

Since January 2020 Elsevier has created a COVID-19 resource centre with free information in English and Mandarin on the novel coronavirus COVID-19. The COVID-19 resource centre is hosted on Elsevier Connect, the company's public news and information website.

Elsevier hereby grants permission to make all its COVID-19-related research that is available on the COVID-19 resource centre - including this research content - immediately available in PubMed Central and other publicly funded repositories, such as the WHO COVID database with rights for unrestricted research re-use and analyses in any form or by any means with acknowledgement of the original source. These permissions are granted for free by Elsevier for as long as the COVID-19 resource centre remains active.

ELSEVIER

Contents lists available at ScienceDirect

Journal of Infection and Public Health

journal homepage: http://www.elsevier.com/locate/jiph



Review Article

New perspective towards therapeutic regimen against SARS-CoV-2 infection



Vartika Srivastava^a, Aijaz Ahmad^{a,b,*}

- ^a Clinical Microbiology and Infectious Diseases, School of Pathology, Faculty of Health Sciences, University of the Witwatersrand, Johannesburg, 2193, South Africa
- b Infection Control, Charlotte Maxeke Johannesburg Academic Hospital, National Health Laboratory Service, Johannesburg, 2193, South Africa

ARTICLE INFO

Article history:

Received 28 September 2020 Received in revised form 5 May 2021 Accepted 16 May 2021

Keywords:
Coronaviruses
nCOVID-19
Drug targets SARS-CoV-2
SARS-Cov-2 infection and co-infections
Vaccine

ABSTRACT

The ongoing enormous loss of human life owing to Severe Acute Respiratory Syndrome Coronavirus-2 (SARS-CoV-2), has led to a global crisis ranging from the collapse of health – care systems to socioeconomic instability. As SARS-CoV-2 is a novel virus, very little information is available from researchers and therefore, a rigorous effort is required to decode its pathogenicity. There are no licenced treatment options available for treating SARS-CoV-2 infections and the development of a new antiviral drug targeting coronavirus cannot happen soon. Consequently, drug repurposing is a promising solution for combating the present pandemic. In this review, we have thoroughly discussed all the proteins encoded by the SARS-CoV-2 genome; their importance in pathogenicity and their potential role in drug discovery. Also, the budding threat of co-infections by other pathogenic microbes has been highlighted. Furthermore, the advances made in the medicinal field for the treatment and prevention of this viral infection is explained. Altogether, this review will provide some insightful discussions about this infectious disease and will meet certain of the knowledge gaps which exist by presenting an exhaustive and extensive scientific report on the ongoing mission for COVID-19 drug discovery.

© 2021 Published by Elsevier Ltd on behalf of King Saud Bin Abdulaziz University for Health Sciences. This is an open access article under the CC BY-NC-ND license (http://creativecommons.org/licenses/by-nc-nd/4.0/).

Contents

Introduction	853
Aim of this review	853
Study selection	853
SARS-CoV-2	854
Structural proteins	854
Spike protein (S)	854
Nucleocapsid protein (N)	854
Membrane protein (M)	854
Envelope protein (E)	854
Non-structural proteins	854
Accessory proteins	854
SARS-Cov-2 infection and co-infections	854
Potential targets for drug discovery in SARS-Cov-2	855
Targeting viral structural protein and its interactions	855
Targeting virus RNA synthesis and replication	
C-like protease (3CLpro/Nsp5)	856
Papain-like proteases (PLpro)	

E-mail addresses: Aijaz.Ahmad@wits.ac.za, Aijaz.Ahmad@nhls.ac.za (A. Ahmad).

Corresponding author.

RNA-dependent RNA polymerases (RdRp/Nsp12)	856
Helicase/NTPase	
Targeting virulence factors	858
Targeting host-specific receptors	858
Available regimens	
Drug repurposing for SARS-Cov-2	858
Vaccines under development for COVID-19	859
Summary	859
Funding	859
Competing interests	
Ethical approval	860
References	860

Introduction

Coronaviruses (CoVs) are a large family of RNA viruses, belongs to the family Coronaviridae and order Nidovirales [1]. These viruses are protected by a membrane envelope and possess a large RNA genome (26–32 kb), which is single-stranded, with positive (mRNA) polarity (ssRNA+) and a nucleocapsid of helical symmetry [2]. The club-shaped glycoprotein spikes projecting from the virus surface give an electron micrograph image that resembles the solar corona; hence its name "coronavirus" [3]. Both human beings and animals are targeted groups of Covs; however, bats have been found to host the largest variety of Covs [4]. There are four classes of coronaviruses, namely: alpha, beta, gamma, and delta. The beta-Covs class harbours Severe Acute Respiratory Syndrome (SARS) virus (SARS-CoV), Middle East Respiratory Syndrome (MERS) virus (MERS-CoV), and the COVID-19 causative agent SARS-CoV-2. Coronaviruses attack the host's lower respiratory system, resulting in viral pneumonia. However, SARS-CoV-2 may also affect different organ systems including the central nervous system, leading to multiple organ failure [5,6]. SARS-CoV-2 has also been identified as more contagious than other betaCovs [7].

The first worldwide coronavirus outbreak, recorded between 2002–2004, was caused by SARS-CoV. The disease caused around 8098 SARS positive cases with a highly infectious mortal form of pneumonia and 774 reported deaths [8]. The second coronavirus outbreak was caused by MERS-CoV towards the end of 2012, which affected around 27 countries, resulting in 2494 positive cases and 858 reported deaths [9]. The disease symptoms ranged from mild to acute respiratory distress syndrome [9].

Coronavirus disease 2019 (COVID-19), reported in late 2019, is the third ruthless coronavirus outburst; caused by SARS-CoV-2. Symptoms range from mild (fever, cough and shortness of breath) to acute (severe pneumonia resulting in multi-organ failure) [10]. As reported by the World Health Organization (WHO), as of 09 September 2020 there were a total 27,417,497 confirmed cases and 894,241 mortalities globally [11]. There was increasing concern due to the fast spread of this disease, and therefore a global emergency was declared by WHO on January 31, 2020, and on March 11, 2020 the disease was documented as a pandemic. The world is desperate to uncover ways that can control the spread of novel coronaviruses and to find successful treatment options. In search of effective treatments or vaccines, more than 200 clinical trials for COVID-19 are either being conducted, or patient recruitment is in process [12]. However, every day new studies are being added, as the number of cases is increasing multi-fold globally. The treatment options currently being explored vary from decade-old malaria drugs to unsuccessful Ebola treatments, to repurposing flu drugs. An antiviral drug named EIDD-2801 has been claimed by scientists to combat SARS-CoV-2 in a better way than remdesivir [13]. Another antiviral drug, favipiravir or avigan, used against influenza in Japan, was also found effective against SARS-CoV-2 infections [14]. Malaria

treatments options, namely, chloroquine and hydroxychloroquine were initially reported to be an effective option against COVID-19 [15]; however, the results from clinical trial (NCT04332991) [16] and prophylaxis data recorded by Boulware and coworkers [17] were not promising. Additionally, the withdrawal of chloroquine and hydroxychloroquine from chief investigations denotes the termination of attempts to repurpose these drugs for combating SARS-CoV-2 infection [18]. Remdesivir, an unsuccessful Ebola drug, has also shown potential and is being repurposed for SARS-CoV-2 infections; however, additional clinical trials are still ongoing to ensure the effectiveness of this drug in COVID-19 patients. Despite all this, a standard treatment for SARS-CoV-2 infections is still lacking and increasing incidences of asymptomatic infections, excessive transmissibility, and a long incubation period have made COVID-19 a competent and challenging pathogen which is very difficult to contain.

Most importantly, the occurrence of microbial co-infections in COVID-19 patients complicates the situation by increasing the hitches in management of diagnosis of and prognosis for SARS-CoV-2. Therefore, clinicians cannot ignore the high chance and risk of co-infections caused by all groups of microbes, bacteria, viruses and fungi among COVID-19 patients that may further result in serious disease symptoms and raise the mortality rate [19]. However, it cannot be denied that coinfected microorganisms can bring hope for developing new strategies against SARS-CoV-2 infection. Therefore, in the present review we have highlighted this crucial aspect to emphasise the importance of microbial coinfection in SARS-CoV-2 infection.

Aim of this review

Although there are no targeted antiviral agents available for treatment of novel SARS-CoV-2, a remarkable amount of research is in progress to find potential treatments that can save humankind and develop vaccines that can secure our future. This review aims to strengthen the intellectual foundation of ongoing research for advances in therapeutic sciences aiming at potential drugs as well as preventive vaccines for combating of SARS-CoV-2 and other coronavirus diseases. The present review provides a cumulative description of recent information on the SARS-CoV-2 structural morphology; its characteristics, related co-infections, potential drug targets and treatment options available.

Study selection

The PubMed, ScienceDirect and Scopus databases have been exhaustively searched up till September 09, 2020, using the keywords: SARS-CoV-2/coronavirus infection, including updates on SARS-CoV-2 treatments and vaccine development, challenges in coronavirus treatments, research and development on therapeutic agents and vaccines for COVID-19 and related human coronavirus

diseases and drug targets for corona viruses. Non-English articles were excluded from the study. In total 87 published articles were accessed and important cross-references were also retrieved and included in the present study. Moreover, updates from WHO, CDC, currently ongoing trials from ClinicalTrials.gov reports and other authentic sources were grouped and presented in this review article.

SARS-CoV-2

The SARS-CoV-2 genome consists of 29,811 nucleotides and is organized into a 5' untranslated region (UTR), replicase complex (ORF1ab), spike gene (ORF2), envelope gene (ORF3a, ORF4), membrane gene (ORF5), nucleocapsid genes (ORF6, ORF7a, ORF7b, ORF8, ORF9), 3' UTR, and other open reading frames (ORFs) yet to be characterized [20,21]. The genome encodes 29 different proteins that are divided into three main groups, namely: structural proteins (spike, envelope, membrane and nucleocapsid), non-structural proteins (NSP) and accessory proteins [22].

Structural proteins

SARS-CoV-2 encodes four main structural glycoproteins – spike (S), envelope (E), membrane (M) and nucleocapsid (N).

Spike protein (S)

S proteins are transmembrane proteins (around 150 kDa) and are projected outwards from the surface of the virus. These proteins bind the angiotensin-converting enzyme 2 (ACE2) expressed in cells of the lower respiratory tract and play a crucial role in attachment, fusion, entry, and transmission of viral particles into host cells. Furin-like protease present in the host cell cleaves S glycoproteins into two sub-units, namely the S1 part (Nterminal), responsible for virus-host receptor binding and the S2 part (C-terminal), which mediates virus-cell membrane fusion in transmitting host cells [22,23]. Therefore, the process of SARS-CoV-2 infection starts with the binding of S1- receptor-binding domain (S1-RBD) to the cell membrane receptors of the host cell, causing a structural change in the S2 part which results in fusion and the entry of the viral particle into the target cell [24,25].

Nucleocapsid protein (N)

N glycoprotein binds to the viral genome to make up nucleocapsid protein and is involved in processes such as the viral replication cycle and the host cells response to viral infections [26–28]. Although, the nucleocapsid protein N-terminal domain of this virus is structurally similar to other known Covs, but the surface electrostatic potential characteristics between them are different [29].

Membrane protein (M)

M proteins are important and most abundant proteins, which specify the outline of the viral envelope and are moreover considered as a principal organizer of CoV assembly [24]. In silico studies revealed that SARS-CoV-2 M protein has a triple helix bundle, forms a single 3-transmembrane domain (TM) and is homologous to the prokaryotic sugar-transport protein semi-SWEET. However, the advantage and role of sugar transporter-like structures in viruses are still unknown [30]. The membrane protein assists S protein during attachment and admission of the virus to the host cell. It also helps in forming a stable N protein-RNA complex and supports the completion of the viral assembly inside the virion [31].

Envelope protein (E)

E proteins are small membrane proteins (8.4–12 kDa size) that interact with M proteins to form the viral envelope and accounts for

SARS-CoV-2 assembly, budding and pathogenesis [26,28]. They are found to be highly conserved among the beta coronaviruses. There are predominantly two structural domains: a hydrophobic domain and a charged cytoplasmic tail. The presence of heme agglutininesterase protein has been reported in some of the coronaviruses.

Non-structural proteins

The second group of proteins, non-structural proteins (NSP), play a vital role and control the assembly of the viral particle as well as its escape from the host defence system. The RNA genome that encodes these proteins is replicase complex, containing two ORFs (ORF1a and ORF1b), complete expression of which is accomplished via ribosomal frameshifting [32]. The translation of ORF1a and ORF1b produces two huge overlapping polyproteins, pp1a and pp1ab. These polyproteins are then cleaved into 16 mature smaller proteins by the papain-like protease (PLpro) and the 3-chymotrypsin-like protease (3CLpro), also known as the main protease (Mpro). From 16 proteins, the first 11 are transcribed from ORF1a and the remaining five from ORF1b [20,23]. A summary of the non-structural proteins as well as and their roles, are outlined in Table 1 below:

Accessory proteins

There are eight accessory proteins reported to date. While they are not essential for replication (suggested by in vitro studies), some of them are proved to be important for virus-host interactions. The group include ORF3a, ORF3b, ORF6, ORF7a, ORF7b, ORF8a, ORF8b, and ORF9b [51]. The details of accessory proteins reported in SARS CoV-2 and their functions are outlined in Table 2 below:

SARS-Cov-2 infection and co-infections

The individuals carrying SARS-CoV-2 are more vulnerable to co-infection with pathogens such as Aspergillus flavus, Candida species, Streptococcus pneumoniae, Staphylococcus aureus, Klebsiella pneumoniae, Haemophilus influenzae, Mycoplasma pneumoniae, Chlamydia pneumonia, Legionella pneumophila, Acinetobacter baumannii, influenza coronavirus, rhinovirus/enterovirus, parainfluenza virus, metapneumovirus, and human immunodeficiency virus (HIV) [61]. Cases of SARS-CoV-2 co-infection with human metapneumovirus (hMPV), human orthopneumovirus (human respiratory syncytial virus/HRSV/RSV), Mycoplasma pneumoniae (MP), rhinovirus/enterovirus, non-SARS-CoV-2 Coronaviridae [62,63], and influenza A virus [64–66] have been reported. According to current medical literature, co-infection between SARS-CoV-2 and bacteria/fungi appear to be low. 2 out of 9 (22%) clinical studies reported bacterial co-infection in SARS-CoV-2 cases, whereas 62 out of 806 (8%) cases of bacterial/fungal co-infection were reported. These patients were put on broad-spectrum antibiotic treatment, with 72% of cases getting antibacterial therapy alone [67]. Co-infections with bacteria such as S. pneumoniae, followed by K. pneumoniae and H. influenzae were commonly reported in SARS-CoV-2 patients [68]. There are also studies reporting SARS-CoV-2 patients suffering with severe invasive pulmonary aspergillosis [69,70]. Among SARS-CoV-2 patients the proportion of bacterial co-infection was highest, followed by bacterial-viral, viral-fungal and viral-bacterial-fungal co-infections [68].

Altogether rates of co-infections in SARS-CoV-2 patients are increasing above what was previously reported. Although the number is small, the chance of co-infection is higher, which needs to be investigated in detail.

Table 1Non-structural proteins of SARS CoV-2 and their function.

NSP	Amino acid (aa)	Function ^a	Accession number	Reference
NSP1 (leader proteins)	180 aa	Blocks host innate immune response and suppresses IFN signalling by binding to host 40S ribosome, halting translation and thereby selectively degrading host mRNA.	YP_009725297.1	[20]
NSP2	638 aa	Found conserved in another coronavirus, SARC-CoV. This protein binds with prohibitin 1(PHB1) and prohibitin 2 (PHB2) present in host, and thus is responsible for disrupting the host cell environment.	YP_009725298.1	[33]
NSP3	1945 aa	The largest protein encoded by coronavirus, it is around 200 KDa in size and a papain-like proteinase protein. This protein also eases release of NSP1, NSP2, and NSP3 from the N-terminal region. NSP3 shuts down host enzymes called PARPs, which prevent viruses from replicating.	YP_009725299.1	[34,35]
NSP4	500 aa	Accommodates transmembrane domain and interacts with NSP3 as well as host proteins and assists reorganization of SARS CoV membrane. However, both NSP4 and NSP3 are involved together for their role in viral replication.	YP_009725300.1	[36]
NSP5	306 aa	The main protease promoting cytokine expression and cleavage of viral polyprotein. SARS CoV-2 NSP5 is highly homologous to SARS NSP5 (identity, 96%; similarity, 98%).	YP_009725301.1	[37]
NSP6	290 aa	This, similarly to other CoVs, presents putative trans-membrane helices and interacts with NSP3 and NSP4. This protein is involved in generation of autophagosome from the endoplasmic reticulum and enable assembly of replicase proteins.	YP_009725302.1	[38,39]
NSP7 and NSP8	83 aa	Both NSP7 and NSP8 form a heterodimer that efficiently performs de novo initiation and primer extension. Both NSP7 and NSP8 are found conserved among 2019-nCoV, BetaCoV.RaTG, and BatSARS-like Cov.	NSP7, YP_009725303.1	[40,41]
	198 aa		NSP8, YP_009725304.1	
NSP9 NSP10	113 aa 139 aa	Suggested involvement in viral replication and virulence. Previous studies of SARS coronavirus demonstrate that NSP10 interacts and stimulates activity of NSP14 [S-adenosylmethionine (SAM)-dependent (guanine-N7) methyl transferase (N7-MTase)]. Additionally, NSP10 also has a crucial role in activating NSP16 (2'-O-methyltransferase).	YP.009725305.1 YP.009725306.1	[42] [43,44]
NSP11	13 aa	Function is still unknown.	YP_009725312.1	[20]
NSP12	932 aa	RNA dependent RNA polymerase (RdRp), along with cofactors NSP7 and NSP8, assists in coping viral RNA. Current studies suggest that SARS-CoV-2 NSP12 is almost identical to that of the SARS-CoV (identity, 96%; similarity, 98%).	YP_009725307.1	[45]
NSP13	601 aa	A helicase enzyme responsible for unwinding viral genome. As reported previously, the overall structure of SARS-CoV-2 NSP13 is composed of five domains giving a triangular pyramid shape similar to SARS and MERS-Nsp13.	YP_009725308.1	[46]
NSP14	527 aa	Current reports suggest it is a proofreading protein with 3' to 5' exoribonuclease (NSP14-ExoN). This activity is an important factor of both viral replication and recombination.	YP_009725309.1	[47]
NSP15	346 aa	It has been characterized as RNA uridylate-specific endoribonuclease carrying catalytic domain at C-terminal. The NSP15 protein prevents uncovering of virus within host system. This is achieved by degrading the viral polyuridine sequences.		[48]
NSP16	298 aa	It is a N7-GpppA-specific, S-adenosyl-L-methionine (SAM)-dependent, 2'-O-MTase and is activated by binding to NSP10. NSP16-NSP10 complex cap viral mRNA transcripts protecting it from degradation by 5'-exoribonucleases, promote efficient translation and assist host innate immunity surveillance.	YP.009725311.1	[49,50]

^a Note: The functions of NSPs are reported for SARS-CoV and are considered to be similar in SARS-CoV-2.

Potential targets for drug discovery in SARS-Cov-2

SARS-CoV-2 infection has already proven to be a devastating disease worldwide. Therefore, the most urgent timely requirement now is to advance treatment options against this disease. The strategy of drug development for tackling this global disaster can be broadly classified on the basis of specific pathways into the following categories: (a) blocking viral structural proteins and thereby inhibiting virus-host interaction and viral entry; (b) inhibiting viral RNA synthesis and replication by targeting different viral enzymes or functional proteins; (c) targeting viral virulence

factors mediating escape from the host immune system; (d) targeting host-specific receptors such as Angiotensin-converting enzyme 2 (ACE2), which serves as an entry point for CoVs.

Targeting viral structural protein and its interactions

Spike protein is a crucial structural protein of CoVs and forms a trimeric structure on the surface and mediates the invasion and virulence of the virus. The S protein is also responsible for activating the host immune response toward the viral particle [71]. There-

Table 2Accessory proteins of SARS CoV-2 and their function.

Accessory proteins	Amino acid (aa)	Function ^a	Reference
ORF3a	275 aa	Involved in formation of ion channels, virulence, infectivity and virus release	[52,20]
ORF3b	22 aa	Strong INF-1 antagonist.	[53]
ORF6	61 aa	Interacts with viral NSP8 (enhancing RNA polymerase activity) and involved in viral pathogenesis	[20]
ORF7a	121 aa	Inhibits activity of bone marrow stromal antigen 2 (BST-2) by blocking its glycosylation.	[54]
ORF7b	43 aa	Both accessory protein and structural component of the SARS virion.	[55]
ORF8	121 aa	Important for adaptation in human host following interspecies transmission and virus replicative efficiency.	
ORF9b	97 aa	Interacts with a mitochondrial import receptor, Tom70, which acts as an essential adaptor linking MAVS to TBK1/IRF3; resulting in the activation of IRF-3	
ORF9c	XX aa	Interacts with multiple proteins that modify IkB kinase and NF-kB signalling pathway, including NLRX1, F2RL1 and NDFIP2.	[60]
ORF10	38 aa	Function is undefined	[20]

^a Note: The functions of accessory proteins are reported for SARS-CoV and are considered to be similar in SARS-CoV-2.

fore, targeting S proteins or specific host cell receptors is a valuable therapeutic strategy for antiviral drug development.

The receptor-binding domain (RBD) is the main target for designing drugs against SARS-CoV-2. Available literature suggested a few potential targets that hamper S1-RBD: ACE-2 binding and therefore, block the entry of SARS-CoV. The inhibitors are: OC43-HR2P (peptide derived from HCoV-OC43) showed pan-CoV fusion inhibition property [72], chloroquine (antimalarial agent, elevates endosomal pH and modifies ACE-2 binding site, thus inhibiting virus receptor binding) [73], SSAA09E2 (block the S-ACE2 binding), SSAA09E1 (blocks viral entry), SSAA09E3 (inhibits host and viral cell membrane fusion) [74], the S230 antibody nullifies various isolates of SARS-CoV [75], m396 (monoclonal antibody) competes for RBD [76], 80R and CR301 (monoclonal antibodies) are spike-specific antibodies that nullify viral infection by preventing S-ACE-2 binding [77].

The other structural proteins in SARS-CoV-2 are E protein and N protein. E protein (E-channel) owns the central function for maintaining the structural and viral pathogenicity, whereas NRBD and CRBD are the important domains of Cov N protein and they are required for an efficient host-viral interaction. Therefore, collectively, these structural proteins can be thoroughly targeted for antiviral drug discovery [78].

Targeting virus RNA synthesis and replication

Non-structural proteins are important for virus replication along with infecting the host. The most potential drug targets among them are: papain-like protease (PLpro), helicase/NTPase, 3C-like protease (3CLpro), haemagglutinin esterase and RNA-dependent RNA polymerase (RdRp), because of their strong vital role and functional enzyme active site.

C-like protease (3CLpro/Nsp5)

The 3CLpro/(Nsp5 is a homodimer protease with an active site consisting of the cys-his dyad responsible for protease activity [79]. It releases mature Nsp4-Nsp16 by cleaving Nsps present downstream at 11 sites, and facilitates production of advanced protein-mediating replication/transcription complex [80,81]. Due to important catalytic activity 3CLpro is an attractive target for developing antiviral drugs and mainly small-molecules and peptide inhibitors are used for screening [82].

An in silico study [83] indicated that antibacterial medications (oxytetracycline, demeclocycline, doxycycline and lymecycline), conivaptan (used for hyponatremia) and anti-hypertensive drugs (nicardipine and telmisartan) were inhibitors of 3CLpro. Other In silico studies suggested potential 3CLpro inhibitors among

presently available drugs (aprepitant, icatibant, colistin, bepotastine, perphenazine, valrubicin, epirubicin, and caspofungin. These drugs also bind to the antiretroviral-binding site on SARS-CoV [84]. Small molecules, phthalhydrazide-substituted ketoglutamine analogs, arylboronic acids, thiophenecarboxylate and quinolinecarboxylate derivatives have been explored and proved to inhibit 3CLpro [85]. The inhibitors of HIV protease, lopinavir and ritonavir also inhibit 3CLpro [84]. Various natural products and their derivatives have been reported to show high binding affinity to 3CLpro [83].

Papain-like proteases (PLpro)

These function by slicing the N-terminus of the replicase poly protein (PP) and produce three NSPs (NSP1, NSP2, and NSP3) which are critical for virus replication [86]. Being vital enzymes for CoV replication and host infection, PLpro are becoming a well-accepted focus for drug advances against SARS-CoV-2. However, there is no candidate yet approved by the FDA as a drug. Zinc and its conjugates at higher doses were found to inhibit PLpro [87]. Benzodioxole [88] and a new lead compound (6577871) [89] were identified as strong inhibitors by in silico studies. Lopinavir-ritonavir combinations are also being used for treating SARS-CoV-2 infection [90]. Wu and coworkers (2020) have discussed a series of available drugs as well as natural products with strong affinity towards PLpro. The major drugs include antibacterial drugs (chloramphenicol, cefamandole and tigecycline), and antiviral drugs (ribavirin, valganciclovir and thymidine) [83].

RNA-dependent RNA polymerases (RdRp/Nsp12)

These are crucial enzymes of the replication/transcription complex and are found conserved in coronavirus. The RdRp domain of the RNA polymerase consists of a conserved motif (Ser-Asp-Asp) present at the C-terminus [91]. However, in previous beta coronavirus epidemics Nsp12-RdRp was considered as a significant drug target because inhibition of this enzyme significantly reduces toxicity and side effects in host cells [92]. Remdesivir and Ribavirin (antiviral agents) have the potential to serve as drug candidates that can block this enzyme [93]. Several other existing compounds are also presented as probable inhibitors of this enzyme, namely, itraconazole, novobiocin, chenodeoxycholic acid, cortisone, idarubicin, silybin and pancuronium bromide [83].

Helicase/NTPase

Helicase (NSP13) is a vital protein which has a critical role in the viral central dogma, with an ability to untangle double-stranded DNA and RNA in an NTP-dependent manner [94]. The SARS-Nsp13 sequence has been found conserved, and is a vital

Table 3 Drug repurposing for SARS-Cov-2.

Possible targets	Ongoing therapeutic options and their functions	Ongoing clinical trials for SARS-CoV-2
Targeting the RdRp	Remdesivir (GS-5734) [115] commonly known as Veklury; broad-spectrum antiviral drug against SARS-CoV, MERS-CoV, and various other RNA viruses.	NCT04252664, NCT04257656, NCT04252664 NCT04292899, NCT04292730, NCT04302766 NCT04323761, NCT04280705, NCT04321616 NCT04315948, NCT04314817 and
	Favipiravir (T-705) [117]; antiviral drug used to treat broad range of RNA viruses.	NCT04349410 ChiCTR2000029600, NCT04358549, NCT04346628, NCT04310228, NCT04349241 NCT04336904, NCT04319900, NCT04359615 NCT04333589, NCT04303299, NCT04351295 NCT04356495 and NCT04345419
	Galidesivir (BCX4430) [118]; broad-spectrum antiviral drug.	NCT04556495 and NCT04545419 NCT03891420
	β-D-N4-hydroxycytidine/NHC/EIDD-1931; strong inhibitory effect against MERS-CoV, SARS-CoV, and SARS-CoV-2 [119].	Data not available
	Ribavirin [118]; broad-spectrum antiviral, primarily used for treatment of hepatitis C.	NCT04356677
nhibiting the viral protease	Ivermectin [120] (Stromectol/Soolantra cream); drug used to treat parasitic infections.	NCT04343092
	Lopinavir-Ritonavir [121]; used for treatment and prevention of HIV/AIDS.	NCT04252885 and ChiCTR2000029308
	Darunavir and Cobicistat [119, 122]; antiretroviral drug against HIV/AIDS	NCT04252274, NTC04303299 and NCT04366089
locking virus cell entry	Recombinant human angiotensin-converting enzyme 2 (RhACE2 APN01) [123]. Used for treating cancer and related problems.	NCT04287686 and NCT04335136
	Arbidol (Umifenovir) [124]. Used for the treatment of influenza and hepatitis C virus. Natural killer cells (NK cells) [125]; play important role	NCT04260594, NCT04255017 and IRCT20180725040596N2 NCT04280224
Enhancing the innate immune system	in cancer immunotherapy. Recombinant interferon [126]; used as an antiviral or	NCT04293887
	antineoplastic drug. Mesenchymal stem cells (MSCs) [127]; found to be effective on acute lung injury (ALI)/acute respiratory distress syndrome (ARDS) occurred by both infectious	NCT04293692, NCT04269525, NCT04288102 and NCT04302519
Attenuating the inflammatory response	and noninfectious diseases. Intravenous immunoglobulin (IVIG) [128]; used to treat a number of health conditions.	NCT04261426
	Neutralizing antibodies (nAbs) [129]; used for anti-retroviral treatment.	Data not available
	Anti-C5a monoclonal antibody [130]; used for treating paroxysmal nocturnal hemoglobinuria and atypical hemolytic uremic syndrome (aHUS).	Data not available
	Blocking the interleukin (IL)-6 Pathway [131]; IL-6 inhibitors are approved for treatment of rheumatoid arthritis.	NCT04315298
	Thalidomide (Thalomid) [132]; used to treat or prevent Hansen's disease (leprosy). Also used in cancer treatment.	NCT04273529 and NCT04273581
	Methylprednisolone (Medrol) [133]; eases inflammation, used to treat arthritis and cancer.	NCT04273321 and NCT04263402
	Fingolimod (Gilenya) [134]; mostly used for treating multiple sclerosis.	NCT04280588
ymptomatic control	Bevacizumab (Avastin) [135]; used to treat cancer and eye disease. Lipid nanoparticle (LNP)-encapsulated mRNA [136].	NCT04275414 NCT04283461
/accine	mRNA-1273, a novel LNP-encapsulated mRNA-based vaccine, encoding full-length, prefusion stabilized	NC104265401
	spike (S) protein of SARS-CoV-2. DNA vaccine (INO-4800) [137], being tested to prevent COVID-19 infection.	NCT04336410
	AZD1222 (ChAdOx1 nCoV-19) [138]; under clinical trials for COVID-19.	NCT04324606
	Nanoparticle-based vaccines [139]; candidate vaccines against various viral infections.	Data not available
Pathogen-specific artificial antigen-presenting cells	Clinical trials are evaluating the safety and immunogenicity of artificial antigen-presenting cells (aAPCs) alone and in combination with antigen-specific cytotoxic T cells [140]	NCT04299724 and NCT04276896

factor for the replication of CoV. Therefore, NSP13 has been recognized as a potential druggable target [95,96]. However, toxicity due to the non-specificity of inhibitors is considered to be the biggest hurdle [97]. On the basis of in silico studies drugs namely, lymecy-

cline, cefsulodine, rolitetracycline, itraconazole and saquinavir were expected to be NTPase inhibitors [83].

Apart from the abovementioned drug targets, some NSPs that are critical players in viral RNA synthesis and replication, namely:

NSP3b, NSP3e, NSP6, NSP7-8 complex, NSP9, NSP10, NSP12 and NSP14-16, need further investigation for anti-viral drug discovery [60].

Targeting virulence factors

The SARS-CoV virulence factors that help the virus to escape the host immune system as well as interfere with the host's innate immunity are Nsp1, Nsp3c and ORF7. (a) Nsp1 inhibits type-I interferon production and is also responsible for degradation of the host mRNA [98,99]. (b) Nsp3c supports the in virus to resisting host innate immunity by binding with host's ADP-ribose [100]. (c) ORF7a binds and inhibits activity of the bone marrow matrix antigen 2 (BST-2) by blocking its glycosylation [54]. These effects of the virulence factors therefore advocate their potential for anti-viral drug advances.

Targeting host-specific receptors

Many studies have already unambiguously proved ACE2 as a receptor for S-RBD of coronavirus [101]. Recent work proves that the SARS-CoV-2 host receptor is constant among SARS-CoVs, therefore, the sequence of S-RBD in SARS-CoV-2 is similar, and central linking exists between the RBD receptor-binding motif and ACE2 receptors [102]. Therefore, ACE2 is considered as viable drug target for handling this infection. Arbidol (a broad-spectrum antiviral agent), which works against the influenza virus by inhibiting virushost cell fusion and preventing virus entry into host cells, is under clinical trial for the treatment of SARS-CoV-2. Camostat mesvlate. an existing TMPRSS2 (host cell protease, responsible for processing of spike protein and facilitating ACE2 binding) inhibitor, stops the entry of SARS-CoV-2 into host cells; thus, indicating its use as a prospective drug against this infection [93]. Despite several challenges and disagreements about targeting host receptors, several professional societies have recommended using this strategy for the treatment in COVID-19 patients.

Available regimens

Drug repurposing for SARS-Cov-2

To date, no medication has been approved for treating SARS-Cov-2 infection. Prevailing therapeutic options are not effective against this infection; however, a variety of possible treatments options are being explored by scientists [103]. In this situation the best way to tackle this infection will be drug repurposing. Considering the information obtained from genomic sequence along with in silico protein modelling, researchers have been working hard to find a way to defeat this menace.

Recently published work has recognized the interaction of SARS CoV-2 proteins with 332 human proteins. Out of these 332 protein-protein interactions, 66 were targetable by different antiviral compounds. All these compounds were further screened by multiple viral assays, resulting in the identification of two classes of antiviral compounds: (i) protein translation inhibitors (i.e., zotatifin, ternatin-4, and PS3061) and (ii) regulators Sigma1 and Sigma2 receptors (i.e., approved drugs: clemastine, cloperastine, progesterone and PB28) [60]. Some of these drugs have been reported to be more effective than hydroxychloroguine. In another study, researchers have screened about 12,000 FDA-approved drugs in clinical trials in Vero-E6 cells (African Green Monkey kidney) against SARS CoV-2 infection. Compounds, namely: apilimod (PIKfyve kinase inhibitor), cysteine protease inhibitors (MDL-28170, Z LVG CHN2, VBY-825, and ONO 5334), and a CCR1 antagonist (MLN-3897), were identified as potential druggable candidates against COVID-19 [104].

Severity of SARS CoV-2 infection is mostly among elderly patients, which is may be due to weak immune response due to the age factor. Therefore, adapting ways to boost innate immunity against viral attack shows great potential. A previous study has shown a promising role of macrophages and NK cells in the clearance of SARS-CoV after their pulmonary migration and thereby raising the levels of chemokines and cytokines [105]. There are several multinational companies who are utilizing this approach and aiming to reuse their NK-based products to combat COVID-19 infection. The most promising step has been taken by Cellularity (a USA-based company) by developing CYNK-001. Also, Type I interferons, used alone or in combination, give a broad-spectrum protection against viral infections including MERS-CoV [106] and SARS-CoV [107].

The SARS viral infection is majorly correllated with a high inflammatory response in the respiratory tract [108,109]. Hence, targeting mesenchymal stem cells (MSC) for therapeutic use in viral treatment has been proposed by various researchers, as these cells are acknowledged to exert anti-inflammatory responses and initiate the tissue repair mechanism [110]. The purpose of MSCs in treatment of COVID-19 pneumonia is still being investigated. Similarly, intravenous immunoglobulin (IVIG) is gaining attention for the treatment [111]. However, a more to the point approach for SARS-CoV treatment could be generation of surface specific epitope-targeting neutralizing antibodies [112]. Unfortunately, this is a time-consuming process and requires a lot of effort. The anti-C5a monoclonal antibodies may attenuate the degree of lung damage caused by COVID-19 by lowering the neutrophil influx and vascular leakage into the alveolar space [113]. When blocking the interleukin (IL)-6 pathway, as previously reported, a high level of IL-6 in blood rapidly reduces lung elasticity, resulting in acute bronchoalveolar inflammation. Thus, specific blocking of the IL-6-regulated signalling cascade may be considered as a valuable approach towards treatment [114]. The drug thalidomide which shows anti-inflammatory and anti-fibrotic effects, may reduce lung injury, and therefore it is also in the pipeline for COVID-19 patients. Moreover, clinical evaluation of methylprednisolone (a synthetic glucocorticoid) and fingolimod (an immunomodulating drug) for suppressing the undesired immune responces caused by SARS-CoV, is still in progress.

Every day knowledge about the COVID-19 virus is being updated and supporting the vaccine development process. However, that is a long-term stratagem to combat future outbreaks of SARS-CoV. There are various proposed vaccine candidates which are nucleic acid-based and in which the core revolves around the sequence-coding S protein. For instance, mRNA-1273 (Moderna) is developed to stimulate antiviral reaction, particularly against Sprotein of COVID-19. Unlike traditional vaccines, development of this lipid nanoparticle (LNP)-encapsulated mRNA vaccine is free from any inactivated virus particle or subunit of live virus. The INO-4800 (Inovio Pharmaceuticals), is a genetic vaccine candidate that triggers immune response after being delivered to host cells. Additionally, these vaccines offer low cost and simple purification procedures as compared with traditional vaccines, and their simple structure prevents the chance of incorrect folding, which is a possibility in protein-based recombinant vaccines [106,110]. But the potency will depend upon the administration route and the numbers and intervals of plasmid doses delivered inside the body. Another candidate under evaluation is AZD1222/ChAdOx1 nCoV-19 (University of Oxford) constituting an adenovirus vector (non-replicating) carrying the COVID-19-S protein genetic sequence, making it comparatively safer in elderly individuals and children. The wide tissue (gastrointestinal and respiratory epithelium) range covered by adenovirus-based vectors increases their likelihood of forming an effective vaccine. However, the dominating immunogenicity for the vector will always be a concern [115].

Table 4Patented vaccines against SARS-CoV and MERS-CoV [93].

Type of vaccine	Patent application	Target
Attenuated virus vaccines	US20060039926	The vaccine incorporates a mutation at specific tyrosine residue (Y6398H) into the viral genome encoding a p59 protein. Showing completely attenuated growth and pathogenicity of mouse coronavirus (MHV-A59).
DNA-based vaccines	WO2005081716	DNA vaccine targeting antigens of SARS-CoV, epitopes of the Membrane (M), Envelope (E), Spike (S) and Nucleocapsid (N) proteins of the virus. Stimulates immune responses (antigen-specific CD8+T cell mediated) against SARS-CoV antigens.
	WO2015081155	DNA-based vaccine comprised of MERS-CoV antigen (consensus spike protein).
Protein-based vaccines	WO2010063685	The vaccine comprises an immunogenic SARS spike protein and an adjuvant comprising an oil-in-water emulsion. Induces a protective immunity against the virus.
	US20070003577	The vaccine comprises purified trimeric S protein of SARS CoV, showing specific binding to ACE2 receptor.
	US20060002947	li-key/antigenic epitope hybrid peptide vaccines under clinical trials against COVID-19.
VIRUS-like particle vaccines	W02015042373	The vaccine is composed of nanoparticles containing MERS virus proteins in polymer structures. Induces a neutralizing antibody response to MERS that reduces or prevents infection in mice and transgenic cattle.
mRNA-Based vaccines	WO2017070626	RNA vaccine and combination vaccine, composed of at least one mRNA encoding antigenic viral full-length S, S1, or S2 proteins from SARS-CoV and MERS-CoV virus, formulated in a cationic lipid nanoparticle.
	WO2018115527	mRNA based vaccine, encoding at least one antigen derived from a MERS-CoV and induces humoral immune responses.

Researchers are also putting effort into evaluating the efficacy of certain genetically modulated artificial antigen-presenting cells (aAPCs) specifically presenting the SARS-CoV structural proteins (conserved domains), and probably helping cells to endure the penetration of COVID-19 [116].

Moreover, there are several existing antiviral drugs which are under clinical trials for their potential against SARS-Cov-2 infection. Table 3 summarises the drugs that are in clinical trial which help in combating novel SARS-CoV2 infection.

Vaccines under development for COVID-19

Owing to the high morbidity and mortality associated with SARS-CoV-2 infection there is an urgent need for mitigation methods, and one of such method is vaccine development. Thorough research suggested that there is a significant sequence homology between SARS-CoV-2 and other beta-coronaviruses (SARS and MERS). The vaccines identified for these lethal coronaviruses can therefore be of high value in preparing vaccines against SARS-CoV-2 [141].

The viral S-protein based vaccine development approach has drawn the attention of many scientists in the fight against COVID-19. These subunit vaccines can elicit protective immunity, producing higher neutralizing antibodies as compared with DNA-based S protein vaccines, live-attenuated coronavirus and full-length S protein [142]. Presently, 188 well-established patents are present in the Chemical Abstracts Service (CAS) collection related to anti-SARS and anti-MERS vaccines. Most of them are associated with the S protein subunit vaccine and vaccines specifically targeting S-RBD [90]. Therefore, the preferred target for vaccine development against beta-coronaviruses is the S protein/gene, and applying the same strategy and knowledge will be beneficial in developing vaccines against SARS-CoV-2 [23,141]. Table 4 comprehends the list of SARS and MERS vaccines that have been patented.

Moreover, the prospect of short-term immunogenicity resulting from neutralising antibodies should be addressed. Along with B cell response, T cell response should also be considered, because these responses are protective and persistent in humans. Strategies for augmenting immunogenicity and preventing undesired side effects should be explored [141].

Summary

RNA viruses (SARS-CoV, MERS-CoV and SARS-CoV-2), causing severe mortal infections, will continue to be a serious global threat in future. These viruses have a high rate of mutation, genetic recombination and the capability of cross-species transmission, which make them a menace to mankind. The present outbreak of COVID-19 should direct us towards uplifting our knowledge and expertise in combating stubborn microbial pathogens and solving global health problems.

The present review summarizes recent research and developmental information published on international platforms related to SARS-CoV-2 infection, current therapeutic options and preventive vaccines. It includes a complete overview of the SARS-CoV-2 morphology, pathogenesis and antiviral strategies. Also, the drugs under clinical trials for COVID-19 have been discussed in detail, with the main focus on drug-repurposing which may be the best way out of this tragic situation. Literature strongly relates sequence similarity between SARS-CoV-2 and other beta-coronaviruses, and about 77.1% of the proteins found in SARS-CoV-2 are also reported in SARS CoV [143]. Mere ignorance of previous research on SARS and MERS proteins will increase vulgarity in drug and vaccine development for SARS-CoV-2. Therefore, utilizing the literature to forge a better understanding of SARS CoV-2 infection will enable us to design better antiviral drugs/vaccines against the virus. Furthermore, to speed up this drug development process, additional structural biological information including life-cycle details of the virus are required. At the same time, action should be taken to enhance SARS-CoV-2 surveillance systems, and individuals with symptoms (fever, cough or sore throat, diarrhea, body ache and rashes) should be screened for COVID-19.

Funding

No funding sources.

Competing interests

None declared.

Ethical approval

Not required.

References

- [1] ICTV Master Species List 2009-v10, International Committee on Taxonomy of Viruses
- [2] Gorbalenya AE, Enjuanes L, Ziebuhr J, Snijder EJ. Nidovirales: evolving the largest RNA virus genome. Virus Res 2006;117:17–37, http://dx.doi.org/10. 1016/j.virusres.2006.01.017.
- [3] Almeida JD, Berry DM, Cunningham CH, Hamre D, Hofstad MS, Mallucci L, et al. Virology: coronaviruses. Nature 1968;220:650, http://dx.doi.org/10. 1038/220650b0.
- [4] Anthony SJ, Johnson CK, Greig DJ, Kramer S, Che X, Wells H, et al. Global patterns in coronavirus diversity. Virus Evol 2017;3:vex012, http://dx.doi. org/10.1093/ve/vex012.
- [5] Su S, Wong G, Shi W, Liu J, Lai ACK, Zhou J, et al. Epidemiology, genetic recombination, and pathogenesis of coronaviruses. Trends Microbiol 2016;24:490–502, http://dx.doi.org/10.1016/j.tim.2016.03.003.
- [6] Zhu N, Zhang D, Wang W, Li X, Yang B, Song J, et al. A novel coronavirus from patients with pneumonia in China, 2019. N Engl J Med 2020;382:727–33, http://dx.doi.org/10.1056/NEJMoa2001017.
- [7] Tang B, Bragazzi NL, Li Q, Tang S, Xiao Y, Wu J. An updated estimation of the risk of transmission of the novel coronavirus (2019-nCov). Infect Dis Model 2020;5:248–55, http://dx.doi.org/10.1016/j.idm.2020.02.001.
- [8] NHS, SARS (severe acute respiratory syndrome). [Online]. https://www.nhs. uk/conditions/sars/. [Accessed 5 June 2020].
- [9] CDC. MERS clinical features, Middle East respiratory syndrome (MERS); 2019[Online]. https://www.cdc.gov/coronavirus/mers/clinical-features.html.[Accessed 5 June 2020].
- [10] WHO [Online]. https://www.who.int/news-room/q-a-detail/q-a-coronaviruses. [Accessed 5 June 2020].
- [11] WHO [Online]. https://covid19.who.int/. [Accessed 9 September 2020].
- [12] Clinical Trials [Online]. https://clinicaltrials.gov/ct2/results?cond=covid19&age_v=&gndr=&type=Intr&rsIt=&Search=Apply.
 [Accessed 9 September 2020].
- [13] Sheahan TP, Sims AC, Zhou S, Graham RL, Pruijssers AJ, Agostini ML, et al. An orally bioavailable broad-spectrum antiviral inhibits SARS-CoV-2 in human airway epithelial cell cultures and multiple coronaviruses in mice. Sci Transl Med 2020;12:eabb5883, http://dx.doi.org/10.1126/scitranslmed.abb5883.
- [14] Livescience [Online]. https://www.livescience.com/flu-drug-could-treat-coronavirus.html. [Accessed 9 September 2020].
- [15] Singh AK, Singh AK, Shaikh A, Singh R, Misra A. Chloroquine and hydrox-ychloroquine in the treatment of COVID-19 with or without diabetes: a systematic search and a narrative review with a special reference to India and other developing countries. Diabetes Metab Syndr 2020;14:241–6, http://dx.doi.org/10.1016/j.dsx.2020.03.011.
- [16] ORCHID: Outcomes Related to COVID-19 treated with Hydroxychloroquine among In-patients with symptomatic disease. ClinicaTrials.gov Identifier: NCT04332991. https://www.clinicaltrials.gov/ct2/show/ NCT04332991?term=NCT04332991&draw=2&rank=1. [Accessed 22 December 2020].
- [17] Boulware DR, Pullen MF, Bangdiwala AS, Pastick KA, Lofgren SM, Okafor EC, et al. A randomized trial of hydroxychloroquine as postexposure prophylaxis for Covid-19. N Engl J Med 2020;383:517–25, http://dx.doi.org/10.1056/NEJMoa2016638.
- [18] Shah RR. Chloroquine and hydroxychloroquine for COVID-19: perspectives on their failure in repurposing. J Clin Pharm Ther 2020;10(September), http://dx. doi.org/10.1111/jcpt.13267.
- [19] Chen X, Liao B, Cheng L, Peng X, Xu X, Li Y, et al. The microbial coinfection in COVID-19. App Microbiol Biotech 2020;104:7777–85, http://dx.doi.org/10. 1007/s00253-020-10814-6.
- [20] Yoshimoto FK. The proteins of severe acute respiratory syndrome coronavirus-2 (SARS CoV-2 or n-CoV19), the cause of COVID-19. Protein J 2020;39:198–216, http://dx.doi.org/10.1007/s10930-020-09901-4.
- [21] da Silva SJR, Alves da Silva CT, Mendes RPG, Pena L. Role of nonstructural proteins in the pathogenesis of SARS-CoV-2. J Med Virol 2020;92:1427–9, http://dx.doi.org/10.1002/jmv.25858.
- [22] Jiang S, Hillyer C, Du L. Neutralizing antibodies against SARS-CoV-2 and other human coronaviruses. Trends Immunol 2020;41:355–9, http://dx.doi.org/10. 1016/i.it.2020.03.007.
- [23] Astuti I, Ysrafil. Severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2): an overview of viral structure and host response [published online ahead of print, 2020 Apr 18]. Diabetes Metab Syndr 2020;14:407–12, http://dx.doi.org/10.1016/j.dsx.2020.04.020.
- [24] Du L, He Y, Zhou Y, Liu S, Zheng BJ, Jiang S. The spike protein of SARS-CoV a target for vaccine and therapeutic development. Nat Rev Microbiol 2009;7:226–36, http://dx.doi.org/10.1038/nrmicro2090.
- [25] Du L, Yang Y, Zhou Y, Lu L, Li F, Jiang S. MERS-CoV spike protein: a key target for antivirals. Expert Opin Ther Targets 2017;21:131–43, http://dx.doi.org/ 10.1080/14728222.2017.1271415.
- [26] Schoeman D, Fielding BC. Coronavirus envelope protein: current knowledge. Virol J 2019;16:69, http://dx.doi.org/10.1186/s12985-019-1182-0.

- [27] Tai W, He L, Zhang X, Pu J, Voronin D, Jiang S. Characterization of the receptor-binding domain (RBD) of 2019 novel coronavirus: implication for development of RBD protein as a viral attachment inhibitor and vaccine. Cell Mol Immunol 2020;17:613–20, http://dx.doi.org/10.1038/s41423-020-0400-4
- [28] Seah I, Su X, Lingam G. Revisiting the dangers of the coronavirus in the ophthalmology practice. Eye (Lond) 2020;34:1155–7, http://dx.doi.org/10.1038/ s41433-020-0790-7.
- [29] Kang S, Yang M, Hong Z, Zhang L, Huang Z, Chen X, et al. Crystal structure of SARS-CoV-2 nucleocapsid protein RNA binding domain reveals potential unique drug targeting sites. Acta Pharm Sin B 2020;10:1228–38, http://dx. doi.org/10.1016/j.apsb.2020.04.009.
- [30] Thomas S. The structure of the membrane protein of SARS-CoV-2 resembles the sugar transporter semiSWEET. Preprints 2020;5:342–63, http://dx.doi. org/10.20944/preprints202004.0512.v1.
- [31] Bianchi M, Benvenuto D, Giovanetti M, Angeletti S, Ciccozzi M, Pascarella S. Sars-CoV-2 envelope and membrane proteins: structural differences linked to virus characteristics? Biomed Res Int 2020;2020:4389089, http://dx.doi. org/10.1155/2020/4389089.
- [32] Masters PS. The molecular biology of coronaviruses. Adv Virus Res 2006;66:193–292, http://dx.doi.org/10.1016/S0065-3527(06)66005-3.
- [33] Cornillez-Ty CT, Liao L, Yates JR, Kuhn P, Buchmeier MJ. Severe acute respiratory syndrome coronavirus nonstructural protein 2 interacts with a host protein complex involved in mitochondrial biogenesis and intracellular signaling. J Virol 2009;83:10314–8, http://dx.doi.org/10.1128/JVI.00842-09.
- [34] Lei J, Kusov Y, Hilgenfeld R. Nsp3 of coronaviruses: structures and functions of a large multi-domain protein. Antiviral Res 2018;149:58–74, http://dx.doi.org/10.1016/j.antiviral.2017.11.001.
- [35] Times of India [Online]. https://timesofindia.indiatimes.com/india/a-covid-guide-understanding-the-s-factor/articleshow/75671621.cms. [Accessed 9 August 2020].
- [36] Sakai Y, Kawachi K, Terada Y, Omori H, Matsuura Y, Kamitani W. Two-amino acids change in the nsp4 of SARS coronavirus abolishes viral replication. Virology 2017;510:165–74, http://dx.doi.org/10.1016/j.virol.2017.07.019.
- [37] Jiang H, Li Y, Zhang H, Wang W, Men D, Yang X, et al. Global profiling of SARS-CoV-2 specific IgG/ IgM responses of convalescents using a proteome microarray. Nat Commun 2020;11:3581, http://dx.doi.org/10.1038/s41467-020-17488-8, 03.20.20039495.
- [38] Benvenuto D, Angeletti S, Giovanetti M, Bianchi M, Pascarella S, Cauda R, et al. Evolutionary analysis of SARS-CoV-2: how mutation of Non-Structural Protein 6 (NSP6) could affect viral autophagy. J Infect 2020;81:e24-7, http://dx.doi.org/10.1016/j.ijinf.2020.03.058.
- [39] Cottam EM, Whelband MC, Wileman T. Coronavirus NSP6 restricts autophagosome expansion. Autophagy 2014;10:1426–41, http://dx.doi.org/ 10.4161/auto.29309.
- [40] te Velthuis AJ, van de Worm SH, Snijder EJ. The SARS-coronavirus nsp7+nsp8 complex is a unique multimeric RNA polymerase capable of both de novo initiation and primer extension. Nucleic Acids Res 2012;40:1737–47, http:// dx.doi.org/10.1093/nar/gkr893.
- [41] Fahmi M, Kubota Y, Ito M. Nonstructural proteins NS7b and NS8 are likely to be phylogenetically associated with evolution of 2019-nCoV. Infect Genet Evol 2020;81:104272, http://dx.doi.org/10.1016/j.meegid.2020.104272.
- [42] Littler DR, Gully BS, Colson RN, Rossjohn J. Crystal structure of the SARS-CoV-2 non-structural protein 9, Nsp9. iScience 2020;23:101258, http://dx.doi.org/ 10.1016/i.isci.2020.101258.
- [43] Ma Y, Wu L, Shaw N, Gao Y, Wang J, Sun Y, et al. Structural basis and functional analysis of the SARS coronavirus nsp14-nsp10 complex. Proc Natl Acad Sci U S A 2015;112:9436-41, http://dx.doi.org/10.1073/pnas.1508686112.
- [44] Wang Y, Sun Y, Wu A, Xu S, Pan R, Zeng C, et al. Coronavirus nsp10/nsp16 methyltransferase can be targeted by nsp10-derived peptide in vitro and in vivo to reduce replication and pathogenesis. J Virol 2015;89:8416–27, http://dx.doi.org/10.1128/IVI.00948-15.
- [45] Shannon A, Le NT, Selisko B, Eydoux C, Alvarez K, Guillemot J-C, et al. Remdesivir and SARS-CoV-2: structural requirements at both nsp12 RdRp and nsp14 Exonuclease active-sites. Antiviral Res 2020;178:104793, http://dx.doi.org/10.1016/j.antiviral.2020.104793.
- [46] Mirza MU, Froeyen M. Structural elucidation of SARS-CoV-2 vital proteins: computational methods reveal potential drug candidates against main protease, Nsp12 polymerase and Nsp13 helicase. J Pharm Anal 2020;10:320-8, http://dx.doi.org/10.1016/j.jpha.2020.04.008.
- [47] Gribble J, Pruijssers AJ, Agostini ML, Anderson-Daniels J, Chappell JD, Lu X, et al. The coronavirus proofreading exoribonuclease mediates extensive viral recombination. bioRxivorg 2020, http://dx.doi.org/10.1101/2020.04.23.057786
- [48] Hackbart M, Deng X, Baker SC. Coronavirus endoribonuclease targets viral polyuridine sequences to evade activating host sensors. Proc Natl Acad Sci U S A 2020;117:8094–103, http://dx.doi.org/10.1073/pnas.1921485117.
- [49] Rosas-Lemus M, Minasov G, Shuvalova L, Inniss NL, Kiryukhina O, Wiersum G, et al. The crystal structure of nsp10-nsp16 heterodimer from SARS-CoV-2 in complex with S-adenosylmethionine. Preprint bioRxiv 2020;2020, http://dx.doi.org/10.1101/2020.04.17.047498, 04.17.047498.
- [50] Decroly E, Debarnot C, Ferron F, Bouvet M, Coutard B, Imbert I, et al. Crystal structure and functional analysis of the Sars-coronavirus RNA cap 2'-O-methyltransferase nsp10/nsp16 complex. PLoS Pathog 2011;7:e1002059, http://dx.doi.org/10.1371/journal.ppat.1002059.

- [51] Liu DX, Fung TS, Chong KK-L, Shukla A, Hilgenfeld R. Accessory proteins of SARS-CoV and other coronaviruses. Antiviral Res 2014;109:97–109, http:// dx.doi.org/10.1016/j.antiviral.2014.06.013.
- [52] Issa E, Merhi G, Panossian B, Salloum T, Tokajian S. SARS-CoV-2 and ORF3a: nonsynonymous mutations, functional domains, and viral pathogenesis. mSystems 2020;5:e00266-20, http://dx.doi.org/10.1128/mSystems.00266-20
- [53] Konno Y, Kimura I, Uriu K, Fukushi M, Irie T, Koyanagi Y, et al. SARS-CoV-2 ORF3b is a potent interferon antagonist whose activity is further increased by a naturally occurring elongation variant. Cell Rep 2020;32:108185, http:// dx.doi.org/10.1016/j.celrep.2020.108185.
- [54] Taylor JK, Coleman CM, Postel S, Sisk JM, Bernbaum JG, Venkataraman T, et al. Severe acute respiratory syndrome coronavirus ORF7a inhibits bone marrow stromal antigen 2 virion tethering through a novel mechanism of glycosylation interference. J Virol 2015;89:11820–33, http://dx.doi.org/10.1128/JVI. 02274-15.
- [55] Schaecher SR, Mackenzie JM, Pekosz A. The ORF7b protein of severe acute respiratory syndrome coronavirus (SARS-CoV) is expressed in virus-infected cells and incorporated into SARS-CoV particles. J Virol 2007;81:718-31, http://dx.doi.org/10.1128/JVI.01691-06.
- [56] Lau SK, Feng Y, Chen H, Luk HK, Yang WH, Li KSM, et al. Severe acute respiratory syndrome (SARS) coronavirus ORF8 protein is acquired from SARS-related coronavirus from greater horseshoe bats through recombination. J Virol 2015;89:10532–47, http://dx.doi.org/10.1128/JVI.01048-15.
- [57] Muth D, Corman VM, Roth H, Binger T, Dijkman R, Gottula LT, et al. Attenuation of replication by a 29 nucleotide deletion in SARS-coronavirus acquired during the early stages of human-to-human transmission. Sci Rep 2018;8:15177, http://dx.doi.org/10.1038/s41598-018-33487-8.
- [58] Su YCF, Anderson DE, Young BE, Zhu F, Linster M, Kalimuddin, S, et al. Discovery of a 382-nt deletion during the early evolution of SARS-CoV-2. mBio 2020;11:987222, http://dx.doi.org/10.1101/2020.03.11.987222.
- [59] Liu XY, Wei B, Shi HX, Shan YF, Wang C. Tom70 mediates activation of interferon regulatory factor 3 on mitochondria. Cell Res 2010;20:994–1011, http:// dx.doi.org/10.1038/cr.2010.103.
- [60] Gordon DE, Jang GM, Bouhaddou M, Xu J, Obernier K, White KM, et al. A SARS-CoV-2-Human protein-protein interaction map reveals drug targets and potential drug-repurposing. Nature 2020;583:459–68, http://dx.doi.org/ 10.1038/s41586-020-2286-9.
- [61] Lai C-C, Wang C-Y, Hsueh P-R. Co-infections among patients with COVID19: The need for combination therapy with non-anti-SARS-CoV-2 agents? J Microbiol Immunol Infect 2020;53:505–12, http://dx.doi.org/10.1016/j.jmii. 2020.05.013
- [62] Kim D, Quinn J, Pinsky B, Shah NH, Brown I. Rates of co-infection between SARS-CoV-2 and other respiratory pathogens. JAMA 2020;323:2085–6, http:// dx.doi.org/10.1001/jama.2020.6266.
- [63] Jiang S, Liu P, Xiong G, Yang Z, Wang M, Li Y, et al. Coinfection of SARS-CoV-2 and multiple respiratory pathogens in children. Clin Chem Lab Med 2020;58:1160-1, http://dx.doi.org/10.1515/cclm-2020-0434.
- [64] Azekawa S, Namkoong H, Mitamura K, Kawaoka Y, Saito F. Co-infection with SARS-CoV-2 and influenza A virus. IDCases 2020;20:e00775, http://dx.doi. org/10.1016/i.idcr.2020.e00775.
- [65] Wu D, Lu J, Ma X, Liu Q, Wang D, Gu Y, et al. Coinfection of influenza virus and severe acute respiratory syndrome coronavirus 2 (SARS-COV-2). Pediatric Infect Dis J 2020;39:e79, http://dx.doi.org/10.1097/INF.0000000000002688.
- [66] Cuadrado-Payan E, Montagud-Marrahi E, Torres-Elorza M, Bordo M, Blasco M, Poch E, et al. SARS-CoV-2 and influenza virus co-infection. Lancet (London England) 2020;395:e84, http://dx.doi.org/10.1016/S0140-6736(20)31052-7.
- [67] Rawson TM, Moore LSP, Zhu N, Ranganathan N, Skolimowska K, Gilchrist M, et al. Bacterial and fungal co-infection in individuals with coronavirus: a rapid review to support COVID-19 antimicrobial prescribing. Clin Infect Dis 2020; 2:ciaa530. http://dx.doi.org/10.1093/cid/ciaa530
- 2020;2:ciaa530, http://dx.doi.org/10.1093/cid/ciaa530.
 [68] Zhu X, Ge Y, Wu T, Zhao K, Chen Y, Wu B, et al. Co-infection with respiratory pathogens among COVID-2019 cases. Virus Res 2020;285:198005, http://dx.doi.org/10.1016/j.virusres.2020.198005.
- [69] Alanio A, Delliere S, Fodil S, Bretagne S, Megarbane B. Prevalence of putative invasive pulmonary aspergillosis in critically ill COVID-19 patients. Lancet Respir Med 2020;8:e48-9, http://dx.doi.org/10.1016/S2213-2600(20)30237-X.
- [70] Koehler P, Cornely OA, Bottiger BW, Dusse F, Eichenauer DA, Fuchs F, et al. COVID-19 associated pulmonary aspergillosis. Mycoses 2020;63:528–34, http://dx.doi.org/10.1111/myc.13096.
- [71] Li F. Structure, function, and evolution of coronavirus spike proteins. Annu Rev Virol 2016;3:237–61, http://dx.doi.org/10.1146/annurev-virology-110615-042301
- [72] Xia S, Yan L, Xu W, Agrawal AS, Algaissi A, Tseng C-TK, et al. A pan-coronavirus fusion inhibitor targeting the HR1 domain of human coronavirus spike. Sci Adv 2019;5:eaav4580, http://dx.doi.org/10.1126/sciadv.aav4580.
- [73] Vincent MJ, Bergeron E, Benjannet S, Erickson BR, Rollin PE, Ksiazek TG, et al. Chloroquine is a potent inhibitor of SARS coronavirus infection and spread. Virol J 2005;2:69, http://dx.doi.org/10.1186/1743-422X-2-69.
- [74] Adedeji AO, Severson W, Jonsson C, Singh K, Weiss SR, Sarafianos SG, et al. Novel inhibitors of severe acute respiratory syndrome coronavirus entry that act by three distinct mechanisms. J Virol 2013;87:8017–28, http://dx.doi.org/10.1128/JVI.00998-13.

- [75] Walls AC, Xiong X, Park YJ, Tortorici MA, Snijder J, Quispe J, et al. Unexpected receptor functional mimicry elucidates activation of coronavirus fusion. Cell 2019;176:1026–39.e15, http://dx.doi.org/10.1016/j.cell.2018.12.028.
- [76] Prabakaran P, Gan J, Feng Y, Zhu Z, Choudhry V, Xiao X, et al. Structure of severe acute respiratory syndrome coronavirus receptor-binding domain complexed with neutralizing antibody. J Biol Chem 2006;281:15829–36, http://dx.doi.org/10.1074/jbc.M600697200.
- [77] Du L, He Y, Zhou Y, Liu S, Zheng BJ, Jiang S, et al. The spike protein of SARS-CoV-a target for vaccine and therapeutic development. Nat Rev Microbiol 2009;7:226–36, http://dx.doi.org/10.1038/nrmicro2090.
- [78] Prajapat M, Sarma P, Shekhar N, Avti P, Sinha S, Kaur H, et al. Drug targets for corona virus: a systematic review. Indian J Pharmacol 2020;52:56–65, http:// dx.doi.org/10.4103/ijp.IJP-115-20.
- [79] Shimamoto Y, Hattori Y, Kobayashi K, Teruya K, Sanjoh A, Nakagawa A, et al. Fused-ring structure of decahydroisoquinolin as a novel scaffold for SARS 3CL protease inhibitors. Bioorg Med Chem 2015;23:876–90, http://dx.doi.org/10. 1016/j.bmc.2014.12.028.
- [80] Hsu MF, Kuo CJ, Chang KT, Chang HC, Chou CC, Ko T-P, et al. Mechanism of the maturation process of SARS-CoV 3CL protease. J Biol Chem 2005;280:31257–66, http://dx.doi.org/10.1074/jbc.M502577200.
- [81] Barretto N, Jukneliene D, Ratia K, Chen Z, Mesecar AD, Baker SC, et al. The papain-like protease of severe acute respiratory syndrome coronavirus has deubiquitinating activity. J Virol 2005;79:15189151–98, http://dx.doi.org/10. 1128/IVI.79.24.15189-15198.2005.
- [82] Pillaiyar T, Manickam M, Namasivayam V, Hayashi Y, Jung SH. An overview of severe acute respiratory syndrome-coronavirus (SARS-CoV) 3CL protease inhibitors: peptidomimetics and small molecule chemotherapy. J Med Chem 2016;59:6595–628, http://dx.doi.org/10.1021/acs.jmedchem.5b0146.1.
- [83] Wu C, Liu Y, Yang Y, Zhang P, Zhong W, Wang Y, et al. Analysis of therapeutic targets for SARS-CoV-2 and discovery of potential drugs by computational methods. Acta Pharm Sin B 2020;10:766–88, http://dx.doi.org/10.1016/j. apsb.2020.02.008.
- [84] Liu X, Wang XJ. Potential inhibitors against 2019-nCoV coronavirus M protease from clinically approved medicines. J Genet Genomics 2020;47:119-21, http://dx.doi.org/10.1016/j.jgg.2020.02.001.
- [85] Liang P-H, et al. Characterization and inhibition of SARS-coronavirus main protease. Curr Top Med Chem 2006;6:361–76, http://dx.doi.org/10.2174/ 156802606776287090
- [86] Harcourt BH, Jukneliene D, Kanjanahaluethai A, Bechill J, Severson KM, Smith CM, et al. Identification of severe acute respiratory syndrome coronavirus replicase products and characterization of papain-like protease activity. J Virol 2004;78:13600–12, http://dx.doi.org/10.1128/JVI.78.24.13600-13612. 2004
- [87] Han YS, Chang GG, Juo CG, Lee HJ, Yeh SH, Hsu JT-A, et al. Papain-like protease 2 (PLP2) from severe acute respiratory syndrome coronavirus (SARS-CoV): expression, purification, characterization, and inhibition. Biochemistry 2005;44:10349–1059, http://dx.doi.org/10.1021/bi0504761.
- [88] Baez-Santos YM, Barraza SJ, Wilson MW, Agius MP, Mielech AM, Davis NM, et al. X-ray structural and biological evaluation of a series of potent and highly selective inhibitors of human coronavirus papain-like proteases. J Med Chem 2014;57:2393-412, http://dx.doi.org/10.1021/jm401712t.
- [89] Ghosh AK, Takayama J, Rao KV, Ratia K, Chaudhuri R, Mulhearn DC, et al. Severe acute respiratory syndrome coronavirus papain-like novel protease inhibitors: design, synthesis, protein-ligand X-ray structure and biological evaluation. J Med Chem 2010;53:4968-79, http://dx.doi.org/10.1021/ ipp.1004489.
- [90] Lim J, Jeon S, Shin HY, Kim MJ, Seong YM, Lee WJ, et al. Case of the index patient who caused tertiary transmission of COVID-19 infection in Korea: the application of Lopinavir/Ritonavir for the treatment of COVID-19 infected pneumonia monitored by quantitative RT-PCR. J Korean Med Sci 2020;35:e79, http://dx.doi.org/10.3346/jkms.2020.35.e79.
- [91] Subissi L, Imbert I, Ferron F, Collet A, Coutard B, Decroly E, et al. SARS-CoV ORF1b-encoded nonstructural proteins 12–16: replicative enzymes as antiviral targets. Antiviral Res 2014;101:122–30, http://dx.doi.org/10.1016/j.antiviral.2013.11.006.
- [92] Chu CK, Gadthula S, Chen X, Choo H, Olgen S, Barnard DL, et al. Antiviral activity of nucleoside analogues against SARS-coronavirus (SARS-COV). Antivir Chem Chemother 2006;17:285–9, http://dx.doi.org/10.1177/095632020601700506.
- [93] Liu C, Zhou Q, Li Y, Garner LV, Watkins SP, Carter LJ, et al. Research and development on therapeutic agents and vaccines for COVID-19 and related human coronavirus diseases. ACS Cen Sci 2020;6:315–31, http://dx.doi.org/10.1021/acscentsci.0c00272.
- [94] Ivanov KA, Ziebuhr J. Human coronavirus 229E nonstructural protein 13: characterization of duplex-unwinding, nucleoside triphosphatase, and RNA 5'-triphosphatase activities. J Virol 2004;78:7833–8, http://dx.doi.org/10. 1128/JVI.78.14.7833-7838.2004.
- [95] Shum KT, Tanner JA. Differential inhibitory activities and stabilisation of DNA aptamers against the SARS coronavirus helicase. Chembiochem 2008;9:3037–45, http://dx.doi.org/10.1002/cbic.200800491.
- [96] Jang KJ, Lee NR, Yeo WS, Jeong YJ, Kim DE. Isolation of inhibitory RNA aptamers against severe acute respiratory syndrome (SARS) coronavirus NTPase/Helicase. Biochem Biophys Res Commun 2008;366:738–44, http:// dx.doi.org/10.1016/j.bbrc.2007.12.020.

- [97] Frick DN, Lam AM. Understanding helicases as a means of virus control. Curr Pharm Des 2006;12:1315–38, http://dx.doi.org/10.2174/ 138161206776361147.
- [98] Kamitani W, Narayanan K, Huang C, Lokugamage K, Ikegami T, Ito N, et al. Severe acute respiratory syndrome coronavirus nsp1 protein suppresses host gene expression by promoting host mRNA degradation. Proc Natl Acad Sci U S A 2006;103:12885–90, http://dx.doi.org/10.1073/pnas.0603144103.
- [99] Narayanan K, Huang C, Lokugamage K, Kamitani W, Ikegami T, Tseng C-TK, et al. Severe acute respiratory syndrome coronavirus nsp1 suppresses host gene expression, including that of type I interferon, in infected cells. J Virol 2008;82:4471–9, http://dx.doi.org/10.1128/JVI.02472-07.
- [100] Forni D, Cagliani R, Mozzi A, Pozzoli U, Al-Daghri N, Clerici M, et al. Extensive positive selection drives the evolution of nonstructural proteins in lineage C betacoronaviruses. J Virol 2016;90:3627–39, http://dx.doi.org/10.1128/JVI. 02988-15.
- [101] Ge XY, Li JL, Yang XL, Chmura AA, Zhu G, Epstein JH, et al. Isolation and characterization of a bat SARS-like coronavirus that uses the ACE2 receptor. Nature 2013;503:535–8, http://dx.doi.org/10.1038/nature12711.
- [102] Wan Y, Shang J, Graham R, Baric RS, Li F. Receptor recognition by novel coronavirus from Wuhan: an analysis based on decade-long structural studies of SARS. J Virol 2020;94:e00127–20, http://dx.doi.org/10.1128/JVI.00127-20.
- [103] Jean S-S, Lee P-I, Hsueh P-R. Treatment options for COVID-19: the reality and challenges. J Microbiol 2020;53:436–43, http://dx.doi.org/10.1016/j.jmii. 2020.03.034.
- [104] Riva L, Yuan S, Yin X, Martin-Sancho L, Matsunaga N, Burgstaller-Muehlbacher S, et al. A large-scale drug repositioning survey for SARS-CoV-2 antivirals. Preprint bioRxiv 2020, http://dx.doi.org/10.1101/2020.04.16.044016.
- [105] Chen J, Lau YF, Lamirande EW, Paddock CD, Bartlett JH, Zaki SR, et al. Cellular immune responses to severe acute respiratory syndrome coronavirus (SARS-CoV) infection in senescent BALB/c mice: CD4+ t cells are important in control of SARS-CoV infection. J Virol 2009;84:1289–301, http://dx.doi.org/10.1128/ IVI.01281-09.
- [106] Sheahan TP, Sims AC, Leist SR, Schafer A, Won J, Brown AJ, et al. Comparative therapeutic efficacy of Remdesivir and combination Lopinavir, Ritonavir, and interferon beta against MERS-CoV. Nat Commun 2020;11:1–14, http://dx.doi. org/10.1038/s41467-019-13940-6.
- [107] Cinatl J, Morgenstern B, Bauer G, Chandra P, Rabenau H, Doerr HW. Treatment of SARS with human interferons. Lancet 2003;362:293–4, http://dx.doi.org/ 10.1016/S0140-6736(03)13973-6.
- [108] Channappanavar R, Perlman S. Pathogenic human coronavirus infections: causes and consequences of cytokine storm and immunopathology. Semin Immunopathol 2017;39:529–39, http://dx.doi.org/10.1007/s00281-017-0639-x
- [109] Huang C, Wang Y, Li X, Ren L, Zhao J, Hu Y, et al. Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. Lancet (London England) 2020;395:497–506, http://dx.doi.org/10.1016/S0140-6736(20)30183-
- [110] Lee J-W, Gupta N, Serikov V, Matthay MA. Potential application of mesenchymal stem cells in acute lung injury. Expert Opin Boil 2009;9:1259-70, http://dx.doi.org/10.1517/14712590903213651.
- [111] Tu YF, Chien CS, Yarmishyn AA, Lin YY, Luo YH, Lin Y-T, et al. A review of SARS-CoV-2 and the ongoing clinical trials. Int J Mol Sci 2020;21:2657, http://dx.doi.org/10.3390/ijms21072657.
- [112] Zhou G, Zhao Q. Perspectives on therapeutic neutralizing antibodies against the Novel Coronavirus SARS-CoV-2. Int J Boil Sci 2020;16:1718–23, http://dx. doi.org/10.7150/iibs.45123.
- [113] Guo R-F, Ward PA. Role of c5a in inflammatory responses. Annu Rev Immunol 2005;23:821–52, http://dx.doi.org/10.1146/annurev.immunol.23. 021704.115835.
- [114] Rose-John S. IL-6 trans-signaling via the soluble IL-6 receptor: importance for the pro-inflammatory activities of IL-6. Int J Boil Sci 2012;8:1237–47, http://dx.doi.org/10.7150/iibs.4989.
- [115] Agostini ML, Andres EL, Sims AC, Graham RL, Sheahan TP, Lu X, et al. Coronavirus Susceptibility to the Antiviral Remdesivir (GS-5734) is mediated by the viral polymerase and the proofreading exoribonuclease. mBio 2018;9, http://dx.doi.org/10.1128/mBio.00221-18.
- [116] Lan L, Xu D, Ye G, Xia C, Wang S, Li Y, et al. Positive RT-PCR test results in patients recovered from COVID-19. JAMA 2020;323:1502-3, http://dx.doi. org/10.1001/jama.2020.2783.
- [117] Agrawal U, Raju R, Udwadia ZF. Favipiravir: a new and emerging antiviral option in COVID-19. Med J Armed Forces India 2020;76:370–6, http://dx.doi.org/10.1016/j.mjafi.2020.08.004.
- [118] Elfiky AA. Ribavirin, Remdesivir, Sofosbuvir, Galidesivir, and Tenofovir against SARS-CoV-2 RNA dependent RNA polymerase (RdRp): a molecular docking study. Life Sci 2020;253:117592, http://dx.doi.org/10.1016/j.lfs.2020.117592.
- [119] Nitulescu GM, Paunescu H, Moschos SA, Petrakis D, Nitulescu GM, Ion GN, et al. Comprehensive analysis of drugs to treat SARS-CoV-2 infection: mechanistic insights into current COVID-19 therapies (Review). Int J Mol Med 2020;46:467–88, http://dx.doi.org/10.3892/ijmm.2020.4608.
- [120] Caly L, Druce JD, Catton MG, Jans DA, Wagstaff KM. The FDA-approved drug ivermectin inhibits the replication of SARS-CoV-2 in vitro. Antiviral Res 2020;178:104787, http://dx.doi.org/10.1016/j.antiviral.2020.104787.
- [121] Cao B, Wang Y, Wen D, Liu W, Wang J, Fan G, et al. A trial of Lopinavir–Ritonavir in adults hospitalized with severe Covid-19. N Engl J Med 2020;382:1787–99, http://dx.doi.org/10.1056/NEJMoa2001282.

- [122] Chen J, Xia L, Liu L, Xu Q, Ling Y, Huang D, et al. Antiviral activity and safety of Darunavir/Cobicistat for the treatment of COVID-19. Open Forum Infect Dis 2020;7:ofaa241, http://dx.doi.org/10.1093/ofid/ofaa241.
- [123] Zoufaly A, Poglitsch M, Aberle JH, Hoepler W, Seitz T, Traugott M, et al. Human recombinant soluble ACE2 in severe COVID-19. Lancet Respir Med 2020;8:1154–8, http://dx.doi.org/10.1016/S2213-2600(20)30418-5.
- [124] Nojomi M, Yassin Z, Keyvani H, Makiani MJ, Roham M, Laali A, et al. Effect of arbidol (Umifenovir) on COVID-19: a randomized controlled trial. BMC Infect Dis 2020;20:954, http://dx.doi.org/10.1186/s12879-020-05698-w.
- [125] Ahmed F, Jo D-H, Lee S-H. Can natural killer cells be a principal player in anti-SARS-CoV-2 immunity? Front Immunol 2020;11:586765, http://dx.doi.org/ 10.3389/fimmu.2020.586765.
- [126] Brzoska J, von Eick H, Hundgen M. Interferons in the therapy of severe coronavirus infections: a critical analysis and recollection of a forgotten therapeutic regimen with interferon beta. Drug Res (Stuttg) 2020;70:291–7, http://dx.doi.org/10.1055/a-1170-4395.
- [127] Sahu KK, Siddiqui AD, Cerny J. Mesenchymal stem cells in COVID-19: a journey from bench to bedside. Lab Med 2021;52:24–35, http://dx.doi.org/10.1093/ labmed/lmaa049.
- [128] Gharebaghi N, Nejadrahim R, Mousavi SJ, Ebrahimi S-RS, Hajizadeh R. The use of intravenous immunoglobulin gamma for the treatment of severe coronavirus disease 2019: a randomized placebo-controlled double-blind clinical trial. BMC Infect Dis 2020;20:786, http://dx.doi.org/10.1186/s12879-020-05507-4.
- [129] Jiang S, Zhang X, Yang Y, Hotez PJ, Du L. Neutralizing antibodies for the treatment of COVID-19. Nat Biomed Eng 2020;4:1134–9, http://dx.doi.org/ 10.1038/s41551-020-00660-2.
- [130] Vlaar APJ, de Bruin S, Busch M, Timmermans SAMEG, van Zeggeren IE, Koning R, et al. Anti-CSa antibody IFX-1 (vilobelimab) treatment versus best supportive care for patients with severe COVID-19 (PANAMO): an exploratory, open-label, phase 2 randomised controlled trial. Lancet Rheumatol 2020;2:e764-73, http://dx.doi.org/10.1016/S2665-9913(20)30341-6.
- [131] Han Q. Guo M, Zheng Y, Zhang Y, De Y, Xu C, et al. Current evidence of Interleukin-6 signaling inhibitors in patients with COVID-19: a systematic review and meta-analysis. Front Pharmacol 2020;11:615972, http://dx.doi.org/10.3389/fphar.2020.615972.
- [132] Khalil A, Kamar A, Nemer G. Thalidomide-revisited: are COVID-19 patients going to be the latest victims of yet another theoretical drug-repurposing? Front Immunol 2020;11:1248, http://dx.doi.org/10.3389/fimmu.2020.01248.
- [133] Edalatifard M, Akhtari M, Salehi M, Naderi Z, Jamshidi A, Mostafaei S, et al. Intravenous methylprednisolone pulse as a treatment for hospitalised severe COVID-19 patients: results from a randomised controlled clinical trial. Euro Resp J 2020;56:2002808, http://dx.doi.org/10.1183/13993003.02808-2020.
- [134] Mallucci G, Zito A, Fabbro BD, Bergamaschi R. Asymptomatic SARS-CoV-2 infection in two patients with multiple sclerosis treated with fingolimod. Mult Scler Relat Disord 2020;45:102414, http://dx.doi.org/10.1016/j.msard.2020. 102414.
- [135] Pang J, Xu F, Aondio G, Li Y, Fumagalli A, Lu M, et al. Efficacy and tolerability of bevacizumab in patients with severe Covid-19. Nat Commun 2021;12:814, http://dx.doi.org/10.1038/s41467-021-21085-8.
- [136] McKay PF, Hu K, Blakney AK, Samnuan K, Brown JC, Penn R, et al. Self-amplifying RNA SARS-CoV-2 lipid nanoparticle vaccine candidate induces high neutralizing antibody titers in mice. Nat Commun 2020;11:3523, http://dx.doi.org/10.1038/s41467-020-17409-9.
- [137] Tebas P, Yang S, Boyer JD, Reuschel EL, Patel A, Christensen-Quick A, et al. Safety and immunogenicity of INO-4800 DNA vaccine against SARS-CoV-2: a preliminary report of an open-label, Phase 1 clinical trial. E Clin Med 2021;31:100689, http://dx.doi.org/10.1016/j.eclinm.2020.100689.
- [138] Folegatti PM, Ewer KJ, Aley PK, Angus B, Becker S, Oxford COVID Vaccine Trial Group, et al. Safety and immunogenicity of the ChAdOx1 nCoV-19 vaccine against SARS-CoV-2: a preliminary report of a phase 1/2, single-blind, randomised controlled trial. Lancet 2020;396:467-78, http://dx.doi.org/10. 1016/S0140-6736(20)31604-4.
- [139] Rashidzadeh H, Danafar H, Rahimi H, Mozafari F, Salehiabar M, Rahmati MA, et al. Nanotechnology against the novel coronavirus (severe acute respiratory syndrome coronavirus 2): diagnosis, treatment, therapy and future perspectives. Nanomedicine 2021;16:497–516, http://dx.doi.org/10.2217/nnm-2020-0441.
- [140] Mahmood Z, Alrefai H, Hetta HF, Kader H, Munawar N, Rahman SA, et al. Investigating virological, immunological, and pathological avenues to identify potential targets for developing COVID-19 treatment and prevention strategies. Vaccines (Basel) 2020;8:443, http://dx.doi.org/10.3390/vaccines8030443
- [141] Padron-Regalado E. Vaccines for SARS-CoV-2: lessons from other coronavirus strains. Infect Dis Ther 2020;9:1–20, http://dx.doi.org/10.1007/s40121-020-00300-x
- [142] Buchholz UJ, Bukreyev A, Yang L, Lamirande EW, Murphy BR, Subbarao K, et al. Contributions of the structural proteins of severe acute respiratory syndrome coronavirus to protective immunity. Proc Natl Acad Sci U S A 2004;101:9804–9, http://dx.doi.org/10.1073/pnas.0403492101.
- [143] Ceraolo C, Giorgi FM. Genomic variance of the 2019-nCoV coronavirus. J Med Virol 2020;92:522–8, http://dx.doi.org/10.1002/jmv.25700.