

P2 receptors in cardiovascular regulation and disease

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Abstract The role of ATP as an extracellular signalling molecule is now well established and evidence is accumulating that ATP and other nucleotides (ADP, UTP and UDP) play important roles in cardiovascular physiology and pathophysiology, acting via P2X (ion channel) and P2Y (G protein-coupled) receptors. In this article we consider the dual role of ATP in regulation of vascular tone, released as a cotransmitter from sympathetic nerves or released in the vascular lumen in response to changes in blood flow and hypoxia. Further, purinergic long-term trophic and inflammatory signalling is described in cell proliferation, differentiation, migration and death in angiogenesis, vascular remodelling, restenosis and atherosclerosis. The effects on haemostasis and cardiac regulation is reviewed. The involvement of ATP in vascular diseases such as thrombosis, hypertension and diabetes will also be discussed, as well as various heart conditions. The purinergic system may be of similar importance as the sympathetic and renin-angiotensin-aldosterone systems in cardiovascular regulation and pathophysiology. The extracellular nucleotides and their cardiovascular P2 receptors are now entering the phase of clinical development.

Keywords ATP · Cardiovascular · Heart · Pathophysiology · Purine · Pyrimidine · P2X · P2Y · Receptor

Introduction

Ever since the first proposition of cell surface receptors for nucleotides [1, 2], it has become increasingly clear that, in addition to functioning as an intracellular energy source, the purines and pyrimidines ATP, adenosine diphosphate (ADP), uridine triphosphate (UTP) and uridine diphosphate (UDP) can serve as important extracellular signalling molecules [3, 4] acting on 13 P2X homo- and heteromultimer ionotropic and 8 P2Y metabotropic receptor subtypes [5, 6] (Table 1). To terminate signalling, ectonucleotidases are present in the circulation and on cell surfaces, rapidly degrading extracellular ATP into ADP, AMP and adenosine [7, 8]. Evidence is accumulating suggesting an important role for the purinergic system in cardiovascular regulation [9–15]. It stimulates vasoconstriction and vasodilatation, growth of vascular smooth muscle cells and endothelial cells, angiogenesis, is involved in vascular remodelling, stimulates platelet aggregation, regulates coagulation, inflammation and several aspects of cardiac function. It is involved in blood pressure regulation, development of myocardial infarction, heart failure and xenograft rejection. The physiological effects of the purinergic signalling system are dependent on the release of extracellular nucleotides, the degradation by ectonucleotidases, the type of P2 receptors expressed on the cells, their desensitisation rates and their second messengers. P2 receptors have highly specific organ distributions and they can be rapidly up- or downregulated. The purinergic system may be of similar importance as the sympathetic and renin-angiotensin-aldosterone systems in cardiovascular regulation and pathophysiology. The involvement of purinergic signalling in a variety of different cardiovascular clinical conditions has been addressed to a limited extent previously [15–19] and the large number of new discoveries calls for a review to summarise the most

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Table 1 Receptor classification, intracellular signalling, ligands and selective agonists and antagonists

P2 subtype	I.c. signalling	Ligand	Selective agonist	Selective antagonist	Non-selective antagonist
P2Y ₁	↑IP3	ADP (ATP)	MRS2365	MRS 2179, MRS2500	
P2Y ₂	↑IP3	UTP = ATP	MRS2498, UTPγS, INS3717		Suramin > RB2
P2Y ₄	↑IP3	UTP (=ATP in rodents)	UTPγS, INS3717	–	RB2 > Suramin
P2Y ₆	↑IP3	UDP	MRS2666, MRS2633, UDPβS	MRS2578	
P2Y ₁₁	↑IP3, ↑cAMP	ATP	AR-C67085MX, NF546	NF157	Suramin > RB2
P2Y ₁₂	↓cAMP	ADP	–	Clopidogrel, prasugrel, AZD6140, INS50589, AR-C9931 (cangrelor)	
P2Y ₁₃	↓cAMP	ADP	–	MRS2211	
P2Y ₁₄	IP3	UDP-glucose, UDP-galactose	UDP-glucose, UDP-galactose	–	
P2X ₁	Positive ion channel	ATP	α,β-mATP	NF023, NF449	TNP-ATP, Ip ₅ I
P2X ₂	Positive ion channel	ATP	–	NF770	Suramin, isoPPADS, RB2
P2X ₃	Positive ion channel	ATP	α,β-mATP	A317491, NF110	Suramin
P2X ₄	Positive ion channel	ATP	Ivermectin potentiates	–	TNP-ATP
P2X ₅	Positive ion channel	ATP	–	–	Suramin, PPADS
P2X ₆	Positive ion channel	ATP	–	–	–
P2X ₇	Positive ion channel	ATP	–	KN62, KN04, MRS2427	Coomassie brilliant blue G
Ectonucleotidase			Apyrase, human SolCD39	ARC67156	

important roles of the purinergic system in cardiovascular regulation and disease.

The research field has grown rapidly since the term P2 receptor was coined [2]. In preparation of this review more than 2,700 references on P2 receptors and cardiovascular regulation were found in PubMed. Among the 15 receptor subtypes, one is the target for one of the most widely used medical drugs—the platelet inhibitor clopidogrel (Plavix). As will be reviewed here, several of the other 15 receptors are promising drug targets to prevent cardiovascular disease.

Regulation of vascular tone

Vasoconstriction produced by ATP released as a cotransmitter with noradrenaline (NA) from perivascular sympathetic nerves was recognised early [20, 21]. However, following the seminal discovery of endothelium-dependent vasodilatation by Furchgott in the early 1980s, it was shown that ATP acted on endothelial cells to release endothelial derived relaxing factor [later shown to be largely nitric oxide (NO)]

resulting in vasodilatation [22] and dual purinergic neural and endothelial control of vascular tone established [23, 24].

Neuronal regulation of vascular tone

ATP is, together with NA and neuropeptide Y (NPY), a cotransmitter in sympathetic neurons [11, 25, 26], and sensory-motor nerves during “axon reflex” activity release ATP to dilate or constrict vessels [27, 28] (Fig. 1). The contribution of ATP to sympathetic contraction varies between vascular beds [25]. For example, it may mediate up to 50% of the neurogenic vasoconstriction via P2X₁ receptors as seen in mesenteric arteries [29], about 70% in rabbit saphenous arteries and 100% in rabbit jejunal artery, while NA acts as a prejunctional modulator [30]. In the renal vasculature both P2X₁ and P2Y₂ receptors are important for neurogenic contraction. P2X₁ receptors mediate contraction in afferent but are absent in the efferent renal arterioles [31, 32]. The result is that extracellular nucleotides selectively influence preglomerular resistance without having an effect on postglomerular tone. ATP is also released locally in the

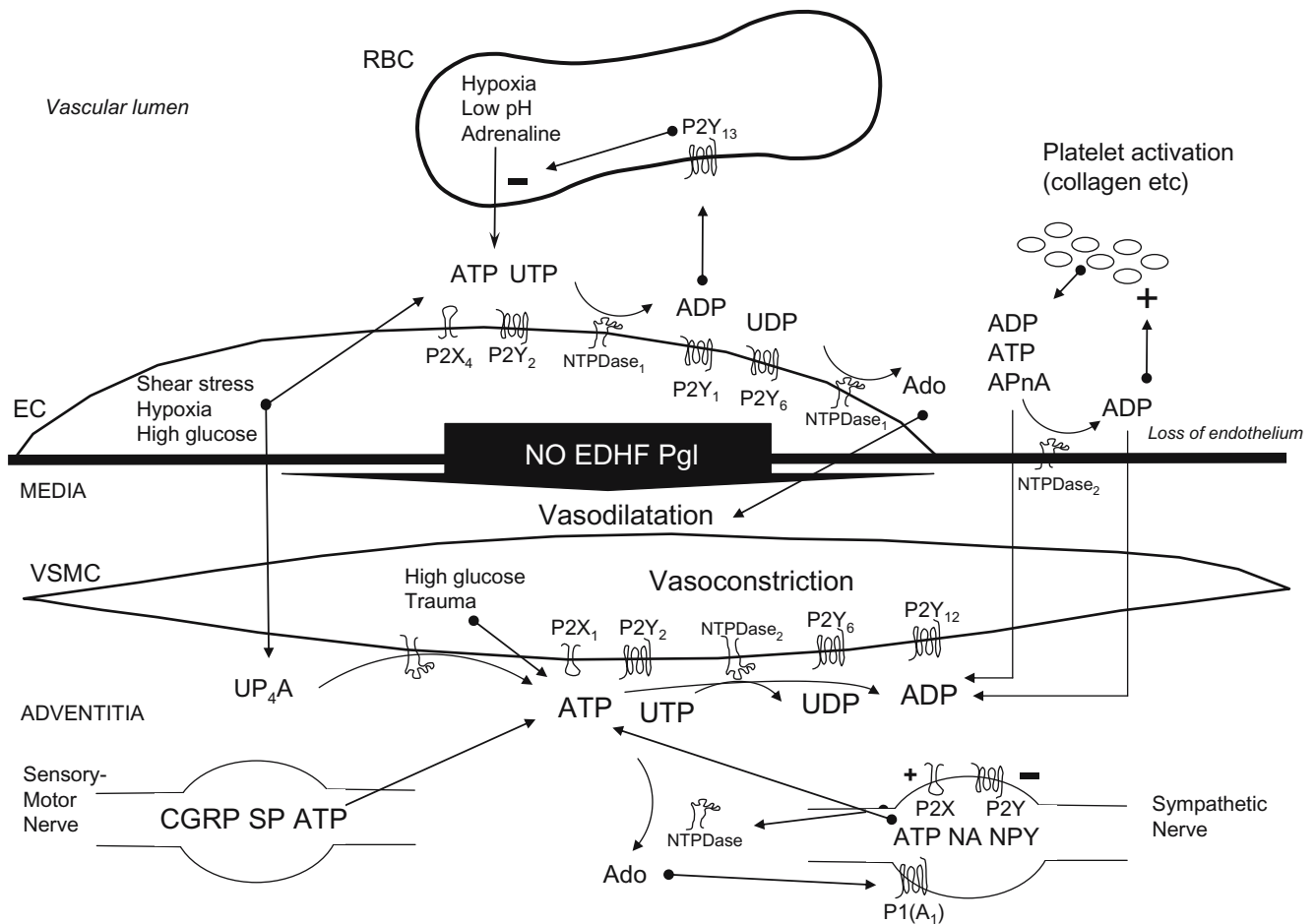


Fig. 1 P2 receptor-mediated regulation of the circulation. See text for details. Purines and pyrimidines are released on the luminal side from endothelial cells, platelets and red blood cells (*RBC*) in response to hypoxia, acidosis, adrenaline, shear stress and other stimuli. When the endothelial cell layer is intact, the response is vasodilatation by the endothelial release of nitric oxide (*NO*), endothelium-derived hyperpolarising factor (*EDHF*) and prostaglandins (*Pgl*). When the endothelium is damaged, platelets accumulate, release ATP and ADP

and mediate vasoconstriction via P2 receptors on the vascular smooth muscle cell (*VSMC*). On the adventitial side sympathetic and sensory nerves mediate vasoconstriction. Extracellular nucleotides are rapidly degraded by NTPDase1 on endothelium reducing ATP to AMP, followed by conversion to adenosine by CD73 (not shown). In the subendothelium NTPDase2 is present, degrading ATP to ADP, maintaining the platelet activating and contractile effects. *Ado* adenosine, *CGRP* calcitonin gene-related peptide, *SP* substance P

kidney in response to increased perfusion pressure, and in P2X₁ receptor knockout mice auto-regulatory responses in kidney afferent arterioles are abolished, indicating an important role in renal glomerular pressure regulation [32, 33].

Sympathetic nerves express inhibitory P2Y and P1 (A₁) receptors indicating a P2 receptor-mediated negative feedback loop both directly by ATP and its degradation product adenosine [34, 35]. Ectonucleotidases are released from sympathetic nerves together with its substrate ATP, as a termination mechanism for the signalling [36]. Another balancing mechanism is the sympathocolytic effect seen in exercising skeletal muscle, which is important for increased blood flow in the working skeletal muscle during exercise, despite sympathetic NA release that normally would reduce blood flow. This sympathocolytic effect is mimicked by injection in the arterial lumen of ATP and UTP, inducing a vasodilatation that overrides sympathetic vasoconstrictor activity in human

skeletal muscle, an effect not obtained by injection of adenosine [37], suggesting that ATP and UTP may mediate this important physiological mechanism.

The first evidence of the presence of P2 receptor subtypes suggested P2X receptors on vascular smooth muscle cells (*VSMC*) and P2Y receptors on endothelial cells [38]. Rat blood vessel immunohistochemistry and human mRNA quantification have shown the P2X₁ receptor to be the highest expressed subtype in smooth muscle cells [39, 40]. It has been difficult to prove the importance of other contractile P2X receptors besides P2X₁ due to the paucity of specific agonists and antagonists. However, contractile effects of UTP suggested that P2Y receptors were present on *VSMCs* and mediated contraction [41]. Using selective pyrimidines resistant to degradation, it has been possible to show that contractile effects are mediated by both UTP-sensitive P2Y₂ and UDP-sensitive P2Y₆ receptors [42]. Ectonucleotidases in

the vessel wall rapidly degrade nucleotides and markedly reduce their contractile effects. Pyrimidine analogues more resistant to ectonucleotidases are powerful vasoconstrictors and up to 1,000-fold more potent than the endogenous ligands [42]. Similarly, a prominent more potent UDP contraction has been seen in NTPDase1 knockout mice, demonstrating the protective effect of ectonucleotidases against nucleotide contraction [43]. In human saphenous vein grafts, used during coronary bypass, a stable UDP analogue stimulates a strong contraction lasting for hours, which is not desensitised [44]. The reason why the P2Y₆ receptor is not desensitised is the lack of serine residues in the C-terminal part of the P2Y₆ receptor [45]. Neither α_1 -adrenoceptors, nor P2Y₆ receptors, are present in human coronary arteries, possibly to avoid deleterious vasospasm during ischaemia [44].

VSMC express P2Y₁₂ receptors that mediate contraction after stimulation with ADP [46]. At the mRNA level, the P2Y₁₂ receptor is the highest expressed ADP receptor and the second highest expressed P2 receptor in human VSMC [46]. The contractions are not inhibited in patients medicated with clopidogrel. However, drugs with antagonistic effects on P2Y₁₂ receptors that reach the peripheral circulation (AZD6140 and INS50589), affecting both platelets and VSMC, could be of double therapeutic benefit in their prevention of both thrombosis and vasospasm [46].

In conclusion, extracellular nucleotides mediate vasoconstriction when released from nerves on the adventitial side, or when released in the lumen when the endothelium is damaged (Fig. 1). The most important contractile receptors on the VSMC are the ATP P2X₁ receptor, the ATP/UTP P2Y₂ receptor, the UDP P2Y₆ receptor and the ADP P2Y₁₂

receptor (Fig. 1 and Table 2). To determine the relative physiological importance of these receptor subtypes in different human vascular beds is an important task for the future.

Endothelial regulation

Shear stress and hypoxia are important stimuli of both ATP and UTP release from endothelial cells [47] (Fig. 1). Extracellular nucleotides have several important effects mediated by activation of endothelial cells. Vasodilatation and decreased blood pressure by release of prostaglandins and NO has been demonstrated in several studies [5], but P2 receptors also mediate release of endothelium-derived hyperpolarising factor (EDHF), which relaxes VSMC by activation of potassium channels, with subsequent hyperpolarisation [48–50]. Both UTP and ATP reduce forearm vascular resistance in a prostaglandin and NO independent way [51], indicating an important role for EDHF in P2 receptor-mediated vasodilatation in man.

The P2Y₁ receptor seems to be of major importance in most vascular beds [11]. However, several other P2 receptors are important for endothelial regulation. Pharmacology of vasodilatation and mRNA quantification in man indicates that P2Y₂ and to a lesser degree P2Y₆ also are important endothelial P2Y receptors [40, 52]. Knockout mice experiments recently confirmed this picture and demonstrated that the P2Y₄ receptor does not mediate dilatation [53]. UTP and ATP are equipotent as vasodilators when infused in the human forearm circulation, indicating a role for both purinergic and pyrimidinergic receptors.

Table 2 P2 receptor expression in the cardiovascular system

P2 subtype	Endothelium	VSMC	Heart	Platelets	Red blood cells	Inflammatory cells	Perivascular sensory nerves	Perivascular sympathetic nerves
P2Y ₁	High	–	–	High	(Turkey)	–	Medium	Medium (neg. feedback)
P2Y ₂	High	High	High	–	–	High	Low	Possible
P2Y ₄	–	Possible	–	–	–	–	–	–
P2Y ₆	Medium	Medium	High	–	–	–	–	–
P2Y ₁₁	Possible	–	High	–	–	High	–	–
P2Y ₁₂	–	Medium	–	High	–	Low	–	–
P2Y ₁₃	–	–	–	–	High	Low	–	–
P2Y ₁₄	–	–	–	–	–	–	–	–
P2X ₁	–	High	Medium	High	–	Medium	–	–
P2X ₂	–	–	–	–	–	–	High	Medium (pos. feedback)
P2X ₃	–	–	–	–	–	–	High	–
P2X ₄	High	–	Low	–	–	High	–	–
P2X ₅	–	–	–	–	–	–	–	–
P2X ₆	–	–	–	–	–	–	–	–
P2X ₇	–	Possible	–	–	Low	High	–	–

Unless specified data refer to human tissue. High, medium or low expression represents an effort to summarise the results of a large number of mRNA, protein and pharmacological studies. Dash (–) denotes lack of convincing evidence for expression. *VSMC* vascular smooth muscle cell

The P2X₁ receptor is not expressed on the vascular endothelium and the evidence for other P2X receptors has been scarce, except for the P2X₄ receptor which is the highest expressed P2 receptor in endothelium [40, 54]. Using antisense oligonucleotides the P2X₄ receptor was shown to be important for shear stress-dependent Ca²⁺ influx via an ATP-dependent mechanism [55]. This indicates that ATP and P2 receptors may be of importance for shear stress-mediated effects, which is in agreement with the well-established release of ATP from endothelial cells during shear stress [56]. Vessel dilation induced by acute increases in blood flow is markedly suppressed in P2X₄ receptor knockout mice [57]. Thus, endothelial P2X₄ channels are crucial to flow-sensitive mechanisms that regulate blood pressure and vascular remodelling [57].

Extracellular ATP in the circulation is rapidly degraded into ADP, AMP and adenosine by ectonucleotidases. Vascular NTPDase1 (CD39) is an endothelial cell membrane protein with both ecto-ATPase and ecto-ADPase activities [8, 58]. Ectonucleotidases are also released by shear stress from endothelial cells [59].

Reactive hyperaemia is the massive increase in blood flow that starts when a blood vessel is opened after a period of ischaemia. It is well known that ATP is released during ischaemia [7, 60]. Adenosine only mediates the late phase of the reactive hyperaemia as has been shown by infusion of adenosine deaminase or a selective A_{2A} antagonist [61, 62]; ADP has been shown to mediate the mid-portion and peak of coronary reactive hyperaemia via endothelial P2Y₁ receptors [63]. It has been proposed that different purines may mediate three phases of the reactive hyperaemia [63]. According to this hypothesis, ATP would mediate the first part of the hyperaemia via endothelial P2Y₂ or P2X₄

receptors. ATP would then be degraded to ADP, which mediates peak hyperaemia via endothelial P2Y₁ receptors, followed by degradation of ADP to adenosine resulting in late phase hyperaemia mediated via A_{2A} receptors on smooth muscle cells.

Red blood cells as regulators of vascular tone

The matching of oxygen supply with demand requires a mechanism that increases blood flow in response to decreased tissue oxygen levels. Several reports suggest that the red blood cell (RBC) acts as a sensor for hypoxia and different mechanisms have been suggested by which the deoxygenated RBC stimulates vasodilatation [37, 64–66]. RBCs contain millimolar amounts of ATP and possess the membrane-bound glycolytic enzymes necessary for its production [67–69]. ATP is released in response to reductions in oxygen tension and pH [37, 64] (see Table 3). It has been shown in vitro that vessels dilate in response to low O₂ levels only when blood vessels are perfused with RBCs [70]. ATP is released in working human skeletal muscle circulation depending on the number of unoccupied haemoglobin O₂ binding sites [64, 71]. Similar results have been shown in the coronary circulation of dogs [72]. The released ATP then binds to P2Y receptors on the endothelium and stimulates vasodilatation. Thus, the RBC functions as an O₂ sensor, contributing to the regulation of blood flow and O₂ delivery, by releasing ATP depending on the oxygenation state of haemoglobin.

ADP activates a negative feedback pathway for ATP release from human RBCs via P2Y₁₃ receptors [73]. Since blood consists of approximately 40% RBCs that contain a 1,000-fold higher ATP concentration than plasma (mmol/

Table 3 Release of purines and pyrimidines in the cardiovascular system

Source	Endothelium	VSMC	Heart	Platelets	Red blood cells	Inflammatory cells	Vascular nerves
Stimuli	Shear stress, hypoxia, high glucose	High glucose, hypoxia	Hypoxia	Collagen, thromboxane, other platelet activators	Hypoxia, adrenaline, deformation	Infl. activators	Sympathetic nerves, sensory nerve collaterals
Mechanism	Vesicular	Unknown	Unknown	Vesicular	Unknown	Unknown or vesicular	Vesicular
Nucleotide	ATP, UTP	ATP, UTP	ATP, UTP	ADP > ATP	ATP	ATP	ATP
Target	Endothelium VSMC	VSMC	Cardiomyocytes, blood vessels, intrinsic cardiac neurons	Platelets; endothelium or VSMC	Endothelium	VSMC, endothelium	VSMC
Degradation	NTPDase1	NTPDase2, NTPDase1	NTPDase1	NTPDase1	NTPDase1	NTPDase1	NTPDase1 (co-released)

Stimuli represents stimuli leading to release. *Mechanism* refers to mechanism of release at the cellular level. *Target* represents the cells that are regulated by the released purine or pyrimidine. *Degradation* refers to the degrading enzyme responsible for removing the purine or pyrimidine after the release. *VSMC* vascular smooth muscle cell

l vs $\mu\text{mol/l}$), even a minor release of ATP from the high intracellular concentrations could have major circulatory effects. A negative feedback system may therefore be of great physiological importance to mitigate ATP release. Another negative feedback system is the mechanism by which NO inhibits ATP release from erythrocytes, illustrating how NO may turn off ATP release [74]. NO is then scavenged by haemoglobin.

Furthermore, the previous view of the RBC as a “passive bag that transports oxygen” is challenged. It now turns out that it releases ATP in response to stimuli and, as with most important signalling systems, it has a negative feedback system to terminate its release. The described negative feedback pathway may be important to avoid high extracellular concentrations of ATP. At levels above 100 $\mu\text{mol/l}$, ATP concentrations may exceed the catalytic capacity of ectonucleotidases and could, in fact, stimulate ATP release by increasing permeability of the RBC [75], probably via P2X₇ receptors [76]. ATP may even release ATP from endothelial cells [77]. At high concentrations of ATP, a self-sustaining process may thus be instigated which may contribute to the irreversible stage of circulatory shock that can develop rapidly in severely ill patients. Similar mechanisms may be of importance in malaria because induction of the osmolyte permeability in *Plasmodium*-infected erythrocytes involves purinoceptor signalling [78].

A mechanism of ATP release from RBCs, which has eluded researchers for many years, has recently been suggested by Locovei and co-workers as being mediated via the gap junction protein pannexin-1 [79]. Erythrocytes do not form gap junctions, instead pannexin-1 forms a mechanosensitive ATP-permeable channel that mediates osmotically induced ATP release.

Hypertension

Blood pressure regulation by purinergic signalling is the net result of balancing contractile and dilatory effects as described above. ATP and UTP released on the luminal side, from endothelial cells and erythrocytes, stimulate vasodilatation, in contrast to release from nerves on the adventitial side, which results in vasoconstriction. ATP may also regulate blood pressure via renal mechanisms or brain stem regulation [31, 32].

ATP plays a significantly greater role as a sympathetic cotransmitter in spontaneously hypertensive rats (SHR) [80, 81], and there is increased responsiveness of the renal vasculature of isolated perfused rat kidneys to α,β -methylene ATP in SHR [82], while mesenteric vascular contractile reactivity to ATP via P2X₁ and P2Y₂ receptors is not altered in deoxycorticosterone acetate (DOCA)-salt hypertension [83]. In the aorta of SHR, endothelium-dependent relaxation to ATP is impaired because of the concomitant generation of

an endothelium-derived contracting factor. Contractions to ATP were significantly potentiated in SHR aorta [84]. In hypertensive mature stroke-prone rats, NPY modulation of release of NA and ATP from sympathetic perivascular nerves in kidney vessels is impaired, which may account for the increased nerve-mediated responses [85]. Thus, animal models of hypertension reveal several alterations in purinergic signalling that may contribute to the development of hypertension.

Diadenosine polyphosphates such as Ap₄A, Ap₅A and Ap₆A are combinations of two adenosine molecules connected with four to six phosphate groups. They have been identified as vasocontractile agents [86], probably via actions on P2X₁ and P2Y₂ receptors. Ap₅A and Ap₆A are stored at higher levels in platelets from patients with hypertension and may contribute to their increased peripheral vascular resistance [87]. Up₄A is a novel endothelium-derived vasoconstrictive factor more potent than endothelin in renal vasoconstriction [88]. It is released upon stimulation of the endothelium by acetylcholine, thrombin and mechanical stress and can be cleaved into either ATP or UTP to stimulate both P2X₁ and P2Y₂ receptors on VSMC resulting in increased blood pressure [88].

As mentioned above, the P2X₄ receptor is the most abundantly expressed P2 receptor in the endothelium and mediates shear stress-stimulated vasodilatation. P2X₄ receptor knockout mice have higher blood pressures and excrete smaller amounts of NO products in their urine than do wild-type mice [57]. The P2X₄ receptor protein is upregulated in the placenta in preeclampsia [89]. Thus, shear stress-stimulated release of ATP acting on P2X₄ receptors may be important for blood pressure regulation.

The importance of P2 receptors for hypertension will not be proven until tested in man. It would be of great interest to perform a clinical study on the effects of P2X₁, P2Y₂ or P2Y₆ receptor antagonists in the treatment of hypertension.

Pulmonary hypertension

Erythrocyte release of ATP appears to regulate pulmonary resistance under some conditions [90], and patients with pulmonary hypertension have been found with impaired release of ATP from RBC [69]. Endothelium-dependent relaxation to ATP has been demonstrated in human pulmonary arteries [91]. On the other hand, ATP is a mitogen for pulmonary artery smooth muscle cells, which may be relevant for the pathophysiological basis of pulmonary hypertension [92]. Again, the balance between endothelial versus smooth muscle stimulation by ATP may regulate blood pressure in different directions and we are far from a full understanding of the role of the purinergic system in pulmonary hypertension.

Migraine and vascular pain

Classic migraine is associated with two distinct vascular changes: an initial vasoconstriction, followed by vasodilatation associated with pain. It has been proposed that purinergic signalling is involved in these changes. ATP released from perivascular sympathetic nerves may participate in producing the initial vasospasm mediated by P2 receptors in vascular smooth muscle. Cerebral arteries are strongly contracted by UTP and especially UDP via P2Y₆ receptors. A UDP agonist might therefore have similar effects as the 5-HT_{1D} agonists frequently used to treat migraine [93]. ATP released by endothelial cells acting on P2 receptors on endothelial cells mediating release of NO and producing vasodilatation may contribute to the phase of reactive hyperaemia following vasospasm [94]. It has been suggested that antagonists to P2X₃ receptors, which are located on nociceptive sensory nerve endings in cerebral vessels, may be promising candidates for anti-migraine drug development [95]. A recent report demonstrates selective upregulation of nociceptive P2X₃ receptors on trigeminal neurons by calcitonin gene-related peptide and suggests that this mechanism might contribute to pain sensitisation in migraine [96]. P2Y receptors on trigeminal sensory nerves have also been implicated in pain in migraine. It has been claimed that migraine attacks are characterised by a relative depletion of sympathetic NA stores in conjunction with an increase in release of ATP [97]. In a review in the *Lancet* in 1996, Burnstock proposed that vascular pain, as in angina, ischaemic muscle and pelvic pain in women, may be initiated by ATP released from microvascular endothelial cells during reactive hyperaemia, diffusing a short distance to activate P2X₃ nociceptive receptors on sensory nerves in the adventitia [26].

Atherosclerosis

Atherosclerosis is the main cause of ischaemic stroke and cardiovascular disease and is now considered to be an inflammatory disease [98]. The formation of a plaque starts with the accumulation of cholesterol followed by invasion of macrophages taking up cholesterol and becoming foam cells. The plaque can be stabilised by smooth muscle cell formation of a fibrous cap that covers the lipid-rich region. However, stimulation of inflammation by oxidised low-density lipoprotein activates macrophages and dendritic cells into antigen-presenting cells, activating T lymphocytes resulting in release of cytokines and metalloproteinases degrading the fibrous cap. The end result is a vulnerable plaque and, when it ruptures, its highly thrombogenic contents activates platelets and causes the formation of a local thrombus occluding the artery or embolising, resulting

in ischaemic stroke or myocardial infarction. Evidence both from basic research and from clinical studies indicates important involvement on several levels for purinergic signalling in the atherosclerotic process (Fig. 2). Interestingly, fish oil components increase the release of ATP [99] and a high cholesterol diet decreases ATP release from arteries [100, 101].

Atherosclerosis—P2 receptor-mediated effects on inflammatory cells

Inflammatory cells express a large number of P2 receptors with multiple effects [102]. The final result is difficult to predict depending on the subtype of receptor expressed by a particular cell type and on the differentiation stage of the cell. The most important inflammatory cells for atherosclerosis are monocytes that differentiate into macrophages or dendritic cells in the plaque, and the T-helper and suppressor lymphocytes that coordinate the inflammatory reaction in the plaque [98].

A large number of P2 receptors are expressed on T lymphocytes and macrophages [14, 103] and have been suggested to be important players in atherosclerosis [14]. Strong evidence for a functional role exists for P2X₇, P2Y₂ and P2Y₁₁ receptors. The P2X₇ receptor is mitogenic and anti-apoptotic for T lymphocytes [104, 105]. P2X₇ is important for release of interleukin (IL)-1 [106], tumour necrosis factor [107] and L-selectin, an adhesion molecule important for lymphocyte binding to endothelium [108]. All of these effects are known to be important for atherosclerosis development. The severity of arthritis is reduced in P2X₇ receptor knockout mice [109] and rheumatoid arthritis is coupled to increased incidence of myocardial infarction. It would be of great interest to examine the P2X₇ receptor knockout mouse in an atherosclerosis model. ATP and UTP are chemotactic for dendritic cells probably via the P2Y₂ receptor and may attract inflammatory cells to the vascular lesion [110]. The P2Y₂ receptor enhances the oxidative burst in human macrophages [111]. P2Y₁ receptor deletion in knockout mice reduces the atherosclerotic lesions and the plaque area occupied by macrophages in ApoE knockout mice [112]. Whether this is due to platelet inhibition, endothelial or an inflammatory mechanism requires further studies.

ATP inhibits CD4⁺ T cell activation via an increase in cyclic AMP (cAMP) probably via a P2Y₁₁ receptor [113]. ATP acting on P2Y₁₁ receptors regulates the maturation of human monocyte-derived dendritic cells and induces immunosuppression by inhibiting T-helper 1 cytokines and promoting T-helper 2 cytokines [114, 115].

A polymorphism in the P2Y₁₁ receptor has been shown to have clinical importance by increasing the risk of myocardial infarction [116]. The G-459-A polymorphism, carried by one fifth of the population, causes an Ala-87-Thr

Atherosclerotic plaque

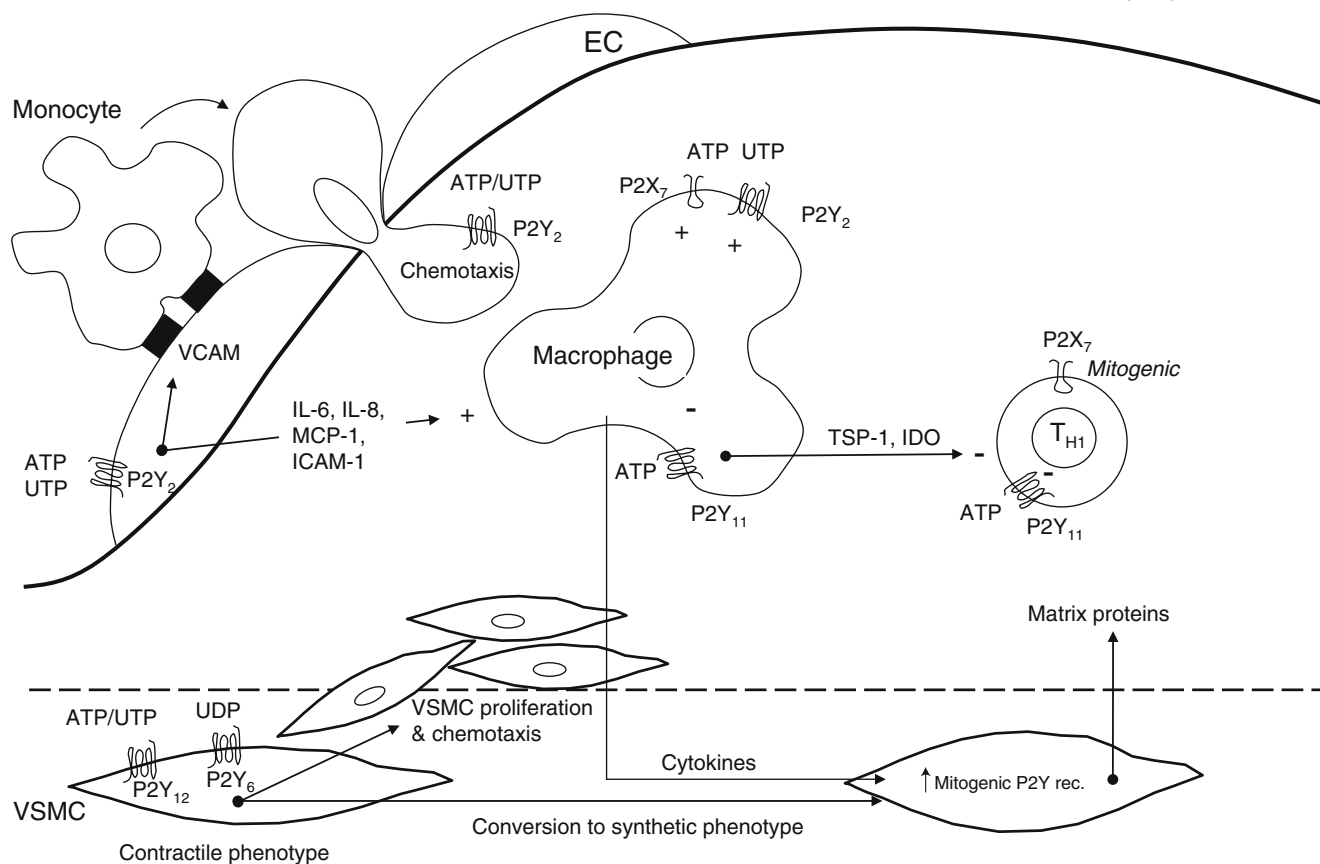


Fig. 2 Functional roles of P2 receptors in the atherosclerotic inflammatory plaque and during restenosis. See text for details. Purines and pyrimidines acting on P2 receptors stimulate vascular inflammation both by actions on the endothelial cell (EC) and by effects on inflammatory cells. Furthermore, they stimulate vascular

smooth muscle cell (VSMC) proliferation, the conversion to synthetic phenotype and production of matrix proteins. Mitogenic P2 receptors are upregulated by growth factors and cytokines. *IL* interleukin, *MCP-1* monocyte chemoattractant protein-1, *ICAM-1* intercellular adhesion molecule-1, *TSP* thrombospondin, *IDO* indoleamine 2,3-dioxygenase

substitution in the P2Y₁₁ ATP receptor and increases the risk of myocardial infarction by 21%. The odds ratio increased stepwise depending on the number of Thr-87 alleles, and in subgroups in which the genetic influence is known to be of increased importance, family history of acute myocardial infarction (AMI), early onset AMI or the combined group of early onset AMI with family history. The mechanism by which the polymorphism causes AMI seems to be coupled to increased inflammation because the Thr-87 variant of the P2Y₁₁ receptor was coupled to elevated C-reactive protein (CRP) levels. CRP is a marker of inflammation and an independent prognostic risk factor for the development of AMI [98]. Thus, the P2Y₁₁ receptor is important in the development of atherosclerosis via modulation of inflammation either via effects on T lymphocytes or macrophage cells.

Proinflammatory effects on the endothelium

Even though P2 receptor-mediated activation of the endothelium stimulates release of NO, which inhibits inflammatory cells, it is now well established that important

proinflammatory endothelial effects may also be triggered. A proinflammatory dysfunctional endothelium is crucial in the recruitment of monocytes to the atherosclerotic plaque and in the general extravasation and loss of peripheral resistance seen in sepsis. ATP stimulates neutrophil adherence to cultured endothelial cells [117]. ATP stimulates release of IL-6, IL-8, monocyte chemoattractant protein-1, growth-regulated oncogene α and increased expression of intercellular adhesion molecule-1 in human microvascular endothelial cells [118]. UTP and ATP stimulate expression of proinflammatory vascular cell adhesion molecule-1 (VCAM-1) in endothelial cells through activation of the P2Y₂ receptor and increases the adherence of monocytic cells to human coronary endothelial cells, an effect that was inhibited by anti-VCAM-1 antibodies [119]. VCAM is important for the recruitment of monocytes and lymphocytes. The VCAM stimulation is caused by P2Y₂ receptor-induced transactivation of the vascular endothelial growth factor receptor-2 [120]. These effects have been confirmed in an in vivo neointima model, in which perivascular infusion of UTP enhanced infiltration by macrophages [121].

In conclusion, ATP and UTP stimulate several inflammatory responses known to be important for atherosclerosis development (Fig. 2).

Restenosis and vascular smooth muscle cell proliferative disease

VSMC proliferation contributes to plaque development, but it is probably beneficial by stabilising the plaque and forming a stronger fibrinous cap. However, there are several situations in which VSMC proliferation is important in vascular disease development: restenosis after balloon angioplasty, diabetic microvascular disease, chronic allograft rejection, pulmonary hypertension and possibly systemic hypertension. The latter is suggested since sympathetic nerves exert a trophic effect on vascular smooth muscle [122]. ATP is a cotransmitter from sympathetic nerves [25] and has been shown to be a more potent mitogen than the other cotransmitters NA and NPY [123].

Extracellular ATP is a potent growth factor for VSMC by activation of G protein-coupled P2Y receptors [12, 123–125]. UTP is also mitogenic, acting on P2Y₂ receptors [123], as is UDP acting on P2Y₆ receptors [124, 126, 127]. ATP is synergistic with polypeptide growth factors (e.g. platelet-derived growth factor, basic fibroblast growth factor) and insulin [123, 124]. The signal transduction is mediated via Gq proteins, phospholipase C and D, diacylglycerol, protein kinase C, extracellular signal-regulated kinase, phosphatidylinositol-3 kinase, MAPK/ERK activity kinase, mitogen-activated protein kinases and Rho [12, 128, 129]. Several immediate early genes are activated and the cell is taken through different phases of the cell cycle [125, 130]. Sometimes, as with UDP acting on P2Y₆ receptors, progression is stimulated to both the S and G₂ phases, that is through the whole cell cycle [127].

Mitogenic effects have been demonstrated in rat, porcine, and bovine VSMC and cells from human coronary arteries, aorta, and subcutaneous arteries and veins [12, 13, 124, 131]. The trophic effects on VSMC and the abundant sources for extracellular ATP in the vessel wall make a pathophysiological role probable in the development of atherosclerosis, neointima formation after angioplasty, chronic allograft rejection, pulmonary hypertension, diabetic microvascular disease and possibly hypertension. In these processes interaction between the VSMC and the matrix is important. It is therefore interesting that the human P2Y₂ receptor contains an integrin-binding domain (RGD) in its first extracellular loop and that interaction with integrins influences P2Y₂ receptor-mediated activation of G proteins [132]. ATP stimulates release of matrix metalloproteinase-2 (MMP-2) from human aortic smooth muscle cells and MMP-2 has been implicated in aortic aneurysm pathogenesis [133].

MMPs open up the matrix for migrating VSMC. ATP and UTP are potent chemotactic agents stimulating VSMC migration via P2Y₂ and P2Y₆ receptors [134]. These mechanisms, together with growth-stimulating effects, explain why perivascular infusion of UTP in a neointima model enhanced neointimal development [121]. Overexpression of the ATP- and UTP-degrading enzyme NTPDase1 reduces neointima development in rat aorta [135]. Augmentation of NTPDase1 activity is an important adaptive response for cardiac allograft survival, with increased inflammation, platelet deposition and infarction rates in NTPDase-deficient mice [136]. The mechanism is dependent on increased expression of the atherosclerotic matrix protein osteopontin [134, 137].

Diabetic microvascular disease

Sympathetic vascular dysfunction in early experimental juvenile diabetes was recognised many years ago. Two weeks after induction of streptozotocin diabetes in rats, there was prejunctional impairment of sympathetic neurotransmission and impaired ATP-mediated endothelial function in mesenteric arteries [138]. Altered relaxant responses to ATP in the corpus cavernosum of men and rats with diabetes have also been reported [139].

As mentioned above, red blood cells may stimulate vasodilatation via release of ATP, which may be important in maintaining perfusion in the microcirculation. Interestingly, in erythrocytes from humans with type 2 diabetes ATP release is impaired, which is consistent with the hypothesis that the defect in erythrocyte physiology could contribute to the vascular disease associated with this clinical condition [140].

High extracellular glucose releases ATP and/or UTP in endothelial cells and pancreatic β cells [141, 142]. An increase in glucose from 5 to 15 mmol/l results in a marked increase in the proatherogenic nuclear factor of activated T cells signalling pathway in VSMC [143]. The effect is mediated via glucose-induced release of ATP and UTP, which subsequently activate P2Y₂ but also P2Y₆ receptors (after degradation to UDP). Thus, nucleotide release is a potential metabolic sensor for the arterial smooth muscle response to high glucose. Diabetic patients experience microvascular disease characterised by increased wall-lumen ratio, mainly because of an increase in VSMC and have higher rates of restenosis after coronary angioplasty. High glucose-induced release of extracellular nucleotides, acting on P2Y receptors to stimulate VSMC growth via NFAT (nuclear factor of activated T cells) activation, may provide a link between diabetes and diabetic vascular disease [143].

In conclusion, ATP/UTP/UDP stimulate VSMC growth, migration, release of MMPs and osteopontin that may contribute to the development of restenosis, diabetic microvascular disease, chronic allograft rejection, pulmo-

nary hypertension and possibly systemic hypertension (Fig. 2).

Vascular remodelling and angiogenesis

Vascular remodelling

P2 receptors may regulate VSMC phenotype and vice versa, and P2 receptor expression is markedly altered during phenotype changes [13]. The shift from a specialised contractile VSMC phenotype into a proliferating, matrix-producing, synthetic phenotype is a prerequisite for VSMC pathogenesis in vascular disease. Pacaud and co-workers found that the Ca^{2+} -mobilising effects of 2-methylthioATP increased in VSMC during culture in serum, indicating upregulation of P2Y₁ receptors in the transition from contractile to synthetic phenotype [144]. This was confirmed at the mRNA level where P2Y₂ receptors were found to be upregulated, while P2X₁ receptors were downregulated [145]. Mitogenic P2Y receptors are upregulated while ion channel receptors, with only contractile effects, are downregulated in the synthetic phenotype [145]. Growth factors, cytokines and interestingly also ATP are potent stimulators of P2Y₂ receptor expression [146, 147]. Thus, factors of importance in the development of vascular disease increase mitogenic P2Y₂ receptors; this is further supported by their upregulation in neointima after balloon angioplasty [148, 149]. ATP has a dual concentration-dependent effect on VSMC phenotype. Low ATP concentrations stimulate expression of genes specific for the contractile phenotype, while high ATP concentrations cause a phenotypic shift from the contractile to the synthetic phenotype, and this shift is dependent on a transient activation of protein kinase A, which inhibits activation of a serum response factor [150]. This previously unrecognised mechanism also appears to be important for the profound mitogenic effect of ATP. In intact human blood vessels examined *in vitro*, shear stress decreased contractile P2X₁ receptor expression, but increased the expression of mitogenic P2Y₂ and P2Y₆ receptors in VSMC [151]. This mechanism could promote vascular growth and remodelling induced by shear stress as an adaptive response to increased flow. P2X₄ receptor knockout mice are incapable of adaptive vascular remodelling, that is, a decrease in vessel size in response to a chronic decrease in blood flow [57].

Angiogenesis

Very little is known about the role of P2 receptors in angiogenesis, but there are some studies suggesting their involvement. Ischaemia stimulates release of ATP [7, 60] and it has been shown to be a growth factor for endothelial

cells [13, 152]. ATP and UTP stimulate cytoskeletal rearrangements with consequent cell migration of human endothelial cells [153] and ATP stimulates vasa vasorum neovascularisation in pulmonary arteries [154]. Newly developed vascular endothelia express very high levels of NTPDase1, also seen under hypoxic conditions [155]. NTPDase1 knockout mice exhibit disordered cellular migration and angiogenesis [156].

Varicose veins

It has been suggested that cell lysis, consequent to P2X₇ receptor-induced pore formation, contributes to the disorganisation and decrease in contractile myocytes in the media of varicose veins [157]. Upregulation of P2Y₁ and P2Y₂ receptors and downregulation of P2X₁ receptors on smooth muscle of varicose veins is associated with a shift from contractile to synthetic and/or proliferative roles; this phenotypic change in smooth muscle leads to weakening of vein walls and may be a causal factor in the development of varicose veins [158], an interesting parallel to the receptor changes seen in response to increased shear stress (see above) [151].

Heart

ATP is released in the heart as a cotransmitter together with catecholamines from sympathetic nerves, but it may also be released from other sources in the heart such as endothelium, platelets, RBC and ischaemic myocardium (Table 3) [7, 60, 159]. P2 receptors are abundantly expressed in the foetal human heart [160] as well as in the adult human heart [60, 161].

Inotropy

In cardiomyocytes, ATP stimulates a pronounced positive inotropic effect and may also act in synergy with β -adrenergic agonists to augment myocyte contractility [60, 162–165]. ATP stimulates an increase in cytosolic calcium and evidence for the involvement of inositol 1,4,5-trisphosphate (IP₃)-coupled P2Y₂ receptors and ion channel P2X receptors has been presented [162, 164–167]. Ap₄A, probably after degradation to ATP, increases force of contraction in human ventricular trabeculae [168].

The inotropic effects of ATP are dependent on both IP₃ and cAMP [169]. The inotropic effects of catecholamines acting on β -receptors are mediated by an increase of cAMP and antagonists of these receptors are important drugs for the treatment of hypertension and reduce mortality in congestive heart failure. Similarly, ATP stimulates an increase in cAMP in cardiac myocytes and may act in

synergy with the β_1 -adrenergic agonist, isoproterenol, by differential activation of adenylyl cyclase isoforms [60, 170, 171]. However, the ATP receptor mediating this increase in cAMP has not been related to any particular P2 receptor subtype in cardiac cells [60]. It could be the P2Y₁₁ receptor that is additionally coupled to G_s and activates adenylyl cyclase [60, 172–174]. mRNA for the P2Y₁₁ receptor has been detected in the human heart at high levels [161, 175].

The unstable agonist, UTP, has been shown to induce a positive inotropic effect in rat atria and in rat and guinea pig ventricular cardiomyocytes [164, 166, 176, 177]. Stable UTP and UDP analogues induce a pronounced inotropic effect on mouse cardiomyocytes [161]. The P2Y₂ receptor is the most abundantly expressed receptor with very low levels of the P2Y₄ receptor in the human heart [161], suggesting that the inotropic effects of UTP are mediated

via P2Y₂ receptors, while the UDP effects are mediated via the P2Y₆ receptor [161]. These mechanisms are mediated via IP₃-mediated signalling.

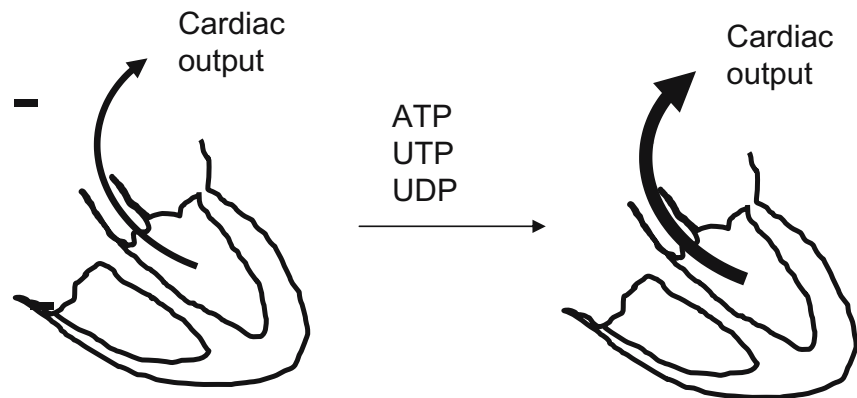
In conclusion, the inotropic effects in man are mediated via P2Y₂ and P2Y₆ receptors and a P2Y₁₁-like receptor (Fig. 3). P2X receptors are probably also involved, since several subtypes are expressed [60, 160], but it has not been possible to perform a clear receptor characterisation due to lack of selective antagonists.

Myocardial infarction

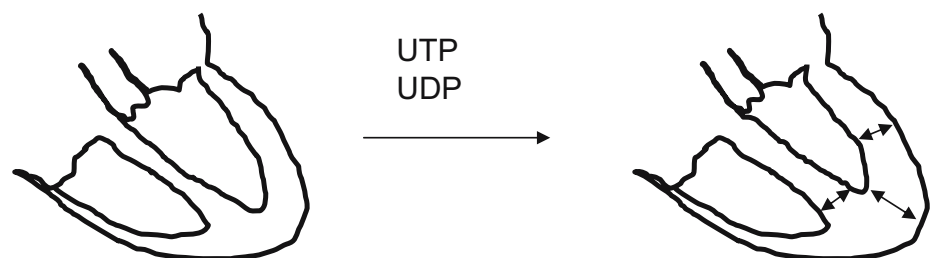
Using microdialysis, ATP in the interstitial space has been estimated to be 40 nmol/l, but the levels may increase markedly during electrical stimulation, ischaemia, challenge with cardiotoxic agents, increase in blood flow, mechanical stretch and increased work load [60]. ATP is released from

Fig. 3 Functional roles of purines and pyrimidines acting on P2 receptors in the regulation of the heart. See text for details. **a** ATP, UTP and UDP exert inotropic effects on cardiomyocytes leading to increased cardiac output. **b** UTP and UDP stimulate hypertrophy of cardiomyocytes, while ATP can have apoptotic effects. **c** UTP protects against ischaemic injury and cardiomyocyte cell death. ATP-degrading enzymes preserve endothelial integrity and protect against allograft rejection. *NTPDase* nucleoside triphosphate diphosphohydrolase

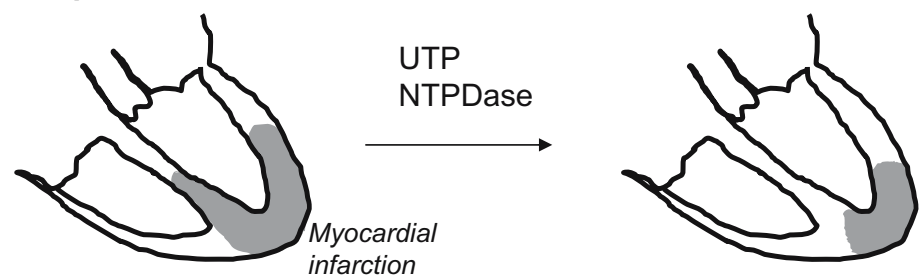
a Inotropic effects



b Hypertrophic effects



c Cardioprotective effects



cardiomyocytes during reduced oxygen tension [60]. UTP and ATP are released from the heart during cardiac ischaemia [178] and patients with myocardial infarction have higher plasma levels of both ATP and UTP [161, 179]. There is a significant increase in ectonucleotidase activity (NTPDase1) in the hearts of patients with ischaemic heart disease [180] that could represent a compensatory mechanism against increased nucleotide levels during chronic ischaemia.

Ischaemia-reperfusion provokes barrier failure of the coronary microvasculature, leading to myocardial oedema, ATP may protect against reperfusion-induced coronary endothelial barrier damage and inhibition of ATP degradation enhances the stabilising effect of ATP on barrier function [181].

The common P2Y₁₁ ATP receptor polymorphism, Ala-87-Thr, is associated with both increased CRP and increased risk of developing myocardial infarction [116]. Based on this association, we hypothesise that the P2Y₁₁ receptor plays an important role in cardiovascular biology and in inflammatory disease (see above), causing myocardial infarction via a proinflammatory effect. However, increased inotropic effects may also contribute.

An interesting yin and yang situation for ATP and UTP has been revealed regarding hypertrophic effects. UTP but not ATP causes hypertrophic growth in neonatal cardiomyocytes [182] (Fig. 3). In contrast, ATP inhibits hypertrophy and may even induce apoptosis and necrosis [163, 183]. The reason for the difference in effects could be due to activation of P2X receptors or cAMP stimulation by ATP [60, 161]. Both UTP and ATP transactivate epidermal growth factor receptors, but only ATP stimulates the hypertrophic marker genes atrial natriuretic peptide and myosin light chain 2 [184]. Similarly, UTP but not ATP protects cultured cardiomyocytes against hypoxic stress [185]. Since UTP is released during preconditioning [178], a role for UTP in the protective effects of preconditioning is plausible. Recently, Yitzhaki and co-workers were able to demonstrate prominent reductions in myocardial infarction size and improved rat heart function *in vivo*, by a single intravenous bolus dose of UTP before ischaemia [186] (Fig. 3).

Congestive heart failure

There are several reports of alterations or adaptations of purinergic signalling during congestive heart failure. The positive inotropic effect of ATP is impaired in heart failure, but reversed by the angiotensin-converting enzyme (ACE) inhibitor imidapril [187]. The contractile responses for the P2Y₁₁ receptor agonist AR-C67085 are decreased in heart failure, suggesting a downregulation of this receptor function in cardiomyocytes in a similar manner as seen for β_1 -receptors in congestive heart failure [169]. In human

hearts, only the P2X₆ receptor was altered in congestive heart failure [188]. In congestive heart failure P2X₁ receptors are downregulated in VSMC in resistance arteries, which could represent a protective response against the increased sympathetic nerve activity and peripheral resistance seen in congestive heart failure [189].

Several P2 receptors have been suggested as targets for pharmacological treatment of congestive heart failure. Overexpression of P2X₄ receptors has a beneficial, life-prolonging effect in a heart failure model [190]. Both ATP and catecholamines are released from sympathetic nerves, acting through cAMP-stimulating receptors to mediate positive inotropic effects, stimulating the same intracellular mechanisms as adrenergic β -receptors. It is possible that agonists for the P2Y₁₁ receptor could be used to improve cardiac output in patients with circulatory shock. However, an even more important drug candidate would be a P2Y₁₁ receptor antagonist that may be beneficial in patients with congestive heart failure. The extracellular pyrimidines UTP and UDP may be inotropic factors in man, acting on P2Y₂ and P2Y₆ receptors stimulating the same intracellular pathways as angiotensin II. Synthetic agonists could thus be used as inotropic agents during circulatory shock and antagonists may have effects similar to angiotensin II receptor blockers, being beneficial in the treatment of hypertension and congestive heart failure.

Chronotropy and arrhythmia

It has been difficult to determine the chronotropic effects of ATP due to the dominating inhibitory effects of adenosine on the AV node. We know that ATP increases the contractile rate in neuron-myocyte co-cultures but the effect is markedly reduced in non-innervated myocyte cultures [191]. ATP could induce arrhythmia based on its increased automaticity and early after-depolarisations [60]. The P2X₁ receptor is colocalised with connexin 43 in gap junctions that transmit the contractile stimulus between cardiomyocytes [192]. However, so far no firm evidence exists of an important role for P2 receptors for chronotropy or arrhythmia.

Platelets and coagulation

Platelets

ADP, released from erythrocytes or produced after ectonucleotidase breakdown of ATP released from erythrocytes and endothelial cells, was found to cause platelet aggregation as early as 1961, before the concept of P2 receptors was conceived [193]. Later, the effects were attributed to a single receptor, designated P2T, but it was not until 1998 that the three receptors involved were characterised [194–196].

Two ADP receptors (P2Y₁₂ and P2Y₁) and one ion channel ATP receptor (P2X₁) are expressed on platelets [197, 198] (Fig. 4). The P2Y₁₂ receptor is coupled to inhibition of cAMP. The molecular identifications of the P2Y₁₂ receptor and generation of knockout mice revealed highly prolonged bleeding times, and their platelets aggregate poorly in response to ADP and display a reduced sensitivity to thrombin and collagen [199]. The P2Y₁ receptor is responsible for ADP-induced shape change and weak, transient aggregation [200–202], while the P2Y₁₂ receptor is responsible for the completion and amplification of the response to ADP and to all platelet agonists, including thromboxane A₂, thrombin and collagen. The platelets also express P2Y₁ receptors that have been shown in knockout mice to be of similar importance as P2Y₁₂ receptors [200, 203].

The physiological importance of the P2X₁ receptor is not fully elucidated. However, recent studies have demonstrated that P2X₁ receptors can generate significant functional platelet responses, independently and in synergy with other receptor pathways. In addition, studies in transgenic animals indicate an important role for P2X₁ receptors in platelet activation, particularly under conditions of shear stress and thus during arterial thrombosis [204]. Synergistic interaction between ATP and NA in stimulating platelet aggregation may have significant clinical implications and suggests a prothrombotic role for ATP in stress [205].

NTPDase1 is an important inhibitor of platelet activation. Unexpectedly, NTPDase1-deficient mice had prolonged bleeding times with minimally perturbed coagulation parameters. Platelet interactions with injured mesenteric vasculature were considerably reduced in vivo and purified mutant platelets failed to aggregate to standard agonists in vitro [206]. This platelet hypofunction was reversible and associated with P2Y₁ receptor desensitisation. In keeping with deficient vascular protective mechanisms, fibrin deposition was found at multiple organ sites in NTPDase1-deficient mice and in transplanted cardiac grafts. Endothelial ADPase activity is lost following ischaemia-reperfusion injury, xenograft rejection and inflammation resulting in increased levels of ATP and ADP [58, 207]. This may cause platelet aggregation but also other effects, such as mitogenic effects on VSMC. Infusion of systemic apyrase inhibits platelet aggregation and prolongs xenograft survival [208]. This has also been shown with adenoviral transfer of NTPDase1 [209]. NTPDase1 is also lost in vascular cardiac grafts subjected to oxidant stress. NTPDase2 only degrades ATP to ADP and has a prothrombotic effect. It is present in the subendothelium and can shift the balance in a prothrombotic direction after endothelial injury [210] (Fig. 1).

ADP acting on P2Y₁₂ receptors is not only important for platelet activation, it also stimulates vasoconstriction. Stable drugs with antagonistic effects on P2Y₁₂ receptors, affecting

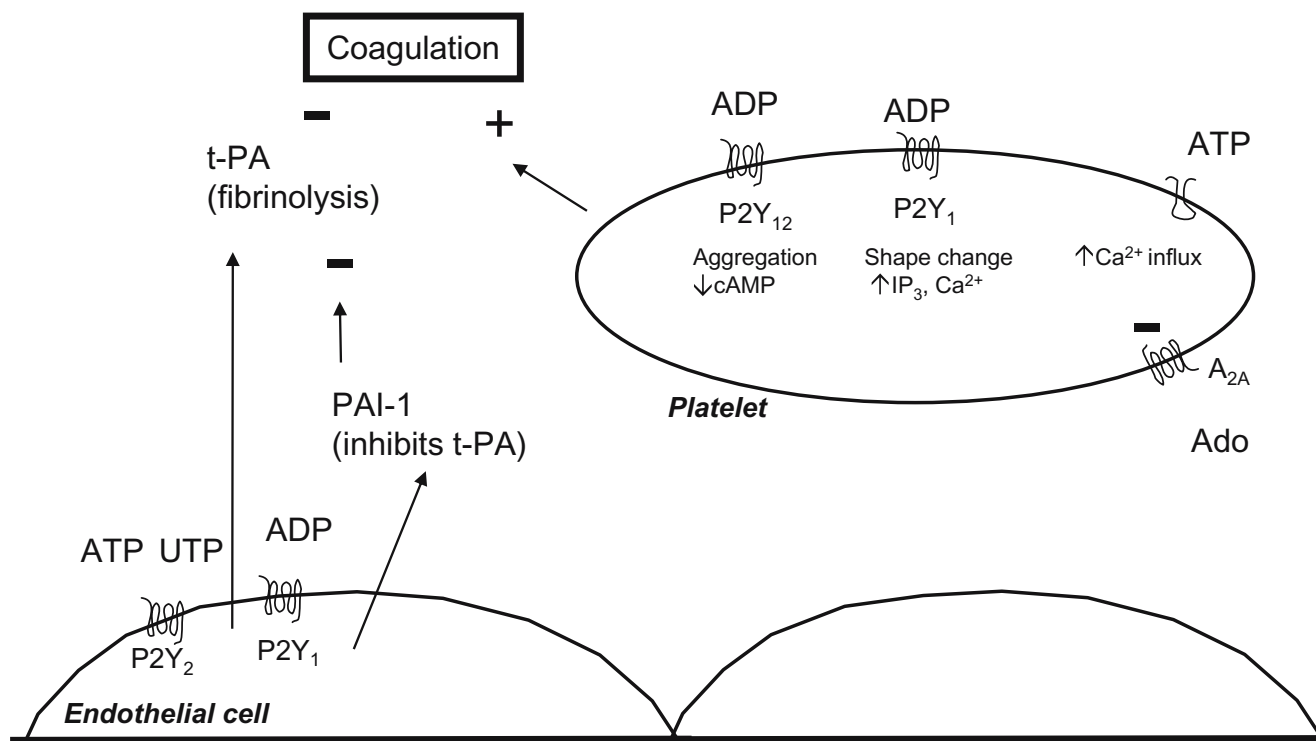


Fig. 4 P2 receptor-mediated regulation of coagulation and platelet aggregation. See text for details. ADP is an important mediator and positive feedback mechanism for platelet aggregation. Activated platelets stimulate coagulation, i.e. formation of a fibrin network. In contrast, ATP, ADP and UTP release tissue plasminogen activator

(*t-PA*) from endothelial cells, resulting in degradation of the fibrin network. At the same time plasminogen activator inhibitor (*PAI-1*) is also released from endothelial cells by ATP stimulation, in turn inhibiting *t-PA*. Thus, P2 receptors regulate several balancing factors in haemostasis. *Ado* adenosine

both platelets and VSMC, could be of double therapeutic benefit in their prevention of both thrombosis and vasospasm [46].

Coagulation

The prothrombotic effect of ADP on the platelets is counteracted by the effects of UTP and ATP on the endothelium that stimulates a substantial release of tissue-type plasminogen activator (tPA) with fibrinolytic effects [51, 211] (Fig. 4). To make the picture even more complex, ATP also releases plasminogen activator inhibitor (PAI-1) which in turn inhibits tPA [212]. Thus, P2 receptors are involved at several stages of haemostasis, which is important for the

development of atherosclerosis and thrombotic occlusions leading to myocardial infarction and stroke.

Clinical importance of P2 receptor-mediated regulation of platelets

The first ADP inhibitors, ticlopidine and clopidogrel, were developed before the cloning and identification of the platelet ADP receptors. Later, it was concluded that they were prodrugs, converted in the liver to a metabolite that binds irreversibly to the P2Y₁₂ receptor resulting in non-competitive antagonism.

Ticlopidine has the disadvantage of a small risk of neutropaenia, so that clinically so far the most important

Table 4 Cardiovascular disease targets

Receptor target	Agonist/antagonist activity of therapeutic drug	Disease
Vascular smooth muscle cell		
P2Y ₂	Antagonist	Hypertension, restenosis, diabetic microvascular disease, subarachnoidal bleeding
P2Y ₆	Antagonist	Hypertension, restenosis, diabetic microvascular disease, subarachnoidal bleeding
P2Y ₁₂	Antagonist	Hypertension, vasospasm
P2X ₁	Antagonist	Hypertension, migraine, subarachnoidal bleeding
P2Y ₆	Agonist	Migraine
SolNTPDase1	Ectonucleotidase	Restenosis, xenograft rejection
Endothelial cell		
P2Y ₂	Antagonist	Atherosclerosis, inflammation
P2X ₄	Antagonist	Vascular remodelling (cancer?), preeclampsia
Heart		
P2Y ₂	Antagonist	Congestive heart failure
P2Y ₆	Antagonist	Congestive heart failure
P2Y ₁₁	Antagonist	Congestive heart failure, myocardial infarction
P2Y ₂	Agonist	Shock, reduced cardiac output, cardiac protection
P2Y ₆	Agonist	Shock, reduced cardiac output
P2Y ₁₁	Agonist	Shock, reduced cardiac output
SolNTPDase1	Ectonucleotidase	Heart transplantation, myocardial infarction
Inflammatory cells		
P2Y ₂	Antagonist	Atherosclerosis
P2Y ₁₁	Antagonist	Atherosclerosis
P2X ₇	Antagonist	Atherosclerosis
Platelets		
P2Y ₁	Antagonist	Myocardial infarction, stroke, peripheral artery disease
P2Y ₁₂	Antagonist	Myocardial infarction, stroke, peripheral artery disease
P2X ₁	Antagonist	Myocardial infarction, stroke, peripheral artery disease
SolNTPDase1	Ectonucleotidase	Myocardial infarction, stroke, peripheral artery disease
Red blood cell		
P2Y ₁₃	Antagonist	Improved peripheral circulation
P2X ₇	Antagonist	Shock
Perivascular nociceptive sensory nerves		
P2Y ₁	Agonist	Migraine, vascular pain
P2X ₃	Antagonist	Migraine, vascular pain
Perivascular sympathetic nerves		
P2Y ₁	Agonist	Hypertension
P2X ₂	Antagonist	Hypertension

The table presents possible targets for the development of new pharmaceutical compounds for the treatment of cardiovascular diseases

contribution of drug development aimed at P2 receptors has been the beneficial effects of the platelet ADP receptor antagonist clopidogrel in atherosclerotic disease. In the CAPRIE trial, clopidogrel was even more effective compared to aspirin in preventing myocardial infarctions [213]. Clopidogrel, as an addition to aspirin, turned out to be necessary to prevent acute in-stent thrombosis in percutaneous coronary interventions. The CURE trial demonstrated that the addition of clopidogrel to aspirin reduced clinical events by 20% in acute coronary syndromes [213, 214]. This has also been shown for ST-elevation myocardial infarctions (COMMIT, CLARITY). However, long-term secondary prevention in high-risk groups with clopidogrel added to aspirin did not have significant benefits (CHARISMA). Clopidogrel combined with aspirin reduces cerebral emboli in patients undergoing carotid endarterectomy [215]. Clopidogrel (Plavix®) is now a blockbuster drug and one of the most sold drugs worldwide to the benefit of atherosclerotic patients. Nevertheless, clopidogrel is a rather weak antagonist at P2Y₁₂ receptors with variable effects, often referred to as clopidogrel resistance. Newer P2Y₁₂ receptor antagonists, such as prasugrel and AZD6140, have a stronger more consistent effect and are currently being studied in major clinical trials (TRITON and PLATO).

Clinical importance of purinergic signalling in cardiovascular disease: future directions

The P2 receptor family provides opportunities for drug development (see Table 4). The large number of receptor subtypes and the increasing knowledge of their tissue distributions may make it possible to target a specific cardiovascular organ with limited side effects. So far, only P2Y₁₂ receptor antagonists have been explored in patients, but with extraordinary success. Clopidogrel is one of the most prescribed drugs in the world for the treatment of thrombosis, stroke and myocardial infarctions in millions of patients. The following compounds, prasugrel and AZD6140, are even more potent P2Y₁₂ receptor antagonists with clinical benefits currently being tested in phase III trials. Improved platelet inhibition can also be achieved with antagonists against the other two P2 receptors on platelets. Studies in P2Y₁ and P2X₁ receptor knockout mice and experimental thrombosis models using selective P2Y₁ and P2X₁ receptor antagonists have shown that, depending on the conditions, these receptors could also be potential targets for new antithrombotic drugs.

ATP, UTP and UDP, acting on P2Y₂ and P2Y₆ receptors, are potent stimulators of inflammation, VSMC proliferation and migration. Antagonists to these receptors may be explored to protect against atherosclerosis, restenosis after

balloon angioplasty, chronic transplant rejection and diabetic microvascular disease.

It is possible that hypertension and pulmonary hypertension could be treated with P2X₁, P2Y₂, P2Y₆ or P2Y₁₂ receptor antagonists. In the heart P2Y₂, P2Y₆ and P2Y₁₁ receptor antagonists may have similar beneficial roles as angiotensin inhibition or β -blockers in the treatment of congestive heart failure. In the acute phase of a myocardial infarction, a UTP analogue could protect cardiomyocytes and limit infarct size. A polymorphism in P2Y₁₁ receptors could be considered as a prognostic marker to identify patients at risk of myocardial infarction and drugs aimed at the receptor could be explored to prevent myocardial infarction.

A promising strategy would be to use soluble NTPDase to prevent thrombosis, restenosis and xenograft transplantation. SolNTPDase1 protects against brain damage in a stroke model and a human genetically engineered solNTPDase has been developed that could be used in clinical trials. Possible clinical indications could be stroke, myocardial infarction and transplantation. P2X₃ receptor antagonists could be used against migraine and vascular pain [see 19, 26].

References

1. Drury A, Szent-Györgyi A (1929) The physiological activity of adenine compounds with special reference to their action upon the mammalian heart. *J Physiol* 68:213–237
2. Burnstock G (1978) A basis for distinguishing two types of purinergic receptor. In: Straub RW (ed) *Cell membrane receptors for drugs and hormones: a multidisciplinary approach*. Raven, New York, pp 107–118
3. Burnstock G (1972) Purinergic nerves. *Pharmacol Rev* 24:509–581
4. Burnstock G (1997) The past, present and future of purine nucleotides as signalling molecules. *Neuropharmacology* 36:1127–1139
5. Ralevic V, Burnstock G (1998) Receptors for purines and pyrimidines. *Pharmacol Rev* 50:413–492
6. Burnstock G (2007) Physiology and pathophysiology of purinergic neurotransmission. *Physiol Rev* 87:659–797
7. Gordon JL (1986) Extracellular ATP: effects, sources and fate. *Biochem J* 233:309–319
8. Zimmermann H (2006) Ectonucleotidases in the nervous system. *Novartis Found Symp* 276:113–128
9. Burnstock G, Kennedy C (1986) A dual function for adenosine 5'-triphosphate in the regulation of vascular tone. Excitatory cotransmitter with noradrenaline from perivascular nerves and locally released inhibitory intravascular agent. *Circ Res* 58:319–330
10. Olsson RA, Pearson JD (1990) Cardiovascular purinoceptors. *Physiol Rev* 70:761–845
11. Ralevic V, Burnstock G (1991) Roles of P2-purinoceptors in the cardiovascular system. *Circulation* 84:1–14
12. Erlinge D (1998) Extracellular ATP: a growth factor for vascular smooth muscle cells. *Gen Pharmacol* 31:1–8
13. Burnstock G (2002) Purinergic signalling and vascular cell proliferation and death. *Arterioscler Thromb Vasc Biol* 22:364–373

14. Di Virgilio F, Solini A (2002) P2 receptors: new potential players in atherosclerosis. *Br J Pharmacol* 135:831–842
15. Gachet C (2006) Regulation of platelet functions by P2 receptors. *Annu Rev Pharmacol Toxicol* 46:277–300
16. Agteresch HJ, Dagnelie PC, van den Berg JW et al (1999) Adenosine triphosphate: established and potential clinical applications. *Drugs* 58:211–232
17. Ralevic V, Burnstock G (2003) Involvement of purinergic signaling in cardiovascular diseases. *Drug News Perspect* 16:133–140
18. Atkinson B, Dwyer K, Enjyoji K et al (2006) Ecto-nucleotidases of the CD39/NTPDase family modulate platelet activation and thrombus formation: potential as therapeutic targets. *Blood Cells Mol Dis* 36:217–222
19. Burnstock G (2006) Pathophysiology and therapeutic potential of purinergic signaling. *Pharmacol Rev* 58:58–86
20. Burnstock G (1976) Do some nerve cells release more than one transmitter? *Neuroscience* 1:239–248
21. Burnstock G (1988) Sympathetic purinergic transmission in small blood vessels. *Trends Pharmacol Sci* 9:116–117
22. Burnstock G (1987) Local control of blood pressure by purines. *Blood Vessels* 24:156–160
23. Burnstock G (1988) Local purinergic regulation of blood pressure (The First John T. Shepherd Lecture). In: Vanhoutte PM (ed) *Vasodilatation: vascular smooth muscle, peptides, autonomic nerves, and endothelium*. Raven, New York, pp 1–14
24. Burnstock G (1990) Local mechanisms of blood flow control by perivascular nerves and endothelium. *J Hypertens Suppl* 8:S95–S106
25. Burnstock G (1990) Noradrenaline and ATP as cotransmitters in sympathetic nerves. *Neurochem Int* 17:357–368
26. Burnstock G (1996) A unifying purinergic hypothesis for the initiation of pain. *Lancet* 347:1604–1605
27. Burnstock G (1993) Introduction: changing face of autonomic and sensory nerves in the circulation. In: Edvinsson L, Uddman R (eds) *Vascular innervation and receptor mechanisms: new perspectives*. Academic, New York, pp 1–22
28. Rubino A, Burnstock G (1996) Capsaicin-sensitive sensory-motor neurotransmission in the peripheral control of cardiovascular function. *Cardiovasc Res* 31:467–479
29. Vial C, Evans RJ (2002) P2X₁ receptor-deficient mice establish the native P2X receptor and a P2Y₆-like receptor in arteries. *Mol Pharmacol* 62:1438–1445
30. Ramme D, Regenold JT, Starke K et al (1987) Identification of the neuroeffector transmitter in jejunal branches of the rabbit mesenteric artery. *Naunyn Schmiedebergs Arch Pharmacol* 336:267–273
31. Chan CM, Unwin RJ, Bardini M et al (1998) Localization of P2X₁ purinoceptors by autoradiography and immunohistochemistry in rat kidneys. *Am J Physiol* 274:F799–F804
32. Inscho EW, Cook AK, Imig JD et al (2004) Renal autoregulation in P2X₁ knockout mice. *Acta Physiol Scand* 181:445–453
33. Inscho EW, Cook AK, Imig JD et al (2003) Physiological role for P2X₁ receptors in renal microvascular autoregulatory behavior. *J Clin Invest* 112:1895–1905
34. Boehm S, Huck S (1997) Receptors controlling transmitter release from sympathetic neurons in vitro. *Prog Neurobiol* 51:225–242
35. Cunha R (2001) Adenosine as a neuromodulator and as a homeostatic regulator in the nervous system: different roles, different sources and different receptors. *Neurochem Int* 38:107–125
36. Todorov LD, Mihaylova-Todorova S, Westfall TD et al (1997) Neuronal release of soluble nucleotidases and their role in neurotransmitter inactivation. *Nature* 387:76–79
37. Rosenmeier JB, Hansen J, Gonzalez-Alonso J (2004) Circulating ATP-induced vasodilatation overrides sympathetic vasoconstrictor activity in human skeletal muscle. *J Physiol* 558:351–365
38. Burnstock G, Kennedy C (1985) Is there a basis for distinguishing two types of P2-purinoceptor? *Gen Pharmacol* 16:433–440
39. Lewis CJ, Evans RJ (2000) Comparison of P2X receptors in rat mesenteric, basilar and septal (coronary) arteries. *J Auton Nerv Syst* 81:69–74
40. Wang L, Karlsson L, Moses S et al (2002) P2 receptor expression profiles in human vascular smooth muscle and endothelial cells. *J Cardiovasc Pharmacol* 40:841–853
41. von Kügelgen I, Haussinger D, Starke K (1987) Evidence for a vasoconstriction-mediating receptor for UTP, distinct from the P2 purinoceptor, in rabbit ear artery. *Naunyn Schmiedebergs Arch Pharmacol* 336:556–560
42. Malmjö M, Adner M, Harden TK et al (2000) The stable pyrimidines UDPβS and UTPγS discriminate between the P2 receptors that mediate vascular contraction and relaxation of the rat mesenteric artery. *Br J Pharmacol* 131:51–56
43. Sévigny J et al (2006) Co-ordinated regulation of P2-receptor signaling by membrane bound NTPDases. *Purinergic Signalling* 2:53
44. Borna C, Wang L, Gudbjartsson T et al (2003) Contractions in human coronary bypass vessels stimulated by extracellular nucleotides. *Ann Thorac Surg* 76:50–57
45. Brinson AE, Harden TK (2001) Differential regulation of the uridine nucleotide-activated P2Y₄ and P2Y₆ receptors. SER-333 and SER-334 in the carboxyl terminus are involved in agonist-dependent phosphorylation desensitization and internalization of the P2Y₄ receptor. *J Biol Chem* 276:11939–11948
46. Wihlborg AK, Wang L, Braun OO et al (2004) ADP receptor P2Y₁₂ is expressed in vascular smooth muscle cells and stimulates contraction in human blood vessels. *Arterioscler Thromb Vasc Biol* 24:1810–1815
47. Burnstock G (1999) Release of vasoactive substances from endothelial cells by shear stress and purinergic mechanosensory transduction. *J Anat* 194:335–342
48. Malmjö M, Edvinsson L, Erlinge D (1998) P2U-receptor mediated endothelium-dependent but nitric oxide-independent vascular relaxation. *Br J Pharmacol* 123:719–729
49. Malmjö M, Erlinge D, Högestätt ED et al (1999) Endothelial P2Y receptors induce hyperpolarisation of vascular smooth muscle by release of endothelium-derived hyperpolarising factor. *Eur J Pharmacol* 364:169–173
50. Mistry H, Gitlin JM, Mitchell JA et al (2003) Endothelium-dependent relaxation and endothelial hyperpolarization by P2Y receptor agonists in rat-isolated mesenteric artery. *Br J Pharmacol* 139:661–671
51. Hrafinkelsdottir T, Erlinge D, Jern S (2001) Extracellular nucleotides ATP and UTP induce a marked acute release of tissue-type plasminogen activator in vivo in man. *Thromb Haemost* 85:875–881
52. Wihlborg AK, Malmjö M, Eyjolfsson A et al (2003) Extracellular nucleotides induce vasodilatation in human arteries via prostaglandins, nitric oxide and endothelium-derived hyperpolarising factor. *Br J Pharmacol* 138:1451–1458
53. Guns PJ, Korda A, Crauwels HM et al (2005) Pharmacological characterization of nucleotide P2Y receptors on endothelial cells of the mouse aorta. *Br J Pharmacol* 146:288–295
54. Yamamoto K, Korenaga R, Kamiya A et al (2000) P2X₄ receptors mediate ATP-induced calcium influx in human vascular endothelial cells. *Am J Physiol Heart Circ Physiol* 279:H285–H292
55. Yamamoto K, Korenaga R, Kamiya A et al (2000) Fluid shear stress activates Ca²⁺ influx into human endothelial cells via P2X₄ purinoceptors. *Circ Res* 87:385–391
56. Bodin P, Bailey D, Burnstock G (1991) Increased flow-induced ATP release from isolated vascular endothelial cells but not smooth muscle cells. *Br J Pharmacol* 103:1203–1205

57. Yamamoto K, Sokabe T, Matsumoto T et al (2006) Impaired flow-dependent control of vascular tone and remodeling in P2X₄-deficient mice. *Nat Med* 12:133–137
58. Kaczmarek E, Koziak K, Sévigny J et al (1996) Identification and characterization of CD39/vascular ATP diphosphohydrolase. *J Biol Chem* 271:33116–33122
59. Yegutkin G, Bodin P, Burnstock G (2000) Effect of shear stress on the release of soluble ecto-enzymes ATPase and 5'-nucleotidase along with endogenous ATP from vascular endothelial cells. *Br J Pharmacol* 129:921–926
60. Vassort G (2001) Adenosine 5'-triphosphate: a P2-purinergic agonist in the myocardium. *Physiol Rev* 81:767–806
61. Saito D, Steinhart CR, Nixon DC et al (1981) Intracoronary adenosine deaminase reduces canine myocardial reactive hyperemia. *Circ Res* 49:1262–1267
62. Zatta AJ, Headrick JP (2005) Mediators of coronary reactive hyperaemia in isolated mouse heart. *Br J Pharmacol* 144:576–587
63. Olivecrona GK et al (2006) Coronary artery reperfusion: the ADP receptor P2Y₁ mediates early reactive hyperemia in vivo in pigs. *Purinergic Signalling* 1:59–64
64. Ellsworth ML, Forrester T, Ellis CG et al (1995) The erythrocyte as a regulator of vascular tone. *Am J Physiol* 269:H2155–H2161
65. McMahon TJ, Moon RE, Lusching BP et al (2002) Nitric oxide in the human respiratory cycle. *Nat Med* 8:711–717
66. Cosby K, Partovi KS, Crawford JH et al (2003) Nitrite reduction to nitric oxide by deoxyhemoglobin vasodilates the human circulation. *Nat Med* 9:1498–1505
67. Bergfeld GR, Forrester T (1992) Release of ATP from human erythrocytes in response to a brief period of hypoxia and hypercapnia. *Cardiovasc Res* 26:40–47
68. Sprague RS (2001) Participation of cAMP in a signal-transduction pathway relating erythrocyte deformation to ATP release. *Am J Physiol Cell Physiol* 281:C1158–C1164
69. Sprague RS, Stephenson AH, Ellsworth ML et al (2001) Impaired release of ATP from red blood cells of humans with primary pulmonary hypertension. *Exp Biol Med (Maywood)* 226:434–439
70. Dietrich HH, Ellsworth ML, Sprague RS et al (2000) Red blood cell regulation of microvascular tone through adenosine triphosphate. *Am J Physiol Heart Circ Physiol* 278:H1294–H1298
71. Gonzalez-Alonso J, Olsen DB, Saltin B (2002) Erythrocyte and the regulation of human skeletal muscle blood flow and oxygen delivery: role of circulating ATP. *Circ Res* 91:1046–1055
72. Farias M, Gorman MW, Savage MV et al (2005) Plasma ATP during exercise: possible role in regulation of coronary blood flow. *Am J Physiol Heart Circ Physiol* 288:H1586–H1590
73. Wang L, Olivecrona G, Götberg M et al (2005) ADP acting on P2Y₁₃ receptors is a negative feedback pathway for ATP release from human red blood cells. *Circ Res* 96:189–196
74. Olearczyk JJ, Ellsworth ML, Stephenson AH et al (2004) Nitric oxide inhibits ATP release from erythrocytes. *J Pharmacol Exp Ther* 309:1079–1084
75. Trams EJ, Kauffman H, Burnstock G (1980) A proposal for the role of ecto-enzymes and adenylates in traumatic shock. *J Theor Biol* 87:609–621
76. Sluyter R, Shemon AN, Barden JA et al (2004) Extracellular ATP increases cation fluxes in human erythrocytes by activation of the P2X₇ receptor. *J Biol Chem* 279:44749–44755
77. Bodin P, Burnstock G (1996) ATP-stimulated release of ATP by human endothelial cells. *J Cardiovasc Pharmacol* 27:872–875
78. Tanneur V, Duranton C, Brand VB (2006) Purinoceptors are involved in the induction of an osmolyte permeability in malaria-infected and oxidized human erythrocytes. *FASEB J* 20:133–135
79. Locovei S, Bao L, Dahl G (2006) Pannexin 1 in erythrocytes: function without a gap. *Proc Natl Acad Sci USA* 103:7655–7659
80. Vidal M, Hicks PE, Langer SZ (1986) Differential effects of α - β -methylene ATP on responses to nerve stimulation in SHR and WKY tail arteries. *Naunyn Schmiedebergs Arch Pharmacol* 332:384–390
81. Brock JA, Van Helden DF (1995) Enhanced excitatory junction potentials in mesenteric arteries from spontaneously hypertensive rats. *Pflugers Arch* 430:901–908
82. Fernandez O, Wangenstein R, Osuna A et al (2000) Renal vascular reactivity to P₂-purinoceptor activation in spontaneously hypertensive rats. *Pharmacology* 60:47–50
83. Galligan JJ, Hess MC, Miller SB et al (2001) Differential localization of P2 receptor subtypes in mesenteric arteries and veins of normotensive and hypertensive rats. *J Pharmacol Exp Ther* 296:478–485
84. Yang D, Gluais P, Zhang JN et al (2004) Endothelium-dependent contractions to acetylcholine, ATP and the calcium ionophore A 23187 in aortas from spontaneously hypertensive and normotensive rats. *Fundam Clin Pharmacol* 18:321–326
85. Vonend O, Turner CM, Chan CM et al (2004) Glomerular expression of the ATP-sensitive P2X receptor in diabetic and hypertensive rat models. *Kidney Int* 66:157–166
86. Schluter H, Offers E, Brüggemann G et al (1994) Diadenosine phosphates and the physiological control of blood pressure. *Nature* 367:186–188
87. Hollah P, Hausberg M, Kosch M et al (2001) A novel assay for determination of diadenosine polyphosphates in human platelets: studies in normotensive subjects and in patients with essential hypertension. *J Hypertens* 19:237–245
88. Jankowski V, Tölle M, Vanholder R et al (2005) Uridine adenosine tetraphosphate: a novel endothelium-derived vasoconstrictive factor. *Nat Med* 11:223–227
89. Roberts VH, Webster RP, Brockman DE et al (2006) Post-translational modifications of the P2X₄ purinergic receptor subtype in the human placenta are altered in preeclampsia. *Placenta* 28:270–277
90. Sprague RS, Olearczyk JJ, Spence DM et al (2003) Extracellular ATP signaling in the rabbit lung: erythrocytes as determinants of vascular resistance. *Am J Physiol Heart Circ Physiol* 285:H693–H700
91. Greenberg B, Rhoden K, Barnes PJ (1987) Endothelium-dependent relaxation of human pulmonary arteries. *Am J Physiol* 252:H434–H438
92. Zhang S, Remillard CV, Fantozzi I et al (2004) ATP-induced mitogenesis is mediated by cyclic AMP response element-binding protein-enhanced TRPC4 expression and activity in human pulmonary artery smooth muscle cells. *Am J Physiol Cell Physiol* 287:C1192–C1201
93. Malmjö M, Hou M, Pendergast W et al (2003) Potent P2Y₆ receptor mediated contractions in human cerebral arteries. *BMC Pharmacol* 3:4
94. Burnstock G (1989) The role of adenosine triphosphate in migraine. *Biomed Pharmacother* 43:727–736
95. Waeber C, Moskowitz MA (2003) Therapeutic implications of central and peripheral neurologic mechanisms in migraine. *Neurology* 61:S9–S20
96. Fabbretti E, D'Arco M, Fabbro A et al (2006) Delayed upregulation of ATP P2X₃ receptors of trigeminal sensory neurons by calcitonin gene-related peptide. *J Neurosci* 26:6163–6171
97. Peroutka SJ (2004) Migraine: a chronic sympathetic nervous system disorder. *Headache* 44:53–64
98. Hansson GK (2005) Inflammation, atherosclerosis, and coronary artery disease. *N Engl J Med* 352:1685–1695
99. Hashimoto M, Shinozuka K, Shahdat HM et al (1998) Antihypertensive effect of all-cis-5,8,11,14,17-icosapentaenoate of aged rats is associated with an increase in the release of ATP from the caudal artery. *J Vasc Res* 35:55–62

100. Hashimoto M, Shinozuka K, Tanabe Y et al (1998) Long-term supplementation with a high cholesterol diet decreases the release of ATP from the caudal artery in aged rats. *Life Sci* 63:1879–1885
101. Karoon P, Burnstock G (1998) Reduced sympathetic noradrenergic neurotransmission in the tail artery of Donryu rats fed with high cholesterol-supplemented diet. *Br J Pharmacol* 123:1016–1021
102. Di Virgilio F, Chiozzi P, Ferrari D et al (2001) Nucleotide receptors: an emerging family of regulatory molecules in blood cells. *Blood* 97:587–600
103. Wang L, Jacobsen SE, Bengtsson A et al (2004) P2 receptor mRNA expression profiles in human lymphocytes, monocytes and CD34+ stem and progenitor cells. *BMC Immunol* 5:16
104. Baricordi OR, Ferrari D, Melchiorri L et al (1996) An ATP-activated channel is involved in mitogenic stimulation of human T lymphocytes. *Blood* 87:682–690
105. Kawamura H, Aswad F, Minagawa M et al (2005) P2X₇ receptor-dependent and -independent T cell death is induced by nicotinamide adenine dinucleotide. *J Immunol* 174:1971–1979
106. Ferrari D, Chiozzi P, Falzoni S et al (1997) Extracellular ATP triggers IL-1 β release by activating the purinergic P2Z receptor of human macrophages. *J Immunol* 159:1451–1458
107. Suzuki T, Hide I, Ido K et al (2004) Production and release of neuroprotective tumor necrosis factor by P2X₇ receptor-activated microglia. *J Neurosci* 24:1–7
108. Gu B, Bendall LJ, Wiley JS (1998) Adenosine triphosphate-induced shedding of CD23 and L-selectin (CD62L) from lymphocytes is mediated by the same receptor but different metalloproteases. *Blood* 92:946–951
109. Labasi JM, Petrushova N, Donovan C et al (2002) Absence of the P2X₇ receptor alters leukocyte function and attenuates an inflammatory response. *J Immunol* 168:6436–6445
110. Idzko M, Dichmann S, Ferrari D et al (2002) Nucleotides induce chemotaxis and actin polymerization in immature but not mature human dendritic cells via activation of pertussis toxin-sensitive P2y receptors. *Blood* 100:925–932
111. Schmid-Antomarchi H, Schmid-Alliana A, Romey G et al (1997) Extracellular ATP and UTP control the generation of reactive oxygen intermediates in human macrophages through the opening of a charybdotoxin-sensitive Ca²⁺-dependent K⁺ channel. *J Immunol* 159:6209–6215
112. Hechler B et al (2006) Reduced atherosclerotic lesions in P2Y₁/ApoE double knockout mice. *Purinergic Signalling* 2:299
113. Duhant X, Schandené L, Bruyns C et al (2002) Extracellular adenine nucleotides inhibit the activation of human CD4⁺ T lymphocytes. *J Immunol* 169:15–21
114. Kaufmann A, Musset B, Limberg SH et al (2005) “Host tissue damage” signal ATP promotes non-directional migration and negatively regulates toll-like receptor signaling in human monocytes. *J Biol Chem* 280:32459–32467
115. Marteau F, Gonzalez NS, Communi D et al (2005) Thrombospondin-1 and indoleamine 2,3-dioxygenase are major targets of extracellular ATP in human dendritic cells. *Blood* 106:3860–3866
116. Amisten S, Melander O, Wihlborg AK et al (2006) Increased risk of acute myocardial infarction and elevated levels of C-reactive protein in carriers of the Thr-87 variant of the ATP receptor P2Y₁₁. *Eur Heart J* 28:13–18
117. Dawicki DD, McGowan-Jordan J, Bullard S et al (1995) Extracellular nucleotides stimulate leukocyte adherence to cultured pulmonary artery endothelial cells. *Am J Physiol* 268:L666–L673
118. Seiffert K, Ding W, Wagner JA et al (2006) ATP γ S enhances the production of inflammatory mediators by a human dermal endothelial cell line via purinergic receptor signaling. *J Invest Dermatol* 126:1017–1027
119. Seye CI, Yu N, Jain R et al (2003) The P2Y₂ nucleotide receptor mediates UTP-induced vascular cell adhesion molecule-1 expression in coronary artery endothelial cells. *J Biol Chem* 278:24960–24965
120. Seye CI, Yu N, González FA et al (2004) The P2Y₂ nucleotide receptor mediates vascular cell adhesion molecule-1 expression through interaction with VEGF receptor-2 (KDR/Flk-1). *J Biol Chem* 279:35679–35686
121. Seye CI, Kong Q, Erb L et al (2002) Functional P2Y₂ nucleotide receptors mediate uridine 5'-triphosphate-induced intimal hyperplasia in collared rabbit carotid arteries. *Circulation* 106:2720–2726
122. Hart MN, Heistad DD, Brody MJ (1980) Effect of chronic hypertension and sympathetic denervation on wall/lumen ratio of cerebral vessels. *Hypertension* 2:419–423
123. Erlinge D, Yoo H, Edvinsson L et al (1993) Mitogenic effects of ATP on vascular smooth muscle cells vs. other growth factors and sympathetic cotransmitters. *Am J Physiol* 265:H1089–H1097
124. Wang DJ, Huang NN, Heppel LA (1992) Extracellular ATP and ADP stimulate proliferation of porcine aortic smooth muscle cells. *J Cell Physiol* 153:221–233
125. Malam-Souley R, Seye C, Gadeau AP et al (1996) Nucleotide receptor P2u partially mediates ATP-induced cell cycle progression of aortic smooth muscle cells. *J Cell Physiol* 166:57–65
126. Erlinge D, You J, Wahlestedt C et al (1995) Characterisation of an ATP receptor mediating mitogenesis in vascular smooth muscle cells. *Eur J Pharmacol* 289:135–149
127. Hou M, Harden TK, Kuhn CM et al (2002) UDP acts as a growth factor for vascular smooth muscle cells by activation of P2Y₆ receptors. *Am J Physiol Heart Circ Physiol* 282:H784–H792
128. Wilden PA, Agazie YM, Kaufman R et al (1998) ATP-stimulated smooth muscle cell proliferation requires independent ERK and PI3K signaling pathways. *Am J Physiol* 275:H1209–H1215
129. Sauzeau V, Le Jeune H, Cario-Toumaniantz C et al (2000) P2Y₁, P2Y₂, P2Y₄, and P2Y₆ receptors are coupled to Rho and Rho kinase activation in vascular myocytes. *Am J Physiol Heart Circ Physiol* 278:H1751–H1761
130. Miyagi Y, Kobayashi S, Ahmed A et al (1996) P2U purinergic activation leads to the cell cycle progression from the G1 to the S and M phases but not from the G0 to G1 phase in vascular smooth muscle cells in primary culture. *Biochem Biophys Res Commun* 222:652–658
131. Erlinge D, Brunkwall J, Edvinsson L (1994) Neuropeptide Y stimulates proliferation of human vascular smooth muscle cells: cooperation with noradrenaline and ATP. *Regul Pept* 50:259–265
132. Erb L, Liu J, Ockerhausen J et al (2001) An RGD sequence in the P2Y₂ receptor interacts with $\alpha_v\beta_3$ integrins and is required for G_o-mediated signal transduction. *J Cell Biol* 153:491–501
133. Robinson WP 3rd, Douillet CD, Milano PM et al (2006) ATP stimulates MMP-2 release from human aortic smooth muscle cells via JNK signaling pathway. *Am J Physiol Heart Circ Physiol* 290:H1988–H1996
134. Chaulet H, Desgranges C, Renault MA et al (2001) Extracellular nucleotides induce arterial smooth muscle cell migration via osteopontin. *Circ Res* 89:772–778
135. Koziak K et al (2006) Adenovirus-mediated over-expression of CD39 inhibits neointima formation in rat aorta. *Purinergic Signalling* 2:Addendum
136. Imai M, Takigami K, Guckelberger O et al (1999) Modulation of nucleoside triphosphate diphosphohydrolase-1 (NTPDase-1) cd39 in xenograft rejection. *Mol Med* 5:743–752
137. Pillois X, Chaulet H, Belloc I et al (2002) Nucleotide receptors involved in UTP-induced rat arterial smooth muscle cell migration. *Circ Res* 90:678–681

138. Ralevic V, Belai A, Burnstock G (1995) Effects of streptozotocin-diabetes on sympathetic nerve, endothelial and smooth muscle function in the rat mesenteric arterial bed. *Eur J Pharmacol* 286:193–199
139. Gür S, Ozturk B (2000) Altered relaxant responses to adenosine and adenosine 5'-triphosphate in the corpus cavernosum from men and rats with diabetes. *Pharmacology* 60:105–112
140. Sprague RS, Stephenson AH, Bowles EA et al (2006) Reduced expression of G_i in erythrocytes of humans with type 2 diabetes is associated with impairment of both cAMP generation and ATP release. *Diabetes* 55:3588–3593
141. Parodi J, Flores C, Aguayo C et al (2002) Inhibition of nitrobenzylthioinosine-sensitive adenosine transport by elevated D-glucose involves activation of P2Y₂ purinoceptors in human umbilical vein endothelial cells. *Circ Res* 90:570–577
142. Hellman B, Dansk H, Grapengiesser E (2004) Pancreatic β -cells communicate via intermittent release of ATP. *Am J Physiol Endocrinol Metab* 286:E759–E765
143. Nilsson J, Nilsson LM, Chen YW et al (2006) High glucose activates nuclear factor of activated T cells in native vascular smooth muscle. *Arterioscler Thromb Vasc Biol* 26:794–800
144. Pacaud P, Malam-Souley R, Loirand G et al (1995) ATP raises [Ca²⁺]_i via different P2-receptor subtypes in freshly isolated and cultured aortic myocytes. *Am J Physiol* 269:H30–H36
145. Erlinge D, Hou M, Webb TE et al (1998) Phenotype changes of the vascular smooth muscle cell regulate P2 receptor expression as measured by quantitative RT-PCR. *Biochem Biophys Res Commun* 248:864–870
146. Hou M, Möller S, Edvinsson L et al (1999) MAPKK-dependent growth factor-induced upregulation of P2Y₂ receptors in vascular smooth muscle cells. *Biochem Biophys Res Commun* 258:648–652
147. Hou M, Möller S, Edvinsson L et al (2000) Cytokines induce upregulation of vascular P2Y₂ receptors and increased mitogenic responses to UTP and ATP. *Arterioscler Thromb Vasc Biol* 20:2064–2069
148. Seye CI, Gadeau AP, Daret D et al (1997) Overexpression of P2Y₂ purinoceptor in intimal lesions of the rat aorta. *Arterioscler Thromb Vasc Biol* 17:3602–3610
149. Shen J, Seye CI, Wang M et al (2004) Cloning, up-regulation, and mitogenic role of porcine P2Y₂ receptor in coronary artery smooth muscle cells. *Mol Pharmacol* 66:1265–1274
150. Hogarth DK, Sandbo N, Taurin S et al (2004) Dual role of PKA in phenotypic modulation of vascular smooth muscle cells by extracellular ATP. *Am J Physiol Cell Physiol* 287:C449–C456
151. Wang L, Andersson M, Karlsson L et al (2003) Increased mitogenic and decreased contractile P2 receptors in smooth muscle cells by shear stress in human vessels with intact endothelium. *Arterioscler Thromb Vasc Biol* 23:1370–1376
152. Van Daele P, Van Coevorden A, Roger PP et al (1992) Effects of adenine nucleotides on the proliferation of aortic endothelial cells. *Circ Res* 70:82–90
153. Kaczmarek E, Erb L, Koziak K et al (2005) Modulation of endothelial cell migration by extracellular nucleotides: involvement of focal adhesion kinase and phosphatidylinositol 3-kinase-mediated pathways. *Thromb Haemostasis* 93:735–742
154. Gerasimovskaya EV, Davie NJ, Ahmad S et al (2005) Extracellular adenosine triphosphate: a potential regulator of vasa vasorum neovascularization in hypoxia-induced pulmonary vascular remodeling. *Chest* 128(6 Suppl):608S–610S
155. Eltzschig HK, Ibla JC, Furuta GT (2003) Coordinated adenine nucleotide phosphohydrolysis and nucleoside signaling in post-hypoxic endothelium: role of ectonucleotidases and adenosine A2B receptors. *J Exp Med* 198:783–796
156. Goepfert C, Sundberg C, Sévigny J et al (2001) Disordered cellular migration and angiogenesis in cd39-null mice. *Circulation* 104:3109–3115
157. Cario-Toumaniantz C, Loirand G, Ladoux A et al (1998) P2X₇ receptor activation-induced contraction and lysis in human saphenous vein smooth muscle. *Circ Res* 83:196–203
158. Metcalfe MJ, Baker DM, Burnstock G (2006) Purinoceptor expression on keratinocytes reflects their function in the epidermis during chronic venous insufficiency. *Arch Dermatol Res* 298:301–307
159. Clemens MG, Forrester T (1981) Appearance of adenosine triphosphate in the coronary sinus effluent from isolated working rat heart in response to hypoxia. *J Physiol* 312:143–158
160. Bogdanov Y, Rubino A, Burnstock G (1998) Characterisation of subtypes of the P2X and P2Y families of ATP receptors in the foetal human heart. *Life Sci* 62:697–703
161. Wihlborg AK, Balogh J, Wang L et al (2006) Positive inotropic effects by uridine triphosphate (UTP) and uridine diphosphate (UDP) via P2Y₂ and P2Y₆ receptors on cardiomyocytes and release of UTP in man during myocardial infarction. *Circ Res* 98:970–976
162. Danziger RS, Raffaelli S, Moreno-Sanchez R et al (1988) Extracellular ATP has a potent effect to enhance cytosolic calcium and contractility in single ventricular myocytes. *Cell Calcium* 9:193–199
163. Zheng JS, Boluyt MO, Long X et al (1996) Extracellular ATP inhibits adrenergic agonist-induced hypertrophy of neonatal cardiac myocytes. *Circ Res* 78:525–535
164. Podrasky E, Xu D, Liang BT (1997) A novel phospholipase C- and cAMP-independent positive inotropic mechanism via a P2 purinoceptor. *Am J Physiol* 273:H2380–H2387
165. Mei Q, Liang BT (2001) P2 purinergic receptor activation enhances cardiac contractility in isolated rat and mouse hearts. *Am J Physiol Heart Circ Physiol* 281:H334–H341
166. Frolidi G, Pandolfo L, Chinellato A et al (1994) Dual effect of ATP and UTP on rat atria: which types of receptors are involved? *Naunyn Schmiedeberg's Arch Pharmacol* 349:381–386
167. Scamps F, Vassort G (1994) Pharmacological profile of the ATP-mediated increase in L-type calcium current amplitude and activation of a non-specific cationic current in rat ventricular cells. *Br J Pharmacol* 113:982–986
168. Vahlensieck U, Boknik P, Gombosová I et al (1999) Inotropic effects of diadenosine tetraphosphate (AP4A) in human and animal cardiac preparations. *J Pharmacol Exp Ther* 288:805–813
169. Balogh J, Wihlborg AK, Isackson H et al (2005) Phospholipase C and cAMP-dependent positive inotropic effects of ATP in mouse cardiomyocytes via P2Y₁₁-like receptors. *J Mol Cell Cardiol* 39:223–230
170. Flitney FW, Singh J (1980) Inotropic responses of the frog ventricle to adenosine triphosphate and related changes in endogenous cyclic nucleotides. *J Physiol* 304:21–42
171. Puecat M, Bony C, Jaconi M et al (1998) Specific activation of adenylyl cyclase V by a purinergic agonist. *FEBS Lett* 431:189–194
172. Communi D, Govaerts C, Parmentier M et al (1997) Cloning of a human purinergic P2Y receptor coupled to phospholipase C and adenylyl cyclase. *J Biol Chem* 272:31969–31973
173. Communi D, Robaye B, Boeynaems JM (1999) Pharmacological characterization of the human P2Y₁₁ receptor. *Br J Pharmacol* 128:1199–1206
174. Zambon AC, Hughes RJ, Meszaros JG et al (2000) P2Y₂ receptor of MDCK cells: cloning, expression, and cell-specific signaling. *Am J Physiol Renal Physiol* 279:F1045–F1052
175. Hou M, Malmjö M, Möller S et al (1999) Increase in cardiac P2X₁- and P2Y₂-receptor mRNA levels in congestive heart failure. *Life Sci* 65:1195–1206
176. Frolidi G, Varani K, Chinellato A et al (1997) P2X-purinoceptors in the heart: actions of ATP and UTP. *Life Sci* 60:1419–1430
177. Frolidi G, Ragazzi E, Caparrotta L (2001) Do ATP and UTP involve cGMP in positive inotropism on rat atria? *Comp Biochem Physiol C Toxicol Pharmacol* 128:265–274

178. Erlinge D, Harnek J, van Heusden C et al (2005) Uridine triphosphate (UTP) is released during cardiac ischemia. *Int J Cardiol* 100:427–433
179. Borna C, Lazarowski E, van Heusden C et al (2005) Resistance to aspirin is increased by ST-elevation myocardial infarction and correlates with adenosine diphosphate levels. *Thromb J* 3:10
180. Kittel A, Kiss AL, Müllner N et al (2005) Expression of NTPDase1 and caveolins in human cardiovascular disease. *Histochem Cell Biol* 124:51–59
181. Gunduz D, Kassekert SA, Härtel FV et al (2006) Accumulation of extracellular ATP protects against acute reperfusion injury in rat heart endothelial cells. *Cardiovasc Res* 71:764–773
182. Pham TM, Morris JB, Arthur JF et al (2003) UTP but not ATP causes hypertrophic growth in neonatal rat cardiomyocytes. *J Mol Cell Cardiol* 35:287–292
183. Zheng JS, Boluyt MO, O'Neill L et al (1994) Extracellular ATP induces immediate-early gene expression but not cellular hypertrophy in neonatal cardiac myocytes. *Circ Res* 74:1034–1041
184. Morris JB, Pham TM, Kenney B et al (2004) UTP transactivates epidermal growth factor receptors and promotes cardiomyocyte hypertrophy despite inhibiting transcription of the hypertrophic marker gene, atrial natriuretic peptide. *J Biol Chem* 279:8740–8746
185. Yitzhaki S, Shneyvays V, Jacobson KA et al (2005) Involvement of uracil nucleotides in protection of cardiomyocytes from hypoxic stress. *Biochem Pharmacol* 69:1215–1223
186. Yitzhaki S, Shainberg A, Cheporko Y et al (2006) Uridine-5'-triphosphate (UTP) reduces infarct size and improves rat heart function after myocardial infarct. *Biochem Pharmacol* 72:949–955
187. Saini HK, Shao Q, Musat S et al (2005) Imidapril treatment improves the attenuated inotropic and intracellular calcium responses to ATP in heart failure due to myocardial infarction. *Br J Pharmacol* 144:202–211
188. Banfi C, Ferrario S, De Vincenti O et al (2005) P2 receptors in human heart: upregulation of P2X₆ in patients undergoing heart transplantation, interaction with TNF α and potential role in myocardial cell death. *J Mol Cell Cardiol* 39:929–939
189. Malmjö M, Bergdahl A, Möller S et al (1999) Congestive heart failure induces downregulation of P2X₁-receptors in resistance arteries. *Cardiovasc Res* 43:219–227
190. Yang A, Sonin D, Jones L et al (2004) A beneficial role of cardiac P2X₄ receptors in heart failure: rescue of the calsequestrin overexpression model of cardiomyopathy. *Am J Physiol Heart Circ Physiol* 287:H1096–H1103
191. Horackova M, Huang MH, Armour JA (1994) Purinergic modulation of adult guinea pig cardiomyocytes in long term cultures and co-cultures with extracardiac or intrinsic cardiac neurones. *Cardiovasc Res* 28:673–679
192. Jiang L, Bardini M, Keogh A et al (2005) P2X₁ receptors are closely associated with connexin 43 in human ventricular myocardium. *Int J Cardiol* 98:291–297
193. Gaarder A, Jonsen J, Laland S et al (1961) Adenosine diphosphate in red cells as a factor in the adhesiveness of human blood platelets. *Nature* 192:531–532
194. Daniel JL, Dangelmaier C, Jin J et al (1998) Molecular basis for ADP-induced platelet activation. I. Evidence for three distinct ADP receptors on human platelets. *J Biol Chem* 273:2024–2029
195. Jin J, Kunapuli SP (1998) Coactivation of two different G protein-coupled receptors is essential for ADP-induced platelet aggregation. *Proc Natl Acad Sci USA* 95:8070–8074
196. Jin J, Daniel JL, Kunapuli SP (1998) Molecular basis for ADP-induced platelet activation. II. The P2Y₁ receptor mediates ADP-induced intracellular calcium mobilization and shape change in platelets. *J Biol Chem* 273:2030–2034
197. Kunapuli SP, Dorsam RT, Kim S et al (2003) Platelet purinergic receptors. *Curr Opin Pharmacol* 3:175–180
198. Gachet C, Hechler B (2005) The platelet P2 receptors in thrombosis. *Semin Thromb Hemost* 31:162–167
199. Foster CJ, Prosser DM, Agans JM et al (2001) Molecular identification and characterization of the platelet ADP receptor targeted by thienopyridine antithrombotic drugs. *J Clin Invest* 107:1591–1598
200. Fabre JE, Nguyen M, Latour A et al (1999) Decreased platelet aggregation, increased bleeding time and resistance to thromboembolism in P2Y₁-deficient mice. *Nat Med* 5:1199–1202
201. Leon C, Alex M, Klocke A et al (2004) Platelet ADP receptors contribute to the initiation of intravascular coagulation. *Blood* 103:594–600
202. Gachet C (2005) The platelet P2 receptors as molecular targets for old and new antiplatelet drugs. *Pharmacol Ther* 108:180–192
203. Leon C, Hechler B, Freund M et al (1999) Defective platelet aggregation and increased resistance to thrombosis in purinergic P2Y₁ receptor-null mice. *J Clin Invest* 104:1731–1737
204. Mahaut-Smith MP, Tolhurst G, Evans RJ (2004) Emerging roles for P2X₁ receptors in platelet activation. *Platelets* 15:131–144
205. Birk AV, Leno E, Robertson HD et al (2003) Interaction between ATP and catecholamines in stimulation of platelet aggregation. *Am J Physiol Heart Circ Physiol* 284:H619–H625
206. Sévigny J, Sundberg C, Braun N et al (2002) Differential catalytic properties and vascular topography of murine nucleoside triphosphate diphosphohydrolase 1 (NTPDase1) and NTPDase2 have implications for thromboregulation. *Blood* 99:2801–2809
207. Robson SC, Kaczmarek E, Siegel JB et al (1997) Loss of ATP diphosphohydrolase activity with endothelial cell activation. *J Exp Med* 185:153–163
208. Koyamada N, Miyatake T, Candinas D et al (1996) Apyrase administration prolongs discordant xenograft survival. *Transplantation* 62:1739–1743
209. Imai M, Takigami K, Guckelberger O et al (2000) Recombinant adenoviral mediated CD39 gene transfer prolongs cardiac xenograft survival. *Transplantation* 70:864–870
210. Mizumoto N, Kumamoto T, Robson SC et al (2002) CD39 is the dominant Langerhans cell-associated ecto-NTPDase: modulatory roles in inflammation and immune responsiveness. *Nat Med* 8:358–365
211. Pearson JD (1993) The control of production and release of haemostatic factors in the endothelial cell. *Baillieres Clin Haematol* 6:629–651
212. Bouchie JL, Chen HC, Carney R et al (2000) P2Y receptor regulation of PAI-1 expression in vascular smooth muscle cells. *Arterioscler Thromb Vasc Biol* 20:866–873
213. CAPRIE Steering Committee (1996) A randomised, blinded, trial of clopidogrel versus aspirin in patients at risk of ischaemic events. *Lancet* 348:1329–1339
214. Mehta SR, Yusuf S, Peters RJ et al (2001) Effects of pretreatment with clopidogrel and aspirin followed by long-term therapy in patients undergoing percutaneous coronary intervention: the PCI-CURE study. *Lancet* 358:527–533
215. Payne DA, Jones CI, Hayes PD et al (2004) Beneficial effects of clopidogrel combined with aspirin in reducing cerebral emboli in patients undergoing carotid endarterectomy. *Circulation* 109:1476–1481