meiosis
[Y]	0	0	0	0	0	0	Nuclear structure
[V]	79	1.82	58	1.15	98	1.94	Defence mechanisms
[T]	151	3.49	197	3.90	198	3.92	Signal transduction mechanisms
[M]	122	2.82	132	2.61	139	2.75	Cell wall/membrane biogenesis
[N]	53	1.22	69	1.36	72	1.42	Cell motility
[Z]	0	0	0	0	1	0.01	Cytoskeleton
[W]	8	0.18	9	0.17	8	0.15	Extracellular structures
[U]	29	0.67	40	0.79	34	0.67	Intracellular trafficking and secretion
[O]	113	2.61	128	2.53	149	2.95	Post-translational modification, protein turnover, chaperones
[X]	26	0.60	32	0.63	66	1.30	Mobilome: prophages, transposons
[C]	167	3.86	254	5.03	208	4.12	Energy production and conversion
[G]	294	6.80	245	4.86	201	3.98	Carbohydrate transport and metabolism
[E]	291	6.73	368	7.30	329	6.52	Amino acid transport and metabolism
[F]	100	2.31	94	1.86	111	2.20	Nucleotide transport and metabolism
[H]	156	3.60	187	3.71	178	3.52	Coenzyme transport and metabolism
[I]	158	3.65	345	6.84	160	3.17	Lipid transport and metabolism
[P]	172	3.97	237	4.70	227	4.49	Inorganic ion transport and metabolism
[Q]	81	1.87	164	3.25	98	1.94	Secondary metabolites biosynthesis, transport and catabolism
[R]	278	6.43	363	7.20	300	5.94	General function prediction only
[S]	230	5.32	220	4.36	239	4.73	Function unknown
-	1582	36.59	1794	35.59	1982	39.28	Not in COGs

COG, cluster of orthologous groups.

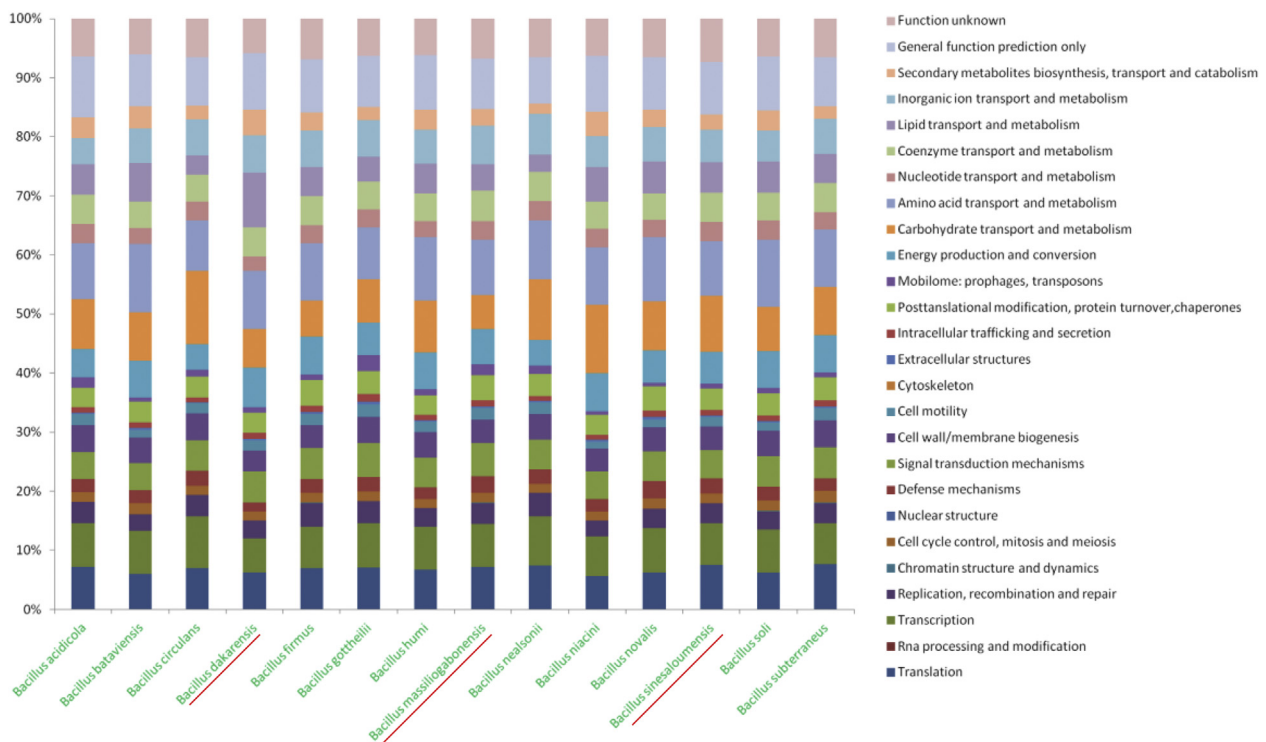
**FIG. 5.** Distribution of functional classes of predicted genes according to the COG of proteins of *Bacillus dakarensis* strain Marseille-P3515^T, *Bacillus sinesaloumensis* strain Marseille-P3516^T and *Bacillus massiliogabonensis* strain Marseille-P2639^T among other related *Bacillus* species.

TABLE 6. Numbers of orthologous proteins shared between genomes (**bold**)^a

Genomes	<i>Bacillus circulans</i>	NCIMB8773	Marseille-P3515	105-2	WCC4585	IAM12464
<i>Bacillus circulans</i>	4950	1829	1857	1842	2031	2037
<i>Bacillus lentus</i> strain NCIMB8773	60.91	4088	1754	1698	1830	1808
<i>Bacillus dakarensis</i> strain Marseille-P3515	61.84	67.91	5040	1816	2099	2244
<i>Bacillus acidicola</i> strain 105-2	60.54	67.96	67.56	4876	1839	1976
<i>Bacillus gottheilii</i> strain WCC4585	61.69	67.52	70.24	67.66	4450	2285
<i>Bacillus firmus</i> strain IAM12464	59.95	58.64	60.36	59.11	61.28	4922
	PBINCIMB	LMG21833	AM31D	LMG22167	Marseille-P3516	IFO15566
<i>Bacillus methanolicus</i> strain PBINCIMB	3410	1704	1524	1627	1628	1715
<i>Bacillus bataviensis</i> strain LMG21833	61.99	5207	1754	2168	2086	2449
<i>Bacillus krulwichiae</i> strain AM31D	53.99	55.98	4596	1774	1793	1880
<i>Bacillus humi</i> strain LMG22167	55.03	58.02	66.45	4842	2316	2198
<i>Bacillus sinesaloumensis</i> strain Marseille-P3516	55.00	57.56	66.84	81.22	4323	2161
<i>Bacillus niacini</i> strain IFO15566	55.76	60.61	66.02	68.48	68.34	5952
	Marseille-P2639	LMG21831	LMG21837	LMG21838	A1-2	DSM13966
<i>Bacillus massiliogabonensis</i> strain Marseille-P2639	5045	2208	2364	2338	2369	1904
<i>Bacillus drenensis</i> strain LMG21831	70.62	5043	2707	2796	2107	1902
<i>Bacillus novalis</i> strain LMG21837	70.47	77.50	5425	3014	2246	1958
<i>Bacillus soli</i> strain LMG21838	70.39	79.26	80.82	5340	2244	1944
<i>Bacillus eiseniae</i> strain A1-2	72.08	69.53	69.15	69.22	5468	1870
<i>Bacillus subterraneus</i> strain DSM13966	60.44	60.27	60.17	60.27	60.07	3465

^aAverage percentage similarity of nucleotides corresponding to orthologous protein shared between genomes and numbers of proteins per genome (**bold**).

Description of *Bacillus sinesaloumensis* sp. nov.

Bacillus sinesaloumensis (si.ne.sa.lou.men'sis, N.L. masc. adj. *sinesaloumensis* of Sine-Saloum, a former administrative region of Senegal where the village of Ndiop is located, from which this strain was sampled). Colonies grow on 5% sheep blood Colombia agar plate after 24 h of incubation under aerobic conditions. They are shiny, beige and 2 mm in diameter. The cells are Gram-positive bacteria, mobile and spore-forming. Oxidase and catalase activities were positive. The DNA G + C content of the type strain is 37.9 mol% in a genome sequence length of 4.52 Mb. Its type strain, Marseille-P3516^T (= CSURP3516^T), was isolated from a 10-year-old girl from Ndiop, a rural area in Senegal.

Description of *Bacillus massiliogabonensis* sp. nov.

Bacillus massiliogabonensis (mas.si.li.ga.bo.nen'sis: NL. masc. adj. a composed name designating Marseille and Gabon, the city and the country where the strain and the stool specimen was characterized and collected, respectively). The strain grows at

temperatures ranging from 23°C to 45°C in aerobic conditions with an optimum temperature of 37°C. Colonies with a white aspect had a mean diameter of 3 mm on blood agar medium. The strain Marseille-P2639^T is a Gram-negative bacterium and exhibits positive catalase and negative oxidase activities. The genome of the Marseille-P2639^T strain was 5.13 Mb with 37.9 mol% of G + C content. The potential pathogenicity of the type strain Marseille-P2639^T (=CSURP2639) is unknown. It was isolated from the stool sample of a healthy 16-year-old Gabonese boy.

Nucleotide sequence accession number

Table 7 shows the 16S rRNA gene and genome sequence accession numbers deposited in GenBank for these three new bacterial species:

Conflict of interest

None to declare.

Funding sources

This study was supported by the Institut Hospitalo-Universitaire (IHU) Méditerranée Infection, the National Research Agency under the programme *Investissements d'avenir*, reference ANR-10-IAHU-03, the Région Provence-Alpes-Côte d'Azur and European funding FEDER PRIMI.

TABLE 7. Accession numbers of *Bacillus dakarensis* strain Marseille-P3515^T, *Bacillus sinesaloumensis* strain Marseille-P3516^T and *Bacillus massiliogabonensis* strain Marseille-P2639^T

Species	Strain number	16S rRNA number	Genome accession number
<i>Bacillus dakarensis</i>	Marseille-P3515	LT671589	FTOZ000000000
<i>Bacillus sinesaloumensis</i>	Marseille-P3516	LT671591	FTOX000000000
<i>Bacillus massiliogabonensis</i>	Marseille-P2639	LT598571	FZJR000000000

Ethics and consent

The study and consent procedures were approved by the ethics committee of the Institut Hospitalo-Universitaire Méditerranée Infection (No. 2011-11), the National ethics committee of Gabon (No. 0023/2013/SG/CNE) and IFR48 of Marseille France (No. 09-022). The volunteers gave a written consent.

Acknowledgements

The authors thank Catherine Robert for sequencing the genome, Aurelia Caputo for submitting the genomic sequence to GenBank and Carine Couderc for producing the MALDI-TOF reference spectrum.

References

- [1] Cohn F. Untersuchungen über Bakterien. Beiträge zur Biologie der Pflanzen Heft 1872;1:127–224.
- [2] Lopez MS, Hodde MK, Chamakura KR, Kutty Everett GF. Complete genome of *Bacillus megaterium* podophage page. *Genome Announc* 2014;2(2):e00332-14.
- [3] Zhang YZ, Chen WF, Li M, Sui XH, Liu HC, Zhang XX, et al. *Bacillus endoradicis* sp. nov., an endophytic bacterium isolated from soybean root. *Int J Syst Evol Microbiol* 2012;62:359–63.
- [4] Jernigan JA, Stephens DS, Ashford DA, Omenaca C, Topiel MS, Galbraith M, et al. Bioterrorism-related inhalational anthrax: the first 10 cases reported in the United States. *Emerg Infect Dis* 2001;7:933–44.
- [5] Bottone EJ. *Bacillus cereus*, a volatile human pathogen. *Clin Microbiol Rev* 2010;23:382–98.
- [6] Turnbaugh PJ, Ley RE, Hamady M, Fraser-Liggett CM, Knight R, Gordon JL. The human microbiome project. *Nature* 2007;449:804–10. Available at: <https://www.nature.com/articles/nature06244>.
- [7] Lagier JC, Armougom F, Million M, Hugon P, Pagnier I, Robert C, et al. Microbial culturomics: paradigm shift in the human gut microbiome study. *Clin Microbiol Infect* 2012;18:1185–93.
- [8] Lagier JC, Hugon P, Khelaifia S, Fournier PE, La Scola B, Raoult D. The rebirth of culture in microbiology through the example of culturomics to study human gut microbiota. *Clin Microbiol Rev* 2015;28:237–64.
- [9] Lagier JC, Khelaifia S, Alou MT, Ndongo S, Dione N, Hugon P, et al. Culture of previously uncultured members of the human gut microbiota by culturomics. *Nat Microbiol* 2016;1:16203.
- [10] Ramasamy D, Mishra AK, Lagier JC, Padhmanabhan R, Rossi M, Sentausa E, et al. A polyphasic strategy incorporating genomic data for the taxonomic description of novel bacterial species. *Int J Syst Evol Microbiol* 2014;64:384–91.
- [11] Fournier PE, Lagier JC, Dubourg G, Raoult D. From culturomics to taxonomogenomics: a need to change the taxonomy of prokaryotes in clinical microbiology. *Anaerobe* 2015;36:73–8.
- [12] Seck EH, Diop A, Armstrong N, Delerce J, Fournier PE, Raoult D, et al. Microbial culturomics to isolate halophilic bacteria from table salt: genome sequence and description of the moderately halophilic bacterium *Bacillus salis* sp. nov. *New Microbe*. *New Infect* 2018;23:28–38.
- [13] Lo CI, Fall B, Sambe-Ba B, Diawara S, Gueye MW, Mediannikov O, Sokhna C, et al. MALDI-TOF mass spectrometry: a powerful tool for clinical microbiology at Hôpital Principal de Dakar, Senegal (West Africa). *PLoS One* 2015;10(12):e0145889.
- [14] Morel AS, Dubourg G, Prudent E, Edouard S, Gouriet F, Casalta JP, et al. Complementarity between targeted real-time specific PCR and conventional broad-range 16S rDNA PCR in the syndrome-driven diagnosis of infectious diseases. *Eur J Clin Microbiol Infect Dis* 2015;34:561–70.
- [15] Meier-Kolthoff JP, Göker M, Spröer C, Klenk HP. When should a DDH experiment be mandatory in microbial taxonomy? *Arch Microbiol* 2013;195:413–8.
- [16] Stackebrandt E, Ebers J. Taxonomic parameters revisited: tarnished gold standards. *Microbiol Today* 2006;33:152–5.
- [17] Lo CI, Sankar SA, Fall B, Ba BS, Diawara S, Gueye MW, et al. High-quality draft genome sequence and description of *Haemophilus massiliensis* sp. nov. *Stand Genom Sci* 2016;11:31.
- [18] Zerbino DR, Birney E. Velvet: algorithms for de novo short read assembly using de Bruijn graphs. *Genome Res* 2008;18:821–9.
- [19] Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, et al. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. *J Comput Biol* 2012;19:455–77.
- [20] Luo R, Liu B, Xie Y, Li Z, Huang W, Yuan J, et al. SOAPdenovo2: an empirically improved memory-efficient short-read de novo assembler. *Gigascience* 2012;1:18.
- [21] Bolger AM, Lohse M, Usadel B. Trimmomatic: a flexible trimmer for Illumina sequence data. *Bioinformatics* 2014;30:2114–20.
- [22] Lee I, Ouk Kim Y, Park SC, Chun J. OrthoANI: an improved algorithm and software for calculating average nucleotide identity. *Int J Syst Evol Microbiol* 2016;66:1100–3.
- [23] Richter M, Rosselló-Móra R. Shifting the genomic gold standard for the prokaryotic species definition. *Proc Natl Acad Sci U S A* 2009;106(45):19126–31.
- [24] Meier-Kolthoff JP, Auch AF, Klenk HP, Göker M. Genome sequence-based species delimitation with confidence intervals and improved distance functions. *BMC Bioinform* 2013;14:60.