

## Research Article

# Molecular Characterization of Culturable Aerobic Bacteria in the Midgut of Field-Caught *Culex tritaeniorhynchus*, *Culex gelidus*, and *Mansonia annulifera* Mosquitoes in the Gampaha District of Sri Lanka

Nayana Gunathilaka <sup>1</sup>, Koshila Ranasinghe <sup>2</sup>, Deepika Amarasinghe <sup>2</sup>,  
Wasana Rodrigo,<sup>3</sup> Harendra Mallawarachchi,<sup>4</sup> and Nilmini Chandrasena<sup>1</sup>

<sup>1</sup>Department of Parasitology, Faculty of Medicine, University of Kelaniya, Ragama, Sri Lanka

<sup>2</sup>Department of Zoology and Environmental Management, Faculty of Science, University of Kelaniya, Colombo, Sri Lanka

<sup>3</sup>Biotechnology Unit, Industrial Technology Institute, Colombo 07, Colombo, Sri Lanka

<sup>4</sup>Department of Parasitology and Medical Entomology, Medical Research Institute, Colombo 08, Colombo, Sri Lanka

Correspondence should be addressed to Nayana Gunathilaka; [n.gunathilaka@kln.ac.lk](mailto:n.gunathilaka@kln.ac.lk)

Received 5 June 2020; Revised 8 August 2020; Accepted 17 September 2020; Published 5 October 2020

Academic Editor: Wen Jun Li

Copyright © 2020 Nayana Gunathilaka et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

**Background.** Larval and adult mosquito stages harbor different extracellular microbes exhibiting various functions in their digestive tract including host-parasite interactions. Midgut symbiotic bacteria can be genetically exploited to express molecules within the vectors, altering vector competency and potential for disease transmission. Therefore, identification of mosquito gut inhabiting microbiota is of ample importance before developing novel vector control strategies that involve modification of vectors. **Method.** Adult mosquitoes of *Culex tritaeniorhynchus*, *Culex gelidus*, and *Mansonia annulifera* were collected from selected Medical Officer of Health (MOH) areas in the Gampaha district of Sri Lanka. Midgut lysates of the field-caught non-blood-fed female mosquitoes were cultured in Plate Count Agar medium, and Prokaryotic 16S ribosomal RNA partial genes of the isolated bacteria colonies were amplified followed by DNA sequencing. Diversity indices were used to assess the diversity and richness of the bacterial isolates in three mosquito species. The distribution pattern of bacterial isolates between different mosquito species was assessed by Distance-Based Redundancy Analysis (dbRDA). **Results.** A total of 20 bacterial species (*Staphylococcus pasteurii*, *Bacillus megaterium*, *Staphylococcus cohnii*, *Pantoea dispersa*, *Staphylococcus chromogenes*, *Bacillus aquimaris*, *Staphylococcus arlettae*, *Staphylococcus sciuri*, *Staphylococcus warneri*, *Moraxella osloensis*, *Enterobacter* sp., *Klebsiella michiganensis*, *Staphylococcus hominis*, *Staphylococcus saprophyticus*, *Streptomyces* sp., *Bacillus niacin*, *Cedecea neteri*, *Micrococcus luteus*, *Lysinibacillus sphaericus*, and *Bacillus licheniformis*) were identified. All of these species belonged to three phyla, *Proteobacteria*, *Firmicutes*, and *Actinobacteria*, out of which phylum *Firmicutes* (71.1%) was the most prominent. The least number of species was recorded from *Actinobacteria*. The relative distribution of midgut microbes in different mosquito species differed significantly among mosquito species (Chi-square,  $\chi^2 = 486.091$ ;  $df = 36$ ;  $P \leq 0.001$ ). Midgut microbiota of *Cx. tritaeniorhynchus* and *Cx. gelidus* indicated a similarity of 21.51%, while *Ma. annulifera* shared a similarity of 6.92% with the cluster of above two species. The gut microbiota of *Cx. tritaeniorhynchus* was also significantly more diverse and more evenly distributed compared to *Ma. annulifera*. Simpson's diversity, Margalef's diversity, and Menhinick's diversity indices were higher in *Cx. gelidus*. Of the recorded species, *P. dispersa* and strains of nonpathogenic species in *Bacillaceae* family (*B. megaterium*, *B. niacini*, *B. licheniformis*, and *L. sphaericus*) can be recommended as potential candidates for paratransgenesis. **Conclusion.** The relative distribution of midgut microbes in different mosquito species differed significantly among the three studied adult mosquito species. The present data strongly encourage further investigations to explore the potential usage of these microbes through paratransgenic approach for novel eco-friendly vector control strategies.

## 1. Background

Mosquitoes transmit a wide range of pathogens that cause diseases in humans and other animals. All mosquito species are aquatic during the immature stages [1]. Studies conducted in the early 1900s noted that larval and adult stages of mosquitoes harbor extracellular microbes in their digestive tract [2, 3]. During the last 10 years, their roles for mosquito biology have been broadly studied. Some findings emphasize that they play important roles in immunity, food digestion, fertility, and fecundity, which ultimately affect larval growth, adult fitness, vector populations, and disease prevalence [4].

Besides, these studies evidenced positive and negative effects of these gut microbial communities on vector competency through host-parasite interaction [5, 6], thereby significantly influencing disease transmission potential [7, 8]. Moreover, mosquito-midgut bacteria are used as vehicles to express molecules in the gut and suppress parasite colonization [9, 10]. This technique is known as paratransgenesis, which emerged as a novel vector controlling strategy. Here, the symbiotic bacteria associated with vectors may genetically modify for the expression of an effector molecule and reintroduced through food to the mosquito midgut.

The identification of suitable midgut bacteria is the fundamental requirement in the paratransgenic techniques. These identified organisms should be able to colonize in the midgut of an interested disease vector and should be transferred to the next generation through transstadial passage [11]. The number of recent studies has used culture-dependent and culture-independent approaches to characterize the microbial communities in the midgut of different mosquito species including *Aedes*, *Culex*, *Anopheles*, *Coquillettidia*, and *Mansonia* [12–17]. However, little is known about the midgut microflora of *Culex* mosquitoes, and very few studies have been conducted on midgut microbiota [11, 18, 19]. In addition, no studies have been carried out to screen the midgut bacteria in *Mansonia annulifera* mosquitoes.

The mosquito *Culex tritaeniorhynchus* is the major vector of Japanese encephalitis (JE) virus throughout Asia and the Pacific islands including Sri Lanka [20–23]. The JE has occurred in Sri Lanka for decades, seriously threatening human life since 1980 [21]. Gampaha was one of the districts that had reported a relatively large number of JE cases in the past [24]. The success of the immunization against JE in Sri Lanka is reflected in the fact that since 1988, the incidence of JE has decreased drastically with the increased coverage of immunization. However, the disease was spreading to new areas with previously low levels of enzootic transmission and report occasional outbreaks in some districts in Sri Lanka [25].

On the other hand, *Ma. annulifera* is a vector of brugian filariasis and dirofilariasis in Sri Lanka [26]. Until recently, brugian filariasis was considered an infection of the past as cases were not reported in Sri Lanka since the late 1960s. However, surveillance activities carried out to evaluate the Programme for Elimination of Lymphatic Filariasis (PELF) in Sri Lanka revealed the sporadic occurrence of brugian filariasis [27, 28] of probable zoonotic origin [29] in endemic

areas. In 2016, Sri Lanka was certified to have eliminated lymphatic filariasis as a public health problem by the World Health Organization [30]. However, the occurrence of zoonotic brugian filariasis raises an issue as mass treatment of the human population with antifilarial drugs would not eliminate the reservoir of infection nor the vector population. Human dirofilariasis is another zoonotic filarial infection, commonly encountered in the country [31–33].

The abundance of disease-transmitting vectors due to limited vector control activities and higher receptivity in these areas have been identified as major challenges to maintain the disease-free status in the country, for JE and filariasis. Therefore, alternative vector control methods are needed as additional tools to be incorporated into the integrated vector management [32]. Paratransgenesis could be a better control tool for vector management. Hence, the present study was conducted to determine the microflora in the midgut of wild-caught *Cx. tritaeniorhynchus*, *Cx. gelidus*, and *Ma. annulifera* which has been limitedly assessed world-wide.

## 2. Methods

**2.1. Selection of Study Area and Sampling Sites.** The study was conducted in the district of Gampaha which is endemic for lymphatic filariasis, dirofilariasis, and JE diseases in Sri Lanka. This district is the second most populous district in Sri Lanka with urban, semiurban, and rural populations in a land area of 1387 km<sup>2</sup> [34]. Sampling sites were selected from Kelaniya (06° 58.426' N, 79° 54.939' E) and Gampaha (07° 06.448' N, 79° 53.049' E) Medical Officer of Health (MOH) areas to carry out mosquito collections (Figure 1). The selection of other sample sites was carried out in consultation with the Regional Entomologist, Gampaha District, based on the vector abundance and potential risk for filariasis with reference to the past prevalence rates of microfilaraemia/antigenaemia.

**2.2. Field Collection of Adult Mosquitoes.** Entomological surveys were conducted at each selected location from January to October 2019 using cattle-baited net trap (CBNT) collections according to the standard guidelines described by the World Health Organization [35]. The cattle-baited net traps were set up at each selected site in the evening (17:00–18:00 h). The trap was made of a white cotton drill (3 m × 3 m × 1.5 m) with net windows (2 m × 1 m) on sides erected using a strong center pole of two-meter height and four side sticks each of the same height (1.5 m). A distance of 15–25 cm gap was allowed between the lower edge of the net and the ground, enabling the mosquitoes to enter the trap. The cattle was introduced into the trap just before the sunset and tethered to the pole fixed to the mid of the trap. The mosquitoes were collected from 21:00–23:00 in the late-night using battery-operated aspirators. Collected adult mosquitoes were transferred safely to the presterilized adult rearing cages. All collected adult mosquito samples were transported and housed at the insectary facility at the Department of Parasitology, Faculty of Medicine, University of Kelaniya, Ragama, Sri Lanka, until taken for further experiments.

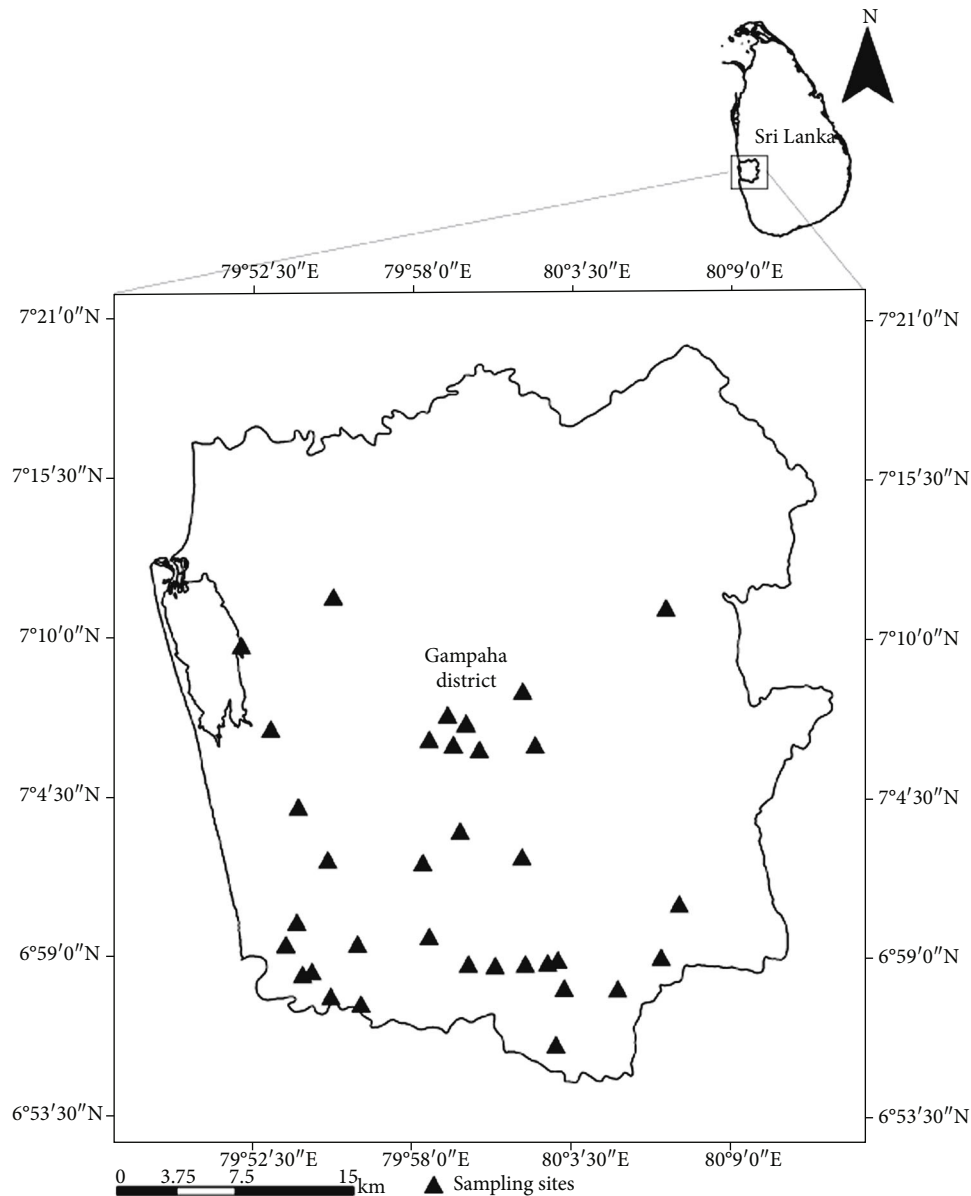


FIGURE 1: The GIS map showing the geographical distribution of mosquito sampling sites in Gampaha District, Sri Lanka.

**2.3. Processing and Identification of Field-Caught Adults.** Unfed (non-blood-fed) live mosquitoes captured from the field that housed at the insectary were sorted within 6-8 hours after collection. The mosquitoes were sacrificed using a clod shock. Mosquito species, namely, *Culex tritaeniorhynchus*, *Culex gelidus*, and *Mansonia annulifera*, were segregated based on key morphological characteristics [36–38]. Mosquito adults were then surface sterilized individually for 30 seconds by tapping in a microcentrifuge tube containing 250  $\mu$ L of 70% ethanol followed by rinsing twice with 250  $\mu$ L of phosphate-buffer saline (PBS). The final discard of washing was used for bacterial analysis to make sure that there was no bacterial contamination from the mosquito surface. For isolation of the midgut bacterial population, the midguts of non-blood-fed adult female mosquitoes were dissected under sterile conditions. Blood or sugar feeding was

not performed during the rearing process before recruitment for midgut analysis, to avoid the potential alterations in the midgut microflora.

Individually dissected midguts of five adults were pooled and transferred to a 1.5 mL microcentrifuge tube containing 250  $\mu$ L of PBS and homogenized with a sterile micropipette. The homogenized lysate was serially diluted in PBS (900  $\mu$ L) to prepare a dilution series from  $10^0$ - $10^{-7}$ . From each mosquito species, a minimum of 250 adults was processed separately to screen for gut flora.

**2.4. Culturing and Isolation of Bacteria.** A volume of 100  $\mu$ L from each dilution was plated on sterile Plate Count Agar (PCA) media and incubated at 35°C for 24-48 hours. To assess microbial growth, the total number of Colony-Forming Units (CFUs) was determined. Bacterial colonies

obtained on plates were differentiated morphologically on colony shape, size, color, margin, opacity, elevation, etc. Morphologically distinct colonies were selected from primary plates and subcultured on nutrient agar plates until a pure colony was obtained. All bacterial isolates were identified up to the genus level by Gram-staining and biochemical testing performed according to Cowan and Steel's manual [39]. Finally, the bacterial isolates differentiated up to the genus level by Gram-staining and biochemical testing were selected further for molecular characterization.

**2.5. Genomic DNA Extractions and Sequencing.** Genomic DNA was isolated from each pure culture using QIAamp DNA mini kit (QIAGEN GmbH, Hilden, Germany) according to the manufacturer's instructions. The PCR amplifications were performed using universal primers 27F (5' AGAGTTTGATCCTGGCTCAG 3') and 1492R (5' TACG GCTACCTTGTTACGACTT 3') [40] targeting 16S rRNA gene sequences. Polymerase Chain Reaction (PCR) was performed with a reaction mixture containing 1x PCR buffer (Invitrogen), 0.5  $\mu$ M of each primer, 2.5 mM MgCl<sub>2</sub>, 200 ng of purified DNA, 0.2 mM dNTPs, and 0.3 units of *Taq* polymerase (Invitrogen). The total volume was adjusted to 25  $\mu$ L. The PCA media and ddH<sub>2</sub>O were used as negative controls. Samples were amplified according to the following cycle: initial denaturation at 94°C for 10 min, followed by 35 cycles of denaturation at 94°C for 30 s, annealing at 55°C for 30 s, and extension at 72°C for 1 min. The final extension was at 72°C for 8 min.

The amplified product was visualized on a 1% agarose gel containing ethidium bromide using a UV transilluminator. The PCR amplicons were then purified using the QIAquick PCR Purification Kit (Qiagen). The purified products were sent to Macrogen, South Korea (Macrogen Inc., 1001, 254 Beotkkot-ro, Geumcheon-gu, Seoul, Republic of Korea) for 16S ribosomal RNA partial gene sequencing by the Sanger method.

**2.6. Phylogenetic Analysis of Midgut Bacteria Isolated from Mosquito species.** Homologous sequences were searched in the GenBank database using BLAST [41]. The isolates were identified when their 16S rRNA gene sequences shared 97% homology with completed 16S rRNA gene sequences found in the GenBank database. The evolutionary history was inferred using the neighbor-joining method, and the evolutionary analyses were conducted in MEGA X. The evolutionary distances which were computed using the Tajima-Nei method were used to infer the phylogenetic tree.

A phylogenetic tree was also constructed for assessing the phylogenetic affiliations of bacterial isolates detected in the midguts of three selected mosquito species and with the usage of some reference sequences available in the literature [19]. Phylogenetic branch length distance (BLD) was estimated based on the maximum likelihood (ML) tree which is estimated on the multiple sequence alignment (MSA). The dissimilarity between a pair of sequences was defined as the total branch length distance on the ML tree between a pair of sequences.

**2.7. Data Analysis.** The percentage relative abundance of each mosquito species was calculated. The Excel spreadsheet tool [42] was used to calculate confidence intervals for the comparison of the presence of each bacterial species in mosquito species. The Chi-square test of independence was used to evaluate the significance in the distribution of different bacteria species in different mosquito species. Diversity indices such as the Simpson Index [43], Shannon Index [44], Margalef diversity index [45], Menhinick's diversity index [46], and evenness of bacterial communities [47] in each tested mosquito were calculated. All the interpretations were done considering 95% significant intervals.

### 3. Results

**3.1. Bacterial Isolates from the Midgut of Different Mosquito Species.** A total of 20 bacterial strains were identified from the midgut of three selected vector mosquitoes. The majority of the midgut isolates (71%; 13/20) were categorized under the phylum *Firmicutes*. The other seven species were from the *Proteobacteria* (18.7%;  $n = 5$ ) and *Actinobacteria* (10.19%;  $n = 2$ ) phyla. The list of bacteria species identified from the midgut lumens of each mosquito species is illustrated in Table 1.

**3.2. *Cx. tritaeniorhynchus*.** A total of eight different bacterial species, namely, *Staphylococcus pasteurii*, *Bacillus megaterium*, *Staphylococcus cohnii*, *Pantoea dispersa*, *Staphylococcus chromogenes*, *Bacillus aquimaris*, *Staphylococcus arlettae*, and *Staphylococcus sciuri*, were identified. According to the phylogenetic tree constructed by the neighbor-joining algorithm, a close relationship was observed between the species of two genera, *Staphylococcus* and *Bacillus*, while the relationship was distant for the genus *Pantoea* (Figure 2(a)).

**3.3. *Culex gelidus*.** Overall, a total of nine bacterial species were isolated (*Staphylococcus sciuri*, *Staphylococcus warneri*, *Moraxella osloensis*, *Enterobacter* sp., *Klebsiella michiganensis*, *Staphylococcus hominis*, *Bacillus megaterium*, *Staphylococcus saprophyticus*, and *Streptomyces* sp.). These were different from the isolates observed from other two mosquito species in this study. It inferred relatively a distant phylogenetic relationship to *K. michiganensis*, *Enterobacter* sp., and *M. osloensis* (Figure 2(b)).

**3.4. *Mansonia annulifera*.** The lowest bacterial diversity was observed from *Ma. annulifera* consisting of only *Bacillus niacin*, *Cedecea neteri*, *Micrococcus luteus*, *Staphylococcus chromogenes*, *Lysinibacillus sphaericus*, and *Bacillus licheniformis*. However, of the six bacteria species identified, five of them were detected only from *Ma. annulifera*, while *S. chromogenes* was observed from *Cx. tritaeniorhynchus* (Table 1). It seems that there is a considerable difference between the gut bacteria in *Ma. annulifera* compared to the other two mosquito species investigated. *Bacillus niacini* and *B. licheniformis* shared a relatively close relationship, while *C. neteri* was having a distant phylogenetic relationship with the all other recorded species (Figure 2(c)).



TABLE 1: The occurrence of different bacterial species in *Culex tritaeniorhynchus*, *Culex gelidus*, and *Mansonia annulifera*.

| Phylum         | Family             | Bacterial species (Accession No.)                                 | Percentage occurrence (95% CI <sup>b</sup> ) |                        |                        |
|----------------|--------------------|---|--|------------------------|------------------------|
|                |                    |   | <i>Cx. tritaeniorhynchus</i>                 | <i>Cx. gelidus</i>     | <i>Ma. annulifera</i>  |
| Proteobacteria | Moraxellaceae      | <i>Moraxella osloensis</i><br>(MT072195)                          | ND   | 5.56<br>(0.025-0.119)  | ND                     |
|                |                    | <i>Klebsiella michiganensis</i><br>(MT072197)                     | ND   | 18.06<br>(0.117-0.267) | ND                     |
|                | Enterobacteriaceae | <i>Enterobacter</i> sp.<br>(MT072198)                             | ND   | 11.11<br>(0.063-0.187) | ND                     |
|                |                    | <i>Cedecea neteri</i><br>(MT072184)                               | ND   | ND                     | 21.05<br>(0.142-0.3)   |
|                | Erwiniaceae        | <i>Pantoea dispersa</i><br>(MT072165/MT072166)                    | 10.44<br>(0.058-0.179)                       | ND                     | ND                     |
|                |                    |   |  |                        |                        |
| Firmicutes     | Staphylococcaceae  | <i>Staphylococcus sciuri</i><br>(MT072175/MT072194)               | 6.71<br>(0.032-0.133)                        | 8.33<br>(0.043-0.154)  | ND                     |
|                |                    | <i>Staphylococcus warneri</i><br>(MT072204/MT072196)              | ND   | 5.56<br>(0.025-0.119)  | ND                     |
|                |                    | <i>Staphylococcus hominis</i><br>(MT072199)                       | ND   | 10.42<br>(0.058-0.179) | ND                     |
|                |                    | <i>Staphylococcus saprophyticus</i><br>(MT072203)                 | ND   | 9.72<br>(0.053-0.171)  | ND                     |
|                |                    | <i>Staphylococcus chromogenes</i><br>(MT072169/MT072170/MT072191) | 16.42<br>(0.104-0.248)                       | ND                     | 11.84<br>(0.069-0.196) |
|                |                    | <i>Staphylococcus pasteurii</i><br>(MT072161)                     | 20.89<br>(0.141-0.298)                       | ND                     | ND                     |
|                |                    | <i>Staphylococcus cohnii</i><br>(MT072164/MT072172)               | 8.95<br>(0.047-0.161)                        | ND                     | ND                     |
|                |                    | <i>Staphylococcus arlettae</i><br>(MT072171/MT072173)             | 13.43<br>(0.081-0.214)                       | ND                     | ND                     |
|                |                    | <i>Bacillus megaterium</i><br>(MT072163/MT072200)                 | 12.69<br>(0.075-0.206)                       | 20.83<br>(0.140-0.298) | ND                     |
|                |                    | <i>Bacillus niacini</i><br>(MT072182)                             | ND   | ND                     | 15.78<br>(0.099-0.242) |
|                | Bacillaceae        | <i>Bacillus aquimaris</i><br>(MT072168)                           | 10.45<br>(0.058-0.179)                       | ND                     | ND                     |
|                |                    | <i>Bacillus licheniformis</i><br>(MT072189)                       | ND   | ND                     | 22.36<br>(0.153-0.315) |
|                |                    | <i>Lysinibacillus sphaericus</i><br>(MT072262)                    | ND   | ND                     | 1.32<br>(0.003-0.059)  |
| Actinobacteria | Actinomycetaceae   | <i>Streptomyces</i> sp.<br>(MT072202)                             | ND   | 10.42<br>(0.058-0.179) | ND                     |
|                | Micrococcaceae     | <i>Micrococcus luteus</i><br>(MT072186)                           | ND   | ND                     | 28.00<br>(0.198-0.371) |

<sup>a</sup>All bacterial species were identified based on a % identity higher than 99% CI. <sup>b</sup>Confidence interval; ND: not detected.

**3.5. Joint Phylogenetic Affiliation of the Midgut Bacterial Isolates in Three Selected Species.** The ML trees estimated from gut microbial sequence alignments showed *Moraxella osloensis* on a long branch and seemed to cluster separately from the majority of the remaining sequences belonging to phylum *Proteobacteria* (Figure 3). Phylum *Firmicutes*, on the other hand, was very compact and contained very short branches. Family *Staphylococcaceae* showed the most variation, with several distinct clusters of sequences. The genetic distance between same gut microbial species (*Pantoea dis-*

*persa*, *Staphylococcus chromogenes*, and *Staphylococcus hominis*) differed between host mosquito species in which they were isolated (*Cx. tritaeniorhynchus* and *Ma. annulifera*) (e.g., 0.347 for *Pantoea dispersa*) (Figure 3).

Genetic distance between some gut microbial strains varied even within the same host mosquito species in which they were isolated (e.g., *Staphylococcus arlettae* isolated from *Cx. tritaeniorhynchus*) (Figure 3). No significant difference was observed for the genetic distances of the same gut microbial strain isolated from the same mosquito species (within-

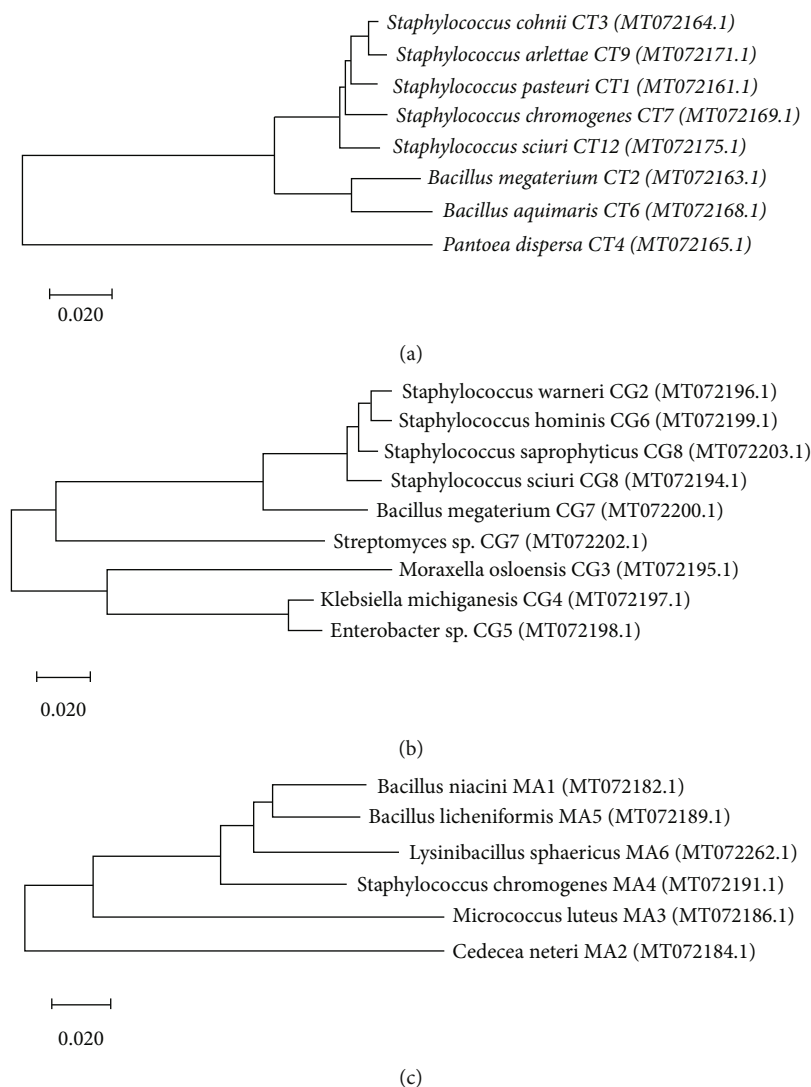


FIGURE 2: Dendrogram showing phylogenetic affiliation of bacterial isolates in the midgut of (a) *Cx. tritaeniorhynchus*, (b) *Cx. gelidus*, and (c) *Ma. annulifera*.

group) or different mosquito species (between groups) ( $P < 0.001$ ), and the genetic distance was not affected by the collection site of the host mosquito. The difference between Sri Lankan isolates from the reference sequences was also not significantly different with cutoffs at  $P \leq 0.05$ .

**3.6. Diversity of Mosquito Microbiota.** The Shannon diversity indices revealed that the gut microbiota of *Cx. gelidus* was the highest diverse followed by *Cx. tritaeniorhynchus*. Simpson's diversity, Margalef's diversity, and Menhinick's diversity indices also advocated the above observation. The diversity indices scored by each mosquito species are given in Table 2.

**3.7. Variation in Midgut Bacterial Communities across Mosquito Species.** The midgut bacteria analysis among the three mosquito species studies were found to be quite similar. The most abundant midgut microbial species were associated with phylum *Firmicutes* with 87.5%, 55.6%, and 66.7% in *Cx.*

*tritaeniorhynchus*, *Cx. gelidus*, and *Ma. annulifera*, respectively.

Other observed phyla found in all field-derived strains of mosquitoes were *Proteobacteria* (12.5%, 33.3%, and 16.7%, respectively, for the three above mosquito species) and *Actinobacteria* (0%, 11.1%, and 16.7%, respectively).

The relative abundance of families of gut microbial species present within three different mosquito species revealed that *Staphylococcaceae* is the most abundant gut microbial family, with 62.5%, 44.4%, and 20.0%, respectively, for three mosquito species followed by family *Bacillaceae* (25.0%, 11.1%, and 60.0%, respectively) and *Enterobacteriaceae* (0%, 22.2%, and 0%, respectively) (Figure 4).

Chi-square analysis revealed that the relative distribution of midgut microbes in different mosquito species differed significantly among three selected mosquito species ( $\chi^2 = 486.091$ ;  $df = 36$ ;  $P \leq 0.001$ ). Midgut microbiota in *Cx. tritaeniorhynchus* and *Cx. gelidus* indicated a similarity

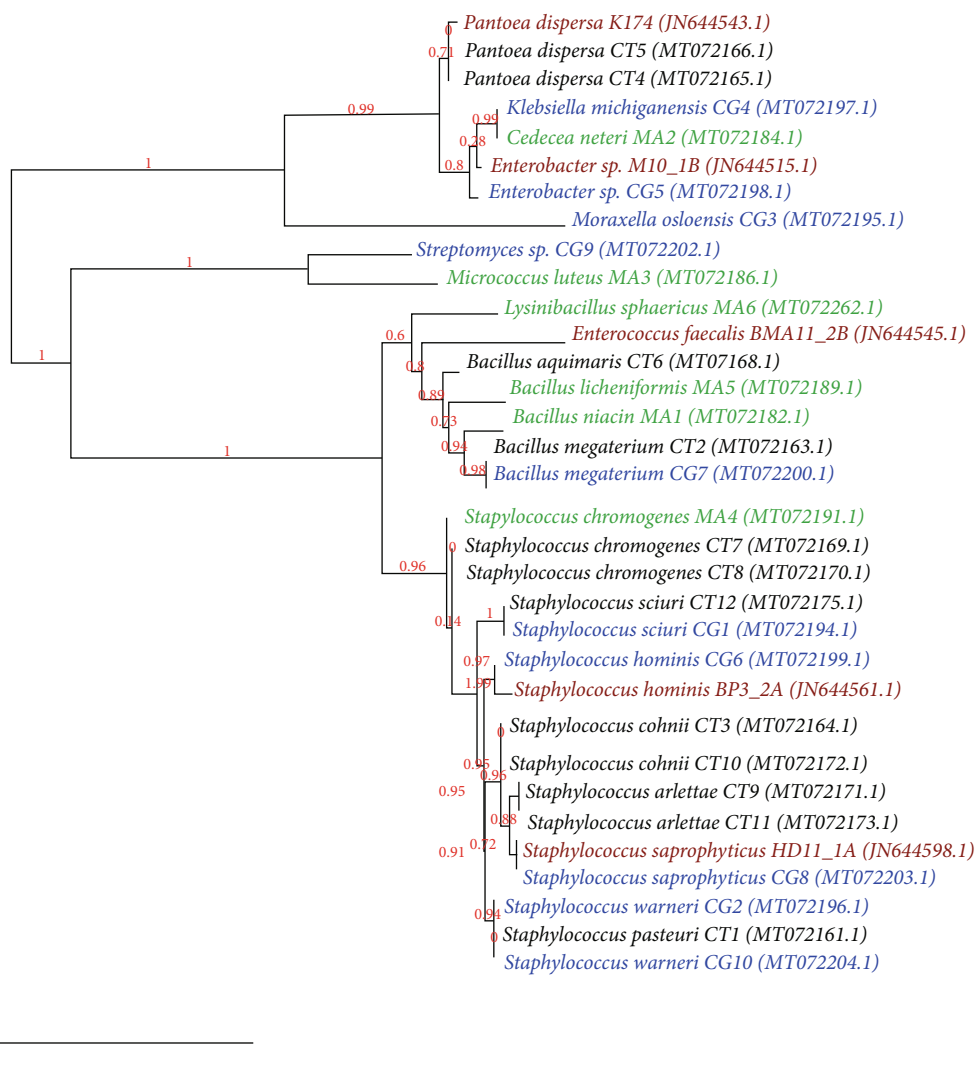


FIGURE 3: Phylogram showing the phylogenetic affiliation of bacterial isolates in the midgut of the three selected mosquito species (*Culex tritaeniorhynchus*, *Culex gelidus*, and *Mansonia annulifera*) along with reference sequences of some mid gut bacterial isolated from *Cx. quinquefasciatus* mosquito collected from India (Chandel et al., 2013).

TABLE 2: Diversity indices and total taxa of midgut bacterial isolates of *Culex tritaeniorhynchus*, *Culex gelidus*, and *Mansonia annulifera*.

| Criteria                    | Mosquito species             |                    |                       |
|-----------------------------|------------------------------|--------------------|-----------------------|
|                             | <i>Cx. tritaeniorhynchus</i> | <i>Cx. gelidus</i> | <i>Ma. annulifera</i> |
| Total taxa identified       | 8                            | 9                  | 6                     |
| Dominance                   | 0.38                         | 0.41               | 0.21                  |
| Simpson's diversity index   | 0.86                         | 0.87               | 0.80                  |
| Shannon's diversity index   | 17.08                        | 20.60              | 8.25                  |
| Evenness                    | 5.80                         | 7.00               | 2.80                  |
| Margalef's diversity index  | 3.29                         | 3.71               | 2.13                  |
| Menhinick's diversity index | 0.69                         | 0.75               | 0.58                  |

of 21.51%, while *Ma. annulifera* shared a similarity of 6.74% with the cluster of above two species.

The midgut microbe assemblage among three mosquito species formed subclusters based on the Bray Curtis similarity. The midgut microbiota in *Cx. tritaeniorhynchus* and *Cx.*

*gelidus* indicated similarity of 21.51%, while *Ma. annulifera* shared a similarity of 6.74% with the cluster of above two species. The dBRDA 1 and dBRDA 2 explained that 64.3% and 35.7% of the total variation of gut microbes in three mosquito species (Figure 5).

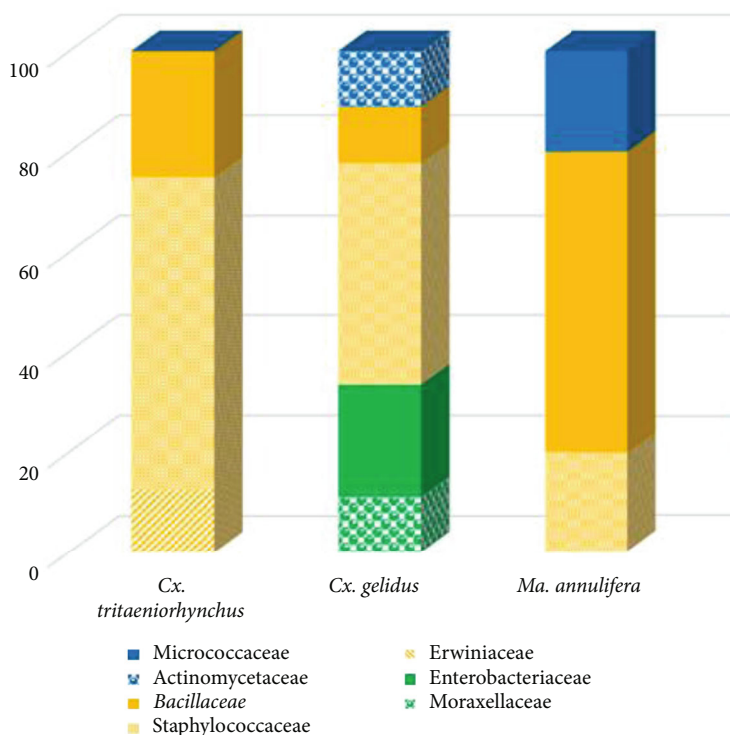


FIGURE 4: Relative abundances of bacterial families (7) of gut microbes present within three studied mosquito species collected at different sites by the 16S rRNA PCR product sequence analysis.

#### 4. Discussion

Microbial symbionts in the insects exist at different organs such as the gut, ovaries, Malpighian tubules, and hemocoel [48]. The microbiota in the gut is of precise interest because it is the first contact point between the parasites and epithelial surfaces [49]. The prevalence of insect gut microflora especially in mosquitoes has been investigated through classical culture-based methods or metagenomic based on 16S rRNA gene sequencing [50–52]. The bacterial composition of mosquitoes sampled from natural habitats is highly variable but often contain a core microbiome that is dominated by a small number of taxa. However, it may vary subject to the insect species, geographical origin, ecological niche, source of food, and gender [50, 51, 53].

This microbiota can influence the capacity of insects to transmit disease-causing pathogens through various mechanisms. Therefore, understanding the interaction between vectors with their microbiota and transmitting pathogens may be useful to limit disease transmissions by exploring the potential role of microbiota in modulating infections that could lead to alternative disease control strategies [53]. Paratransgenesis is one such approach that controls the transmission of pathogens by arthropod vectors using gut microflora. This approach consists of the use of genetically altered symbiotic bacteria that secrete effector molecules that kill the infectious agent, disturb the reproduction, make the vector less competent, or reduce the life span of the host. However, this needs screening of microbiota among locally available disease-transmitting vectors using culture-dependent methods since the availability of gut microflora may vary with the geographical location [19].

The studies have carried out to screen the midgut microflora in mosquito species under different genera including *Aedes*, *Anopheles*, *Culex*, and *Mansonia*. However, only a few studies have been conducted to evaluate the prevalence of gut microflora in *Cx. tritaeniorhynchus*, *Cx. gelidus*, and *Ma. annulifera*. A study conducted by Abalain-Colloc et al. [54] has reported the presence of one microbiota species, *Spiroplasma taiwanense*, from the midgut of field-collected *Cx. tritaeniorhynchus* in Taiwan. This is the only available information regarding the midgut microbes of this species, whereas available information is limited for *Cx. gelidus* and *Ma. annulifera*. Even after 31 years, only a few studies have contributed to compensate for the above limitation, which may be grossly inadequate to get an idea about the diversity of gut microflora of the importance of pursuing control strategies. Thus, the present study contributes to filling the knowledge gap regarding the prevalence of gut bacterial flora in the above mosquito species.

Examination of 16S ribosomal RNA amplicons from culturable microflora isolated from the midgut of adult female *Cx. quinquefasciatus* from India revealed the presence of 83 bacterial species under 31 bacterial genera [19]. All species identified were fit into three Phyla: *Proteobacteria* (37 species), *Firmicutes* (33 species), and *Actinobacteria* (13 species). Similarly, the present study has also isolated the midgut bacterial species that belonged to the same three phyla mentioned above. These three bacterial phyla have commonly been recorded in the gut of many insect species including mosquitoes [12, 14, 16].

However, the diversity of microbes in Phylum *Actinobacteria* was comparatively lower (only 2 species) compared to



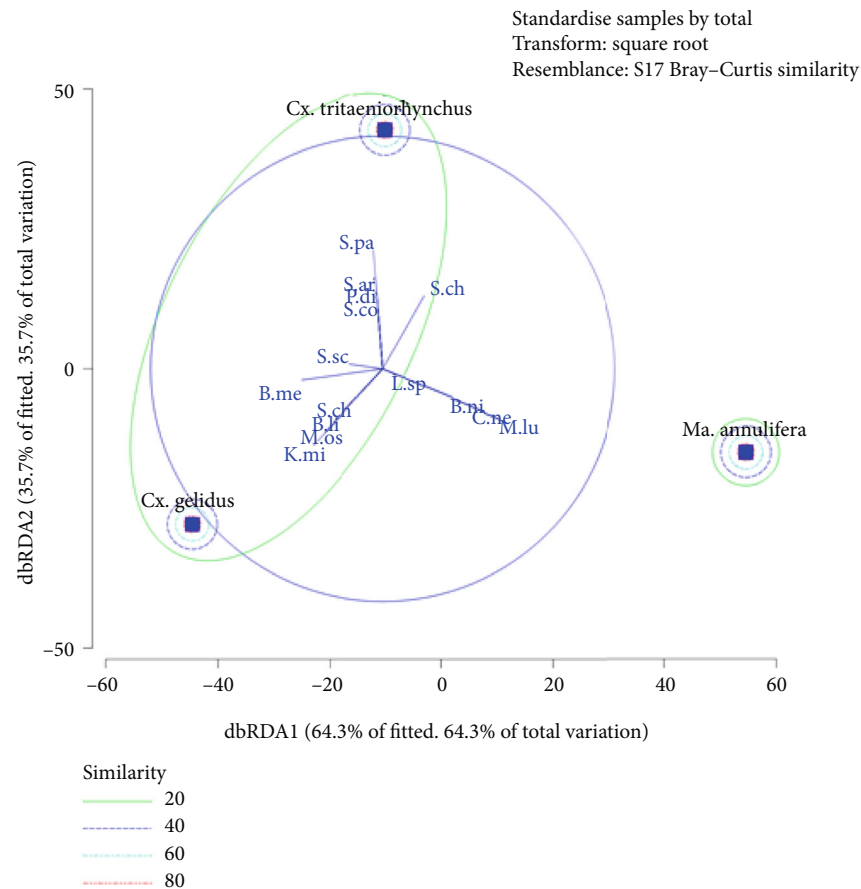


FIGURE 5: dbRDA plot for distribution of midgut microbiota in *Culex tritaeniorhynchus*, *Culex gelidus*, and *Mansonia annulifera*.

the other two Phyla (*Proteobacteria* and *Firmicutes*). About 66.4% and 23.14% of the total isolated bacteria in *Cx. tritaeniorhynchus* belonged to genera *Staphylococcus* and *Bacillus*, respectively, which represented around 89.54% of the total bacterial population. None of the species was detected under phylum *Actinobacteria* from *Cx. tritaeniorhynchus*. Some of the *Staphylococcus* species isolated from *Cx. tritaeniorhynchus* and *Cx. gelidus* during the present study, namely, *S. cohnii*, *S. pasteurii*, *S. saprophyticus*, *S. arlettae*, *S. hominis*, and *S. warneri*, had also been recorded previously among the wild-caught *Aedes albopictus* in India [55]. Therefore, these species may coexist among mosquito species at different geographical isolates, and there could be alterations among the gut microflora in mosquitoes.

The presence of different microbial species as midgut bacteria in three mosquito species could be recognized as the reason for the separation of two distinct clusters in the dbRDA analysis. However, *S. chromogenes* was found as a common bacterial species between *Cx. tritaeniorhynchus* and *Ma. annulifera*. In addition, *S. sciuri* and *B. megaterium* were common in both *Cx. tritaeniorhynchus* and *Cx. gelidus*. The occurrence of *C. neteri*, *B. niacin*, *L. sphaericus*, and *B. licheniformis* was limited only to *Ma. annulifera*. This could be identified as another possible reason for the separation of midgut bacteria in *Ma. annulifera* as a single cluster.

There are some evidence on the usage of gut microbacteria available in the insects for control approaches in the means of vector suppression, inhibition of parasite/pathogenic development in the invertebrate hosts, oviposition attractants in the development of lethal ovitraps, and transformation carriers in expressing molecules. Apart from mosquitoes, achievements were made from kissing bug gut inhabiting microbe, *Rhodococcus rhodnii*, for the prevention of the Chagas 4disease and sand fly gut inhabiting microbe, *Enterobacter cloacae*, for reducing *Leishmania* parasite transmission [56]. *Bacillus megaterium* and *B. licheniformis* recorded during the present study have been identified previously as candidates for paratransgenesis in *Phlebotomus papatasi* [57] while *S. saprophyticus* detected from *Cx. gelidus* is used as a strong oviposition inducer for gravid *P. papatasi* [58]. In addition, some *Enterobacter* and *Klebsiella* species have been previously documented on the ability to reduce the intensity and prevalence of *Plasmodium falciparum* infection [59]. Furthermore, *Klebsiella* sp. serves as an attractant for the selection of oviposition habitat, and it nourishes the most fragile larval stage, first instars to more resilient second instars [60].

Besides, *Enterobacter* sp., *Klebsiella michiganensis*, and *Pantoea dispersa* that were recorded during the present investigation have also been identified as potential candidates for paratransgenesis in *Aedes* mosquitoes [55]. *Lysinibacillus*

*sphaericus* isolated from *Ma. annulifera* has previously been isolated from *An. maculipennis* in Iran and revealed as a potential paratransgenic bacteria to control malaria transmission [61]. Therefore, exploring the feasibility of using these identified bacteria species for the control of different disease vectors in terms of their ability to inhibit the disease pathogen, as paratransgenesis candidates and oviposition attractant/inducer, is of paramount importance for a country like Sri Lanka.

The composition of gut microflora varies with the host defense and immune mechanisms in disease vectors. Recent investigations suggest the significance of microbiota in insects by sustaining its basal immune activity and immune priming [62–64]. Further, the microbiota can modulate the mosquito immune response and influence vector competence to human pathogens. For instance, elimination of the major midgut bacteria in *An. gambiae* and *Ae. aegypti* through antibiotics may increase the susceptibility to *P. falciparum* and dengue virus (DENV) infections, respectively [6, 65]. In addition, bacteria can also influence the mosquito competence by impairing the pathogenic infection through competition for resources or secretion of antipathogen molecules [66–68]. It has been noticed that during the chikungunya infection in *Aedes* mosquitoes, the abundance of bacteria under *Enterobacteriaceae* family is elevated [13]. Therefore, it is deduced that the changes in gut microflora may represent the occurrence of disease pathogens which may be used as reflective organisms to identify the presence of disease pathogens in the wild population of vectors. Further, this phenomenon can be used to suppress vector population through modulation of gut microflora by introducing certain field-derived bacteria into the mosquito gut through an artificial feeding which may increase mosquito resistance to the pathogen [53].

Modification of bacteria species in recognition of pathogens through pattern recognition receptors (PRRs), immune pathway activation results in the systemic production of antimicrobial peptides (AMPs), and other antipathogen immune effectors, as well as stimulating immune defense mechanisms, is one of the applications [53]. *Asaia* sp. which is isolated from the midgut of *Ae. aegypti* was found to be expressing antipathogenic molecules within the mosquito [56]. The genetically modified *Asaia* can be successfully transformed into adults through sugar or blood meals and thereby can be vertically transmitted horizontally or cofeeding [69]. The Anti-Plasmodium effect or molecules can be displayed by genetically modified *Escherichia coli* in the gut of *An. stephensi* mosquitoes by producing effector molecules for inhibition of *P. berghei* development inside the mosquito [70].

*Pantoea dispersa* (phylum *Proteobacteria*) was identified from the midgut of adult *Cx. tritaeniorhynchus* during the present study with a 10.44% of relative abundance. A study has indicated that another symbiotic bacterium in the genus *Pantoea* (*Pantoea agglomerans*) could express and secrete the anti-Plasmodium effect or proteins (SM1, anti-Pbs21, and PLA2) [71]. Further, genus *Pantoea* is a possible candidate for paratransgenesis and is known for the vector competence of *Ae. albopictus* as well [51]. Moreover, genus *Pantoea*,

which was recorded from *Cx. tritaeniorhynchus*, exhibits the transstadial and horizontal transmission properties [72]. Therefore, in paratransgenesis application, the modified bacteria could be provided with the larval diet which can also be transmitted to the adult stage through transstadial and horizontal mechanisms. Therefore, the species in the genus *Pantoea* may be a suitable candidate for the paratransgenesis approach. However, the feasibility of the identified species from the present study for such intervention needs more investigations.

The present study suffers from some limitations. The composition of gut microbiota in different mosquito species varies as a result of complex interactions between environmental characteristics and developmental stages, physiological status, and feeding behaviour [73] which may influence the findings in the present study. Therefore, cattle-baited net traps were set up for mosquito collections with similar environmental settings such as closer to marshy lands, human habitation, and live-stock sheds to minimize the potential bias in the diversity of midgut bacteria that may be caused due to environmental variables. Host-related factors such as nutrition also influence microbiota composition [17]. For instance, blood meal induces an overall decrease of microbiota richness during the first 24 hours after the blood meal [74, 75]. However, the bacterial population increases substantially 48 h after a blood meal. Changes in the composition of the gut microbiota following a blood meal may be due to the oxidative stress associated with the catabolism of the blood meal [12, 76]. Further, some studies have emphasized that sugar feeding also influences the diversity of midgut microbiota in mosquitoes [77]. Therefore, midguts of non-blood-fed field-caught adult female mosquitoes were taken for dissections under sterile conditions for the isolation of the midgut bacterial population. Surface sterilized mosquitoes were taken for dissections immediately within 6-8 hours after field collections. Further, during the rearing of mosquitoes at the insectary, no artificial feeding of blood or sugar source was provided.

When using a culture-dependent approach, only 20% of environmental bacteria can be grown on a growth medium [78]. Thus, the composition of the microbiota is not a direct reflection of richness and abundance of the whole gut bacterial community which rises as a limitation of the culture-dependent microflora analysis. The development of future perspectives for paratransgenesis approach in Sri Lanka is interesting and necessary to examine the potential success of this method. To that end, *P. dispersa* can be proposed as the best candidate, and strains of non-pathogenic species in *Bacillaceae* family such as *B. megaterium*, *B. niacini*, *B. licheniformis*, and *Lysinibacillus sphaericus* were identified as other potential candidates for paratransgenesis. Therefore, while filling the lack of updated information on symbiotic bacteria in the midgut of *Cx. tritaeniorhynchus*, *Cx. gelidus*, and *Ma. annulifera* mosquitoes, the present data strongly encourages further investigations to explore the potential usage of these microbes through paratransgenic approach in consideration with ecological and other safety concerns.

## 5. Conclusion

This is the first attempt to explore the midgut microbiota of *Culex tritaeniorhynchus*, *Culex gelidus*, and *Mansonia annulifera* in Sri Lanka. Since there is only a few published information on the midgut microflora of *Culex* mosquitoes and the absence of any study for *Mansonia annulifera* mosquitoes, the present study provides fundamental information to the literature. The relative distribution of midgut microbes in different mosquito species differed significantly among the three studied mosquito species. The gut microbiota of *Cx. gelidus* was the highest diverse followed by *Cx. tritaeniorhynchus* and *Ma. annulifera*.

For paratransgenic applications, the feasibility of *P. dispersa* could be evaluated, and the suitability of nonpathogenic species in *Bacillaceae* family such as *B. megaterium*, *B. niacini*, *B. licheniformis*, and *Lysinibacillus sphaericus* should be further investigated. Therefore, the present data strongly encourage further investigations to explore the potential usage of these microbes through the paratransgenic approach which is a novel eco-friendly vector control strategy.

## Data Availability

The datasets supporting the conclusions of this article are included in the manuscript.

## Conflicts of Interest

The authors declare that they have no conflict of interests.

## Authors' Contributions

NG designed the research, supervised the research work overall, identified samples, performed molecular assays, and wrote the manuscript. KR conducted the microbial and molecular assays. DA supervised the research work and reviewed the manuscript. WR supervised the molecular assays and data analysis. HM did the field collection of mosquito samples and reviewed the manuscript. NC did the final editing and reviewed the manuscript. All authors read and approved the final manuscript.

## Acknowledgments

The laboratory experiments were at the insectary facility and molecular laboratory developed at the Department of Parasitology, Faculty of Medicine, University of Kelaniya, Ragama, Sri Lanka, from the National Research Council Funded Project NRC 16-142. Therefore, National Research Council, Sri Lanka (Grant No. 16-142) is greatly acknowledged for sharing facilities and consumables. The present research received funds for consumables and stipend for Research Assistant from the National Science Foundation, Sri Lanka (RG/2017/EB/02).

## References

- [1] A. N. Clements, "The biology of mosquitoes," in *Development, nutrition, and reproduction*, vol. 1, Chapman & Hall, New York, 1992.
- [2] E. H. Hinman, "A study of the food of mosquito larvae," *American Journal of Hygiene*, vol. 12, pp. 238–270, 1930.
- [3] J. Chao and G. A. Wistreich, "Microbial isolations from the mid-gut of *Culex tarsalis coquillet*," *Journal of Insect Pathology*, vol. 1, pp. 311–318, 1959.
- [4] R. S. Michael, "Composition and functional roles of the gut microbiota in mosquitoes," *Current Opinion in Insect Science*, vol. 28, pp. 59–65, 2018.
- [5] L. Gonzalez-Ceron, F. Santillan, M. H. Rodriguez, D. Mendez, and J. E. Hernandez-Avila, "Bacteria in midguts of field-collected *Anopheles albimanus* block *Plasmodium vivax* sporogonic development," *Journal of Medical Entomology*, vol. 40, no. 3, pp. 371–374, 2003.
- [6] Y. Dong, F. Manfredini, and G. Dimopoulos, "Implication of the mosquito midgut microbiota in the defense against malaria parasites," *PLoS Pathogens*, vol. 5, no. 5, article e1000423, 2009.
- [7] C. A. M. Finney, S. Kamhawi, and J. D. Wasmuth, "Does the arthropod microbiota impact the establishment of vector-borne diseases in mammalian hosts," *PLoS Pathogens*, vol. 11, no. 4, article e1004646, 2015.
- [8] K. K. Yadav, A. Naglot, K. Chandel et al., "Molecular characterization of midgut bacteria of *Aedes albopictus* exhibiting swarming motility property," *African Journal of Microbiology Research*, vol. 10, no. 25, pp. 949–959, 2016.
- [9] X. Ren, E. Hoiczky, and J. L. Rasgon, "Viral paratransgenesis in the malaria vector *Anopheles gambiae*," *PLoS Pathogens*, vol. 4, no. 8, article e1000135, 2008.
- [10] J. L. Rasgon, "Using infections to fight infections: Paratransgenic fungi can block malaria transmission in mosquitoes," *Future Microbiology*, vol. 6, no. 8, pp. 851–853, 2011.
- [11] V. J. Pidiyar, K. Jangid, M. S. Patole, and Y. S. Shouche, "Studies on cultured and uncultured microbiota of wild *Culex quinquefasciatus* mosquito midgut based on 16s ribosomal RNA gene analysis," *The American Journal of Tropical Medicine and Hygiene*, vol. 70, no. 6, pp. 597–603, 2004.
- [12] Y. Wang, T. M. Gilbreath, P. Kukutla, G. Yan, and J. Xu, "Dynamic gut microbiome across life history of the malaria mosquito *Anopheles gambiae* in Kenya," *PLoS One*, vol. 6, no. 9, article e24767, 2011.
- [13] K. Zouache, F. N. Raharimalala, V. Raquin et al., "Bacterial diversity of field-caught mosquitoes, *Aedes albopictus* and *Aedes aegypti*, from different geographic regions of Madagascar," *FEMS Microbiology Ecology*, vol. 75, pp. 377–389, 2011.
- [14] A. Boissière, M. T. Tchioffo, D. Bachar et al., "Midgut microbiota of the malaria mosquito vector *Anopheles gambiae* and interactions with *Plasmodium falciparum* infection," *PLoS Pathogens*, vol. 8, no. 5, article e1002742, 2012.
- [15] A. R. Chavshin, M. A. Oshaghi, H. Vatandoost et al., "Identification of bacterial microflora in the midgut of the larvae and adult of wild caught *Anopheles stephensi*: a step toward finding suitable paratransgenesis candidates," *Acta Tropica*, vol. 121, pp. 129–134, 2011.
- [16] J. Osei-Poku, C. M. Mbogo, W. J. Palmer, and F. M. Jiggins, "Deep sequencing reveals extensive variation in the gut microbiota of wild mosquitoes from Kenya," *Molecular Ecology*, vol. 21, no. 20, pp. 5138–5150, 2012.

- [17] G. Minard, P. Mavingui, and C. V. Moro, "Diversity and function of bacterial microbiota in the mosquito holobiont," *Parasites & Vectors*, vol. 6, no. 1, p. 146, 2013.
- [18] J. Demaio, C. B. Pumpuni, M. Kent, and J. C. Beier, "The midgut bacterial flora of wild *Aedes triseriatus*, *Culex pipiens*, and *Psorophora columbiae* mosquitoes," *The American Journal of Tropical Medicine and Hygiene*, vol. 54, no. 2, pp. 219–223, 1996.
- [19] K. Chandel, M. J. Mendki, R. Y. Parikh et al., "Midgut microbial community of *Culex quinquefasciatus* mosquito populations from India," *PLoS One*, vol. 8, no. 11, article e80453, 2013.
- [20] L. A. Lacey and C. M. Lacey, "The medical importance of rice land mosquitoes and their control using alternatives to chemical insecticides," *Journal of the American Mosquito Control Association*, vol. 6, pp. 1–93, 1990.
- [21] S. H. P. P. Karunaratne and J. Hemingway, "Insecticide resistance spectra and resistance mechanisms in populations of Japanese encephalitis vector mosquitoes, *Culex tritaeniorhynchus* and *Cx. gelidus* in Sri Lanka," *Medical and Veterinary Entomology*, vol. 14, pp. 430–436, 2000.
- [22] J. S. M. Peiris, F. P. Amerasinghe, P. H. Amerasinghe, C. B. Ratanayake, S. H. P. P. Karunaratne, and T. F. Tsai, "Japanese encephalitis in Sri Lanka- the study of an epidemic: vector incrimination, porcine infection and human disease," *Transactions of the Royal Society of Tropical Medicine and Hygiene*, vol. 86, no. 3, pp. 307–313, 1992.
- [23] B. T. Robert and S. Tom, "Japanese encephalitis, West Nile, and other Flavivirus infections," in *Tropical Infectious Diseases*, pp. 511–514, Elsevier, 2011.
- [24] H. Gunasekera, C. P. Senanayake, L. Mendis, and N. P. S. Chandra, "Japanese encephalitis virus infection in an endemic area: hospital based study 1998 to 2000," *Sri Lankan Journal of Infectious Diseases*, vol. 1, no. 2, pp. 19–27, 2012.
- [25] Epidemiology Unit Ministry of Health, "Sri Lanka Japanese Encephalitis, A manual for Medical Officers of Health," 2020, <http://www.epid.gov.lk/web/attachments/article/141/JE%20book.pdf>.
- [26] S. Ramalingam, "Vectors of *Wuchereria bancrofti* and *Brugia malayi* in South and Southeast Asian region; their distribution and biology and control," *Ceylon Journal of Medical Science*, vol. 24, pp. 1–27, 1975.
- [27] P. E. Fisher, T. Liyanage, R. U. Rao, and G. J. Weil, "Molecular characterization of re-emergent *Brugia malayi* in Sri Lanka," *57th Annual Meeting, American Society of Tropical Medicine and Hygiene*, vol. 79, no. 6, p. 433, 2008.
- [28] T. G. A. N. Chandrasena, R. Premaratna, D. S. Samarasekera, and N. R. de Silva, "Surveillance for transmission of lymphatic filariasis in Colombo and Gampaha districts of Sri Lanka following mass drug administration," *Transactions of the Royal Society of Tropical Medicine and Hygiene*, vol. 5, pp. 620–622, 2016.
- [29] C. H. Mallawarachchi, T. G. A. N. Chandrasena, R. Premaratna, S. M. N. S. M. Mallawarachchi, and N. R. de Silva, "Human infection with sub-periodic *Brugia* spp. in Gampaha District, Sri Lanka: a threat to filariasis elimination status," *Parasites & Vectors*, vol. 11, no. 1, p. 68, 2018.
- [30] WHO Sri Lanka, Annual Report, "About WHO South-East Asia," 2016, [https://www.searo.who.int/srilanka/documents/who\\_srl\\_annual\\_report\\_2016.pdf](https://www.searo.who.int/srilanka/documents/who_srl_annual_report_2016.pdf).
- [31] D. Iddawela, K. Ehambaram, and S. Wickramasinghe, "Human ocular dirofilariasis due to *Dirofilaria repens* in Sri Lanka," *Asian Pacific Journal of Tropical Medicine*, vol. 8, no. 12, pp. 1022–1026, 2015.
- [32] N. Gunathilaka, S. Siriwardana, L. Wijesooriya, G. Gunaratne, and N. Perera, "Subcutaneous dirofilariasis caused by *Dirofilaria* (Nochtiella) *repens* in Sri Lanka: a potential risk of transmitting human dirofilariasis," *SAGE Open Medical Case Reports*, vol. 5, pp. 1–4, 2017.
- [33] T. G. A. N. Chandrasena, R. Premaratna, C. H. Mallawarachchi et al., "The diversity of human dirofilariasis in Western Sri Lanka," *BioMed Research International*, vol. 2019, Article ID 9209240, 7 pages, 2019.
- [34] Department of Census and Statistics, "District Statistical Handbook," *Department of Census and Statistics*, 2018, <http://www.statistics.gov.lk/DistrictStatHBook.asp>.
- [35] World Health Organization (WHO), *Entomological Field Techniques for Malaria Control Part 1 Learner's Guide*, World Health Organization (WHO), Geneva, Switzerland, 1992.
- [36] F. P. Amerasinghe, "Illustrated keys to the genera of mosquitoes (Diptera: Culicidae) in Sri Lanka," *Journal of National Science Council of Sri Lanka*, vol. 23, no. 4, pp. 183–211, 1995.
- [37] R. V. Chelliah, "Keys and illustrations to the genera of mosquitoes of Sri Lanka (Diptera: Culicidae)," *Contributions of the American Entomological Institute*, vol. 7, no. 4, pp. 1–84, 1984.
- [38] R. Rattanarithkul, B. A. Harrison, P. Panthusiri, and R. E. Coleman, "Illustrated keys to the mosquitoes of Thailand 1, background; geographic distribution; list of genera, subgenera and species; and a key to the genera," *The Southeast Asian Journal of Tropical Medicine and Public Health*, vol. 36, no. 1, pp. 1–80, 2005.
- [39] S. T. Cowan and K. J. Steel, "Diagnostic tables for the common medical bacteria," *Journal of Hygiene*, vol. 59, no. 3, pp. 357–373, 1961.
- [40] S. Srivastava and V. Singh, "Identification of regulatory elements in 16S rRNA gene of *Acinetobacter* species isolated from water sample," *Bioinformation*, vol. 3, no. 4, pp. 173–176, 2008.
- [41] National Center for Biotechnology Information, "Basic Local Alignment Search Tool," 2020, <https://www.ncbi.nlm.nih.gov/BLAST>.
- [42] R. Herbert, "Confidence Interval Calculator," 2013, 2020, <https://www.pedro.org.au/english/downloads/confidence-interval-calculator/>.
- [43] E. H. Simpson, "Measurement of diversity," *Nature*, vol. 163, no. 4148, p. 688, 1949.
- [44] C. E. Shannon and W. Weaver, *The mathematical theory of communication*, The University of Illinois Press, Urbana, 1949.
- [45] R. Margalef, "Information theory in ecology," *General Systems*, vol. 3, pp. 36–71, 1958.
- [46] R. H. Whittaker, "Evolution of species diversity in land communities," in *Evolutionary Biology*, vol. 10, pp. 1–67, Springer, 1977, cited in Magurran AE. *Measuring biological diversity*, Blackwell Publishing: Oxford, UK, 2004, p. 256.
- [47] E. C. Pielou, "The measurement of diversity in different types of biological collections," *Journal of Theoretical Biology*, vol. 13, pp. 131–144, 1966.
- [48] I. Eleftherianos, J. Atri, J. Accetta, and J. C. Castillo, "Endosymbiotic bacteria in insects: guardians of the immune system," *Frontiers in Physiology*, vol. 4, p. 46, 2013.
- [49] P. Azambuja, E. S. Garcia, and N. A. Ratcliffe, "Gut microbiota and parasite transmission by insect vectors," *Trends in Parasitology*, vol. 21, no. 12, pp. 568–572, 2005.



- [50] O. Terenius, J. M. Lindh, K. Eriksson-Gonzales et al., "Midgut bacterial dynamics in *Aedes aegypti*," *FEMS Microbiology Ecology*, vol. 80, no. 3, pp. 556–565, 2012.
- [51] C. Valiente Moro, F. H. Tran, F. N. Raharimalala, P. Ravelonandro, and P. Mavingui, "Diversity of culturable bacteria including *Pantoea* in wild mosquito *Aedes albopictus*," *BMC Microbiology*, vol. 13, no. 1, p. 70, 2013.
- [52] B. Chouaia, P. Rossi, M. Montagna et al., "Molecular evidence for multiple infections as revealed by typing of *Asaia* bacterial symbionts of four mosquito species," *Applied and Environmental Microbiology*, vol. 76, no. 22, pp. 7444–7450, 2010.
- [53] N. J. Dennison, N. Jupatanakul, and G. Dimopoulos, "The mosquito microbiota influences vector competence for human pathogens," *Current Opinion in Insect Science*, vol. 3, pp. 6–13, 2014.
- [54] M. L. Abalain-Colloc, L. Rosen, J. G. Tully, J. M. Bove, C. Chastel, and D. L. Williamson, "*Spiroplasma taiwanense* sp. nov. from *Culex tritaeniorhynchus* mosquitoes collected in Taiwan," *International Journal of Systematic Bacteriology*, vol. 38, pp. 103–107, 1988.
- [55] K. K. Yadav, S. Datta, A. Naglot et al., "Diversity of cultivable midgut microbiota at different stages of the Asian tiger mosquito, *Aedes albopictus* from Tezpur, India," *PLoS One*, vol. 11, no. 12, article e0167409, 2016.
- [56] K. K. Yadav, A. Bora, S. Datta et al., "Molecular characterization of midgut microbiota of *Aedes albopictus* and *Aedes aegypti* from Arunachal Pradesh, India," *Parasites & Vectors*, vol. 18, no. 8, p. 641, 2015.
- [57] J. Mukhopadhyay, H. R. Braig, E. D. Rowton, and K. Ghosh, "Naturally occurring culturable aerobic gut flora of adult *Phlebotomus papatasi*, vector of *Leishmania major* in the Old World," *PLoS One*, vol. 7, no. 5, 2012.
- [58] K. Radjame, R. Sirinivasan, and V. Dhanda, "Oviposition response of phlebotomid sandfly *Phlebotomus papatasi* to soil bacteria isolated from natural breeding habitats," *Indian Journal of Experimental Biology*, vol. 35, pp. 59–61, 1997.
- [59] M. T. Tchioffo, A. Boissiere, T. S. Churcher et al., "Modulation of malaria infection in *Anopheles gambiae* mosquitoes exposed to natural midgut bacteria," *PLoS One*, vol. 8, no. 12, article e81663, 2013.
- [60] L. M. Díaz-Nieto, C. D'Alessio, M. A. Perotti, and C. M. Berón, "*Culex pipiens* development is greatly influenced by native bacteria and exogenous yeast," *PLoS One*, vol. 11, no. 4, article e0153133, 2016.
- [61] N. D. Djadid, H. Jazayeri, A. Raz, G. Favia, I. Ricci, and S. Zakeri, "Identification of the midgut microbiota of *An. stephensi* and *An. maculipennis* for their application as a para-transgenic tool against malaria," *PLoS One*, vol. 6, no. 12, article e28484, 2011.
- [62] R. J. Dillon and V. M. Dillon, "The gut bacteria of insects: non-pathogenic interactions," *Annual Review of Entomology*, vol. 49, no. 1, pp. 71–92, 2004.
- [63] B. Charroux and J. Royet, "*Drosophila* immune response: from systemic antimicrobial peptide production in fat body cells to local defense in the intestinal tract," *Fly*, vol. 4, pp. 40–47, 2014.
- [64] J. H. Ryu, S. H. Kim, H. Y. Lee et al., "Innate immune homeostasis by the homeobox gene caudal and commensal-gut mutualism in *Drosophila*," *Science*, vol. 319, no. 5864, pp. 777–782, 2008.
- [65] Z. Xi, J. L. Ramirez, and G. Dimopoulos, "The *Aedes aegypti* toll pathway controls dengue virus infection," *PLoS Pathogens*, vol. 4, no. 7, article e1000098, 2008.
- [66] L. A. Moreira, I. Iturbe-Ormaetxe, J. A. Jeffery et al., "A *Wolbachia* symbiont in *Aedes aegypti* limits infection with dengue, Chikungunya, and *Plasmodium*," *Cell*, vol. 139, no. 7, pp. 1268–1278, 2009.
- [67] C. M. Cirimotich, J. L. Ramirez, and G. Dimopoulos, "Native microbiota shape insect vector competence for human pathogens," *Cell Host & Microbe*, vol. 10, no. 4, pp. 307–310, 2011.
- [68] C. M. Cirimotich, Y. Dong, A. M. Clayton et al., "Natural microbe-mediated refractoriness to *Plasmodium* infection in *Anopheles gambiae*," *Science*, vol. 332, no. 6031, pp. 855–858, 2011.
- [69] G. Favia, I. Ricci, C. Damiani et al., "Bacteria of the genus *Asaia* stably associate with *Anopheles stephensi*, an Asian malarial mosquito vector," *Proceedings of the National Academy of Sciences*, vol. 104, no. 21, pp. 9047–9051, 2007.
- [70] M. A. Riehle, C. K. Moreira, D. Lampe, C. Lauzon, and M. Jacobs-Lorena, "Using bacteria to express and display anti-Plasmodium molecules in the mosquito midgut," *International Journal for Parasitology*, vol. 37, no. 6, pp. 595–603, 2007.
- [71] D. C. Bisi and D. J. Lampe, "Secretion of anti-Plasmodium effector proteins from a natural *Pantoea agglomerans* isolate by using PelB and HlyA secretion signals," *Applied and Environmental Microbiology*, vol. 77, no. 13, pp. 4669–4675, 2011.
- [72] J. M. Lindh, A. K. Borg-Karlson, and I. Faye, "Transstadial and horizontal transfer of bacteria within a colony of *Anopheles gambiae* (Diptera: Culicidae) and oviposition response to bacteria-containing water," *Acta Tropica*, vol. 107, no. 3, pp. 242–250, 2008.
- [73] F. Rosso, V. Tagliapietra, D. Albanese et al., "Reduced diversity of gut microbiota in two *Aedes* mosquito species in areas of recent invasion," *Scientific Reports*, vol. 8, article 1609, 2018.
- [74] P. H. Kelly, S. M. Bahr, T. D. Serafim et al., "The gut microbiome of the vector *Lutzomyia longipalpis* is essential for survival of *Leishmania infantum*," *mBio*, vol. 8, article e01121, 2017.
- [75] D. S. Gusmão, A. V. Santos, D. C. Marini, M. Bacci, M. A. Berbert-Molina, and F. J. A. Lemos, "Culture-dependent and culture-independent characterization of microorganisms associated with *Aedes aegypti* (Diptera: Culicidae) (L.) and dynamics of bacterial colonization in the midgut," *Acta Tropica*, vol. 115, no. 3, pp. 275–281, 2010.
- [76] O. Terenius, C. D. Oliveira, W. D. Pinheiro, W. P. Tadei, A. A. James, and O. Marinotti, "16S rRNA gene sequences from bacteria associated with adult *Anopheles darlingi* (Diptera: Culicidae) mosquitoes," *Journal of Medical Entomology*, vol. 45, no. 1, pp. 172–175, 2008.
- [77] E. J. Muturi, J. J. Bara, A. P. Rooney, and A. K. Hansen, "Midgut fungal and bacterial microbiota of *Aedes triseriatus* and *Aedes japonicus* shift in response to La Crosse virus infection," *Molecular Ecology*, vol. 25, no. 16, pp. 4075–4090, 2016.
- [78] C. A. Molina, J. F. Cana-Roca, A. Osuna, and S. Vilchez, "Selection of a *Bacillus pumilus* strain highly active against *Ceratitis capitata* (Wiedemann) larvae," *Applied and Environmental Microbiology*, vol. 76, no. 5, pp. 1320–1327, 2010.