

# A Third Measure-Metastable State in the Dynamics of Spontaneous Shape Change in Healthy Human's White Cells

Karen A. Selz<sup>1,2\*</sup>

**1** Fetzter Franklin Laboratory of the Cielo Institute, Asheville, North Carolina, United States of America, **2** Emory University School of Medicine, Atlanta, Georgia, United States of America

## Abstract

Human polymorphonuclear leucocytes, PMN, are highly motile cells with average 12-15  $\mu\text{m}$  diameters and prominent, loboid nuclei. They are produced in the bone marrow, are essential for host defense, and are the most populous of white blood cell types. PMN also participate in acute and chronic inflammatory processes, in the regulation of the immune response, in angiogenesis, and interact with tumors. To accommodate these varied functions, their behavior is adaptive, but still definable in terms of a set of behavioral states. PMN morphodynamics have generally involved a non-equilibrium stationary, spheroid *Idling* state that transitions to an activated, ellipsoid translocating state in response to chemical signals. These two behavioral shape-states, spheroid and ellipsoid, are generally recognized as making up the vocabulary of a healthy PMN. A third, "random" state has occasionally been reported as associated with disease states. I have observed this third, *Treadmilling* state, in PMN from healthy subjects, the cells demonstrating metastable dynamical behaviors known to anticipate phase transitions in mathematical, physical, and biological systems. For this study, human PMN were microscopically imaged and analyzed as single living cells. I used a microscope with a novel high aperture, cardioid annular condenser with better than 100 nanometer resolution of simultaneous, mixed dark field and intrinsic fluorescent images to record shape changes in 189 living PMNs. Relative radial roundness,  $R(t)$ , served as a computable order parameter. Comparison of  $R(t)$  series of 10 cells in the *Idling* and 10 in the *Treadmilling* state reveals the robustness of the "random" appearing *Treadmilling* state, and the emergence of behaviors observed in the neighborhood of global state transitions, including increased correlation length and variance (divergence), sudden jumps, mixed phases, bimodality, power spectral scaling and temporal slowing. Wavelet transformation of an  $R(t)$  series of an *Idling* to *Treadmilling* state change, demonstrated behaviors concomitant with the observed transition.

**Citation:** Selz KA (2011) A Third Measure-Metastable State in the Dynamics of Spontaneous Shape Change in Healthy Human's White Cells. PLoS Comput Biol 7(4): e1001117. doi:10.1371/journal.pcbi.1001117

**Editor:** Michael Shlesinger, Physical Sciences Division, Office of Naval Research, United States of America

**Received:** July 9, 2010; **Accepted:** March 4, 2011; **Published:** April 7, 2011

**Copyright:** © 2011 Karen A. Selz. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

**Funding:** The Fetzter Foundation supported this work. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

**Competing Interests:** The author has declared that no competing interests exist.

\* E-mail: selz@cieloinstitute.org

## Introduction

Polymorphonuclear neutrophil granulocytes (PMNs) are the body's most abundant class of white blood cells. At circulating cell levels of  $\sim 10^{11}$ , PMNs make up about 62% of all human white cells [1]. An idling, non-activated neutrophil circulates in the blood for 8 to 12 hours before undergoing apoptosis. When stimulated by inflammatory cytokine signals released by endothelial, mast cells and/or macrophages, PMNs can survive for 24 to 48 hours [2–4].

Once activated, idling PMNs exploit shape change dynamics as they tether to the endothelium of post-capillary venules, making and breaking bonds while rolling along the venule's endothelial bed of selectins. Following concentration gradients of inflammatory ligands, they wriggle through the vessel wall and migrate to the site of the initiating inflammation. The motions accompanying this migration exploit the PMN's cytoskeletal, actin polymerization-depolymerization cycle that configures the dynamics of shape change and translation [5,6].

The behavioral dynamics of two of the shape changing metastable states and their associated translational motions are

well established. PMNs manifest these metastable shape-motional states *in vivo* and *in vitro*. They are: (1) The circular-spherical, *Idling* state manifesting standing waves and fast and fine random fluctuations in the leucocyte's apron edge; (2) The activated, ellipsoid, polarized migrating state, with almost exclusively positive gradient-directed lamellopodia and filopodia formation [7,8].

The complexity of the scenario described above and the necessity of as many as 100 distinct protein/protein interactions to coordinate the actin cytoskeletal apparatus alone, prompts both a phenomenological approach and the consideration of a potentially larger set of shape-motional states which may be transitional, non-stationary and not easily quantified.

In addition to the metastable spherical-round, *Idling* and elliptical-migratory state, I have found a statistically prominent, third, measure metastable state in the PMNs from the fresh blood of healthy human subjects. This non-translating, high amplitude, shape changing state has been previously observed and interpreted as a functionally disordered manifestation of immunological and hematological pathology [9–14]. The goals of these studies are to identify, quantitatively characterize and discriminate this third

## Author Summary

Human white blood cells, polymorphonuclear leucocytes (PMN), were microscopically imaged and analyzed as single living cells. PMN are generally observed in a spheroid *Idling* state transitioning to an activated, egg-shaped, translocating state when triggered by the body's signals of infection or inflammation. Occasionally, PMN are observed in a third behavioral state that looks like dancing in place, with protrusions thrown out and retracted, sometimes several simultaneously, in apparently random directions. This behavior previously had been thought to be associated with disease. Here this third state, that I call *Treadmilling*, is a relatively common way that PMN from healthy people get "stuck" in an intermediate phase. Relative radial roundness,  $R(t)$ , served as a computable order parameter, and time series of  $R(t)$  were derived from microscopic image series of each of 189 PMN. Only  $R(t)$  series from cells that stayed healthy, maintained a single behavioral state and did not have contact with other bodies for the 30 min recording period were analyzed further. Comparison of measures made on the  $R(t)$  series of cells in the *Idling* versus *Treadmilling* states quantitatively distinguish states and suggest behavior in the vicinity of global state transitions. Wavelet transformation of an  $R(t)$  series of a captured state change supports this finding.

shape-motional PMN state from the other two. I call this third dynamical state, *Treadmilling*, the word used to also describe a key part of the underlying F,G-actin dynamics [15]. I have observed the *Treadmilling* state in 12 of the 18 healthy, adult volunteers contributing blood for this study.

## A Statistical Concern

We remind ourselves that there are neurobiological limits on the both the resolution afforded by empirically meaningful partitions [16], and the minimal length of a neurobiologically defensible time series [17]. In these studies I must be concerned with how long the cells being observed maintain their anatomical and functional integrity and within that viability limit, how many observations I can make without the vacuous artifice of over-sampling.

The issue more generally is the selection of an appropriate sample size of an intrinsically non-stationary system. Counter-intuitively, it has been shown that under certain conditions of limited information, repeated too-short sample lengths come to be computationally superior globally [18].

In the past I have dealt with this problem by studying repeated time series derived measures yielding populations of not necessarily convergent estimates [19] with, nonetheless, distributional properties of the measures, such that I can estimate each measure's central and higher moments, range of variation and statistical differences between measures in comparisons of varying observed system state [20]. This is the approach to be taken here.

## Methods

### Relative Radial Roundness, $R(t)$ , as an Order Parameter

It is difficult to find a global quantitative measure on the dynamics of emergent phenomenon with the nice properties of additivity, continuity and differentiability [21,22]. Such a measure has been called an *order parameter*, named for its use in tracking a system's dynamics through transitions in the system's degree of order. In gas-liquid transitions, the order parameter is density [23], while in ferromagnet transitions, for instance, it is net magnetization [23,24]. Perhaps the best known example of an order

parameter is relative phase, the Landau-Ginsberg order parameter [23,24], used in the phenomenological description of thermodynamic and superconducting transitions [25].

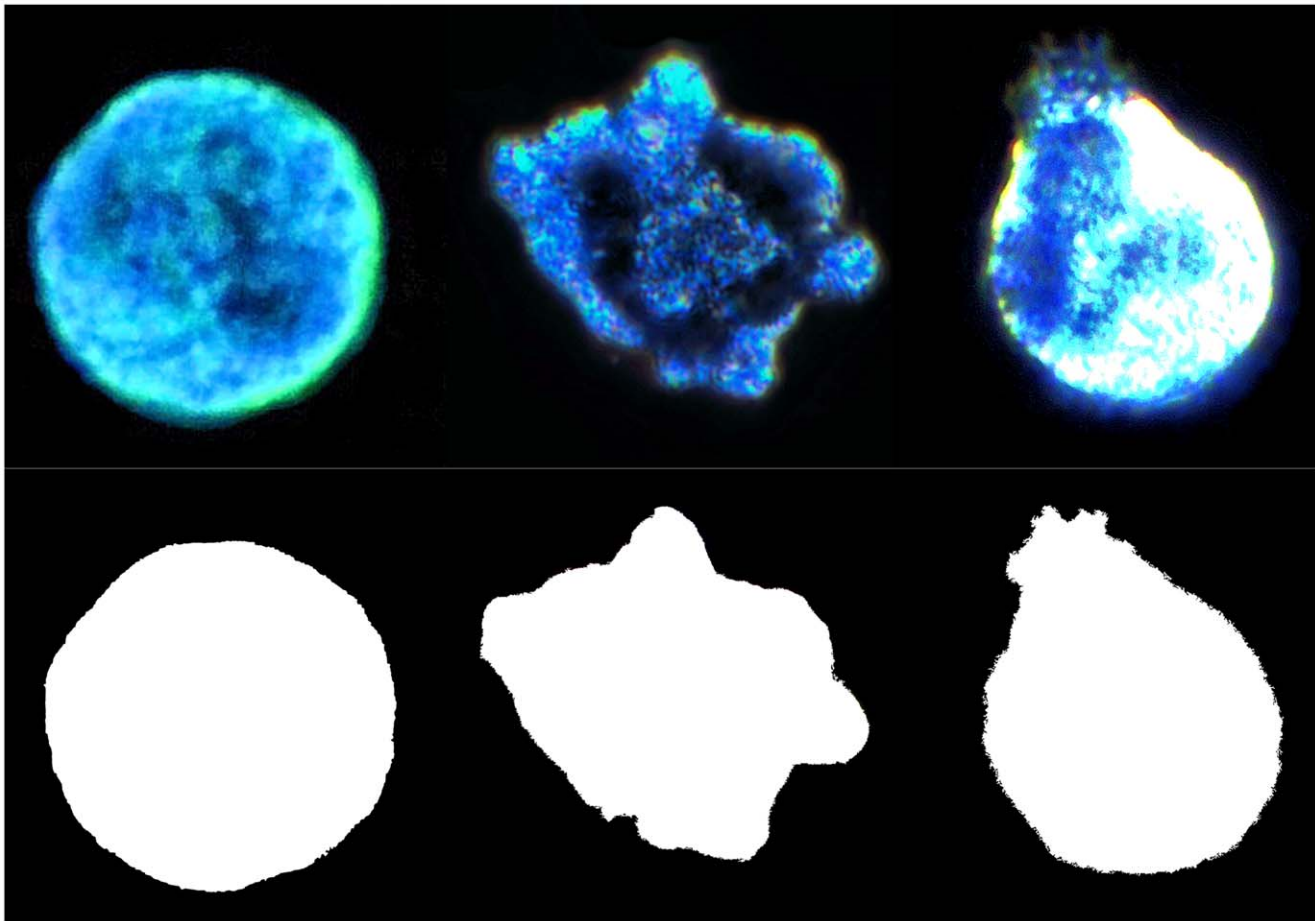
I have examined the autonomous, time-dependent shape changes of individual *Idling* and *Treadmilling* human white cells as real, spatially extended dynamical systems. I use a global measure on the PMN's *relative radial roundness*,  $R(t)$ , as the order parameter.  $R(t) = r_1/r_2$  is computed as the ratio of the radius of the cell assumed to be an ideal circle,  $r_1 = p/2\pi$  in which  $p$  is the sum of the pixels outlining the cell's *perimeter* and  $r_2 = (A/\pi)^{0.5}$  using the sum of the pixels within the cell's silhouette as the cell's *area*,  $A$ .  $R(t) = r_1(t)/r_2(t) = \{p/2\pi/(A/\pi)^{0.50}\}(t)$  computed at each time step. If both  $r_1$  and  $r_2$  were derived from an abstract, idealized circle,  $R(t) = r_1/r_2 = 1.0$ , such that  $\log\{R(t) = r_1/r_2\} = 0$ , the characteristic lower limit of a generic order parameter. Deviations from this reference characterize changes in state [21–23,25]. My use of the global order parameter,  $R(t)$ , contrasts with a previous use of an averaged local measure, the power spectral transformation of a sequence of angles resulting from the piece wise linear segmentation of the cell's circumference [26]. The use of  $R(t)$  more closely resembles a differential geometric pattern map [27].

## Experimental Procedures

One hundred and eighty-nine PMNs from fifty-three peripheral blood samples were collected from 18 healthy adult volunteers, aged 26 to 72. The blood samples were allowed to sediment gravitationally for 40 minutes at room temperature. A population of PMNs (and other white cells and platelets) were removed from the buffy coat by micropipette and, along with associated plasma, placed within a 12 mm ring painted on a glass slide, forming a  $\sim 20$ – $25$   $\mu\text{m}$  deep well, compared to the average  $5.7$   $\mu\text{m}$  vertical space between a plain slide and its cover slip [28]. The  $5.7$   $\mu\text{m}$  gap of standard slides and coverslips is considerably smaller than the average diameter of PMNs, leading to some mechanical compression of the cell contributing to their activation, and allowing the cell to move along the slide substrate and cover slip simultaneously [28]. The slides used in this study do not suffer from these deficits.

PMN autonomous motions were observed using an Olympus BX41 microscope fitted with CytoViva dark field and fluorescent optical illumination systems, which includes a unique, high-aperture, cardioid annular condenser (www.scitech.com.au). The CytoViva condenser makes it possible to visualize objects of below  $100$  nm in diameter in real time, and with the cellular samples in an unfixed, living, active state [29,30]. Because PMNs were treated gently, avoiding perturbations of column separation and elution, it became possible to reliably study a PMN continuously for 30 to 60 minutes before the onset of granular clumping, membrane blebbing and other signs of nascent apoptosis [31]. Data collection continued until ten each *Idling* and *Treadmilling* cells met the conditions for inclusion in the study. Specifically, only cells that maintained healthy, one state behavior, and did not have contact with any extracellular objects for the entire 30 min recording period were retained for analysis.

*Idling* PMNs are characterized by their near spheroid shape (quasi-circular in two dimensions). In this state, the microscopically visible autonomous motions are limited to standing waves on the cell surface and low amplitude fluctuations of the cell's microvillus border. In contrast, *Treadmilling* PMNs demonstrate large and irregular changes in cell shape. Multiple transient, often simultaneously appearing, pseudopodia and lamellopodia emerge from the cell surface, oriented apparently randomly and without significant movement of the PMNs center of mass. The cell's movement was less than 1.5 times the maximum diameter of the cell over the typical  $\sim 30$  minute recording sessions. **Figure 1**



**Figure 1. Representative binary color coded silhouettes of PMNs in round *Idling*, *Treadmilling* and elliptically polarized/translocating shape-states.**

doi:10.1371/journal.pcbi.1001117.g001

portrays binary color coded, characteristic silhouettes of the round *Idling*, *Treadmilling* and elliptically polarized/translocating shape-states of PMNs. Only videos micrographs of mature, segmented neutrophils that did not have contact with any cells or extracellular entities and remained visibly healthy over the thirty minutes of observation, in addition to manifesting stable state behavior were retained for further analysis.

Images were collected every 2 sec for 30 min using an Optronics Microfire 1200×1600 CCD array camera [30] resulting in a 900 point  $R(t)$  series of high resolution images per PMN. The slowness of the cell shape changing motions led to the finding that more frequent sampling within the time limit of cellular integrity was obviously redundant. In the geometric computations, each primary image was used to produce two binary, 0,1, digital daughter images: an area map of  $A$ , and perimeter map of  $p$ . The 0,1 coding of the pixels of the two daughter images were converted into binary arrays and used compute the  $R(t)$  time series.

### Computation of the Measures on $R(t)$

In light of the above discussion of biological constraints on sample length and the intrinsic non-stationarity of the PMNs shape motion series, statistical distributions of often individually non-convergent measures made on each of the cells, serve as the basis for comparisons of *Idling* and *Treadmilling* states. Statistical evaluations are then made on populations of possibly incomplete measures, not on the raw observations. Rules of thumb concerning sample length requirements

for any particular measure [32] though easily attainable in physical and computational systems, often ignore the intrinsic series limits and non-stationarity of real, behaving, biological systems. In addition to the use of the distribution of each particular kind of measure, I study an aggregate of several, often incomplete measures, each reflecting different aspects of the shape-motional dynamics of PMNs.

On the  $R(t)$  of each cell, I study: (1) The central moments of the  $R(t)$  distribution, the mean  $S_1$  and standard deviation,  $S_2$ , as well as

**Table 1. Measure averages for cells in each state group.**

<b>Idling (n = 10)</b>	<b>Treadmilling (n = 10)</b>	<b>t(df); <math>\rho</math></b>
Mean = 2.935	Mean = 2.932	$t_{(18)} = 0.004$ ; $\rho < 0.4983$
SD = 0.1023	SD = 0.3882	$t_{(18)} = 4.816$ ; $\rho < 0.0001$
Skew = 0.2993	Skew = 0.7273	$t_{(18)} = 2.419$ ; $\rho < 0.0132$
Kurtosis = 0.7229	Kurtosis = 1.165	$t_{(18)} = 0.652$ ; $\rho = 0.2612$
$\lambda_1 = 0.6242$	$\lambda_1 = 0.5066$	$t_{(18)} = 2.592$ ; $\rho < 0.0090$
$\alpha = -0.4561$	$\alpha = -1.0290$	$t_{(18)} = 10.600$ ; $\rho < 0.0001$

**Table 1** reports the results of measures made on the  $R(t)$  series of ten *Idling* PMN and ten *Treadmilling* PMN, all of which had remained healthy and in a single behavioral state for the 30 min recording period and made no contact with extracellular bodies during that time.

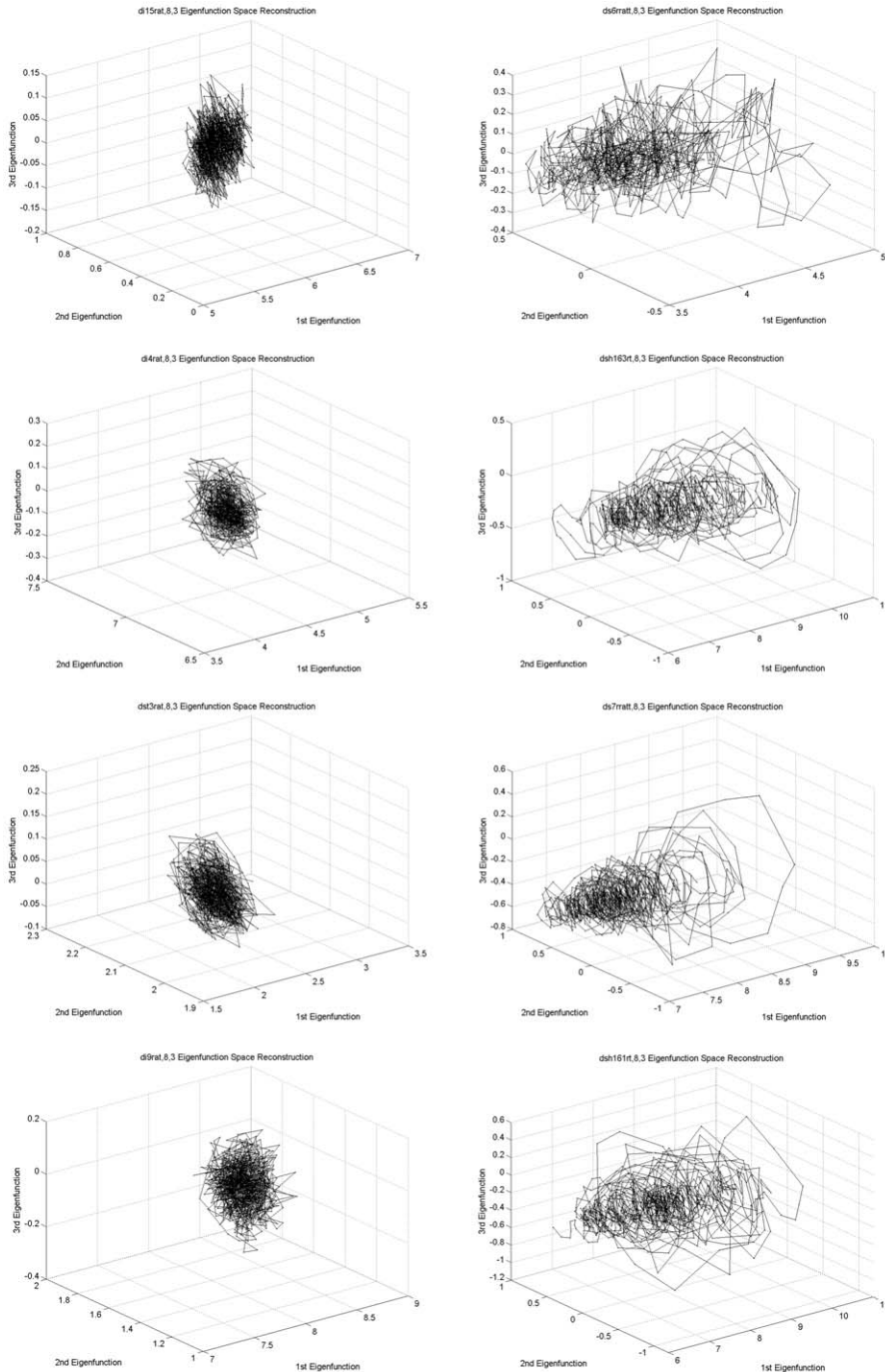
doi:10.1371/journal.pcbi.1001117.t001

the skewness,  $S_3$ , indicating the asymmetry of the density distribution of  $R(t)$ , estimated using the third moment,  $m_3$ , divided by the cubed root of the variance squared,  $S_3 = m_3 / \text{var}^{3/2}$ . The kurtosis,  $S_4$ , of  $R(t)$  is computed using the relation,  $S_4 = m_4 / \text{variance}^2 - 3$  [33]; (2) An estimate of the  $R(t)$ 's orbital divergence, its sensitivity to initial conditions, in a three dimensional embedding space, was computed using a generic algorithm for the leading Lyapounov exponent,  $\Lambda_1$  [34]; (3) Differences in a hierarchical scaling property of  $R(t)$ , by computing the scaling exponent  $\alpha$  derived from its power (frequency)spectrum, as the slope of the

middle third of the linear best fit of the log power-log frequency relation [35]; (4) An example of the time dependence of the scaling of  $R(t)$  was estimated from a Morlet continuous wavelet transformation using standard algorithms [36–38].

### Visualizing Phase Space Behavior of $R(t)$

To visualize the phase space behavior I used relatively denoised, three dimensional Broomhead-King, B/K, eigenfunction,  $\Psi_i$  embedding of the  $R(t)$ s. To do this, I computed and plotted  $\Psi_1$ ,  $\Psi_2$ , and  $\Psi_3$  with respect to each other [39,40]. Each  $R(t)$  series



**Figure 2. Shows the eigenfunction space embedding (see text) for four representative Idling cells (left column) and four Treadmilling cells (right column).**

doi:10.1371/journal.pcbi.1001117.g002

generated an  $M$ -lagged data matrix on which an  $M \times M$  Hermitian autocovariance,  $C_M$ , matrix was computed, with  $M=8$ , a typical correlation decay interval.  $C_M$  was then decomposed into its eigenvalue-ordered eigenvectors. The eigenvectors associated with the three largest eigenvalues were each composed with the original  $R(t)$  series to form B/K eigenfunctions  $\Psi_1$ ,  $\Psi_2$ , and  $\Psi_3$ . These formed the axes of the B/K eigenspace reconstruction. Because  $R(t)$  behavior attributable to the lower, excluded, eigenvectors accounts for the trivial, “noise” component of the variance, the resulting *eigenfunction space embedding* (each successive point being a triple) is relatively denoised compared with the more commonly used phase delay space construction [41].

Another graphical representation of the orbital behavior of  $R(t)$  is its two dimensional,  $i = \tau_1, j = \tau_2$ , Recurrence Plot,  $RP[R(t)]_{i,j}$ , introduced by Eckmann [42]. Graphical representations of  $RP[R(t)]_{i,j}$  are two dimensional lattices, each point computed as  $RP_{i,j} = \Theta(\varepsilon \|\vec{x}_i - \vec{x}_j\|)$ ,  $i, j = 1 \dots N$ , where  $\vec{x}_i$  in  $R^2$  represents the location of the orbit in phase space at time  $i$ .  $\varepsilon$  is the static distance defining the “closeness” threshold, and  $\Theta$  is the Heavyside function. The resulting binary series, each point  $\varepsilon$ - close or not to the previous value, is coded in black and white. Here, a standard time delay three dimensional embedding was used, with delay  $\tau = 1$  [43]. If  $\vec{x}_j$  falls within the distance  $\varepsilon$  of  $\vec{x}_i$ ,  $\vec{x}_j$  is considered to be a recurrence of  $\vec{x}_i$ , otherwise not. Clustering in  $RP_{i,j}$  has been used to discriminate among three characteristic patterns of intermittency [44].

## Results

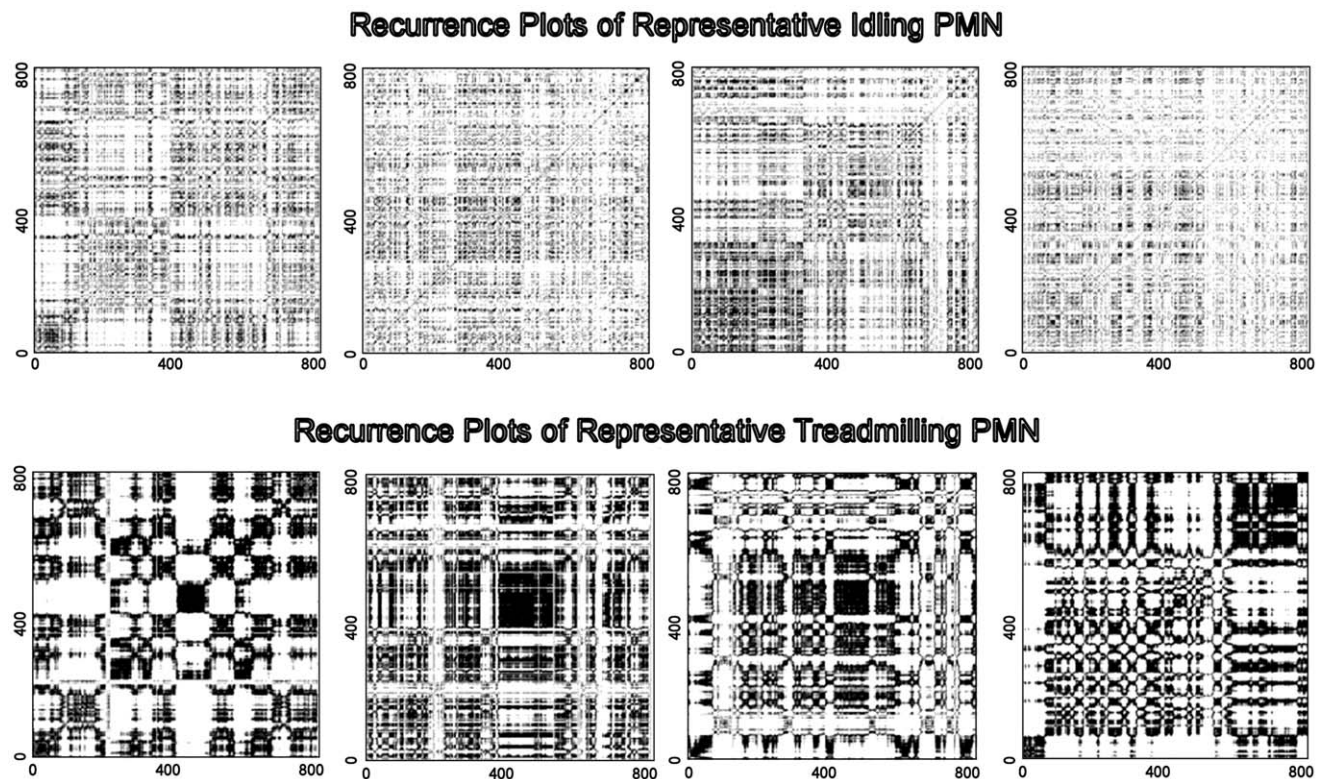
There were highly significant differences between the measures  $S_2, S_3, \Lambda_1$ , and  $\alpha$  that were made on the  $R(t)$  series of the PMNs in

the *Idling* versus the *Treadmilling* state, see **Table 1**. No significant differences were found between the two distributions of  $S_1$  or  $S_4$ .

The qualitative differences in the shape-motional patterns implied by the statistically significant differences in the measures in **Table 1** are consistent with behavior that was observed microscopically in the two pre-polarized states: (1) The small, stochastically wavy border fluctuations in cell shape of the generic *Idling* PMNs; (2) A range of large, simultaneously multiscale motions in cell shape variations of  $R(t)$  in the *Treadmilling* state. For examples, compared with *Idling*, the increase in asymmetric amplitude in *Treadmilling* is reflected in increases in  $S_2$  and  $S_3$ , and the increase in shape-motional order in *Treadmilling* is seen in the statistically significant decrease in  $\Lambda_1$ , the leading Lyapounov index of expansive, orbital mixing [45]. The larger, smoother, more correlated shape motions of the *Treadmilling* state are seen in statistically significant increases in  $\alpha$  in the *Treadmilling* versus *Idling* states. Without a significant difference in the means of  $R(t)$ , the variational measures make the discrimination between *Idling* and *Treadmilling* states.

Consistent with the differences in behavior described by direct observation and the aggregate of measures (see **Table 1**), **Figure 2** portrays the previously described  $\{\Psi_1, \Psi_2, \Psi_3\}_{1 \dots 900}$  B/K eigenfunctions embedding of four representative *Idling* cells (left column) and four *Treadmilling* cells (right column). The phase portraits of the *Idling* cells reflect symmetric, small, random fluctuations around a near stationary state. *Treadmilling* cells manifest larger, more irregular, asymmetric phase space motions which occupy almost an order of magnitude larger volume than that by the *Idling* state.

Another geometric, graphical treatment of the cell’s shape motional behavior is displayed in **Figure 3**. We see the recurrence

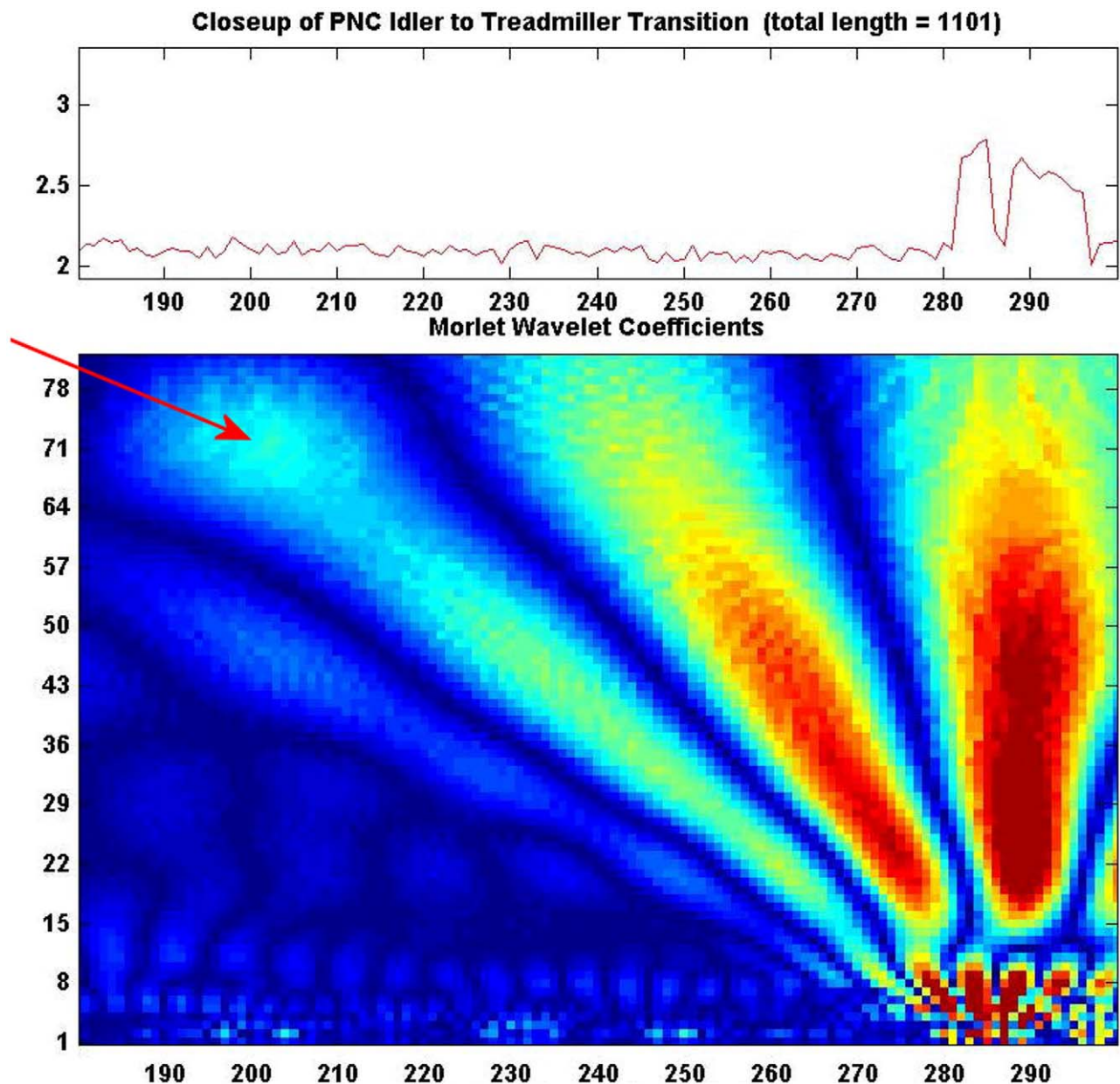


**Figure 3.** Contains recurrence plots,  $RP_{i,j}$ , of four representative PMNs in the *Idling* state (top row) and four in the *Treadmilling* state (bottom row).  $\varepsilon = 1$  for all plots.  
doi:10.1371/journal.pcbi.1001117.g003

plots,  $RP[R(t)]_{i,j}$  of the four representative PMNs in the *Idling* state (top row) and four in the *Treadmilling* state (bottom row), in which  $\varepsilon = 1$  for all plots. The  $RP[R(t)]_{i,j}$  of the *Idling* cells demonstrate more homogeneous temporal distributions of returns typical of more random data with shorter correlation lengths/relaxation times. The square patches of only lightly increased density overlaid on the more uniform surround are consistent with both the visualized small amplitude oscillations in  $R(t)$  in the *Idling* state and with the statistical results reported in **Table 1**. The  $RP[R(t)]_{i,j}$  of cells in the *Treadmilling* state are, as expected, less homogeneously distributed, manifesting clustering in the return times across multiple times scales, as well as apparent discontinuous changes in their phase space patterns. For example, short interval “bursting”

interleaved with low amplitude, long interval behavior is seen in the *Treadmilling* cells’  $RP[R(t)]_{i,j}$ . *Treadmilling* PMNs  $RP[R(t)]_{i,j}$  portraits are consistent with recurrence patterns of intermittency [44,46].

Four of the seven order parameter measures demonstrated statistically significant differences between the *Idling* and *Treadmilling* PMNs, **Table 1**. While observing and recording the real-time behavior of 189 PMNs, I witnessed many cells transitioning from one state to another among my three defined behavioral regimes. Data series including such transitions were plagued by the same complications as were the single state series (e.g., cell-cell interactions, apoptotic behavior) in addition to too short times in one or more behavioral state to allow any analysis. I was finally



**Figure 4. Shows a “close-up” of a PMN during a phase transition.** The upper panel shows the  $R(t)$  series, while the lower panel depicts the continuous 1D Morlet wavelet moduli for that time interval (x-axis). Scale of the basis function increases up the y-axis. The colormap passes through the visible spectrum; blues representing low Morlet moduli amplitudes to high valued reds. The cell is initially *Idling*, and begins *Treadmilling* at data point  $t = 283$ . Note that anticipatory slow high amplitude fluctuations begin to appear at about point  $t = 198$ .  
doi:10.1371/journal.pcbi.1001117.g004

**Table 2.** Measures made on a neutrophil before and after idling to treadmill state change.

Idling	Treadmilling
Mean = 2.0969	Mean = 2.1614
SD = 0.0348	SD = 0.1620
Skew = -0.1942	Skew = 2.0076
Kurtosis = 2.8552	Kurtosis = 8.0831
$\lambda_1 = 0.6520$	$\lambda_1 = 0.498$
$\alpha = -0.5681$	$\alpha = -1.0486$

**Table 2** reports the results of the same measures listed in **Table 1**, this time made on the  $R(t)$  series of a single PMN first in an *Idling* state and then in a *Treadmilling* state. This PMN also remained healthy for the 30 min recording period and made no contact with extracellular bodies during that time.  
doi:10.1371/journal.pcbi.1001117.t002

able to make sufficient observations portraying a single PMN shape motion transformation in real time. **Figure 4** is a Morlet wavelet graph, in continuous time along the x-axis, and scale ( $\sim$ wavelength) along the Y axis. **Figure 4** contributes evidence for a continuous transition in shape motion state, here from *Idling* to *Treadmilling*. **Table 2** lists measures before and after this single cell transition. Note that the direction and approximate magnitude of change resemble those of the population of statistically significant values in **Table 1**.

## Discussion

There are established physiological mechanisms and behavior that are consistent with both our qualitative microscopic observations and quantitative aggregate measure descriptions. PMNs are known to oscillate on multiple time and space scales, from 7 sec, 70 sec, and 260 sec membrane potential fluctuations [47] and 25 sec calcium flux oscillations [48], to the  $\sim$ 8 sec bound/unbound actin oscillations [49], to 21.6 sec and 230 sec glycolytic cycles producing NAD(P)H oscillations [47], and 10 sec and 20 sec pericellular proteolysis fluctuations [48], among many others. The  $R(t)$  series in this study evidenced scaling, broad band power spectra with multiple resonances [50]. It is likely some reported modes contribute to the cell shape fluctuations directly and others contribute to the emergence of other dynamical patterns.

The slowest Fourier mode in  $S_\omega [R(t)]$  of the *Idling* state had an average 8.457 minutes oscillation, whereas that of the slowest  $S_\omega [R(t)]$  of the *Treadmilling* state averaged a 4.201 minutes oscillation. It is interesting that these characteristic times correspond roughly to the results of studies of the characteristic remodeling times composed of actin filament diffusion, polymerization and then turnover coordinated with cellular migratory motions [51,52]. It appears that the transition from *Idling* into the intermittent *Treadmilling* regime occurs as the *Idling* state loses some of its dynamical structural stability, and its shape motion scenario becomes driven by several quasiperiodic, multi-periodic metabolic and physiological cellular oscillator mechanisms [53,54].

As listed in **Table 1** and **Table 2**, a comparison of *Idling* with *Treadmilling* PMNs reveals significantly different  $R(t)$  order parameter dynamics. Projected to a two dimensional plane (**Figure 1**), one sees an associated difference in the underlying

**Table 3.** Observed cell state transitions.

From To	Idling	Treadmilling	Translocating
Idling	X	X	X
Treadmilling	X	X	
Translocating	X		X

doi:10.1371/journal.pcbi.1001117.t003

planar geometry, with the *Idling* PMNs manifesting one centroids in their circularity, and the *Treadmilling* PMNs with two point defined, barycentric ellipses.

Many characteristics of the changes in measures in the distinct single state observations and in the computable, real-time transition from *Idling* to *Treadmilling* suggest the typical signs of a phase transition [21–23,25]. These included: (1) Increasing amplitude of  $R(t)$  variability seen in the  $S_2$  and  $S_3$  of the cell shape fluctuations; (2) Decreased leading  $\Lambda_1$  becoming less positive in the direction of zero, shadowing the leading eigenvalue of the unknown underlying partial differential equation; (3) An increase in the log-log power spectral scaling index,  $\alpha$ , reflecting a “less white” spectral pattern of  $R(t)$  fluctuations, also consistent with slowing; (4) The Morlet wavelet transformation of a continuous time  $R(t)$ , evidenced anticipatory, high amplitude slowing and a mixed phase regimes in the neighborhood of a real-time PMN shape fluctuation transition. The *eigenfunction space embedding* of the sequence of triples,  $\{\Psi_1, \Psi_2, \Psi_3\}_i$  demonstrated directly the space-time morphogenic transformation undergone by  $R(t)$  in the *Treadmilling* state with reference to that of the *Idling* cell state. Recurrence plots,  $RP_{i,j}$  depicted increased phase space clustering consistent with the more hierarchical, intermittent dynamics of the *Treadmilling* PMNs in contrast with the more randomly distributed and metrically transitive space of the *Idling*  $RP_{i,j}$ . It should be noted that the action spaces of less uniform intermittency and those of more uniform transitivity reflect common metastable alternatives in the dynamics of some biological sciences [48].

Finally, I have spent hundreds of hours microscopically tracking 189 individual PMN cells in the hopes of answering these questions about state and state transitions. While only one such transition was recorded with sufficient observations in both the *Idling* and *Treadmilling* states to allow statistical analyses, many transitions were observed. I have seen *Idling* cells transition to *Treadmilling*, and *Treadmilling* cells ball up and *Idle* (although with slightly ragged aprons). I have also observed numerous instances of *Idling* cells polarizing and *Translocating* until they reach some point at which point they *Idle* again. The only transitions that were not observed were from the *Treadmilling* to the polarized, single lamelopod, *Translocating* state or vice versa. In either case the cells ball-up briefly into an *Idling* appearance before changing again. See **Table 3**.

## Acknowledgments

I wish to thank Prof. Arnold Mandell and Dr. Paul Gailey for their helpful comments and suggestions.

## Author Contributions

Conceived and designed the experiments: KAS. Performed the experiments: KAS. Analyzed the data: KAS. Contributed reagents/materials/analysis tools: KAS. Wrote the paper: KAS. Designed and wrote computer programs used in data transformation and analysis: KAS.

## References

- Segal AW (2005) How neutrophils kill microbes. *Ann Rev Immunology* 23: 192–223.
- Underhill DM, Ozinsky A (2002) Phagocytosis of microbes, complexity in action. *Ann Rev Immunology* 20: 825–852.
- Kubes P (2002) Introduction: The complexities of leukocyte recruitment. *Semin Immunology* 14: 65–72.
- Galligan C, Yoshimura T (2003) Phenotypic and functional changes of cytokine-activated neutrophils. *Chem Immunol Allergy* 85: 25–45.
- Pollard TD (2003) The cytoskeleton, cellular motility and the reductionist agenda. *Nature* 422: 741–745.
- Pollard TD, Borisy GG (2003) Cellular motility driven by assembly and disassembly of actin filaments. *Cell* 112: 453–465.
- Xu J, Wang F, Van Keymeulen A, Herzmark P, Straight A, et al. (2003) Divergent signals and cytoskeletal assemblies regulated self-organizing polarity in neutrophils. *Cell* 114: 201–214.
- Vicente-Manzanares M, Webb DJ, Horwitz AR (2005) Cell migration at a glance. *J Cell Sci* 118: 4917–4919.
- Keller HU, Wilkinson PC, Abercrombie M, Becker EL, Hirsch JG, et al. (1977) A proposal for the definition of terms related to locomotion of leucocytes and other cells. *Clin Exp Immunol* 27: 377–380.
- Ehrengruber MU, Deranleau DA, Coates TD (1996) Shape oscillations of human neutrophil leukocytes: characterization and relationship to cell motility. *J Exp Biol* 199: 741–747.
- Korzynska A (2002) Neutrophils movement in vitro. *Ann NY Acad Sci* 972: 139–143.
- Rabson AR, Andersen RA, Glover A (1978) Defective neutrophil motility and recurrent infection. *Clin Exp Immunol* 33: 142–149.
- Lakshman R, Finn A (2001) Neutrophil disorders and their management. *J Clin Pathol* 54: 7–19.
- Dinauer MC (2006) Disorders of neutrophil function. In: M. T. Quinn, F. R. DeLeo, G. M. Bokoch, eds. *Methods in Molecular Biology*, Vol. 412. M.T. Quinn, F.R. DeLeo, G.M. Bokoch, eds. Totowa, New Jersey: Humana Press. pp 489–504.
- Hamill OP, Martinac B (2001) Molecular basis of mechanotransduction in living cells. *Physiol Rev* 81: 685–740.
- Selz KA, Mandell AJ (1991) Bernoulli partition equivalence of intermittent neuronal discharge patterns. *Int J Bifurcation Chaos* 1: 717–722.
- Selz KA, Mandell AJ (1992) Critical coherence and characteristic times in brain stem neuronal discharge patterns. In: McKenna T, Davis J, Zornetzer SF, eds. *Single Neuron Computation*. San Diego, Ca: Academic Press. pp 525–560.
- Vul E, Goodman ND, Griffiths TL, Tenenbaum JB (2009) One and done? Optimal decisions from very few samples. In: *Proceedings of the 31st Annual Conference of the Cognitive Science Society*; 148–153. 21 July–1 August 2009; Amsterdam, Netherlands. *CogSci2009*.
- Mantegna RN, Stanley JP (1994) Stochastic process with ultraslow convergence to a Gaussian: the truncated Lévy flight. *Phys Rev Lett* 73: 2946–2949.
- Selz KA, Mandell AJ, Anderson CM, Smotherman WP, Teicher M (1995) Distribution of local Mandelbrot–Hurst exponents: motor activity in cocaine treated fetal rats and manic depressive patients. *Fractals* 3: 893–904.
- Reichl LA (1980) *Modern Course in Statistical Physics*. Texas University Press: Austin. pp 367–9.
- Haken H (1977) *Synergetics; Nonequilibrium Phase Transitions and Self Organization in Physics, Chemistry and Biology*. Berlin: Springer-Verlag. pp 125–6, 274.
- Stanley ER (1971) *Introduction to Phase Transitions and Critical Phenomena*. Oxford, UK: Oxford University Press. 63 p.
- Haken H (1993) *Advanced Synergetics; Instability Hierarchies of Self-Organizing Systems and Devices*. Berlin: Springer-Verlag. pp 321–2.
- De Gennes PG (1992) *Simple Views of Condensed Matter*. Singapore: World Scientific.
- Covello R, Saccio M, Maffioni A, Pennaro M, Mitolo V (2002) Power spectral analysis of the shape of fM LP stimulated granulocytes. *Immunopharmacol Immunotoxicol* 24: 139–163.
- Dunn GA, Brown AF (1987) A unified approach to analyzing cell motility. *J Cell Biol* 8: 81–102.
- Malawista SE, deBoisfleury Cheveance A (1997) Random locomotion and chemotaxis of human blood polymorphonuclear leukocytes (PMN) in the presence of EDTA: PMN in close quarters require neither leukocyte integrins nor external divalent cations. *Proc Natl Acad Sci U S A* 94: 11577–82.
- Samoylov AM, Samoylova TI, Pustovyy OM, Samoylov AA, Toivio-Kinnunen MA, et al. (2005) Novel metal clusters isolated from blood are lethal to cancer cells. *Cells Tissues Organs* 179: 115–124.
- Vainrub A, Pustovyy O, Vodyanov V (2006) Resolution of 90 nm ( $\lambda/5$ ) in an optical transmission microscope with an annular condenser. *Optics Lett* 31: 2855–2857.
- Lodish HF (2005) *Molecular and Cell Biology*. New York: W.H. Freeman. pp 62, 84.
- Ott E (2009) *Chaos in Dynamical Systems* Cambridge University Press. pp 122–6.
- de Gennes PG (1979) *Scaling Concepts in Polymer Physics*. Cornell Univ Press. 312 p.
- Wolf A, Swift J, Swinney HL, Vastano JA (1985) Determining Lyapunov exponents from a time series. *Physica D* 16: 285–317.
- Shlesinger M (1988) Fractal time in condensed matter. *Ann Rev Phys Chem* 39: 269–290.
- Feller W (1950) *An Introduction to Probability Theory and Its Applications*. New York: John Wiley.
- Wickerhauser MV (1994) *Adapted Wavelet Analysis from Theory to Software*. Wellesley, Massachusetts: A.K. Peters.
- Strang G (1993) Wavelet transformation versus Fourier transformation. *Bull AMS* 28: 288–305.
- Broomhead DS, King GP (1986) Extracting qualitative dynamics from experimental data. *Physica D* 20: 217–236.
- Selz KA, Mandell AJ, Shlesinger MF, Arguagi V, Owens MF (2004) Designing human  $m_1$  muscarinic receptor-targeted hydrophobic eigenmode matched peptides as functional modulators. *Biophys J* 86: 1308–1331.
- Sternberg S (2010) *Dynamical Systems*. New York: Dover. pp 203–214.
- Eckmann J-P, Kamphorst SO, Ruelle D (1987) Recurrence plots of dynamical systems. *Europhys Lett* 4: 973–977.
- Takens F (1981) Detecting strange attractors in turbulence. *Lecture Notes in Mathematics*. Berlin: Springer-Verlag. pp 366–387.
- Klimaszewska K, Zebrowski JJ (2009) Detection of the type of intermittency using characteristic patterns in recurrence plots. *Phys Rev E* 80: 026214-1-14.
- Pollicott M, Yuri M (1998) *Dynamical Systems and Ergodic Theory*. Cambridge, UK: Cambridge University Press.
- Pomeau P, Manneville Y (1980) Intermittent transition to turbulence in dissipative dynamical systems. *Comm Math Phys* Vol74, Number 2: 189–197.
- Jäger U, Gruler H, Bültmann B (1988) Morphological changes and membrane potential of human granulocytes under influence of chemotactic peptide and/or echo-virus type 9. *Klin Wochenschr* 66: 434.
- Marks PW, Maxfield FR (1990) Local and global changes in cytosolic free calcium in neutrophils during chemotaxis and phagocytosis. *J Cell Biol* 110: 43.
- Mandell AJ (1983) From intermittency to transitivity in neuropsychobiological flows. *Am J Physiol* 245: R484–R494.
- Pollicott M, Yuri M (1998) *Dynamical Systems and Ergodic Theory*. Cambridge, UK: Cambridge University Press. pp 308–311.
- Petty HR (2001) Temporal spatial aspects of neutrophil behavior. *Immunol Res* 23: 85–94.
- Ehrengruber MU, Deranleau DA, Coates TD (1996) Shape oscillations of human neutrophil leukocytes: characterization and relationship to cell motility. *J Exp Biol* 199: 741–747.
- Goldbeter A (1996) *Biochemical Oscillators and Cellular Rhythms*. Cambridge, UK: Cambridge University Press. pp 6769–71.
- Maroto M, Monk N (2009) *Cellular Oscillatory Mechanisms*. *Advances in Experimental Medicine and Biology*, Volume 164. New-York: Springer. 299 p.