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Vascular Dysfunction Associated with Type 2 **Diabetes and Alzheimer's Disease: A Potential Etiological Linkage**

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The endothelium performs a crucial role in maintaining vascular integrity leading to whole organ metabolic homeostasis. Endothelial dysfunction represents a key etiological factor leading to moderate to severe vasculopathies observed in both Type 2 diabetic and Alzheimer's Disease (AD) patients. Accordingly, evidence-based epidemiological factors support a compelling hypothesis stating that metabolic rundown encountered in Type 2 diabetes engenders severe cerebral vascular insufficiencies that are causally linked to long term neural degenerative processes in AD. Of mechanistic importance, Type 2 diabetes engenders an immunologically mediated chronic pro-inflammatory state involving interactive deleterious effects of leukocyte-derived cytokines and endothelial-derived chemotactic agents leading to vascular and whole organ dysfunction. The long term negative consequences of vascular pro-inflammatory processes on the integrity of CNS basal forebrain neuronal populations mediating complex cognitive functions establish a striking temporal comorbidity of AD with Type 2 diabetes. Extensive biomedical evidence supports the pivotal multi-functional role of constitutive nitric oxide (NO) production and release as a critical vasodilatory, anti-inflammatory, and anti-oxidant, mechanism within the vascular endothelium. Within this context, we currently review the functional contributions of dysregulated endothelial NO expression to the etiology and persistence of Type 2 diabetes-related and co morbid ADrelated vasculopathies. Additionally, we provide up-to-date perspectives on critical areas of AD research with special reference to common NO-related etiological factors linking Type 2 diabetes to the pathogenesis of AD.

MeSH Keywords: Nitric Oxide • Diabetes Mellitus, Type 2 • Free Radicals • Endothelial Cells • Etiology • **Alzheimer Disease**

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Introduction: Diabetes

Diabetes mellitus (DM), one of the major leading chronic morbidities worldwide, is continually increasing with a high prevalence from 100 million in 1999 to 382 million in 2013, and to over 592 million by 2035 estimated by the International Diabetes Federation (IDF) [1,2]. Maturity-onset DM (Type 2 DM or T2DM), the most common form of diabetes, accounts for over 85% cases in all diagnosed patients, and tends to increase in children and adolescents [1]. In the United States, approximately 10% to 20% of the population older than 45 years of age were diagnosed with T2DM in 1999 [1]; by the year 2013, a total of 25.8 million Americans (some 8.3% of the population) were diabetic, of which, approximately 90–95% have T2DM [2]. Great progress has been made regarding the pathogenesis of T2DM in the past decade with particular attention on the intertwining relationship between vasopathies and T2DM.

Diabetes-associated vascular dysfunction in multiple organs has long been considered as the sequelae of the progressive disease [3], whereas some other disorders like hypertension, hypercholesterolemia, retinopathy, etc. were regarded as comorbidities of diabetes [4], and more recently, new evidence indicated that the genesis of diabetes is most likely a result of vascular dysfunction [5], and the heterogeneity property of diabetes was also highlighted [6]. Although the risk factors of T2DM have been known for many years, the vascular dysfunction-associated aspect has become a new factor, considered to be an essential contributor to the new-onset T2DM.

A well-functioning vascular system plays a pivotal role in keeping the organs healthy under both non- and genetic conditions. However, when this normal state is affected by different extrinsic or/and intrinsic factors, and then some pathologic changes occur, especially when predisposing factors exist, they work chronically to enhance the pathology. One example for this relationship is that the major four classes of drugs that are commonly used for cardiovascular risk reduction (statins, niacin, thiazide diuretics, and β -blockers) have been shown to increase the risk of new-onset diabetes by 9% to 43% [5]. The stability of vascular function largely depends on the prompt action of different mediators working precisely on the whole. Solid evidence has confirmed the importance of a functionally intact endothelium for preventing vascular dysfunction [7].

Dysfunction of multiple aspects of the endothelium has been identified in diabetes mellitus. In human retinal endothelial cells, modulation of plasminogen activator synthesis by insulin-like growth factor-1 (IGF-1), epidermal growth factor (EGF) or acidic fibroblast growth factor (AFGF) is known to be influenced by diabetes [8]. Alterations in the synthesis and release of von Willebrand factor (vWF), an important factor for efficient platelet adhesion, are common in diabetic endothelium [9]. Abnormal glycosylation of intracellular and plasma proteins occurs and affects endothelial function in diabetic patients [10,11]. Recently, cumulating evidence indicated that endothelium-dependent vasomotion was affected by DM and also contributed to the new onset DM.

In this review, we will summarize the new developments on the interrelationship between T2DM and vascular dysfunction, and provide evidence that vascular-functioning mediators are involved in the pathogenetic regulation of T2DM, and discuss complicated disorders with vascular dysfunction that are strongly associated with T2DM.

Causal Relationship Between Vascular Dysfunction and Type 2 Diabetes

It is ascertained, at least in part, that a correlation exists between vascular dysfunction and the occurrence of T2DM, but we still do not know the precise causal relationship between them. Indeed, the pathology may be initiated metabolic dysregulation, leading to poor energy production in the microenvironment. Thus, the association may have existed in the beginning of the disorder and in Alzheimer's disease (AD) as well.

Vascular dysfunction, including microvascular and macrovascular, results from DM affecting several organs (muscle, skin, heart, brain, eyes, and kidneys, et al.) [3]. The common etiology link for the different types of diabetes-associated vascular diseases is the chronic hyperglycemia that evokes pathologic responses in the vasculature, which finally cause constitutive nitric oxide (cNO) inhibition, smooth muscle cell dysfunction, overproduction of vascular endothelial growth factor (VEGF), chronic inflammation, hemodynamic dysregulation, impaired fibrinolytic ability, and enhanced platelet aggregation. In these situations, diabetes, specifically T2DM, was considered as the initiating factor for the series of vascular diseases with the contribution of common risk factors like hypertension, tobacco use, and obesity.

Although it has been confirmed that the on-going diabetes plays a crucial part in inducing subsequent vascular dysfunction through affecting endothelium [3,12], the concerns regarding the priority (i.e. which comes first – vascular dysfunction or diabetes) was raised recently. The question was derived from the evidence that statin therapy increased the risk of new onset diabetes (NOD), of which belongs to T2DM, by 9% to 21% [13,14], and this was validated by a more recent study, in which if the patients had 2–4 risk factors for NOD, atorvastatin 80 mg increased the risk of NOD by 24% compared with standard therapy [15]. From these studies, at least in part, it can be deduced that vascular dysfunction itself could initiate diabetes even though the underlying mechanisms are not yet unknown.



Figure 1. Interrelationship between diabetes and vascular system. Diabetes originally evokes disturbance in vasoendothelium function that eventually changes vascular responses, which in turn form a positive feedback circuit with diabetes. In normal patients, vascular drugs can function as an initiator of new onset diabetes through mediating vascular and vaso-endothelium function. Almost simultaneously, pathologies associated with Alzheimer's Disease may emerge given the discussed commonalities since all can be linked to mitochondrial responses.

One potential mutual link between diabetes and vascular endothelium exists which determines the outcomes of the patients. In this proposed link, DM affects vascular endothelium functions through serious vaso-associated systems and pathways (see details on "Diabetes Endothelium" below), which results in changes in vascular function. Significantly, the altered endothelium and blood vessels positively are deteriorating in diabetes, i.e. a positive feedback formed. Moreover, exogenous cardiovascular medications may cause new onset diabetes through impacting endothelium and overall vascular function (Figure 1).

Dynamic Diabetic Endothelium

Endothelium is the major part of the vasculature in regulating vascular function. In the context of T2DM, chronic stimulation of hyperglycemia activates clusters of intrinsic vasoregulating systems or pathways in vascular endothelium [16]. Understanding the dynamic underlying mechanisms is critical for understanding the diabetes-associated vascular dysfunction so that preventive strategies can be developed.

Renin-angiotensin system

The renin-angiotensin system (RAS) is linked to local regulatory mechanisms that contribute to a great number of homeostatic pathways, including cellular growth, extracellular matrix formation, and apoptosis, whereas particular concerns were raised on its role in endothelium and vascular function. Blockade of the RAS can improve visual acuity and a short-term reduction in central macular thickness in patients with refractory diabetic macular edema [17], and blockade of angiotensin II attenuates VEGF-mediated blood-retinal barrier breakdown in diabetic retinopathy [18]. Furthermore, inhibition of RAS may ameliorate islets function of diabetic rats by increasing the microvessel density in islets [19], and can increase the resistance to streptozotocin-induced diabetes, reduce inflammatory markers, and improve islet cell function [20]. Additionally, evidence accumulated to date indicates that reduction in RAS activity can prevent NOD, and the blockers are considered as the first choice drugs in hypertensive patients with diabetes to prevent the occurrence and progression of complications in diabetes [21].

Reactive oxygen species

Reactive oxygen species (ROS) are important secondary messengers for signaling pathways associated with apoptosis, proliferation, damage and inflammation. Diabetes is a condition of increased oxidative stress and requires antioxidants [22]. In diabetic condition, the delicate equilibrium between ROS production and antioxidant capability is distorted resulting in oxidative stress and further tissue injury. Vascular cells function as the major part of reactive oxygen species production, which underlies the pathogenic progression of diabetes [23]. Increased generation of reactive oxygen species from multiple enzymatic sources promotes insulin resistance, specifically at the level of the endothelium, and leads to acceleration of atherosclerosis in areas with disturbed flow patterns [24]. Based on these findings, counter regulating of ROS was studied to reverse associated vascular dysfunction. α-Melanocyte-stimulating hormone, a naturally occurring endogenous peptide hormone of the melanocortin family, can normalize oxidative stress, reduce apoptosis and ultrastructural injuries, and correct gene expression levels in early diabetic retinas [25]. Augmented superoxide dismutase 2 (SOD2) ubiquitination leads to the increase in mitochondrial ROS concentration in coronary endothelium from T2DM mice and attenuates coronary vascular relaxation [26]. Moreover, direct anti-ROS treatment can prevent exacerbation of inflammation and insulin resistance that alleviates diabetic pathologies [27]. Amelioration and/or prevention of vascular endothelial and contractile dysfunction by doxycycline is strongly associated with a clear reduction in oxidative stress markers of diabetes, suggesting a therapeutic intervention for amelioration and/or prevention of vascular disorders in diabetic subjects [28].

Protein kinase C pathway

The regulatory enzyme protein kinase C (PKC) is known to play a key role in vascular tone regulation in physiological and pathological conditions[30]. The association of protein kinase C (PKC) with vascular alterations such as increases in permeability, contractility, extracellular matrix synthesis, cell growth and apoptosis, angiogenesis, leukocyte adhesion, and cytokine activation and inhibition has been confirmed [30]. These

perturbations in vascular cell homeostasis, caused by different PKC isoforms (PKC- α , - β 1/2, and PKC- δ), are linked to the development of pathologies affecting large vessel (atherosclerosis, cardiomyopathy) and small vessel (retinopathy, nephropathy and neuropathy) complications in the context of diabetes [29]. Activated PKC in diabetes increases VEGF expression and activates NADPH oxidases leading to raised ROS production. In addition, PKC in DM is involved in enhancement of vascular contractility in an endothelium-independent manner by inactivation of K+ channels and Ca2+ sensitization of myofilaments in vascular smooth muscle cells [30]. PKC-B phosphorylates occludin regulating tight junction trafficking in VEGF-induced permeability *in vivo* [31]. Silencing of the PKC-δ gene expression using siRNAs led to restoration of vasodilator potential in rats with diabetes [32]. PKC inhibition ameliorates functional endothelial insulin resistance and smooth muscle cell hypersensitivity to insulin, but does not restore acetylcholine-activated endothelium-dependent vasodilation in diabetic rats [33]. The function of PKC pathway in diabetic context is intricate. How to reach an ideal function of PKC in the regulation of vascular function needs in-depth investigation.

Heme oxygnase-1/carbon monoxide

The intracellular levels of carbon monoxide (CO) can increase under stressful conditions following the induction of heme oxygnase-1 (HO-1), a ubiquitous enzyme responsible for the catabolism of heme. Although CO does not contain free electrons, it may still be involved in oxidative stress, and Heme oxygenase-1 (HO-1) was found to be a key defense mechanism against oxidative injury [34]. HO-1 possesses a protective effect against aortic endothelial dysfunction in the insulin resistance (IR) state by inducing antioxidation and promoting regulative effect of vasoactive substances [35]. Induction of HO-1 with hemin ameliorates the abnormality of endothelium-dependent vascular relaxation in T2D rats through suppressing reactive oxygen species production and inhibiting COX-2 upregulation induced by diabetes mellitus [36], and ameliorating exaggerated vascular contractility by reducing TNF- α and aortic ROS levels [37] and pulmonary artery by involving a reduction in inducible NO synthase-derived NO production [38].

Further studies found the enhancement of the HO system facilitates insulin sensitivity and glucose metabolism in diabetic animals [39,40]. HO-1-induced increase in eNOS and decrease in iNOS are potentially contributing to restoration of vascular responses in diabetic rats [41], and HO-1 up regulation in diabetic rats brings about an increase in serum bilirubin, a reduction in O*-2 production, and a decrease in endothelial cell sloughing [42], and amelioration in postprandial and fasting hyperglycemia in T2DM rats [43]. Therefore, attention is being paid to the use of selective HO inhibitors that were considered very important tools to clarify the role of the HO system and the mechanisms underlying its physiological effects and pathological involvement; due to the inducible nature of HO-1, selective inhibition of HO-1 isoforms is generally preferable. Notably, HO-1 inhibitors may be also beneficial in therapeutic applications.

Nitric oxide system

The vascular endothelium is a multifunctional organ and is critically involved in modulating vascular tone and structure. Nitric oxide (NO) is a short-lived gaseous signaling molecule in mammals. The regulation of NO bioavailability is critical to maintain blood vessel function. Endothelium-derived NO has been demonstrated to mediate many important endothelial properties [44]. NO inhibits the adhesion and aggregation of platelets and the release of their contained growth factors, the chemotaxis and activation of mononuclear leukocytes, the expression of leukocyte adhesion molecules by activated vascular endothelium, and the migration and proliferation of smooth muscle cells. It decreases endothelial permeability for macromolecules and lipoproteins [44-46]. In addition, NO is essential for the maintenance of basal vascular tone and its regulation in response to various physiologic and pathologic stimuli [47,48]. Therefore it has been surmised that impaired activity of NO diminishes the resistance of the vascular wall to disease and disrupts vascular homeostasis.

Initially, studies in animals were performed which found both conduit and resistance arteries of chemically induced diabetes to have attenuated endothelium-dependent relaxation [49,50]. Subsequently, evidence of impaired endotheliummediated smooth muscle relaxation in humans was discovered in the penile corpora cavernosa [51]. To date, numerous studies have confirmed that endothelium-dependent relaxation is impaired in DM. Early reports have examined NO activity in diabetic endothelium, but contradicting results have emerged [52–56]. One possible reason was that all of these studies have used indirect methods in an attempt to determine NO's presence and thus its role. As a result, no conclusive direct evidence was reached on the levels of NO release in diabetic endothelium.

One indirect method is the administration of NO trapping agents, which were used to assess the basal NO activity in the rat aorta [57,58]. These scavengers bind and eliminate NO after its release without altering NOS activity. Analysis of tension in rings pre-contracted with phenylephrine revealed that the application of NO trapping agents produced an additional increment in tension that was greater in the control than the diabetic rings. These findings were thought to suggest that the basal NO activity in diabetics is less than that in non-diabetics. Another indirect means used in assessing information regarding potential deficits in NO synthesis is the measurements of cGMP in vascular tissue [59]. Decreases in acetylcholine-stimulated cGMP production were observed in multiple diabetic animal models. No apparent intrinsic change in either guanylate cyclase or phosphodiesterase activity of the vascular smooth muscle was found to account for defective cGMP production in these blood vessels. Therefore, the conclusion was suggested that the basal NO bioactivity in experimental diabetes was decreased. Unfortunately, all of these investigations in both animal models and human studies make the assumption that endotheliumdependent relaxation in both control and diabetic blood vessels is exclusively mediated via NO to reach their conclusions.

Beside these developments, we first, in 2002, reported on the NO stimulated cNOS activation by morphine in diabetic human saphenous vein [7]. In this early study [7], we first employed the real-time NO release using an amperometric probe, real-time technology, to conclusively provide information on NO levels, from which we found that diabetic endothelium exhibits a diminished basal NO release in comparison to nondiabetic controls. Through the application of a NO synthase inhibitor, further information on the actual basal level of NO release was obtained. Diabetic endothelium showed no decrease in its basal release level. We believed this signifies that the actual level of NO release is negligible. By exposing the tissues to superoxide dismutase (SOD) we confirmed that the depressed levels of released NO seen were not the result of extracellular NO scavengers. Unfortunately, though we could not exclude the possibility of intracellular scavengers. Lastly, we evaluated the capacity of diabetic endothelium to produce NO when stimulated by an agonist - morphine sulfate. The results obtained were consistent with the premise that the previous findings support, NO metabolism is impaired in DM. A lower peak level and shorter duration of stimulated release, as well as a decreased expression of mu opiate receptors was seen in diabetic compared to non-diabetic tissue.

In considering the consequences of these findings in DM, a negligible basal level and a diminished capacity for stimulated endothelial NO release, some of the possible potentially pathologic resultant alterations in cellular metabolism should be mentioned. Firstly, NO has been shown to have antioxidant actions [59-62]. This scavenging property gives NO a major intracellular and extracellular action against oxidative stress [61,63–70]. Emerging evidence attested the role of endothelial NO in diabetic vascular pathologies. First, endogenous NO suppresses vascular inflammation, via inhibition of p47(phox) expression, leading to attenuation of NADPH oxidase-dependent superoxide production [71]. However, selective endothelial insulin resistance itself is sufficient to induce a reduction in NO bioavailability and endothelial dysfunction that is secondary to increased generation of reactive oxygen species [72]. Third, diabetes produces a cascade of events involving production of reactive oxygen species from the NADPH oxidase leading to oxidation of BH4 and uncoupling of NOS, which promotes the

oxidative inactivation of NO with subsequent formation of peroxynitrite [73]. As thus, in the absence of NO, reactive chemical species may occur and cause tissue damage [7].

Heme proteins (e.g., hemoglobin, cytochromes, etc.) reacting with H_2O_2 results in ferryl cation (FE⁴⁺=O), a toxic substance [74]. However, once in contact with NO, this compound is reduced (FE³⁺ + NO₂⁻)[61,75], suggesting the antioxidant action of NO. NO also has the potential to diminish the formation of OH[•], demonstrating once more an antioxidant action [60,76]. Here again we note that in the absence of NO, as potentially occurring in diabetes, these reactive chemical species may cause the tissue damage associated with diabetes's pathological progression.

Meanwhile, NO interacts with heme proteins to activate soluble guanylyl cyclase and cyclooxygenase and also has the ability to inhibit lipoxygenase [77-80]. Inducible nitric oxide synthase (iNOS) driven NO production regulates expression of iNOS, suppresses cyclooxygenase-2 (COX-2) levels, and maintains Hemeoxygenase-1 (HO-1) levels in nephritic animals [81]. However, in the context of pouchitis, another type of inflammatory disease, the increased COX-2 and NOS were not associated with HO-1 induction [82]. A more recent study showed that HO-1 regulates lipopolysaccharide (LPS) and nicotine induced the production of NO and PGE2 through affecting the expression of iNOS and COX-2 in human periodontal ligament cells [83]. These data suggest, at least in part, that altered NO functions through connecting with COX and HO-1 systems, which is involved in the mediation of subsequent inflammatory processes. One of the mechanisms is the altered formation and release of the induced substances secondary to the impaired NO levels could adversely impact the important regulation of the pro-inflammatory processes.

Additionally, NO is an inhibitor of cytochrome c oxidase, the terminal enzyme in the electron transport chain, in a manner that is reversible and competitive with oxygen, by reacting with both the two-electron and single-electron reduced active sites [84-93]. Diminished NO levels could alter the modulation of oxygen utilization, in a similar manner to how the inhibition of c-NOS derived NO increases oxygen consumption in many animal species [94–98]. Besides, the endogenously released endothelial NO, either basal or stimulated, can modulate oxygen consumption both throughout the thickness of conductance vessels and in the microcirculation, which may be implicated in the origin of vascular pathologies such as atherosclerosis [99]. Moreover, inflammatory mediators such as IL-1 β hampers glucose-stimulated insulin secretion (GSIS) in Cohen diabetic rat islets through mitochondrial cytochrome c oxidase inhibition by NO. Reduced islet COX activity renders vulnerability to GSIS inhibition on high-sucrose, low-copperdiet (HSD) through inducing the expression of iNOS and nitric



oxide-mediated COX inhibition [100] (Figure 2). Intriguingly, the blunted endothelial NO in patients suffering from insulin resistance T2DM played an interaction between endothelial insulin sensitivity, of which the increased insulin signaling in endothelium increases the generation of superoxide anion via activation of NOX2 NADPH oxidase and reduced NO production in response to insulin due to increased endothelial proline-rich tyrosine kinase (PYK2) activity leading to a pro-atherosclerotic state [101].

Therefore, we surmise that basal NO levels promote the health of the endothelium by limiting its immune and vascular activating potential, i.e., decreasing the appearance of an uncalled for pro-inflammatory response [7,44]. Thus, in diabetic individuals, the decrease in the capacity for this vital action leads to both enhanced vascular and immune activity, as noted by increased platelet-derived plaque formation for example [44]. The fact that stimulated NO levels are significantly diminished, but still exist, denotes the progressive nature of the vascular pathology associated with diabetes. The remaining capacity for stimulated NO release may help down-regulate mediators in the vascular and immune tissues [44]. However, since it is not continuous and diminished in the diabetic, its effect is probably only partial, allowing for a progressive decrease in the ability to down-regulate these tissues over the long term. Certainly, this may not be the only explanation for the vascular abnormalities found in diabetes mellitus given the complexity of the pathological processes, however, we do believe that this impaired NO metabolism plays a significant role.

NO and superoxide free radicals in diabetes

Additionally, NO is emerging as a central regulator of energy metabolism and body composition. In considering the consequence of a general lack of NO in severe diabetics on the cellular level, we surmise an alteration of endothelial cellular Figure 2. Illustration of the interconnectivity of health and pathology and their dependence, in part, on mitochondrial associated processes. Given the cells. Michochondria's position in its hierarchy of immediate response its significance is highlighted. It's significance is also enhanced by the number of processes that can be associated with it that can "dampen" the immediate/acute effect/stimulus of most abnormalities, making it prone to chronic debilitation. This hypothesis is further highlighted by the fact that each condition has a co morbidity that is related to a prime pathology. Thus co morbidities represent the scope of the disorder while documenting other components, which can be considered holistically as a medical target.

metabolism. NO can interact with oxygen, metals and other free radicals [44,102,103]. NO can form peroxynitrite (ONOO-) and dinitrogen trioxide (N₂O₃), following an interaction with the superoxide radical (O2-) and oxygen, respectively [61,104]. NO and ONOO- have inhibitory effects on purified cytochrome b5 reductase (Cb5R), a pleiotropic flavoprotein that catalyzes multiple one-electron reduction reactions with various redox partners, providing the basis for a feedback cellular protection mechanism through modulation of excessive extramitochondrial superoxide anion production by Cb5R [105]. In this regard, NO's actions are directly felt when its level is low and of short duration, occurring under physiological conditions [61]. For example, NO interaction with the heme proteins represents the activation of soluble guanylyl cyclase (sGC) and/or COX [77-79]. The sGC-cGMP signaling is one of the major pathways of NOrelated vascular function mediation [106]. NO-associated COX activation is of importance in the regulation of a pro-inflammatory process [79,107]. Additionally, at low NO concentrations it modulates the redox form of COX, converting the ferrous iron to the ferric active form, acting also as a scavenger of superoxide [61]. Meanwhile, NO functions as a weak ligand to ferric heme, which, at the same time, forms a strong Fe-NO bond to regulate the reactivities of ferrous and ferric hemenitrosyls [108]. NO also has the ability to inhibit lipoxygenase [80,109]. It can also reversibly inhibit the heme moiety of cytochrome P-450, preventing the binding of oxygen to the catalytic sites [110,111]. However, a peculiar P450, P450nor, can receive electrons directly from NADH for the reduction of NO [112]. Interestingly, at low NO levels H₂O₂ can be consumed to yield HNO₂ [61,113], suggesting that H₂O₂ might serve to control NO levels that contribute to the development of vascular diseases [61,114]. In this regard, if NO were absent, H₂O₂ may generate tissue damage and energy metabolism may proceed impaired as appears to occur in diabetes [115]. A more interesting finding is that endothelium-derived hyperpolarizing factor (EDHF) compensates for diminished NO-dependent dilation in IL-6-induced endothelial dysfunction by the activation of H_2O_2 or a K+ channel in T2DM, suggesting that the interaction between NO and H_2O_2 is not as simple as figuring out which depends upon which [116]. We suppose that the superoxide products in diabetic endothelium confound each other to induce vascular pathologies.

Furthermore, mitochondria represent a NO target since NO is an inhibitor of cytochrome oxidase of the electron transport process [84–89,117], suggesting again a NO role in modulating oxygen utilization [85,118]. The inhibition of cNOS-derived NO increases oxygen consumption in many animal species [94– 98]. This last fact is critical to our NO hypothesis concerning diabetes (see earlier discussion). Furthermore, a NOS isoform, mtNOS, is present in mitochondria [84,119–121] supporting an important modulatory function as well.

Alzheimer's Disease and Type 2 Diabetes

Chronic vascular diseases are the major complications that account for over 90% of mortality from diabetes. Alzheimer's disease (AD), which affects 36 million people worldwide, is generally considered as an age-related degenerative disease. However, the actual etiology of AD is yet unknown. Emerging evidence indicates that the pathogenesis of AD is attributable to the chronic vascular pathologies [122].

Vasculopathy and Alzheimer's disease

Our previous work disclosed a causal relationship between vascular pathologies and AD [122-133], and we evidenced that AD may be a vascular disorder with neurodegenerative consequences rather than a neurodegenerative disorder with vascular consequences. Based on this proposal, two factors are needed to be present for AD to develop: 1) advanced ageing, 2) presence of a condition that lowers cerebral perfusion, such as a vascular-risk factor. The first factor introduces a normal but potentially insidious process that lowers cerebral blood flow in inverse relation to increased ageing; the second factor adds a crucial burden which further lowers brain perfusion and places vulnerable neurons in a state of high energy compromise leading to a cascade of neuronal metabolic turmoil. Convergence of these two factors, will culminate in a Critically Attained Threshold of Cerebral Hypoperfusion (CATCH) [122-133]. CATCH is a hemodynamic microcirculatory insufficiency that will destabilize neurons, synapses, neurotransmission and cognitive function, creating a neurodegenerative state characterized by the formation of senile plagues, neurofibrillary tangles, amyloid angiopathy and in some cases, Lewy bodies. Since any of a considerable number of vascular-related conditions must be present in the ageing individual for cognition to be disturbed, CATCH identifies an important aspect of the heterogenic disease profile assumed to be present in the AD syndrome [122].

It is proposed that CATCH initiates AD by distorting regional brain capillary structure, involving endothelial cell shape changes and impairment of NO release, which affects signaling between the immune, cardiovascular and nervous systems. Evidence indicates that in many tissues there is a basal level of NO being produced and that the actions of several signaling molecules may initiate increases or decreases in basal NO levels [44]. Moreover, these temporary alterations in basal NO levels exert inhibitory cellular actions, via cellular conformational changes. Findings indicate that 1) constitutive NO is responsible for a basal or "tonal" level of NO; 2) this NO keeps particular types of cells in a state of inhibition and 3) activation of these cells occurs through disinhibition. Consequently, tissues not maintaining a basal NO level are more prone to excitatory, immune, vascular and neural influences. Under such circumstances, these tissues cannot be down-regulated to normal basal levels, thus prolonging their excitatory state [44]. Thus, the clinical convergence of advanced ageing in the presence of a chronic, pre-morbid vascular risk factor, can, in time, contribute to an endotheliopathy involving basal NO deficit, to the degree where regional metabolic dysfunction leads to cognitive meltdown and to progressive neurodegeneration characteristic of AD [122].

The conventional view proposes that AD precedes vascular dysfunction. In this regard, an increase in the concentration of amyloid beta (A β) above its normal physiological level in the circulation results in decreased NO production and vessel sensitivity to endothelium-dependent vasodilation, that could lead to constricted blood vessels and ischemia in the surrounding tissue [134]. Furthermore, increases in A β lead to cell death and decreased maximum vasodilator response of cerebral vessels in the context of AD [134]. Nonetheless, this conception has been challenged by our previous compelling evidence discussed above, and along with our evidenced findings, other supportive data emerged from peer institutes upon the vascular originality of AD's pathogenesis. A number of review articles have provided an excellent view on the vascular contribution to the genesis of AD by focusing on the role of chronic hypoperfusion in triggering mitochondrial dysfunction in vascular cells, which, in turn, enhances the production of ROS and reactive nitrogen species (RNS) that were driven by NOS [122,135-138]. Therefore, NO-associated oxidative responses of the vascular system were considered as an initiator of brain lesions during the development of AD [122,139]. Further, NO production is IFN- γ dependent both in cognitive impairment and Alzheimer's patients, and the high levels of NO, probably iNOS driven since the condition is chronic, are

associated with an elevation of TNF- α levels in severe stages of AD [140], suggesting the existence of a causal interaction between NO and inflammation that underlies the severity of AD and cognition impairment. Meanwhile, the causal relationship among NO, mitochondrial dysfunction, oxidative stress, and AD has been highlighted as a therapeutic target [122] (Figure 2). Widespread cerebral amyloid angiopathy and arteriosclerosis/lipohyalinosis are associated with the development of cognitive deficits in AD and a combination of them may contribute to neurodegeneration in AD, suggesting that small vessel disease due to arteriosclerosis and fibrolipohyalinosis is a potential target for the treatment of AD [141]. Elimination of the imbalance seen in energy production and restoration of the normal cellular function, make the antioxidants powerful alternates for treating vascular pathologies as well as neurodegenerative diseases [142].

Alzheimer's Disease and Diabetes Type 2 Commonalities

Strong evidence demonstrates that AD shares characteristics and possible origins with both cardiovascular disorders and diabetes [44,122,143]. Given the discussion in the earlier reports noted, which emphasized energy metabolism due to hypoperfusion, it is not difficult to speculate that DM may be a major component in the pathogenesis of AD as reflected in the associated vasculopathy, resulting in damaging the microvascular environment. In this regard, it has been suggested that DM is a risk factor for AD [44,122,144,145]. Thus, the progression of events includes hyperglycemia leading to diabetic vasculopathy via glyceraldehyde-derived AGEs Glycer-AGE. Moreover, in human AD brain, Glycer-AGE is distributed in the cytosol of neurons in the hippocampus [145].

Furthermore, hyperglycemia-induced mitochondrial dysfunction and oxidative stress have been associated with AB induced proinflammatory responses [44,122,146], which is expected, given the damage to the vascular conduit of glucose utilization. Interestingly, high glucose and A\beta1-40 reduced cell and enhanced mitochondrial O2 - and H,O, production. This situation creates an environment promoting a higher susceptibility to the deleterious actions of A β -40 [146] (Figure 2). Semicarbazide-sensitive amine oxidase/vascular adhesion protein-1 (SSAO/VAP-1) also are implicated in AD and DM [147]. These findings support the hypothesis that DM predisposes to cerebrovascular alterations, cognitive decline, and development of AD [148]. In mice, cultured endothelial cells exposed to both glucose and Aß generate oxygen reactive products and glycation entities, which also have been associated with cognitive decline [149]. Thus, glucose associated presence and regulation are coupled to this pathological process and its alteration may represent the first step in this deteriorating energy pathway in both DM and AD.

Recently, Grammas et al. [143] proposed that in AD animal models, the cerebrovasculature is activated and overexpresses A β . Importantly, we proposed a similar vascular theory in 2000 (see previous section) [44,122,144]. We demonstrated that hypoperfusion of brain areas caused endothelial cells to loosen their juxtaposed borders, creating "gaps" whereby greater brain access occurred, i.e., allowing A β and immunocyte entry. Given A β immunostimulant activities a proinflammatory reaction would go from acute to chronic over time due to the "gaps" [122]. In part, this inflammatory state was, we surmise, initiated by a dysfunctional nitric oxide release from compromised endothelia, which constitutively stabilizes endothelia and immune cells [44,122]. Supporting this concept is the fact that rat hippocampal neurons exposed to CATCH exhibit an altered nitric oxide release [150]. In summary, an impaired microvasculature is present in both DM and AD as well as vascular dementia [44,122,144]. Additionally, it would appear that the vasculopathy emerges early on, remaining below "detection", leading to the neurodegenerative processes associated with AD.

Hayden et al. [151] proposes that cognitive decline in AD results from a combination of factors some of which have long been associated with the disorder (aging, genetic, and lifestyle), which result in multiple injurious metabolic and immunologic toxicities such as dysfunctional immune responses, oxidative stress, inflammation, insulin resistance, and dysglycemia (systemically and in the brain). These converging abnormalities may lead to endothelial blood-brain barrier tight junction/adherens junction complex remodeling and microglia activation, which may result in neurodegeneration, impaired cognition, and dementia. Indeed, on an individual basis, variations in susceptibility to these alterations probably exist, leading to the variations in the expression of characteristics associated with the disorders. The fact that similar patterns of vascular anatomy and functional dysfunction are quite similar in DM and AD, suggests that intrinsic susceptibility to both or either of these disorders also exists.

Conclusions

The early development of cardiovascular complications, including accelerated atherosclerosis and microangiopathy, which occurs with T2DM, is responsible for a high morbidity and mortality. Endothelial dysfunction plays an essential role in the initiation of cellular events, evolving into the development of vascular complications in diabetes. Diminished production and function of endothelium-derived NO and corresponding factors, in combination with the overproduction of pro-inflammatory mediators and vasoconstrictors, eventually result in endothelial dysfunction, which finally elevates vascular tone that culminates in changes to micro- and macro-vascular dysfunction. The diabetic context-associated NO imbalance and the subsequent cascades of free radicals underlie the vascular dysfunction,

which has been considered as the initiator of T2DM. In considering the potential commonalities of DM and AD, i.e. the NO-related oxidative alterations in vascular endothelium, new concerns have arisen on the preceded occurrence of DM and AD. The NO-oxidative links underlying vascular dysfunction appears to contributes to the genesis and progression of T2DM. Therapeutics targeting of ROS using antioxidants, inhibitors of the RAS, mediators increasing eNOS activity, or methods affecting vaso-function-related signaling pathways might assist in reversing the endothelial dysfunction that has been proposed. This strategy may aid in reducing the related vascular morbidity and mortality in diabetes, AD and possibly atherosclerosis.

References:

- 1. Stoffel M: Molecular genetics of non-insulin dependent diabetes mellitus. In: Chein KR (ed.), Molecular Basis of Cardiovascular Disease. Philadelphia, PA: W.B. Saunders, 1999; 477–501
- 2. American Diabetes Association. Fast Facts: Data and Statistics about Diabetes. 2013
- 3. Cade WT. Diabetes-related microvascular and macrovascular diseases in the physical therapy setting. Phys Ther, 2008; 88: 1322–35
- McNeely MJ, Boyko EJ: Diabetes-related comorbidities in Asian Americans: results of a national health survey. J Diabetes Complications, 2005; 19: 101–6
- 5. Ong KL, Barter PJ, Waters DD: Cardiovascular drugs that increase the risk of new-onset diabetes. Am Heart J, 2014; 167: 421–28
- Tuomi T, Santoro N, Caprio S et al: The many faces of diabetes: a disease with increasing heterogeneity. Lancet, 2014; 383: 1084–94
- 7. Bilfinger TV, Vosswinkel JA, Cadet P et al: Direct assessment of diminished production of morphine stimulated NO by diabetic endothelium from saphenous vein. Acta Pharmacologica Sinica, 2002; 23: 97–102
- Grant MB, Guay C: Plasminogen activator production by human retinal endothelial cells of nondiabetic and diabetic origin. Invest Ophthalmol Vis Sci, 1991; 32: 53–64
- 9. Pearson JD: Vessel wall interactions regulating thrombosis. Br Med Bull, 1994; 50: 776–88
- Lee TS, Saltsman KA, Ohashi H, King GL: Activation of protein kinase C by elevation of glucose concentration: proposal for a mechanism in the development of diabetic vascular complications. Proc Natl Acad Sci USA, 1989; 86: 5141–45
- Schmidt AM, Hasu M, Popov D et al: Receptor for advanced glycation end products (AGEs) has a central role in vessel wall interactions and gene activation in response to circulating AGE proteins. Proc Natl Acad Sci USA, 1994; 91: 8807–11
- 12. Tang X, Luo YX, Chen HZ, Liu DP: Mitochondria, endothelial cell function, and vascular diseases. Front Physiol, 2014; 5: 175
- 13. Sattar N, Preiss D, Murray HM et al: Statins and risk of incident diabetes: a collaborative meta-analysis of randomised statin trials. Lancet, 2010; 375: 735–42
- Preiss D, Seshasai SR, Welsh P et al: Risk of incident diabetes with intensive-dose compared with moderate-dose statin therapy: a meta-analysis. JAMA, 2011; 305: 2556–64
- Waters DD, Ho JE, Boekholdt SM et al: Cardiovascular event reduction versus new-onset diabetes during atorvastatin therapy: effect of baseline risk factors for diabetes. J Am Coll Cardiol, 2013; 61: 148–52
- Wong WT, Wong SL, Tian XY, Huang Y: Endothelial dysfunction: the common consequence in diabetes and hypertension. J Cardiovasc Pharmacol, 2010; 55: 300–7
- Abu El-Asrar AM, Al-Mezaine HS: Advances in the treatment of diabetic retinopathy. Saudi J Ophthalmol, 2011; 25: 113–22
- Kim JH, Kim JH, Yu YS et al: Blockade of angiotensin II attenuates VEGFmediated blood-retinal barrier breakdown in diabetic retinopathy. J Cereb Blood Flow Metab, 2009; 29: 621–28

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Conflict of interests

None.

- 19. Li X, Yuan L, Xu G et al: Effect of renin angiotensin system blockade on the islet microvessel density of diabetic rats and its relationship with islet function. J Huazhong Univ Sci Technolog Med Sci, 2009; 29: 684–88
- Li X, Yuan L, Li J et al: Blockade of renin angiotensin system increased resistance to STZ-induced diabetes in rats with long-term high-fat diet. Exp Diabetes Res, 2012; 2012: 618923
- 21. Saitoh S, Takeishi Y: Pleiotropic effects of ARB in diabetes mellitus. Curr Vasc Pharmacol, 2011; 9: 136–44
- 22. Rani AJ, Mythili SV: Study on total antioxidant status in relation to oxidative stress in type 2 diabetes mellitus. J Clin Diagn Res, 2014; 8: 108–10
- 23. Singh R, Devi S, Gollen R: Role of free radical in atherosclerosis, diabetes and dyslipidemia: larger-than-life. Diabetes Metab Res Rev, 2014 [Epub ahead of print]
- Gage MC, Yuldasheva NY, Viswambharan H et al: Endothelium-specific insulin resistance leads to accelerated atherosclerosis in areas with disturbed flow patterns: a role for reactive oxygen species. Atherosclerosis, 2013; 230: 131–39
- Zhang L, Dong L, Liu X et al: alpha-Melanocyte-stimulating hormone protects retinal vascular endothelial cells from oxidative stress and apoptosis in a rat model of diabetes. PLoS One, 2014; 9: e93433
- Cho YE, Basu A, Dai A et al: Coronary endothelial dysfunction and mitochondrial reactive oxygen species in type 2 diabetic mice. Am J Physiol Cell Physiol, 2013; 305: C1033–40
- Miao H, Ou J, Ma Y et al: Macrophage CGI-58 deficiency activates ROSinflammasome pathway to promote insulin resistance in mice. Cell Rep, 2014; 7: 223–35
- Zeydanli EN, Kandilci HB, Turan B: Doxycycline ameliorates vascular endothelial and contractile dysfunction in the thoracic aorta of diabetic rats. Cardiovasc Toxicol, 2011; 11: 134–47
- 29. Geraldes P, King GL: Activation of protein kinase C isoforms and its impact on diabetic complications. Circ Res, 2010; 106: 1319–31
- Kizub IV, Klymenko KI, Soloviev AI: Protein kinase C in enhanced vascular tone in diabetes mellitus. Int J Cardiol, 2014; 174: 230–42
- Murakami T, Frey T, Lin C, Antonetti DA: Protein kinase cbeta phosphorylates occludin regulating tight junction trafficking in vascular endothelial growth factor-induced permeability *in vivo*. Diabetes, 2012; 61: 1573–83
- Klymenko K, Novokhatska T, Kizub I et al: PKC-delta isozyme gene silencing restores vascular function in diabetic rat. J Basic Clin Physiol Pharmacol, 2014, 1–9 [Epub ahead of print]
- 33. Lu X, Bean JS, Kassab GS, Rekhter MD: Protein kinase C inhibition ameliorates functional endothelial insulin resistance and vascular smooth muscle cell hypersensitivity to insulin in diabetic hypertensive rats. Cardiovasc Diabetol, 2011; 10: 48
- Rochette L, Cottin Y, Zeller M, Vergely C: Carbon monoxide: mechanisms of action and potential clinical implications. Pharmacol Ther, 2013; 137: 133–52
- Chen YS, Zhu XX, Zhao XY et al: Hemin, a heme oxygenase-1 inducer, improves aortic endothelial dysfunction in insulin resistant rats. Chin Med J (Engl), 2008; 121: 241–47

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- 36. Wang Y, Ying L, Chen YY et al: Induction of heme oxygenase-1 ameliorates vascular dysfunction in streptozotocin-induced type 2 diabetic rats. Vascul Pharmacol, 2014; 61: 16–24
- Hassan N, El-Bassossy HM, Zakaria MN: Heme oxygenase-1 induction protects against hypertension associated with diabetes: effect on exaggerated vascular contractility. Naunyn Schmiedebergs Arch Pharmacol, 2013; 386: 217–26
- El-Bassossy HM, El-Maraghy NN, El-Fayoumi HM, Watson ML: Haem oxygenase-1 induction protects against tumour necrosis factor alpha impairment of endothelial-dependent relaxation in rat isolated pulmonary artery. Br J Pharmacol, 2009; 158: 1527–35
- Ndisang JF, Jadhav A: Heme oxygenase system enhances insulin sensitivity and glucose metabolism in streptozotocin-induced diabetes. Am J Physiol Endocrinol Metab, 2009; 296: E829–41
- Ndisang JF, Lane N, Jadhav A: The heme oxygenase system abates hyperglycemia in Zucker diabetic fatty rats by potentiating insulin-sensitizing pathways. Endocrinology, 2009; 150: 2098–108
- Ahmad M, Turkseven S, Mingone CJ et al: Heme oxygenase-1 gene expression increases vascular relaxation and decreases inducible nitric oxide synthase in diabetic rats. Cell Mol Biol (Noisy-le-grand), 2005; 51: 371–76
- 42. Quan S, Kaminski PM, Yang L et al: Heme oxygenase-1 prevents superoxide anion-associated endothelial cell sloughing in diabetic rats. Biochem Biophys Res Commun, 2004; 315: 509–16
- Ndisang JF, Lane N, Jadhav A: Upregulation of the heme oxygenase system ameliorates postprandial and fasting hyperglycemia in type 2 diabetes. Am J Physiol Endocrinol Metab, 2009; 296: E1029–41
- 44. Stefano GB, Goumon Y, Bilfinger TV et al: Basal nitric oxide limits immune, nervous and cardiovascular excitation: Human endothelia express a mu opiate receptor. Prog Neurobiol, 2000; 60: 513–30
- Bilfinger TV, Hartman AR, Liu Y et al: Cryopreserved veins in myocardial revascularization: Possible mechanism for their increased failure. Ann Thorac Surg, 1997; 63: 1063–69
- Duran WN, Beuve AV, Sanchez FA: Nitric oxide, S-nitrosation, and endothelial permeability. IUBMB Life, 2013; 65: 819–26
- 47. Gimbrone MA Jr, Topper JN: Biology of the vessel wall: endothelium. In: Chein KR, editors. Molecular basis of cardiovascular disease. Philadelphia: W.B. Saunders, 1999; 336
- Tousoulis D, Kampoli AM, Tentolouris C et al: The role of nitric oxide on endothelial function. Curr Vasc Pharmacol, 2012; 10: 4–18
- Oyama Y, Kawasaki H, Hattori Y, Kanno M: Attenuation of endothelium-dependent relaxation in aorta from diabetic rats. Eur J Pharmacol, 1986; 132: 75–78
- Takiguchi Y, Satoh N, Hashimoto H, Nakashima M: Changes in vascular reactivity in experimental diabetic rats: comparison with hypothyroid rats. Blood Vessels, 1988; 25: 250–60
- Saenz de Tejada I, Goldstein I et al: Impaired neurogenic and endothelium-mediated relaxation of penile smooth muscle from diabetic men with impotence. N Engl J Med, 1989; 320: 1025–30
- 52. Nitenberg A, Valensi P, Sachs R et al: Impairment of coronary vascular reserve and ACh-induced coronary vasodilation in diabetic patients with angiographically normal coronary arteries and normal left ventricular systolic function. Diabetes, 1993; 42: 1017–25
- 53. Karasu C, Soncul H, Altan VM: Effects of non-insulin dependent diabetes mellitus on the reactivity of human internal mammary artery and human saphenous vein. Life Sci, 1995; 57: 103–12
- Avogaro A, Piarulli F, Valerio A et al: Forearm nitric oxide balance, vascular relaxation, and glucose metabolism in NIDDM patients. Diabetes, 1997; 46: 1040–46
- 55. Wakabayashi I, Hatake K, Kimura N et al: Modulation of vascular tonus by the endothelium in experimental diabetes. Life Sci, 1987; 40: 643–48
- Furman BL, Sneddon P: Endothelium-dependent vasodilator responses of the isolated mesenteric bed are preserved in long-term streptozotocin diabetic rats. Eur J Pharmacol, 1993; 232: 29–34
- Pieper GM, Lai CS: N-Methyl-D-glucamine dithiocarbamate-Fe2+ discriminates basal vs. agonist-stimulated nitric oxide activity in control vs. diabetic rat aorta. Pharmacologist, 1997; 39: 52
- Pieper GM, Siebeneich W: Use of a nitronyl nitroxide to discriminate the contribution of nitric oxide radical in endothelium-dependent relaxation of control and diabetic blood vessels. J Pharmacol Exp Ther, 1997; 283: 138–47

- Abiru T, Watanabe Y, Kamata K et al: Decrease in endothelium-dependent relaxation and levels of cyclic nucleotides in aorta from rabbits with alloxan-induced diabetes. Res Commun Chem Pathol Pharmacol, 1990; 68: 13–25
- 60. Kanner J, Harel S, Granit R: Nitric oxide as an antioxidant. Arch Biochem Biophys, 1991; 289: 130–36
- Wink DA, Mitchell JB: Chemical biology of nitric oxide: Insights into regulatory, cytotoxic, and cytoprotective mechanisms of nitric oxide. Free Radic Biol Med, 1998; 25: 434–56
- Stefano GB, Kream RM: Reciprocal regulation of cellular nitric oxide formation by nitric oxide synthase and nitrite reductases. Med Sci Monit, 2011; 17(10): RA221–26
- 63. Hogg N, Kalyanaraman B: Nitric oxide and lipid peroxidation. Biochim Biophysi Acta, 1999; 1411: 378–84
- Arstall MA, Bailey C, Gross WL et al: Reversible S-nitrosation of creatine kinase by nitric oxide in adult rat ventricular myocytes. J Mol Cell Cardiol, 1998; 30: 979–88
- Kaasik A, Minajeva A, De Sousa E et al: Nitric oxide inhibits cardiac energy production via inhibition of mitochondrial creatine kinase. FEBS Letters, 1999; 444: 75–77
- 66. Molina y Vedia L, McDonald B, Reep B et al: Nitric oxide-induced S-nitrosylation of glyceraldehyde-3-phosphate dehydrogenase inhibits enzymatic activity and increases endogenous ADP-ribosylation [published erratum appears in J Biol Chem, 1993; 268(4): 3016]. J Biol Chem, 1992; 267: 24929–32
- Mohr S, Stamler JS, Brune B: Posttranslational modification of glyceraldehyde-3-phosphate dehydrogenase by S-nitrosylation and subsequent NADH attachment. J Biol Chem, 1996; 271: 4209–14
- Mohr S, Hallak H, de Boitte A et al: Nitric oxide-induced S-glutathionylation and inactivation of glyceraldehyde-3-phosphate dehydrogenase. J Biol Chem, 1999; 274: 9427–30
- Schuppe-Koistinen I, Moldeus P, Bergman T, Cotgreave IA: S-thiolation of human endothelial cell glyceraldehyde-3-phosphate dehydrogenase after hydrogen peroxide treatment. Eur J Biochem, 1994; 221: 1033–37
- Frenzel J, Richter J, Eschrich K: Pyruvate protects glucose-deprived Muller cells from nitric oxide-induced oxidative stress by radical scavenging. Glia, 2005; 52: 276–88
- Harrison CB, Drummond GR, Sobey CG, Selemidis S: Evidence that nitric oxide inhibits vascular inflammation and superoxide production via a p47phox-dependent mechanism in mice. Clin Exp Pharmacol Physiol, 2010; 37: 429–34
- Duncan ER, Crossey PA, Walker S et al: Effect of endothelium-specific insulin resistance on endothelial function *in vivo*. Diabetes, 2008; 57: 3307–14
- Bitar MS, Wahid S, Mustafa S et al: Nitric oxide dynamics and endothelial dysfunction in type 2 model of genetic diabetes. Eur J Pharmacol, 2005; 511: 53–64
- Jourd'Heuil D, Mills L, Miles AM, Grisham MB: Effect of nitric oxide on hemoprotein-catalyzed oxidative reactions. Nitric Oxide, 1998; 2: 37–44
- Van DS, Desmet F: The power of using continuous-wave and pulsed electron paramagnetic resonance methods for the structure analysis of ferric forms and nitric oxide-ligated ferrous forms of globins. Methods Enzymol, 2008; 437: 287–310
- Rapoport RM: Acute nitric oxide synthase inhibition and endothelin-1-dependent arterial pressure elevation. Front Pharmacol, 2014; 5: 57
- 77. Murad F: Nitric oxide signaling: would you believe that a simple free radical could be a second messenger, autacoid, paracrine substance, neurotransmitter, and hormone? Recent Prog Horm Res, 1998; 53: 43–59
- Denninger JW, Marletta MA: Guanylate cyclase and the NO/cGMP signaling pathway. Biochim Biophys Acta, 1999; 1411: 334–50
- Salvemini D: Regulation of cyclooxygenase enzymes by nitric oxide. Cell Mol Life Sci, 1997; 53: 576–82
- Grisham MB, Granger DN, Lefer DJ: Modulation of leukocyte-endothelial interactions by reactive metabolites of oxygen and nitrogen: relevance to ischemic heart disease. Free Radic Biol Med, 1998; 25: 404–33
- Datta PK, Dhupar S, Lianos EA: Regulatory effects of inducible nitric oxide synthase on cyclooxygenase-2 and heme oxygenase-1 expression in experimental glomerulonephritis. Nephrol Dial Transplant, 2006; 21: 51–57
- Leplingard A, Brung-Lefebvre M, Guedon C et al: Increase in cyclooxygenase-2 and nitric oxide-synthase-2 mRNAs in pouchitis without modification of inducible isoenzyme heme-oxygenase-1. Am J Gastroenterol, 2001; 96: 2129–36

- 83. Pi SH, Jeong GS, Oh HW et al: Heme oxygenase-1 mediates nicotine- and lipopolysaccharide-induced expression of cyclooxygenase-2 and inducible nitric oxide synthase in human periodontal ligament cells. J Periodontal Res, 2010; 45: 177–83
- Brown GC: Nitric oxide and mitochondrial respiration. Biochim Biophys Acta, 1999; 1411(2–3): 351–69
- Brown GC, Cooper CE: Nanomolar concentrations of nitric oxide reversibly inhibit synaptosomal respiration by competing with oxygen at cytochrome oxidase. FEBS Letters, 1994; 356: 295–98
- Cleeter MW, Cooper JM, Darley-Usmar VM et al: Reversible inhibition of cytochrome c oxidase, the terminal enzyme of the mitochondrial respiratory chain, by nitric oxide. Implications for neurodegenerative diseases. FEBS Letters, 1994; 345: 50–54
- Schweizer M, Richter C: Nitric oxide potently and reversibly deenergizes mitochondria at low oxygen tension. Biochem Biophys Res Commun, 1994; 204: 169–75
- Takehara Y, Kanno T, Yoshioka T et al: Oxygen-dependent regulation of mitochondrial energy metabolism by nitric oxide. Arch Biochem Biophys, 1995; 323: 27–32
- Nishikawa M, Sato EF, Kuroki T, Inoue M: Role of glutathione and nitric oxide in the energy metabolism of rat liver mitochondria. FEBS Letters, 1997; 415: 341–45
- Nishida CR, Ortiz de Montellano PR: Autoinhibition of endothelial nitric-oxide synthase. Identification of an electron transfer control element. J Biol Chem, 1999; 274: 14692–98
- Giuffre A, Barone MC, Brunori M et al: Nitric oxide reacts with the singleelectron reduced active site of cytochrome c oxidase. J Biol Chem, 2002; 277: 22402–6
- 92. Moncada S, Higgs EA: Nitric oxide and the vascular endothelium. Handb Exp Pharmacol, 2006; 213–54
- Erusalimsky JD, Moncada S: Nitric oxide and mitochondrial signaling: from physiology to pathophysiology. Arterioscler Thromb Vasc Biol, 2007; 27: 2524–31
- 94. Shen W, Hintze TH, Wolin MS: Nitric oxide. An important signaling mechanism between vascular endothelium and parenchymal cells in the regulation of oxygen consumption. Circulation, 1995; 92: 3505–12
- 95. Shen W, Xu X, Ochoa M et al: Role of nitric oxide in the regulation of oxygen consumption in conscious dogs. Circ Res, 1994; 75: 1086–95
- 96. Laycock SK, Vogel T, Forfia PR et al: Role of nitric oxide in the control of renal oxygen consumption and the regulation of chemical work in the kidney. Circ Res, 1998; 82: 1263–71
- 97. King CE, Melinyshyn MJ, Mewburn JD et al: Canine hindlimb blood flow and O2 uptake after inhibition of EDRF/NO synthesis. J Appl Physiol, 1994; 76: 1166–71
- Ishibashi Y, Duncker DJ, Zhang J, Bache RJ: ATP-sensitive K+ channels, adenosine, and nitric oxide-mediated mechanisms account for coronary vasodilation during exercise. Circ Res, 1998; 82: 346–59
- 99. Victor VM, Nunez C, D'Ocon P et al: Regulation of oxygen distribution in tissues by endothelial nitric oxide. Circ Res, 2009; 104: 1178–83
- 100. Weksler-Zangen S, Aharon-Hananel G, Mantzur C et al: IL-1beta hampers glucose-stimulated insulin secretion in Cohen diabetic rat islets through mitochondrial cytochrome c oxidase inhibition by nitric oxide. Am J Physiol Endocrinol Metab, 2014; 306: E648–57
- 101. Viswambharan H, Sukumar P, Sengupta A et al: Increasing insulin sensitivity in the endothelium leads to reduced nitric oxide bioavailability. Heart, 2014; 100(Suppl.3): A98
- 102. Beckman JS, Koppenol WH: Nitric oxide, superoxide, and peroxynitrite: the good, the bad, and ugly. Am J Physiol, 1996; 271: C1424–37
- 103. Valko M, Morris H, Cronin MT: Metals, toxicity and oxidative stress. Curr Med Chem, 2005; 12: 1161–208
- Poderoso JJ: The formation of peroxynitrite in the applied physiology of mitochondrial nitric oxide. Arch Biochem Biophys, 2009; 484: 214–20
- 105. Samhan-Arias AK, Gutierrez-Merino C: Purified NADH-cytochrome b reductase is a novel superoxide anion source inhibited by apocynin: sensitivity to nitric oxide and peroxynitrite. Free Radic Biol Med, 2014; 73C: 174–89
- 106. Qian J, Fulton D: Post-translational regulation of endothelial nitric oxide synthase in vascular endothelium. Front Physiol, 2013; 4: 347
- 107. Kim SF: The role of nitric oxide in prostaglandin biology; update. Nitric Oxide, 2011; 25: 255–64

- Goodrich LE, Paulat F, Praneeth VK, Lehnert N: Electronic structure of hemenitrosyls and its significance for nitric oxide reactivity, sensing, transport, and toxicity in biological systems. Inorg Chem, 2010; 49: 6293–316
- Rubbo H, O'Donnell V: Nitric oxide, peroxynitrite and lipoxygenase in atherogenesis: mechanistic insights. Toxicology, 2005; 208: 305–17
- 110. Veihelmann A, Brill T, Blobner M et al: Inhibition of nitric oxide synthesis improves detoxication in inflammatory liver dysfunction *in vivo*. Am J Physiol, 1997; 273: G530–36
- 111. Takemura S, Minamiyama Y, Imaoka S et al: Hepatic cytochrome P450 is directly inactivated by nitric oxide, not by inflammatory cytokines, in the early phase of endotoxemia. J Hepatol, 1999; 30: 1035–44
- 112. Omura T: Structural diversity of cytochrome P450 enzyme system. J Biochem, 2010; 147: 297–306
- 113. Brown GC: Reversible binding and inhibition of catalase by nitric oxide. Eur J Biochem, 1995; 232: 188–91
- 114. Cai H: Hydrogen peroxide regulation of endothelial function: origins, mechanisms, and consequences. Cardiovasc Res, 2005; 68: 26–36
- 115. Sivitz WI, Yorek MA: Mitochondrial dysfunction in diabetes: from molecular mechanisms to functional significance and therapeutic opportunities. Antioxid Redox Signal, 2010; 12: 537–77
- 116. Park Y, Capobianco S, Gao X et al: Role of EDHF in type 2 diabetes-induced endothelial dysfunction. Am J Physiol Heart Circ Physiol, 2008; 295: H1982–88
- 117. Toledo JC Jr, Augusto O: Connecting the chemical and biological properties of nitric oxide. Chem Res Toxicol, 2012; 25: 975–89
- Nordquist L, Stridh S: Effects of proinsulin C-peptide on oxygen transport, uptake and utilization in insulinopenic diabetic subjects – a review. Adv Exp Med Biol, 2009; 645: 193–98
- 119. Bates TE, Loesch A, Burnstock G, Clark JB: Mitochondrial nitric oxide synthase: a ubiquitous regulator of oxidative phosphorylation? Biochem Biophys Res Commun, 1996; 218: 40–44
- 120. Finocchietto PV, Franco MC, Holod S et al: Mitochondrial nitric oxide synthase: a masterpiece of metabolic adaptation, cell growth, transformation, and death. Exp Biol Med (Maywood), 2009; 234: 1020–28
- 121. Zaobornyj T, Ghafourifar P: Strategic localization of heart mitochondrial NOS: a review of the evidence. Am J Physiol Heart Circ Physiol, 2012; 303: H1283–93
- 122. de la Torre JC, Stefano GB: Evidence that Alzheimer's disease is a microvascular disorder: The role of constitutive nitric oxide. Brain Res Rev, 2000; 34: 119–36
- 123. de la Torre JC: Cerebromicrovascular pathology in Alzheimer's disease compared to normal aging. Gerontology, 1997; 43: 26–43
- 124. de la Torre JC: Hemodynamic consequences of deformed microvessels in the brain in Alzheimer's disease. Ann NY Acad Sci, 1997; 826: 75–91
- 125. de la Torre JC, Cada A, Nelson N et al: Reduced cytochrome oxidase and memory dysfunction after chronic brain ischemia in aged rats. Neurosci Lett, 1997; 223: 165–68
- 126. de la Torre JC, Mussivand T: Can disturbed brain microcirculation cause Alzheimer's disease? Neurol Res, 1993; 15: 146–53
- 127. de la Torre JC, Butler K, Kozlowski P et al: Correlates between nuclear magnetic resonance spectroscopy, diffusion weighted imaging, and CA1 morphometry following chronic brain ischemia. J Neurosci Res, 1995; 41: 238–45
- 128. de la Torre JC: Critical threshold cerebral hypoperfusion causes Alzheimer's disease? Acta Neuropathol, 1999; 98: 1–8
- 129. de la Torre JC: Cerebrovascular changes in the aging brain. Adv Cell Aging Gerontol, 1997; 2: 77–107
- de la Torre JC, Fortin T, Park GA et al: Aged but not young rats develop metabolic, memory deficits after chronic brain ischaemia. Neurol Res, 1992; 14: 177–80
- 131. de la Torre JC, Fortin T, Park GA et al: Chronic cerebrovascular insufficiency induces dementia-like deficits in aged rats. Brain Res, 1992; 582: 186–95
- 132. de la Torre JC: ls Alzheimer's disease a neurodegenerative or a vascular disorder? Data, dogma, and dialectics. Lancet Neurol, 2004; 3: 184–90
- 133. de la Torre JC. Alzheimer's disease is a vasocognopathy: a new term to describe its nature. Neurol Res, 2004; 26: 517–24
- Price JM, Chi X, Hellermann G, Sutton ET: Physiological levels of beta-amyloid induce cerebral vessel dysfunction and reduce endothelial nitric oxide production. Neurol Res, 2001; 23: 506–12

- 135. Aliev G, Priyadarshini M, Reddy VP et al: Oxidative stress mediated mitochondrial and vascular lesions as markers in the pathogenesis of Alzheimer disease. Curr Med Chem, 2014; 21: 2208–17
- 136. Orsucci D, Mancuso M, Ienco EC et al: Vascular factors and mitochondrial dysfunction: a central role in the pathogenesis of Alzheimer's disease. Curr Neurovasc Res, 2013; 10: 76–80
- 137. Grammas P: Neurovascular dysfunction, inflammation and endothelial activation: implications for the pathogenesis of Alzheimer's disease. J Neuroinflammation, 2011; 8: 26
- 138. Humpel C: Chronic mild cerebrovascular dysfunction as a cause for Alzheimer's disease? Exp Gerontol, 2011; 46: 225-32.
- 139. Aliev G, Palacios HH, Lipsitt AE et al. Nitric oxide as an initiator of brain lesions during the development of Alzheimer disease. Neurotox Res, 2009; 16: 293–305
- 140. Belkhelfa M, Rafa H, Medjeber O et al: IFN-gamma and TNF-alpha Are Involved During Alzheimer Disease Progression and Correlate with Nitric Oxide Production: A Study in Algerian Patients. J Interferon Cytokine Res, 2014 [Epub ahead of print]
- 141. Thal DR, Ghebremedhin E, Orantes M, Wiestler OD: Vascular pathology in Alzheimer disease: correlation of cerebral amyloid angiopathy and arteriosclerosis/lipohyalinosis with cognitive decline. J Neuropathol Exp Neurol, 2003; 62: 1287–301
- 142. Aliev G, Li Y, Palacios HH, Obrenovich ME: Oxidative stress induced mitochondrial DNA deletion as a hallmark for the drug development in the context of the cerebrovascular diseases. Recent Pat Cardiovasc Drug Discov, 2011; 6: 222–41

- 143. Grammas P, Martinez J, Sanchez A et al: A new paradigm for the treatment of Alzheimer's disease: targeting vascular activation. J Alzheimers Dis, 2014; 40: 619–30
- 144. Taguchi A: Vascular factors in diabetes and Alzheimer's disease. J Alzheimers Dis, 2009; 16: 859–64
- 145. Takeuchi M, Yamagishi S: Involvement of toxic AGEs (TAGE) in the pathogenesis of diabetic vascular complications and Alzheimer's disease. J Alzheimers Dis, 2009; 16: 845–58
- 146. Carvalho C, Katz PS, Dutta S et al: Increased susceptibility to amyloid-beta toxicity in rat brain microvascular endothelial cells under hyperglycemic conditions. J Alzheimers Dis, 2014; 38: 75–83
- 147. Valente T, Gella A, Sole M et al: Immunohistochemical study of semicarbazide-sensitive amine oxidase/vascular adhesion protein-1 in the hippocampal vasculature: pathological synergy of Alzheimer's disease and diabetes mellitus. J Neurosci Res, 2012; 90: 1989–96
- 148. Carvalho C, Machado N, Mota PC et al: Type 2 diabetic and Alzheimer's disease mice present similar behavioral, cognitive, and vascular anomalies. J Alzheimers Dis, 2013; 35: 623–35
- 149. Burdo JR, Chen Q, Calcutt NA, Schubert D: The pathological interaction between diabetes and presymptomatic Alzheimer's disease. Neurobiol Aging, 2009; 30: 1910–17
- 150. de la Torre JC, Pappas BA, Prevot V et al: Hippocampal nitric oxide upregulation precedes memory loss and A beta I-40 accumulation after chronic brain hypoperfusion in rats. Neurol Res, 2003; 25: 635–41
- 151. Hayden MR, Banks WA, Shah GN et al: Cardiorenal metabolic syndrome and diabetic cognopathy. Cardiorenal Med, 2013; 3: 265–82