

# Chemical nature and structure of organic coating of quantum dots is crucial for their application in imaging diagnostics

Rumiana Bakalova<sup>1</sup>  
Zhivko Zhelev<sup>1</sup>  
Daisuke Kokuryo<sup>1</sup>  
Lubomir Spasov<sup>2</sup>  
Ichio Aoki<sup>1</sup>  
Tsuneo Saga<sup>1</sup>

<sup>1</sup>Molecular Imaging Center, National Institute of Radiological Sciences, Chiba, Japan; <sup>2</sup>Medical Faculty, Sofia University, Sofia, Bulgaria

**Background:** One of the most attractive properties of quantum dots is their potential to extend the opportunities for fluorescent and multimodal imaging in vivo. The aim of the present study was to clarify whether the composition and structure of organic coating of nanoparticles are crucial for their application in vivo.

**Methods:** We compared quantum dots coated with non-crosslinked amino-functionalized polyamidoamine (PAMAM) dendrimers, quantum dots encapsulated in crosslinked carboxyl-functionalized PAMAM dendrimers, and silica-shelled amino-functionalized quantum dots. A multimodal fluorescent and paramagnetic quantum dot probe was also developed and analyzed. The probes were applied intravenously in anesthetized animals for visualization of brain vasculature using two-photon excited fluorescent microscopy and visualization of tumors using fluorescent IVIS<sup>®</sup> imaging (Caliper Life Sciences, Hopkinton, MA) and magnetic resonance imaging.

**Results:** Quantum dots coated with non-crosslinked dendrimers were cytotoxic. They induced side effects in vivo, including vasodilatation with a decrease in mean arterial blood pressure and heart rate. The quantum dots penetrated the vessels, which caused the quality of fluorescent imaging to deteriorate. Quantum dots encapsulated in crosslinked dendrimers had low cytotoxicity and were biocompatible. In concentrations <0.3 nmol quantum dots/kg bodyweight, these nanoparticles did not affect blood pressure and heart rate, and did not induce vasodilatation or vasoconstriction. PEGylation (PEG [polyethylene glycol]) was an indispensable step in development of a quantum dot probe for in vivo imaging, based on silica-shelled quantum dots. The non-PEGylated silica-shelled quantum dots possessed low colloidal stability in high-salt physiological fluids, accompanied by rapid aggregation in vivo. The conjugation of silica-shelled quantum dots with PEG1100 increased their stability and half-life in the circulation without significant enhancement of their size. In concentrations <2.5 nmol/kg bodyweight, these quantum dots did not affect the main physiological variables. It was possible to visualize capillaries, which makes this quantum dot probe appropriate for investigation of mediators of vasoconstriction, vasodilatation, and brain circulation in intact animals in vivo. The multimodal silica-shelled quantum dots allowed visualization of tumor tissue in an early stage of its development, using magnetic resonance imaging.

**Conclusion:** The present study shows that the type and structure of organic/bioorganic shells of quantum dots determine their biocompatibility and are crucial for their application in imaging in vivo, due to the effects of the shell on the following properties: colloidal stability, solubility in physiological fluids, influence of the basic physiological parameters, and cytotoxicity.

**Keywords:** quantum dot, organic shell, biocompatibility, in vivo imaging, two-photon excited fluorescent microscopy, magnetic resonance imaging

Correspondence: Rumiana Bakalova  
Molecular Imaging Center,  
National Institute of Radiological  
Sciences, 4-9-1 Anagawa,  
Inage-ku, Chiba 263-8555, Japan  
Tel +81 43 206 3274  
Fax +81 43 206 4682  
Email bakalova@nirs.go.jp; ra\_bakalova@yahoo.com

## Introduction

One of the most attractive properties of quantum dots is their potential to extend the opportunities for fluorescent and multimodal imaging in vivo. The high quantum yield

(over 50%) and large excitation/emission Stokes shift (up to 300 nm) allow minimization of tissue autofluorescence, especially when using quantum dots with deep-red or near-infrared emission. The high intensity of multiphoton excited fluorescence of quantum dots and the possibility for time-resolved detection of fluorescence additionally increase the signal-to-background ratio in the living organism. Using quantum dots it is possible to visualize complex structures, cells, and even molecular targets at significantly greater depth *in vivo*, in comparison with that achieved by conventional organic fluorophores.<sup>1-3</sup> The chemically functionalized surface of quantum dots, encapsulated in silica or other organic/bioorganic shells, also opens up an opportunity for development of multimodal probes, applicable in two or more imaging techniques, eg, magnetic resonance imaging/optical, positron emission tomography/optical, and positron emission tomography/magnetic resonance imaging/optical).<sup>1-4</sup>

In the development of quantum dot probes for *in vivo* multimodal imaging, it is necessary to follow a few rules.<sup>4</sup> The most important are that the quantum dot nanocrystal has to be encapsulated in a bioinert and biocompatible shell to isolate it from the environment and to minimize its toxicity, and that the capsulated quantum dot nanocrystal has to be conjugated with biocompatible substances (usually polymers), decreasing its rate of accumulation in the liver and spleen and increasing the half-life in the bloodstream, facilitating access of quantum dots to the respective target and its visualization, and preventing opsonization and induction of a nonspecific immune response.<sup>5</sup> The bioinert polymer shells may also increase the water solubility of quantum dots, which facilitates their excretion from the organism through renal filtration and decreases their toxicity.<sup>6,7</sup>

The idea of encapsulation of nanoparticles in biocompatible shells and their conjugation with biocompatible polymers emerged in the mid 1990s in the context of development of water-soluble paramagnetic and ferromagnetic nanoparticles for bioanalyses,<sup>8,9</sup> as a result of earlier experience in the synthesis of biocompatible polymers for drug encapsulation and delivery.<sup>10-12</sup> The earliest and most frequently used polymers for encapsulation of nanoparticles are polyethylene glycol (PEG) and dextran, as well as silica/silicagel shells.<sup>8,9</sup> Later, biocompatible dendrimers and diblock/triblock copolymers were included in the composition.<sup>13,14</sup>

Dextran and PEG make the nanoparticles hydrophilic and increase their half-life in the circulation. However, in some cases, the dextran can induce an acute immune response as anaphylaxis.<sup>15</sup> Currently, PEG or PEG derivatives are the polymers most commonly used to stabilize nanoparticles

and to extend their half-life in the bloodstream due to the “sterically stabilizing effect of PEG”.<sup>16</sup> PEG forms a protective hydrophilic layer on the surface, preventing interaction between blood lipoproteins and cells (eg, macrophages) and significantly delays accumulation of the nanoparticles in the liver and spleen.<sup>5</sup> This is an essential condition for development of contrast nanoprobe for imaging diagnostics, because it enables a longer scan time and accurate localization and visualization of the target of interest (tissues, cells, or expression of specific receptors or molecules). Therefore, PEGylation seems to be an indispensable step in the development of quantum dot probes for *in vivo* diagnostic imaging. The longer half-life of PEGylated quantum dots in the circulation also contributes to their effectiveness in target-specific diagnostic imaging. Penetration of quantum dots into the vasculature and their localization in target tissue is known as the “enhanced permeability and retention effect.” There are also other polymers, including poly(oxazoline), poly(glycerol), and poly-N-vinylpyrrolidone, which have a sterically stabilizing action on nanoparticles.<sup>17,18</sup>

In the present study, we demonstrate the importance of the chemical nature and structure of the organic/bioorganic shells of quantum dots for their application in *in vivo* diagnostic imaging. Several quantum dot probes were developed and their biocompatibility was investigated and compared in experimental animals, using conventional physiological tests and two-photon excited fluorescent microscopy.

## Materials and methods

### Quantum dots and quantum dot probes

CdSe/ZnS quantum dots were synthesized using the methods described by Peng and Peng<sup>19</sup> and Zlateva et al.<sup>20</sup> Nanocrystals were characterized by ultraviolet light/ultraviolet and fluorescent spectroscopy, X-ray analysis (Rigaku RINT-2100, Tokyo, Japan), and high-resolution transmission electron microscopy (JEOL JEM-3010, Tokyo, Japan).

The concentration of quantum dots was calculated using the method of Yu et al, with an extinction coefficient of  $1 \times 10^5/\text{cm}/\text{M}$ .<sup>21</sup> Quantum dots were additionally coated with silica as described by Bakalova et al,<sup>22</sup> polyamidoamine (PAMAM) dendrimers as described by Wu et al<sup>23</sup> (with a slight modification of using the second-generation PAMAM-C12 dendrimer [Sigma-Aldrich, St Louis, MO] instead of octylamine-modified polyacrylic acid), and the second-generation PAMAM-C12 dendrimer (Sigma-Aldrich) crosslinked by a PAMAM-succinamic acid dendrimer (Sigma-Aldrich), with N-(3-dimethylaminopropyl)-N'-ethyl-carbodiimide hydrochloride used as a zero-length crosslinker.

The conjugation of silica- or dendrimer-coated quantum dots with PEG was carried out using the method described in Hermanson.<sup>24</sup> Methyl-PEG<sub>n</sub>-NHS esters or methyl-PEG<sub>n</sub> amines were used for conjugation with amino-functionalized or carboxy-functionalized quantum dots, respectively. The coated quantum dots and quantum dot-PEG conjugates were purified by spin ultrafiltration and kept at 4°C. The amino-functionalized silica-shelled quantum dots were also conjugated with DOTA-phenyl isothiocyanate (DOTA-Bn-NCS/gadolinium, Macrocyclics Company, Dallas, TX) and PEGylated using methods described by Hermanson.<sup>24</sup> The quantum dot/DOTA/PEG conjugates were purified by dialysis using a Slide-A-Lyzer<sup>®</sup> (Pierce Biotechnology Inc, Rockford, IL). The multimodal quantum dots were kept at 4°C. Their T<sub>1</sub> and T<sub>2</sub> contrast properties were characterized using 7 Tesla magnetic resonance imaging (Kobelco, Kobe, Japan).<sup>25</sup> All reagents used in this study were of analytical or high-pressure liquid chromatographic grade.

## Animals and experimental models

Sprague-Dawley rats weighing 325 ± 15 g were used for investigation of the biocompatibility of the quantum dot probes and their usefulness for fluorescent imaging *in vivo*. Balb6 nude mice (22 ± g) were used for investigation of the usefulness of quantum dot probes for multimodal imaging of experimental tumors *in vivo*. Colon26 cells (1 × 10<sup>5</sup> in 10 µL phosphate-buffered saline, pH 7.4) were inoculated subdermally in the left/right hindpaw. All measurements were performed five or ten days after inoculation, when the tumor was 1–2 mm or 5–6 mm in size, respectively.

All experiments were conducted in accordance with the guidelines of the Physiological Society of Japan and were approved by the Animal Care and Use Committee of the National Institute of Radiological Sciences, Chiba, Japan.

## Two-photon imaging of brain circulation *in vivo*

The rat was anesthetized with isoflurane (Abbott, Tokyo, Japan) 4% for induction, 2% for surgery, and 1.4% during measurements. Tracheal intubation was performed, and the femoral vein was catheterized for drug administration. The tail artery was catheterized for blood pressure monitoring and blood sampling. The rat was fixed in a stereotactic frame, the parietal bone was thinned to translucency over the somatosensory cortex (3 mm × 3 mm), and the dura was kept intact. Body temperature was maintained at 37°C, and systemic arterial blood pressure was periodically monitored at the tail artery. Heart rate and end-tidal gas levels were

monitored continuously. To visualize the brain microcirculation, the quantum dot probe was injected (intravenously as a single dose, with an injected volume of 0.3 mL, containing different quantum dot concentrations), and the brain area was excited by an 810 nm pulse laser (Maitai HP, Spectra-Physics, Irvine, CA) under microscope (SP5, Leica) equipped with a 10× objective lens (0.30 numerical aperture). The vessel diameter was then determined by measuring the width of the vessel image projected across the 15 Z images (up to nearly 0.3 mm from the cortical surface) in each vessel segment. The experimental scheme is shown in Supplementary Figure S1.

## IVIS<sup>®</sup> imaging *in vivo*

The mouse was anesthetized with 1.5% isoflurane (Abbott). The tail vein was catheterized for drug administration and the mouse was fixed in the camera of the IVIS<sup>®</sup> imaging system (Caliper Life Sciences, Hopkinton, MA). Body autofluorescence was registered using an excitation filter of 450 ± 30 nm and an emission filter of 650 nm. The quantum dot probe was injected (intravenously as a single dose of 1.6 nmol/kg bodyweight, 100 µL volume) and body fluorescence was registered at a series of time intervals over 90 minutes. The data were analyzed by Living Image<sup>®</sup> software (version 2.50.1, Xenogen Corporation, Alameda, CA).

## Magnetic resonance imaging *in vivo*

Magnetic resonance imaging measurements were performed on a 7.0 Tesla horizontal magnet (Kobelco and Jastec, Tokyo, Japan) interfaced to a Bruker Avance I console (Bruker BioSpin, Karlsruhe, Germany) and controlled with ParaVision 4.0.1 (Bruker BioSpin). The animal (a Balb6 mouse) was anesthetized using isoflurane 1.5% and held in a body cradle (Rapid Biomedical, Rimpf, Germany) in the prone position. Rectal temperature was continuously monitored and automatically controlled at 36.5 ± 0.5°C using a nonmagnetic temperature probe (FOT-M and FTI-10, FISO Technology, Olching, Germany) and an electric temperature controller (E5CN, Omron, Kyoto, Japan) during magnetic resonance imaging measurements. The tail vein was catheterized using a polyethylene tube (PE-10, Becton-Dickinson, Franklin Lakes, NJ) for drug administration. The mouse was then placed in a proton volume radiofrequency resonator (Bruker BioSpin) with a surface radiofrequency receiver (Rapid Biomedical) previously warmed using a body temperature controller (Rapid Biomedical). The resonator units, including the mouse, were placed in the magnet bore. Magnetic resonance imaging was performed using the following parameters: T<sub>1</sub>-weighted

imaging; spin echo sequence with fat suppression preparation pulse; repetition time 400 msec; echo time 9.574 msec; field of view  $32 \times 32$  mm; matrix size  $256 \times 256$ ; slice thickness 1.0 mm; spatial resolution  $125 \times 125 \times 1000$   $\mu\text{m}$ ; four averages; and a total scan time of six minutes and 50 seconds. The magnetic resonance images were obtained before and after injection of the quantum dot probe (intravenously, single dose of  $1 \mu\text{mol/kg}$  bodyweight,  $100 \mu\text{L}$  volume) using the same gain level. The magnetic resonance imaging data were reconstructed by ParaVision and analyzed using MRVision image analysis software (version 1.5.8, MRVision Co, Winchester, MA).

### Flow-cytometric cell viability assay

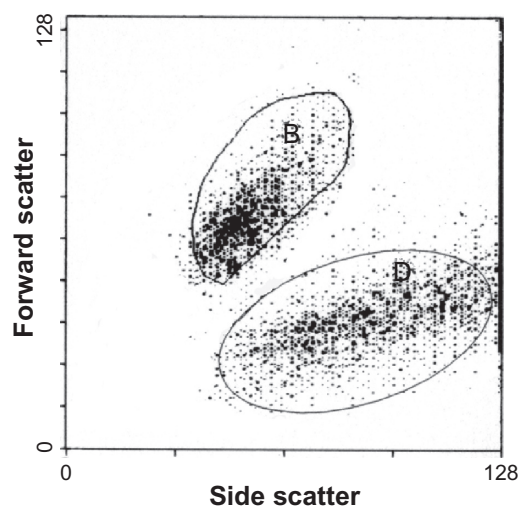
The cell suspension ( $5 \times 10^5$  cells/mL) was incubated with quantum dots at a series of concentrations over 24 hours in a humidified atmosphere and washed three times with phosphate-buffered saline. Five hundred microliters of the same cell suspension was incubated with  $20 \mu\text{L}$  of propidium iodide (Sigma-Aldrich) for 10 minutes at room temperature. The cells were washed three times by phosphate-buffered saline, using centrifugation ( $1000 \times g/10$  minutes), and finally resuspended in  $500 \mu\text{L}$  of phosphate-buffered saline. The propidium iodide entered the dead cells and did not enter the live ones. The cell suspension was subjected to flow cytometric analysis, using a Beckman Coulter-Epics XL flow cytometer (Fullerton, CA), operated in accordance with the manufacturer's recommendations after fine adjustments for optimization of the measurements. The forward-scattered and side-scattered parameters were adjusted to accommodate the inclusion of viable and dead cells within the acquisition data. No cells were excluded from the analysis, and about 10,000 cells were counted. Data were collected and analyzed using XL System II software.

The results were presented as a dot plot of cell fluorescence with quadrant markers drawn to distinguish viable (nonlabeled) and dead (propidium iodide-labeled) cells (Figure 1).

Cell viability after treatment with quantum dots was calculated by the following equation:

$$\text{cell viability (\%)} = \frac{B_p}{B_p + (D_p - D_c)} \times 100$$

where  $B_p$  is the number of viable cells in quadrant B of the sample, treated by quantum dots;  $D_p$  is the number of dead cells in quadrant D of the sample, treated by quantum dots;  $D_c$  is the number of dead cells in quadrant D of the control



**Figure 1** A flow cytometric histogram of cell fluorescence with quadrant markers, drawn to distinguish viable (nonlabeled) from dead (propidium iodide-labeled) cells. Quadrant B shows viable cells and quadrant D shows dead cells.

sample (not treated with quantum dots). Quantum dot<sup>485</sup> probes were used in all flow cytometric analyses.

### Mitochondrial potential assay

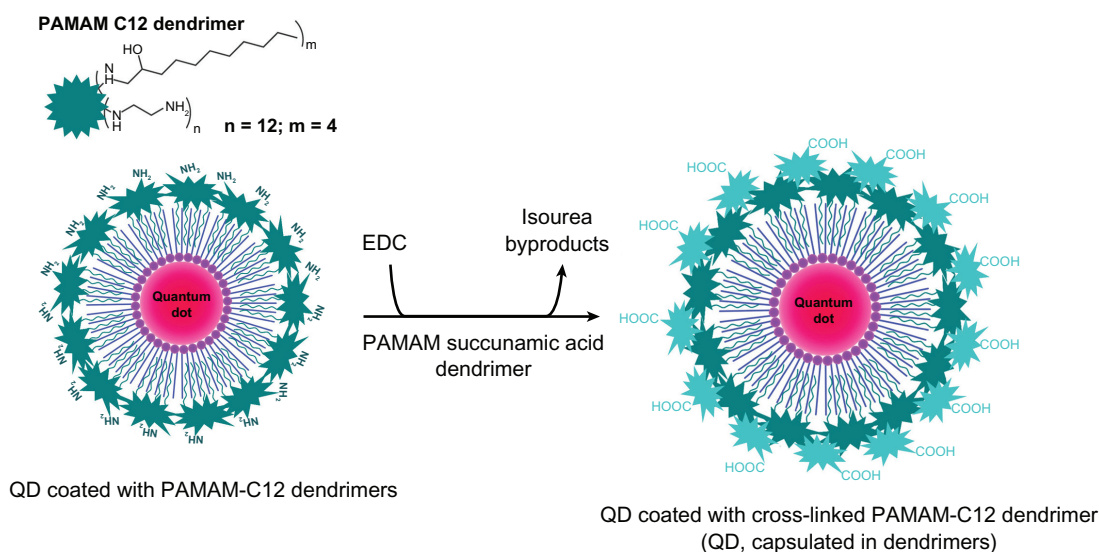
The mitochondrial potential was analyzed using the method described by Wong and Cortopassi<sup>26</sup> using 48-well plates and tetramethylrhodamine ethyl ester as a fluorescent marker (TMRE, Immunochem Diagnostic Technologies Ltd, Truro, Nova Scotia). Briefly,  $50 \text{ nM}$  of TMRE was added to  $500 \mu\text{L}$  of cell suspension ( $1 \times 10^6$  cells/mL), and the fluorescence was measured using a BioRad microplate reader (BioRad Laboratories, Hercules, CA) equipped with a temperature controller. The temperature of the cell suspension was maintained at  $37^\circ\text{C}$ . The fluorescence of each well was detected over two cycles every five minutes at  $\lambda_{\text{ex}} = 540 \text{ nm}$  and  $\lambda_{\text{em}} = 575 \text{ nm}$ . A quantum dot<sup>485</sup> probe was added to each well and the fluorescence detection was continued over seven cycles every five minutes. The data were normalized to the autofluorescence of the quantum dot and the cell suspension was not treated with TMRE (baseline fluorescence).

## Results and discussion

### Biocompatibility of dendrimer-shelled quantum dots

Two quantum dot probes were developed using PAMAM dendrimers as an organic shell (Figure 2). For the first probe, a quantum dot nanocrystal was coated with a non-crosslinked second generation PAMAM C12 dendrimer). The carbohydrate chains (C12) of dendrimer interact with





**Figure 2** Model structure of quantum dot probes, consisting of CdSe/ZnS quantum dots coated with non-crosslinked PAMAM dendrimers or encapsulated in crosslinked PAMAM dendrimers.

**Abbreviations:** PAMAM, polyamidoamine; EDC, 1-ethyl-3(3-dimethylamino propyl)-carbodiimide; QD, quantum dot.

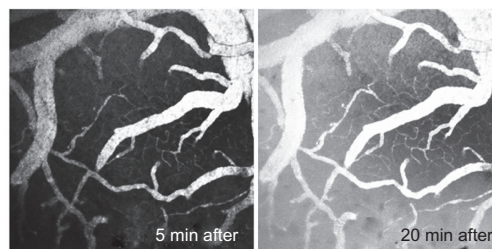
the hydrophobic coordinating ligands of the quantum dot via van der Waals interactions. As a result, the surface of this dendrimer-coated quantum dot was amino-functionalized and positively charged. The second quantum dot probe was developed from the first one through a crosslink between the surface amino groups of the PAMAM C12 dendrimer and the carboxyl groups of the second-generation PAMAM succinamic acid dendrimer, with formation of amide bonds (Figure 2). Thus, in the second quantum dot probe, the nanocrystal was capsulated in crosslinked PAMAM dendrimers, and the surface of the nanoparticles was carboxyl-functionalized and negatively charged.

The zeta potential of amino-functionalized dendrimer-coated quantum dots was about +7.0 mV at pH 7.4, and for carboxyl-functionalized dendrimer-capsulated quantum dots was about -18.7 mV at pH 7.4. Both measurements were made in phosphate-buffered saline. Both quantum dot probes were characterized with excellent solubility in distilled water, and low-salt and high-salt physiological fluids (eg, buffers, saline solution, phosphate-buffered saline, cell cultured medium, plasma). The quantum dot probes were used for in vivo fluorescent imaging of the brain circulation in anesthetized rats, using two-photon excited fluorescent microscopy.

Figure 3 shows two-photon excited fluorescent images of brain vasculature after intravenous injection of dendrimer-coated quantum dots<sup>525</sup>. The images were obtained at  $\lambda_{ex} = 810$  nm with an emission filter of  $525 \pm 50$  nm, in a horizontal scanning plane at a depth of 150  $\mu\text{m}$  from the surface of the skull.

In the case of quantum dots coated with non-crosslinked dendrimers, turbidity of the images increased during the scan time (Figure 3A). This impeded measurement and calculation of the average diameter of the arterioles and venuoles. Concentration of the injected quantum dot probe

**A CdSe/ZnS QD, capsulated in PAMAM dendrimers (0.3 nmol/kg bodyweight)**



**B CdSe/ZnS QD, capsulated in crosslinked dendrimers (0.3 nmol/kg bodyweight)**



**Figure 3** Two-photon excited fluorescent images of brain vasculature in horizontal planes (at 150  $\mu\text{m}$  depth from brain surface) after intravenous injection of QD<sup>525</sup> (coated with non-crosslinked PAMAM dendrimers [A] or capsulated in crosslinked dendrimers [B] in the anesthetized rat).

**Notes:** All images were obtained at  $\lambda_{ex} = 810$  nm and a mean laser power of about 2.7 W. Detectors: filter/band-pass 525/50 nm (via 560/10 nm beam splitter).

**Abbreviations:** PAMAM, polyamidoamine; QD, quantum dot.

**Table 1** Physiological characteristics of rats before and after intravenous injection of QDs (coated with noncrosslinked PAMAM dendrimers)

Parameters	Before injection	About 20 minutes after injection	P value
Mean arterial blood pressure (mmHg)	102.5 ± 2.5	75.5 ± 7.1	<0.05
Heart rate (beats per minute)	378 ± 29	310 ± 15	<0.05
Mean diameter of detected arteries (µm)	28 ± 9	35 ± 10 (vasodilatation)	<0.05
Mean diameter of detected veins (µm)	42 ± 23	45 ± 18 (slight vasodilatation)	NS

**Notes:** Means ± standard deviations from six independent experiments. QD concentration 0.3 nmol/kg bodyweight.

**Abbreviations:** QD, quantum dot; NS, not significant; PAMAM, polyamidoamine.

was 0.3 nmol quantum dots/kg bodyweight. Increasing the concentration also made it difficult to define the outlines of the large blood vessels (arteries and veins). Intravenous injection of this quantum dot probe changed the basic physiological characteristics of the animal (Table 1). Mean arterial blood pressure and heart rate decreased, accompanied by vasodilatation.

PAMAM C12 dendrimers (without quantum dots) had similar effects (see Table S1 for supplementary information). Even low concentrations of the PAMAM C12 dendrimer (0.1 nmol/kg bodyweight injected intravenously in the rat) decreased blood pressure and heart rate and induced vasodilatation.

In the case of quantum dots encapsulated in crosslinked dendrimers, the turbidity of the fluorescent images was negligible (Figure 3B). The images did not change during the scan time. The large and small blood vessels were well outlined and it was easy to measure their average diameter. This quantum

**Table 2** Physiological characteristics of rats before and after intravenous injection of QDs (encapsulated in crosslinked PAMAM dendrimers)

Parameters	Before injection	About 20 minutes after injection	P value
Mean arterial blood pressure (mmHg)	96.8 ± 4.9	100.5 ± 8.4	NS
Heart rate (beats per minute)	340 ± 21	336 ± 23	NS
Mean diameter of detected arteries (µm)	23 ± 10	Constant	NS
Mean diameter of detected veins (µm)	41 ± 20	Constant	NS

**Notes:** Means ± standard deviations from six independent experiments. QD concentration 0.3 nmol/kg bodyweight.

**Abbreviations:** QD, quantum dot; NS, not significant; PAMAM, polyamidoamine.

**Table 3** Effect of CdSe/ZnS QD<sup>485</sup> (coated with noncrosslinked PAMAM dendrimers) on cell viability in vitro (% from control)

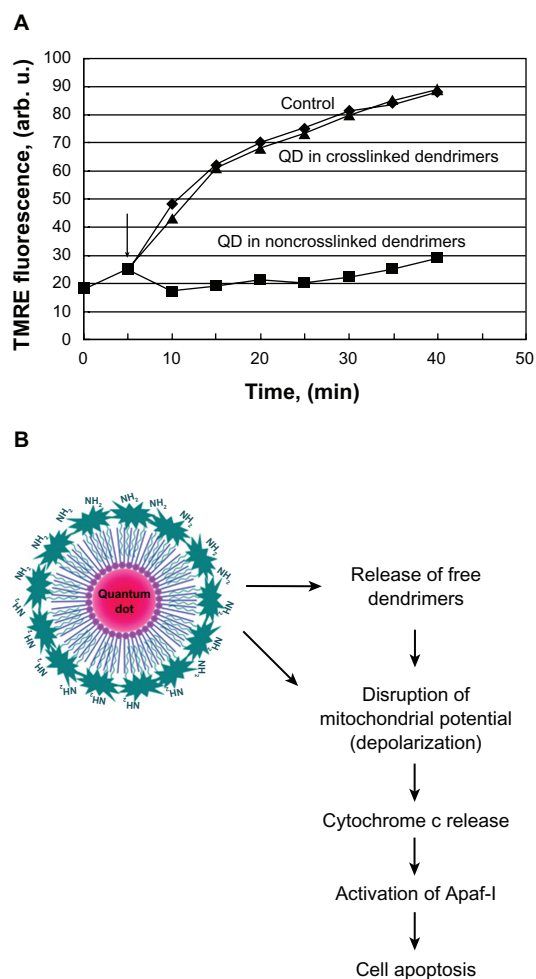
Cell line	10 nM QD	50 nM QD	100 nM QD	200 nM QD	500 nM QD
Jurkat	99 ± 7	97 ± 5	62 ± 5	30 ± 7	21 ± 4
K562	101 ± 6	98 ± 9	62 ± 9	37 ± 5	16 ± 3
A549	103 ± 9	101 ± 5	65 ± 7	39 ± 5	28 ± 5
HeLa	99 ± 3	100 ± 5	60 ± 8	32 ± 3	20 ± 4

**Notes:** The cell suspension ( $5 \times 10^5$  cells/mL) was incubated with QDs over 24 hours in a humidified atmosphere, and cell viability was analyzed by flow cytometry. Cell viability in the absence of QD was considered 100% (control). The data are mean ± standard deviation from six independent experiments.

**Abbreviations:** QD, quantum dot; PAMAM, polyamidoamine.

dot probe did not affect mean arterial blood pressure or heart rate (Table 2) even one hour after injection.

The dendrimer-encapsulated quantum dots were also PEGylated using PEG1100. However, PEGylation did not significantly increase the half-life of the quantum dots in the

**Figure 4** (A) Dynamics of tetramethylrhodamine ethyl ester (TMRE) fluorescence of Jurkat cells in the absence (control) or presence of dendrimer-coated QD<sup>485</sup> 100 nM. (B) Hypothetical mechanism of cytotoxicity of QD, coated with noncrosslinked polyamidoamine dendrimers.

**Abbreviations:** Apaf-1, apoptotic peptidase-activating factor 1; QD, quantum dot.





**Table 4** Physiological characteristics of rats before and after intravenous injection of PEG 100-grafted silica-shelled QDs

Parameters	Before injection	About 20 minutes after injection	P value
Mean arterial blood pressure (mmHg)	96.2 ± 5.4	95.3 ± 12.8	NS
Heart rate (beats per minute)	320 ± 55	345 ± 51	NS
Mean diameter of detected arteries (µm)	24 ± 9	Constant	NS
Mean diameter of detected veins (µm)	41 ± 22	Constant	NS

**Notes:** Means ± standard deviations from six independent experiments. QD concentration 0.4 nmol/kg bodyweight.

**Abbreviations:** QD, quantum dot; NS, not significant; PEG, polyethylene glycol.

not shown), whereas the quantum dots coated with non-crosslinked dendrimers were highly cytotoxic in concentrations >50 nM.

The cytotoxicity of quantum dots coated with non-crosslinked dendrimers can be explained with the intrinsic instability of the micelle type of dendrimer coat. The micelle structure is a result of weak hydrophobic interactions, and its partial destruction in the cell suspension is highly possible. Thus, cytotoxic monomers can be released into the intracellular and extracellular environment. Several groups have also shown that cationic PAMAM dendrimers are cytotoxic in their monomer form.<sup>33–38</sup>

The micelle type of organic coat could also be destroyed in the bloodstream, and the monomers can have side effects on blood pressure and heart rate (Table 1 and Table 2). It has been established in vitro that the cationic PAMAM dendrimers modulate the release of nitric oxide, one of the major vasodilators in vivo,<sup>39</sup> while the anionic PAMAM-COOH dendrimers significantly reduce this process.<sup>36</sup> In our study, we observed vasodilatation of the brain arterioles and venuoles after injection of quantum dots coated by non-crosslinked positively charged dendrimers (Table 1). The quantum dots encapsulated

**Table 5** Effect of silica-shelled CdSe/ZnS QDs (conjugated with PEG 100) on cell viability in vitro (% from control)

Cell line	10 nM QD	50 nM QD	100 nM QD	200 nM QD	500 nM QD
Jurkat	101 ± 9	99 ± 7	100 ± 7	103 ± 10	98 ± 7
K562	100 ± 5	102 ± 8	98 ± 5	96 ± 8	98 ± 7
A549	98 ± 4	101 ± 5	98 ± 7	98 ± 9	96 ± 10
HeLa	102 ± 6	103 ± 4	99 ± 9	98 ± 5	96 ± 8

**Notes:** The cell suspension ( $5 \times 10^5$  cells/mL) was incubated with QDs for 24 hours in a humidified atmosphere, and cell viability was analyzed by flow cytometry. Cell viability in the absence of QDs was considered to be 100% (control). The data are means ± standard deviations from six independent experiments.

**Abbreviations:** QD, quantum dot; PEG, polyethylene glycol.

in crosslinked carboxyl-functionalized dendrimers did not manifest such effects (Table 2). The encapsulated organic coat is stable because of covalent bonds, and release of monomers is impossible without a specific metabolic pathway.

It is widely accepted that the cytotoxicity of dendrimers is due to their high cellular permeability and colocalization in the mitochondria and lysosomes.<sup>35,40</sup> We investigated the effect of both quantum dot probes on the mitochondrial potential ( $\Delta\psi_m$ ) of living cells (Figure 4). The mitochondrial potential is one of the major markers for induction of apoptosis. For this purpose, we used the quantum dot<sup>485</sup> and TMRE ( $\lambda_{em} = 575$  nm) as a fluorescent sensor. The fluorescence of TMRE changes proportionate to mitochondrial potential. Loss of the mitochondrial potential is accompanied by a significant decrease or total loss of TMRE fluorescence.<sup>26</sup> The kinetic curves in Figure 4A show that quantum dots coated with non-crosslinked PAMAM dendrimers induce depolarization of the mitochondrial membrane and loss of the mitochondrial potential in living cells. The quantum dots encapsulated in crosslinked dendrimers did not affect the mitochondrial potential.

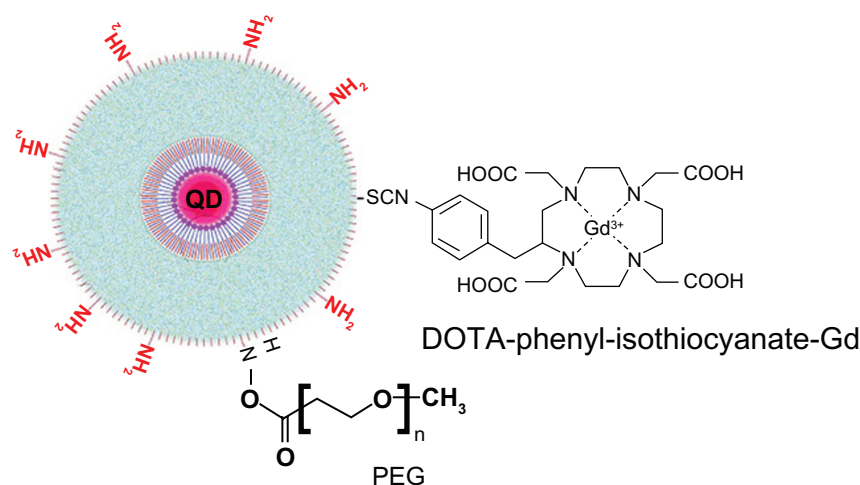
It is widely accepted that loss of mitochondrial potential induces expression of cytochrome c and apoptotic peptidase-activating factor 1. This leads to activation of caspases, induction of apoptosis, and cell death.<sup>41–43</sup> This molecular mechanism could explain, at least partially, the cytotoxic effect of quantum dots coated with non-crosslinked PAMAM dendrimers (Figure 4B).

## Biocompatibility of silica-shelled quantum dots

Silica materials are one of the most bioinert and biocompatible materials for coating. It has been found that silica nanoparticles with diameters below 50 nm do not activate macrophages<sup>44–46</sup> and should not induce an immune response after intravenous injection. However, silica shells do not provide sufficiently high solubility of quantum dots in high-salt physiological fluids, such as saline solution, phosphate-buffered saline, blood, and plasma.<sup>47</sup>

In 2006, we developed silica-shelled single CdSe/ZnS quantum dots with an average diameter of  $17.42 \pm 2.12$  nm, a size distribution of about 12%, and a quantum yield of 30%–45% (depending on the size of the silica-shelled quantum dots, and measured in distilled water, Figure 5).<sup>48</sup> However, without PEGylation, these amino-functionalized nanoparticles possessed good colloidal stability in distilled water and low-salt buffer, but not in saline solution, phosphate-buffered saline, or body fluids. For example, if mice were inoculated intranasally with non-PEGylated





**Figure 8** Model structure of PEGylated multimodal silica-shelled quantum dots.<sup>48</sup>

**Note:** Reprinted with permission from Zhelev Z, Ohba H, Bakalova R. Single quantum dot-micelles coated with silica shell as potentially non-cytotoxic fluorescent cell tracers. *J Am Chem Soc.* 2006;128:6324–6325. Copyright 2006 American Chemical Society.

**Abbreviations:** PEG, polyethylene glycol; DOTA, tetraazacyclododecanetetraacetic acid.

silica-shelled quantum dots, we observed a lot of aggregates in the nasal mucosa (Figure 6). Therefore, PEGylation and stabilization were necessary before these nanoparticles could be applied for imaging in vivo and especially in the case of intravenous and intranasal administration. The PEGylated silica-shelled quantum dots possessed high colloidal stability and did not aggregate in high-salt physiological fluids (eg, 100 mM phosphate-buffered saline and plasma).

PEG1100-grafted silica-shelled quantum dots injected intravenously into the rat and brain vasculature were visualized using two-photon excited fluorescent microscopy (Figure 7). The images were obtained at  $\lambda_{\text{ex}} = 810$  nm and an emission filter of  $525 \pm 50$  nm, with scanning in the horizontal plane at a depth of 150  $\mu\text{m}$  from the surface of the skull.

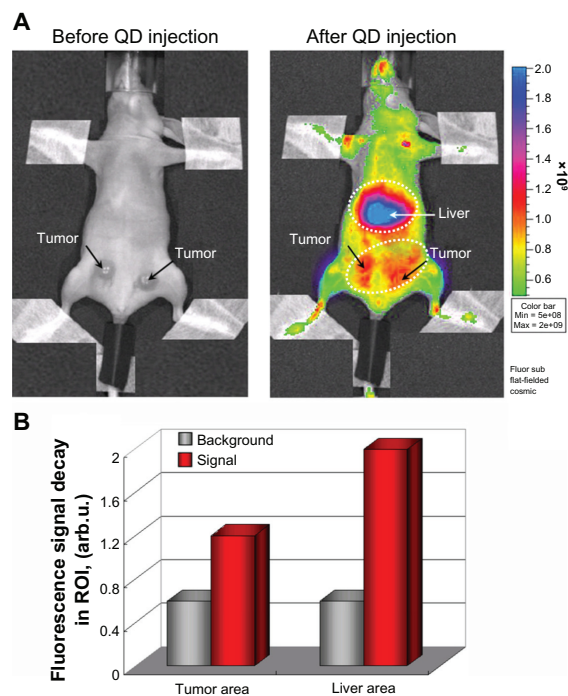
**Table 6** Spectral characteristics of PEG1100-grafted multimodal silica-shelled QDs

Spectral characteristic	Value
QY in 10 mM PBS, pH 7.4 (%)	35–48
QY in 100 mM PBS, pH 7.4 (%)	32–45
Longitudinal relaxation time, $T_1$ (msec)	$293.4 \pm 23.5$
Transverse relaxation time, $T_2$ (msec)	$128.5 \pm 5.3$

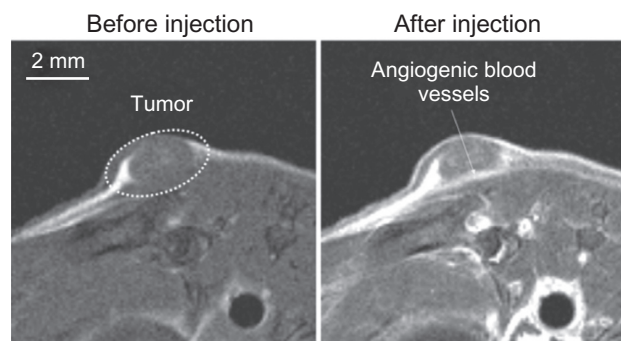
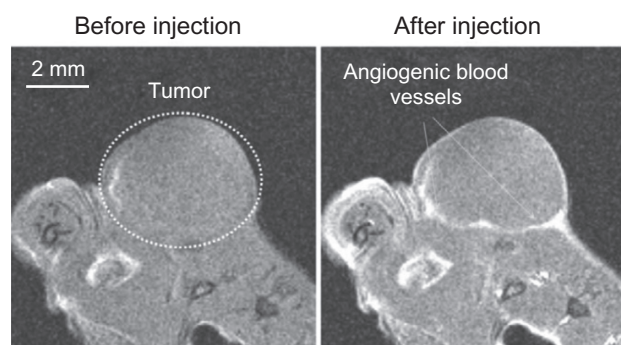
**Notes:**  $T_1$  and  $T_2$  data are means  $\pm$  standard deviations from six independent experiments.  $T_1$  and  $T_2$  data were calculated at a QD concentration of 500 nM. The magnetic resonance imaging measurements were performed in a 4.7 T magnet (General Electric, Fairfield, CT) interfaced to a Bruker Avance console (Bruker Medical GmbH, Germany). A 30 mm diameter Litz coil (Doty Scientific Inc, Columbia, SC) was used for measurement of the samples. The sample temperature was maintained at room temperature (approximately 22–24°C). The measurements were performed in the following order:  $T_1$ -weighted imaging using conventional spin echo sequence; multiecho spin echo imaging for  $T_2$  calculations; and inversion recovery spin echo imaging for  $T_2$  calculations. For calculation of fluorescence quantum yield, the spectrally integrated emission of particle dispersion in different aqueous solutions was compared with the integrated emission of an ethanol solution of rhodamine G (Fluka) of identical optical density ( $<0.015$ ) at an excitation wavelength of 365 nm.

**Abbreviations:** QD, quantum dot; PEG, polyethylene glycol; PBS, phosphate-buffered saline; QY, quantum yield.

The fluorescent contrast was constant during 30 minutes of scanning. The PEG1100-grafted silica-shelled quantum dots gave clear images even at high concentrations (2.5 nmol quantum dots/kg bodyweight, Figure 7). On the other hand, 0.2 pmol quantum dots/kg bodyweight was enough



**Figure 9** (A) Fluorescent imaging of tumor by angiogenesis in the anesthetized mouse injected intravenously with PEG1100-grafted multimodal QD<sup>655</sup> (1.6 nmol/kg bodyweight, single dose). Dashed lines indicate liver area and tumor area (including angiogenic network). The images were obtained 15 minutes after injection using an IVIS<sup>®</sup> imaging system, with excitation at  $450 \pm 30$  nm and emission at 650 nm (DsRed filter). The images were obtained on day 10 after inoculation ( $1 \times 10^5$  cells in 10  $\mu\text{L}$ ). (B) Fluorescence intensity in tumor area, liver area, and other parts of the body (background fluorescence) after intravenous injection of PEG1100-grafted multimodal QD<sup>655</sup> (1.6 nmol/kg bodyweight, single dose). Data were calculated from the images in A. **Abbreviations:** QD, quantum dot; PEG, polyethylene glycol.

**A Three days after inoculation (tumor diameter ~2 mm)****B Ten days after inoculation (tumor diameter ~6 mm)**

**Figure 10**  $T_1$ -weighted magnetic resonance images of colon cancer in the anesthetized mouse injected intravenously with PEG1100-grafted multimodal QD<sup>655</sup> embedded with gadolinium (1  $\mu\text{mol/kg}$  bodyweight, single dose). Dashed lines indicate tumor area (including angiogenic vasculature). The images were obtained immediately after injection, using 7.0 Tesla magnetic resonance imaging. Imaging parameters: spin echo sequence with fat suppression preparation pulse; repetition time 400 msec; echo time 9.574 msec; field of view  $32 \times 32$  mm; matrix size  $256 \times 256$ ; slice thickness 1.0 mm; number of averages 4.

**Abbreviations:** QD, quantum dot; PEG, polyethylene glycol.

to visualize the large-sized and middle-sized blood vessels at a depth of 150  $\mu\text{m}$ . Increasing the concentration of the quantum dot probe, it was possible to visualize the arteries and veins at a depth of 600–700  $\mu\text{m}$ . These quantum dots did not affect blood pressure or heart rate, and did not provoke vasodilatation or vasoconstriction up to 60 minutes after injection (Table 4). The PEGylated silica-shelled quantum dots were noncytotoxic in concentrations up to 500 nM, as determined in cultured cells (Table 5).

The PEG1100-grafted silica-shelled quantum dots<sup>655</sup> were conjugated with DOTA/ $\text{Gd}^{3+}$ , and the probe was applied for multimodal imaging of cancer in mice. This is a classical scheme for the multimodal quantum dot probes developed by other researchers (Figure 8).<sup>1,4</sup>

The fluorescent and magnetic resonance imaging characteristics of our multimodal quantum dots are shown in Table 6. The fluorescent quantum yield was about 40% in high-salt buffer. The longitudinal and transverse relaxation

times ( $T_1$  and  $T_2$ ) were about 290 msec and 120 msec, respectively.

The fluorescent images in Figure 9A show that the multimodal silica-shelled quantum dots visualized the tumor by angiogenesis. The experimental tumor was developed within 10 days after inoculation of Colon32 cells in the left and right hind paws of mice. Fluorescence intensity in the tumor area was approximately three times higher than that in the surrounding tissues (Figure 9B). Maximum fluorescence intensity was detected in the liver area. It seems that these nanoparticles accumulated predominantly in the liver area within 90 minutes of their injection into the tail vein. Other authors have also reported that polymer-coated quantum dots accumulate predominantly in the liver, partially in the spleen, lung, and kidney, and are almost never observed in the heart and brain.<sup>49</sup>

The paramagnetic characteristics of the quantum dot probe also enabled visualization of tumors using magnetic resonance imaging (Figure 10). The short longitudinal relaxation time ( $T_1$ ) of gadolinium in the silica spheres allowed visualization of the tumor on the third day after inoculation, when tumor size was about 2 mm in diameter and the angiogenic network was not well developed.

## Conclusion

The data presented here show that the type and structure of organic/bioorganic shells of quantum dots are crucial for their colloidal stability and solubility in physiological fluids, and their biocompatibility and cytotoxicity. All these parameters have to be identified and controlled before application of organic-coated quantum dots for in vivo imaging. CdSe/ZnS quantum dots, coated with non-crosslinked PAMAM dendrimers and amino-functionalized, are cytotoxic and nonbiocompatible. These quantum dots have side effects after intravenous administration in healthy animals, ie, decreased blood pressure and heart rate, as well as vasodilatation. Presumably this is as a result of instability of the dendrimer coat in the bloodstream and release of free monomers, inducing cytotoxic mechanisms in the organism. These quantum dots are not appropriate for in vivo imaging of vasculature structure, because the nanoparticles seem to penetrate the vessels, affecting the quality of the images.

CdSe/ZnS quantum dots encapsulated in crosslinked carboxyl dendrimers have low cytotoxicity and comparatively good biocompatibility in concentrations up to 0.3 nmol/kg bodyweight. This quantum dot probe is appropriate for in vivo imaging. PEGylation is not required to increase the half-life of dendrimer-coated quantum dots in the bloodstream.

However, PEGylation is an indispensable step in development of quantum dot probes for in vivo imaging, based on silica-shelled quantum dots. Non-PEGylated and amino-functionalized silica-shelled quantum dots have low colloidal stability in high-salt physiological fluids, which is accompanied with rapid aggregation. The aggregates could induce activation of macrophages and a nonspecific immune response in vivo. The conjugation of silica-shelled quantum dots with PEG1100 increases their half-life in the circulation without significant enhancement of their size. PEG1100-grafted silica-shelled quantum dots are appropriate for visualization of blood vasculature and circulation in vivo, using two-photon excited fluorescent microscopy. In concentrations up to 2.5 nmol/kg bodyweight injected intravenously, these quantum dots do not affect physiological variables, eg, blood pressure, heart rate, and blood vessel diameter. Using these quantum dots, it was possible to visualize capillaries, which makes them appropriate for investigation of mediators of vasoconstriction, vasodilatation, and brain circulation in intact animals in vivo.

It should be noted that our results are related to use of quantum dots for structural fluorescent imaging of blood vessels and tumors. Quantum dot probes with sensing properties also have potential for functional imaging diagnostics in vivo.<sup>50</sup>

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## Disclosure

The authors report no conflicts of interest in this work.

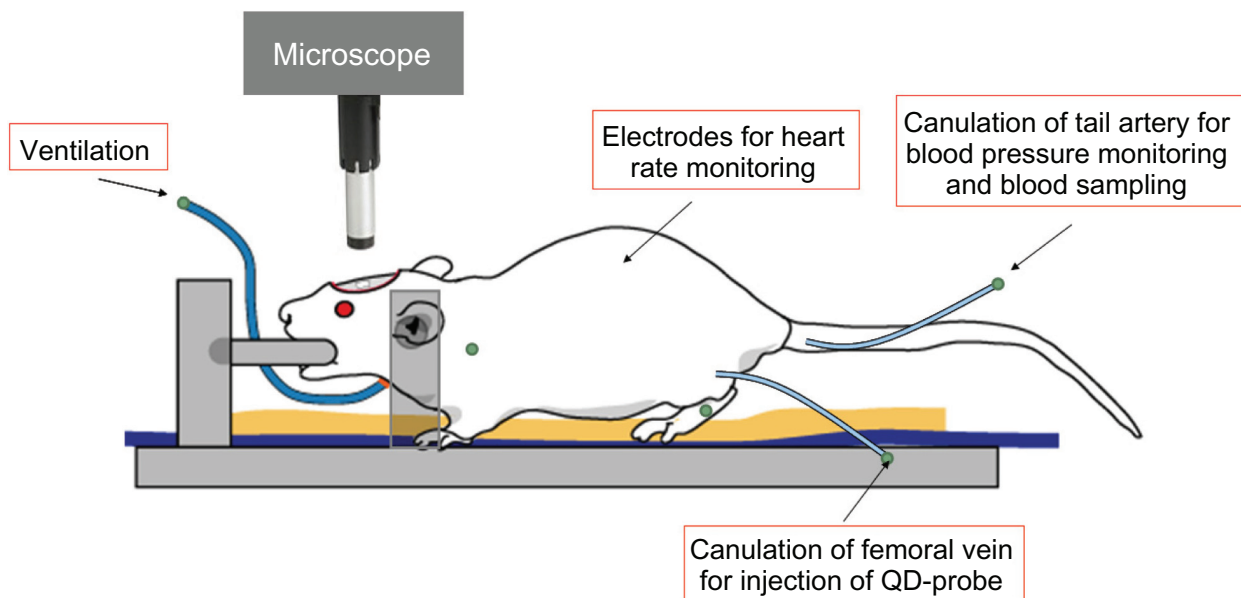
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## Supplementary data



**Figure S1** Experimental scheme of imaging of two-photon excited fluorescence in rats.  
**Abbreviation:** QD, quantum dot.

**Table S1** Physiological characteristics of rats before and after intravenous injection of polyamidoamine C12 dendrimer 0.1 nmol/kg bodyweight

Parameters	Before injection	About 20 minutes after injection	P value
Mean arterial blood pressure (mmHg)	114.0 ± 5.5	68.3 ± 8.5	<0.01
Heart rate (beats per minute)	392 ± 24	317 ± 18	<0.05
Mean diameter of detected arteries (μm)	29 ± 7	39 ± 11 (vasodilatation)	<0.05
Mean diameter of detected veins (μm)	42 ± 21	47 ± 14 (vasodilatation)	<0.05

**Notes:** Data are means ± standard deviations from five independent experiments. The experimental conditions are the same as in Table 1.

**Table S2** Effect of PAMAM C12 dendrimers on cell viability in vitro (% from control)

Cell line	50 nM dendrimer	100 nM dendrimer	500 nM dendrimer
Jurkat	65 ± 5	37 ± 4	8 ± 4
K562	57 ± 9	31 ± 5	3 ± 1
A549	68 ± 9	40 ± 8	5 ± 1
HeLa	70 ± 8	42 ± 4	12 ± 4

**Notes:** The cell suspension ( $5 \times 10^5$  cells/mL) was incubated with dendrimers over 24 hours in a humidified atmosphere, and cell viability was analyzed by flow cytometry. Cell viability in the absence of dendrimer was considered 100% (control). The data are means ± standard deviations from six independent experiments.

**Table S3** Effect of water-soluble amino-functionalized CdSe/ZnS QD<sup>465</sup> without additional organic shell on cell viability in vitro (% from control)

Cell line	10 nM QD	50 nM QD	100 nM QD	200 nM QD	500 nM QD
Jurkat	104 ± 5	98 ± 8	100 ± 2	97 ± 6	96 ± 8
K562	100 ± 3	101 ± 5	103 ± 4	97 ± 5	95 ± 5
A549	100 ± 6	103 ± 5	99 ± 8	100 ± 4	97 ± 4
HeLa	102 ± 4	98 ± 7	97 ± 6	98 ± 5	94 ± 9

**Notes:** The cell suspension ( $5 \times 10^6$  cells/mL) was incubated with QDs within 24 hours at humidified atmosphere, and cell viability was analyzed by flow cytometry. The cell viability in absence of QD was considered 100% (control). Data are means ± standard deviations from six independent experiments.

**Abbreviation:** QD, quantum dot.

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