



# Genes down-regulated in spaceflight are involved in the control of longevity in *Caenorhabditis elegans*

## SUBJECT AREAS:

MOLECULAR  
NEUROSCIENCE

GENE EXPRESSION  
PHYSIOLOGY  
SENESCENCE

Yoko Honda<sup>1</sup>, Akira Higashibata<sup>2</sup>, Yohei Matsunaga<sup>3</sup>, Yukiko Yonezawa<sup>4</sup>, Tsuyoshi Kawano<sup>3</sup>, Atsushi Higashitani<sup>5</sup>, Kana Kuriyama<sup>6</sup>, Toru Shimazu<sup>6</sup>, Masashi Tanaka<sup>1</sup>, Nathaniel J. Szewczyk<sup>7</sup>, Noriaki Ishioka<sup>2</sup> & Shuji Honda<sup>1</sup>

Received  
15 February 2012

Accepted  
26 April 2012

Published  
5 July 2012

Correspondence and requests for materials should be addressed to Y.H. (yhonda@center.tmig.or.jp) or S.H. (hondas@center.tmig.or.jp).

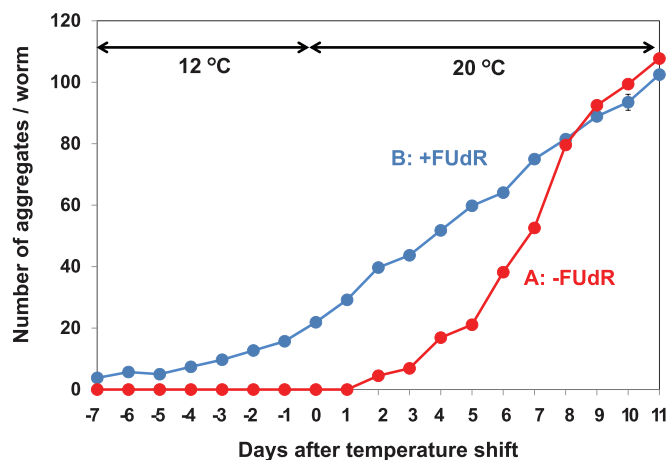
<sup>1</sup>Genomics for Longevity and Health, Tokyo Metropolitan Institute of Gerontology, Tokyo, 173-0015, Japan, <sup>2</sup>Japan Aerospace Exploration Agency, Tsukuba, 305-8505, Japan, <sup>3</sup>Department of Bioresources Science, The United Graduated School of Agricultural Sciences, Tottori University, Tottori, 680-8553, Japan, <sup>4</sup>Department of Bioresource Sciences, Faculty of Agriculture, Tottori University, Tottori, 680-8553, Japan, <sup>5</sup>Graduate School of Life Science, Tohoku University, Sendai, 980-8577, Japan, <sup>6</sup>Japan Space Forum, Tokyo, 101-0062, Japan, <sup>7</sup>MRC/Arthritis Research UK Centre for Musculoskeletal Ageing Research, University of Nottingham, Derby, DE22 3DT, UK.

How microgravitational space environments affect aging is not well understood. We observed that, in *Caenorhabditis elegans*, spaceflight suppressed the formation of transgenically expressed polyglutamine aggregates, which normally accumulate with increasing age. Moreover, the inactivation of each of seven genes that were down-regulated in space extended lifespan on the ground. These genes encode proteins that are likely related to neuronal or endocrine signaling: acetylcholine receptor, acetylcholine transporter, choline acetyltransferase, rhodopsin-like receptor, glutamate-gated chloride channel, shaker family of potassium channel, and insulin-like peptide. Most of them mediated lifespan control through the key longevity-regulating transcription factors DAF-16 or SKN-1 or through dietary-restriction signaling, singly or in combination. These results suggest that aging in *C. elegans* is slowed through neuronal and endocrine response to space environmental cues.

Lifespan and aging rate in metazoans are influenced by environmental factors, including temperature<sup>1</sup>, oxygen<sup>2</sup>, pheromone<sup>3</sup>, and food intake<sup>4</sup>. Accordingly, perturbation of sensory perception or signaling of mechanical, chemical, or osmotic stimuli changes the lifespan of *Caenorhabditis elegans*<sup>5</sup>. Microgravity has been shown to induce several physiological or pathological changes including disturbance of the sense of equilibrium and loss of muscle and bone mass, mainly from the observations in spaceflight<sup>6</sup>. However, how microgravity affects organismal aging and lifespan has not been well understood. This issue is not only intriguing from a basic scientific perspective but also important from a space medical aspect, because the duration of space missions will be increasing such as those for human exploration of other planets or colonization of the moon. To obtain some basic insights into aging in space, we examined an aging marker in space-flown *C. elegans* and explored the involvement of the genes whose expression was changed during spaceflight, in the control of lifespan.

## Results

During the International *C. elegans* Experiment First (ICE 1st) project<sup>7</sup>, we investigated the effect of spaceflight on the formation of aggregates of a 35-glutamine repeat (Q35) in *C. elegans* transgenically expressing the (CAG)<sub>35</sub>-yellow fluorescent protein (YFP) gene in muscle, which normally increases with advancing age<sup>8</sup> (Fig. 1). Q35 aggregate formation expressed as the number of aggregates per worm was found to be lower in worms flown in space from the L1 larva stage and L4/young adult stage than in matched ground control worms (Fig. 2A, B). This difference may be because of the possible changes in growth rate induced by spaceflight. However, growth of worms has been reported to be unaffected by spaceflight<sup>7,9</sup>. Moreover, the number of aggregates per body length was lower in space-flown worms than in ground control worms (Fig. 2D, E). This indicated that the spaceflight-induced suppression of Q35 aggregates was not ascribed to the spaceflight-induced changes in growth rate. Numbers of Q35 aggregates per total YFP fluorescence intensity in each worm, an indicator of Q35 expression, was also lowered by spaceflight (Fig. 2F, G). This showed that the spaceflight-induced suppression of Q35



**Figure 1 | Age-dependent increases in Q35 aggregate formation on ground: simulation of procedures of spaceflight experiment.** (A) Eggs obtained by treatment of gravid-adult Q35–YFP transgenic worms with hypochlorite were allowed to hatch and maintained at L1 larva stage in CeMM for 7 days at 12°C. Then, temperature was shifted to 20°C. Worms grew to adult stage and started to lay eggs at 7 days after the temperature shift, on the other hand, the progenies produced by these worms did not grow to adult stage by 11 days after the temperature shift. Therefore, at 9 days after the temperature shift, adults seen in culture could be considered as the worms originally inoculated. (B) L4/young adult worms were incubated in CeMM with 40  $\mu$ M FUdR for 7 days at 12°C and temperature was shifted to 20°C. Average numbers of Q35 aggregates/worm of more than 40 worms were plotted against time after the temperature shift to 20°C with  $\pm$  S.E. Absent error bars indicate that errors fell within symbols. These figures show one of two trials, each of which gave similar results.

aggregates was not due to spaceflight-induced changes in Q35 expression. These results suggest that biomarkers of aging are expressed more slowly in space-flown *C. elegans* in than ground control worms.

Further, these findings led us to propose a working hypothesis that the space environment changes the expression of genes involved in the control of aging. To identify the possible longevity-control genes, we first used the data from a DNA microarray experiment conducted to examine changes in gene expression in response to spaceflight<sup>10</sup> (The data set was deposited in the Gene Expression Omnibus (GEO) database (accession number: GSE36358) and WormBase (www.wormbase.org)), which showed that the expression of 48 genes increased by more than two-fold, and that of 199 genes decreased to less than half in the spaceflight conditions relative to the ground control<sup>11</sup>. Among these genes, we noticed that eleven genes likely related to neuronal or endocrine signaling were down-regulated in space-flown worms. We confirmed these observations and quantitatively evaluated the extent of decreased mRNA expression using real-time RT-PCR (Fig. 3). Second, we examined the effects of the inactivation of these eleven genes by loss- or reduction-of-function mutations and/or feeding RNA interference (RNAi) on the lifespan under ground laboratory conditions. We found that the inactivation of each of seven genes among these eleven genes extended lifespan on NGM agar covered with killed or live bacteria as food (Fig. 4A, Supplementary Tables S1 and S2 online). These included *gar-3*, acetylcholine receptor; *unc-17*, acetylcholine transporter; *cha-1*, choline acetyltransferase; F57A8.4, rhodopsin-like G-protein coupled receptor (GPCR); *glc-4*, glutamate-gated chloride channel; *shk-1*, shaker family of potassium channel; and *ins-35*, insulin-like peptide. We also ascertained whether these lifespan extensions could be seen in the liquid culture medium (CeMM) that had been used to culture worms in the spaceflight experiment. The mutations in all of these genes, except *unc-17(e245)* whose lifespan could

not be assessed because it could not grow in CeMM for presently unknown reasons, were also found to extend lifespan (Fig. 5).

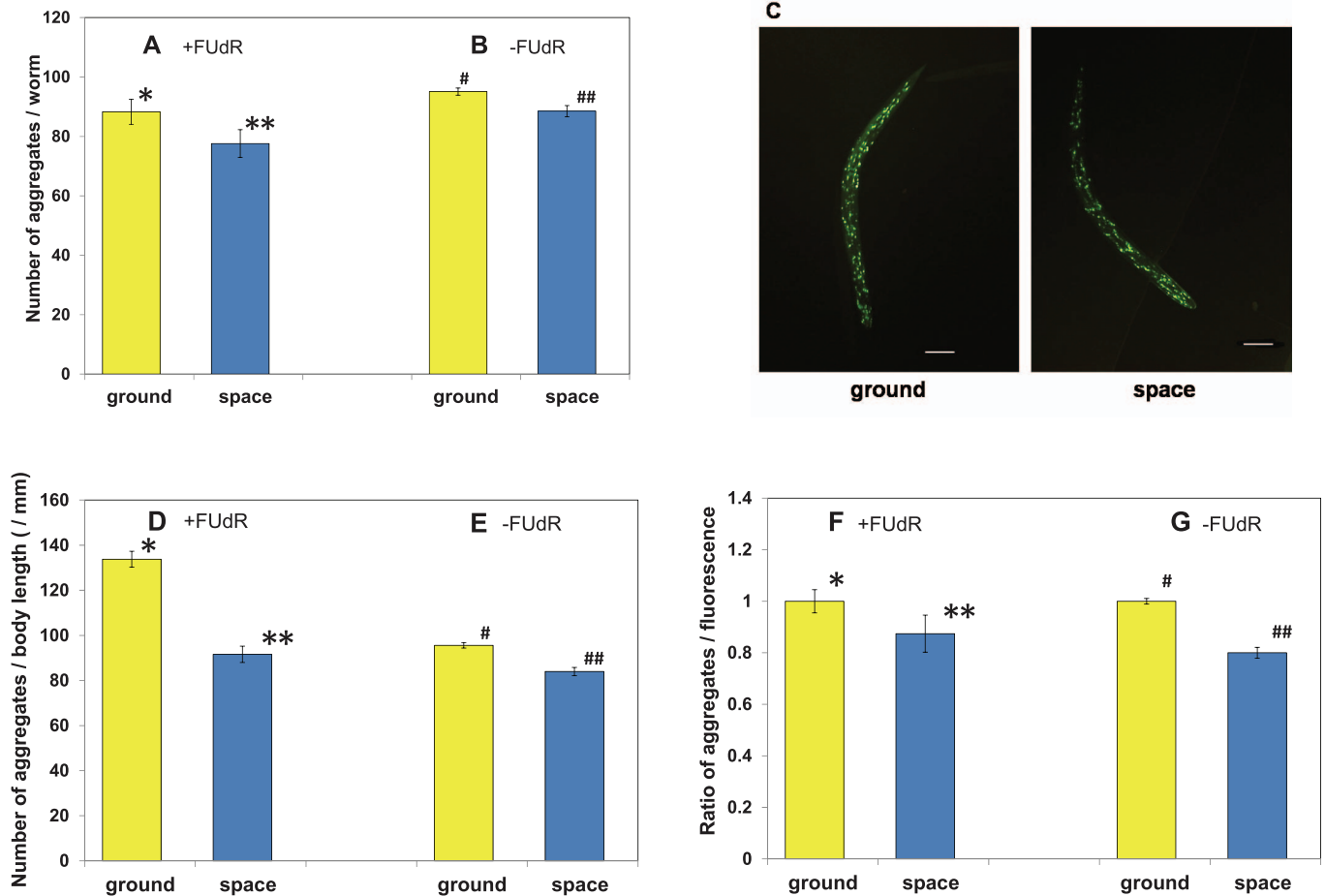
To further explore the mechanism of lifespan extension by the inactivation of each of these genes, we investigated whether lifespan extensions are mediated through the DAF-16/FOXO transcription factor, which is a key factor in lifespan extension by reduction of insulin/IGF-1 like signaling (IIS)<sup>12</sup>, or through SKN-1, an Nrf-like xenobiotic-response factor, which is the other key factor in lifespan extension both by reduction of IIS<sup>13</sup> and by dietary-restriction signaling<sup>14</sup>. Lifespan extensions caused by mutations either in *ins-35*, *glc-4*, *unc-17*, or *shk-1* were totally abolished by *daf-16* RNAi inactivation, whereas mutations either in *gar-3*, F57A8.4, or *cha-1* still lived longer than wild-type worms under *daf-16* RNAi (Fig. 4B, Supplementary Table S2 online). These results suggest that INS-35, GLC-4, UNC-17, and SHK-1 control lifespan through IIS/DAF-16 signaling. Further, the *skn-1* RNAi completely abolished the lifespan extension induced by mutations in *ins-35*, *glc-4*, and *shk-1*, whereas mutants in *unc-17*, *gar-3*, F57A8.4, and *cha-1* still lived longer than wild-type worms under *skn-1* RNAi (Fig. 4C, Supplementary Table S2 online). These results suggest that INS-35, GLC-4, and SHK-1 control lifespan through SKN-1. To determine whether these lifespan extensions are mediated through dietary-restriction signaling, we examined the lifespan under the inactivation of *eat-2*, which induces feeding impairment-based dietary restriction<sup>15</sup>. The *eat-2* RNAi further enhanced the extension of lifespan by mutations in *ins-35*, *glc-4*, *unc-17*, or *gar-3*, whereas the *eat-2* RNAi shortened the lifespan of mutants of *shk-1*, *cha-1*, or F57A8.4 (Fig. 4D and Supplementary Table S2 online). These results suggest that SHK-1, CHA-1, and the F57A8.4 protein share a common lifespan control mechanism with dietary-restriction signaling.

In order to explore the involvement of these seven genes in the suppression of Q35 aggregate formation during spaceflight, we examined the effect of RNAi inactivation of some of these genes on Q35 aggregation. RNAi inactivation of *gar-3*, *cha-1*, and *shk-1* reduced Q35 aggregation (Fig. 6), suggesting that GAR-3, CHA-1, and SHK-1 control Q35 aggregate formation.

We investigated whether these seven genes relate to the formation of dauer larvae, a long-lived growth arrest state under harsh environmental conditions. Mutation and RNAi of *ins-35* and mutation of *shk-1* were found to enhance pheromone-induced dauer formation, whereas mutations in *glc-4*, *unc-17*, or F57A8.4 suppressed it (Fig. 7A, B). These results suggest that INS-35 and SHK-1 may be related to dauer-associated life maintenance. Alternatively, some of these five genes may be involved in sensory perception or signaling related to dauer induction.

## Discussion

We identified seven genes, which were down-regulated in space and whose inactivation extended lifespan under ground conditions. These genes encode proteins that are conserved across animal phylogeny and that are likely related to neuronal or endocrine signaling. How do these genes play a role in longevity control? One possibility is that they are involved in longevity control through sensory perception or signaling of space environmental cues. Mutations that cause defects in sensory perception and signaling systems, related to mechanical, chemical, thermal, and osmotic stimuli and pheromones, have been reported to extend the lifespan of *C. elegans*<sup>5</sup>. SHK-1<sup>16</sup> and GLC-4<sup>17</sup> are expressed in sensory neurons and UNC-17 in interneurons in sensory signaling<sup>18</sup>. F57A8.4 encodes a homolog of rhodopsin that is known to sense light<sup>19</sup>, suggesting that it may be involved in sensing stimuli in space environments. Pheromone-induced dauer formation was inhibited by the inactivation of *unc-17*, *glc-4*, or F57A8.4 and enhanced by that of *ins-35* or *shk-1* (Fig. 7A, B), suggesting that some of these genes are involved in lifespan control through sensory perception or signaling processes overlapping with dauer-switching processes. Alternatively, it may be possible that

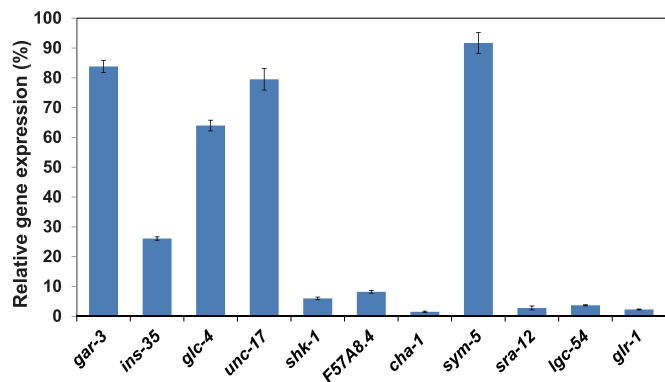


**Figure 2 | Spaceflight reduced Q35 aggregate formation.** Q35-YFP transgenic worms were inoculated into CeMM with 40  $\mu$ M FUDR at L4/young adult stage (A, D, F) or without FUDR at egg stage (B, E, G), and incubated on ground for 5 days at 12°C. Then they were space-flown or on ground for 2 days at 12°C and for next 9 days at 20°C. In B, E, and G, only adult worms in the samples containing L1, L2 and adult stage worms were measured as adult worms were considered as the worms originally inoculated (See Legend for Fig. 1). Numbers of Q35 aggregates were counted from the photographs of worms obtained by a fluorescence microscope (as in (C)) using ImageJ software (Analyze Particles). (A, B) Numbers of Q35 aggregates per worm are shown. \* vs \*\* and # vs ##:  $p < 0.001$  (Student's t test). (C) Photographs of YFP fluorescence of the worms flown in space from L1 stage and matched ground control are shown. Bars show 100  $\mu$ m. (D, E) Numbers of Q35 aggregate formation per body length are shown. The lengths of individual worms were measured using the ImageJ software. \* vs \*\* and # vs ##:  $p < 0.001$  (Student's t test). (F, G) Ratio of Q35 aggregate formation of space-flown worms to those of ground control worms, per total body fluorescence intensity are shown. Whole-worm YFP fluorescence was quantified using the measurement and analysis software VH-HIA5 (Keyence). \* vs \*\* and # vs ##:  $p < 0.001$  (Student's t test). Numbers of worms examined were as follows, A, D and F: space-flown ( $n = 15$ ) and ground control worms ( $n = 47$ ); B, E and G: space-flown ( $n = 138$ ) and ground control worms ( $n = 367$ ). Data are expressed as the mean  $\pm$  S.E.

the inactivation of *ins-35* or *shk-1* extends lifespan, as some abnormally efficient dauer formation mutants such as *daf-2* display adult-lifespan extension, and it has been hypothesized that these mutants ectopically express a dauer-related efficient life maintenance capability in the adult stage<sup>20</sup>. Meanwhile, *ins-35* is likely involved in endocrine signaling in the control of dauer formation and lifespan, as some of the 40 insulin-like peptides participate in regulating dauer formation and lifespan in *C. elegans* through a cell-nonautonomous mechanism<sup>21</sup>. The inactivation of *ins-35* as well as *glc-4* and *shk-1* extended lifespan through the DAF-16 transcription factor (Fig. 4B). Reduction in IIS is known to extend lifespan through DAF-16<sup>12</sup> similar to some sensory signaling mutants<sup>5</sup>. Together with the observation that the space environments and IIS reduction shared some common features in terms of their effects on gene expression<sup>22</sup> and that a hypergravity environment induces DAF-16 localization to nuclei we suggest that DAF-16 also mediates responses to altered gravity levels (the hypergravity experiment used *Escherichia coli* as food instead of CeMM<sup>23</sup>). On the other hand, the lifespan extension by the inactivation of *glc-4*, *ins-35*, *shk-1*, or *cha-1* required SKN-1

(Fig. 4C). SKN-1 in sensory neurons was reported to mediate lifespan determination during dietary-restriction signaling<sup>14</sup>, although SKN-1 in intestine also plays a role in lifespan determination through IIS<sup>13</sup>.

A second possibility is that the genes in the present study are involved in longevity control through motor neuron-muscle signaling. SHK-1 functions in muscle<sup>24</sup> and is expressed in neurons<sup>16</sup>. CHA-1 and UNC-17, which are expressed in motor neurons<sup>25</sup>, and GAR-3, which is expressed in motor neurons and muscle<sup>26</sup>, play roles in acetylcholine transmission in motor neuron-muscle signaling. *cha-1* and *unc-17* are encoded by a single polycistronic cluster. They share the first noncoding exon; therefore, a common promoter termed as “cholinergic promoter” regulates the expression of both genes<sup>27</sup>. Increased acetylcholine signaling enhanced polyglutamine aggregation in post-synaptic muscle cells in *C. elegans*, suggesting that excess neuronal stimuli disrupt a balance in protein homeostasis in the target muscle<sup>28</sup>. Conversely, reduction in neuromuscular signaling may enhance protein structure stabilization, which has been reported to be associated with the enhancement of longevity<sup>29</sup>. Consistent with these studies, several mutations and RNAi



**Figure 3 | The genes likely related to neuronal and endocrine signaling were down-regulated by spaceflight.** Relative gene expression levels in the mixed population of space-flown or ground control N2 worms are shown. *gar-3*: acetylcholine receptor, *unc-17*: acetylcholine transporter, *cha-1*: choline acetyltransferase, *F57A8.4*: rhodopsin-like GPCR, *glc-4*: glutamate-gated chloride channel, *shk-1*: shaker family of potassium channel, and *ins-35*: insulin-like peptide, *lgc-54*: ligand-gated ion channel, *glr-1*: glutamate receptor, *sym-5*: synthetic lethal with *mec*, *sra-12*: serpentine receptor. Real-time RT-PCR was performed in triplicate for one sample each from the spaceflight experiment and ground control. The mRNA levels of each gene were adjusted to that of *gpd-2* (glyceraldehyde-3-phosphate dehydrogenase) mRNA, which was used as the internal standard. Bar graphs are ratios of the mRNA levels in spaceflight to those on ground (%). Data are expressed as the mean  $\pm$  S.E. ( $n=3$ ). All of them were significantly different ( $p<0.05$ ) compared with ground control (Student's t-test).

treatments recently predicted to disrupt depolarization of the muscle membrane have been shown to alter protein homeostasis within muscle<sup>30</sup>. In fact, the inactivation of *gar-3*, *cha-1*, and *shk-1* suppressed Q35 aggregation (Fig. 6), suggesting that GAR-3, CHA-1, and SHK-1 play a role in protein structure stability in muscle during motor neuron-muscle signaling. In case of *cha-1*, it has been shown that loss-of-function mutants evoke muscle protein degradation, which is suppressed by inhibitors of the ubiquitin-proteasome activity<sup>31</sup>. PolyQ aggregation was also reported to be regulated by genes related to the ubiquitin-proteasome system<sup>32</sup>. These neuromuscular signaling changes may affect the muscle protein metabolism observed during spaceflight<sup>10</sup>. Alternatively, the possibility remains that SHK-1, which is expressed in sensory neurons<sup>16</sup>, controls polyQ aggregation through a mechanism such as thermosensory neuronal circuit nonautonomously modulates protein misfolding in muscle and intestine cells via  $Ca^{2+}$ -dependent dense core vesicle neurosecretion<sup>33</sup>. The  $Ca^{2+}$ -dependent dense core vesicle neurosecretion mechanism modulates the HSP-70 levels in muscle to affect polyQ aggregation<sup>33</sup>, and HSP-70 levels are elevated in response to spaceflight<sup>22</sup>. In either case, the suppression of Q35 aggregation observed in space might be explained in part by reduced expression of *gar-3*, *cha-1*, and/or *shk-1*. Given that polyglutamine aggregation is considered to be regulated by protein synthesis, degradation, or transport<sup>32</sup>, it seems likely that reduction of *gar-3*, *cha-1*, and/or *shk-1* affect proteostasis in muscle; this also may partially account for the decreased expression of muscle proteins during spaceflight<sup>10</sup>.

Lifespan extension by *eat-2* inactivation inducing the dietary-restriction state<sup>15</sup> was totally abolished by the inactivation of either *cha-1*, *shk-1* or *F57A8.4* (Fig. 4D), suggesting that CHA-1, SHK-1, or *F57A8.4* protein function in processes related to sensory perception of food or signaling in response to food. Alternatively, some of these may be involved in motor neuron-muscle signaling related to food intake. In support of this idea, CHA-1 was reported to participate in determining pharyngeal pumping rate to affect food intake<sup>34</sup>.

The combined results suggest that environmental factors associated with spaceflight, including microgravity, modulate neuronal or

endocrine signaling to induce some of “longevity-promoting” processes, including dietary-restriction signaling, stabilization of protein structure, or dauer-related efficient life maintenance during harsh conditions (Fig. 8). However, it should be noted that whether suppression of Q35 aggregation and the expression of the seven studied genes is due to microgravity or other environmental factors such as radiation is still uncertain, because the necessary experiments under the artificial gravitational field of the earth (1 G) in space environments have not been performed. Although further investigation regarding the effects of a decrease in the expression of the present genes comparable to the level in spaceflight and a simultaneous decrease in the expression of all these genes on lifespan is required, the present findings suggest that space-flown worms age more slowly compared with ground control worms and further predict that spaceflight extends worm lifespan. Male *Drosophila* lived shorter on the ground after spaceflight compared to controls that maintained on the ground through their lifetimes<sup>35</sup>. Further investigation is needed to compare aging and lifespan under microgravity and 1 G in space environments.

## Methods

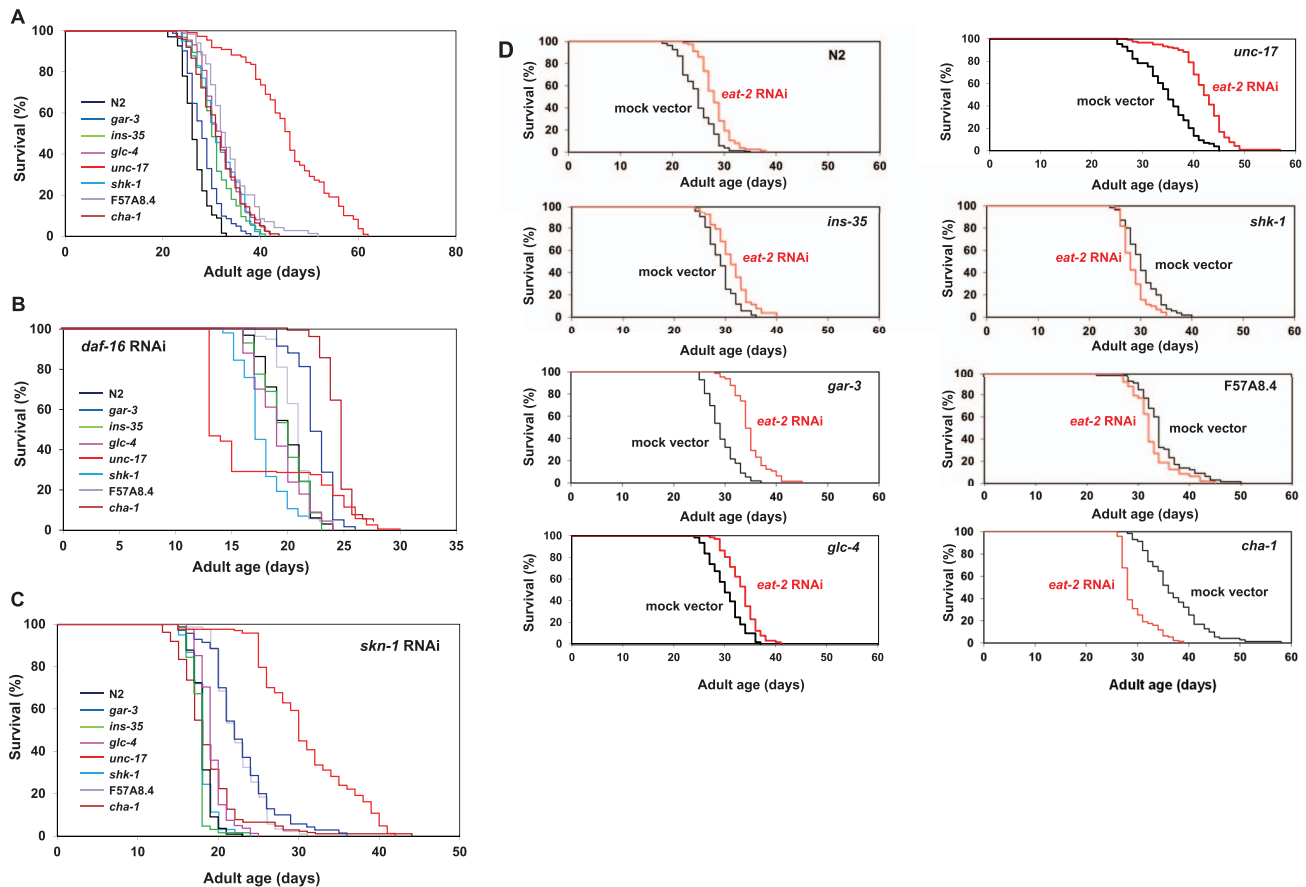
**Nematode strains and culture conditions.** Unless otherwise stated, the *C. elegans* strains were maintained at 20°C on nematode growth medium (NGM) agar with *E. coli* OP50 as a food source, as previously described<sup>36</sup>. The N2 Bristol strain was used as the wild-type *C. elegans*. The mutant strains used in this study were JD31: *glc-4(ok212)* II, NL2099: *rrf-3(pk1426)* II, RB1392: *shk-1(ok1581)* II, PR1152: *cha-1(p1152)* IV, TY1652: *cha-1(y226)* IV, CB933: *unc-17(e245)* IV, *F57A8.4(tm4341)* V, VC670: *gar-3(gk337)* V, TM290 (x5 outcrossed): *ins-35(ok3297)* V, and Q35-YFP strain.

**Spaceflight experiment.** The spaceflight experiment was conducted as described previously<sup>7</sup>. Briefly, eggs isolated after treatment of gravid adult worms with hypochlorite were incubated in a chemically defined liquid medium, *C. elegans* Maintenance Medium (CeMM)<sup>37</sup> and allowed to hatch and grow. Approximately 70 worms at the L4 larva/young adult stage were placed into a gas-permeable culture bag<sup>7</sup> containing 2.5 mL CeMM with 40  $\mu$ M 5-fluoro-2'-deoxyuridine (FUDU, Sigma Aldrich, St. Louis, MO, USA) which was used to prevent self-fertilization. Approximately 100 hatched L1 worms were also placed in a culture bag containing 2.5 mL CeMM without FUDU. Both of them were incubated for 5 days at 12°C on the ground. After the launch, they were flown for 2 days at 12°C and on board the International Space Station for 9 days at 20°C. When samples were returned to earth, they were flash frozen (freezing time <1 min) in liquid nitrogen within 2 h of landing<sup>7</sup>. Control worms underwent the same procedures at the same time on the ground.

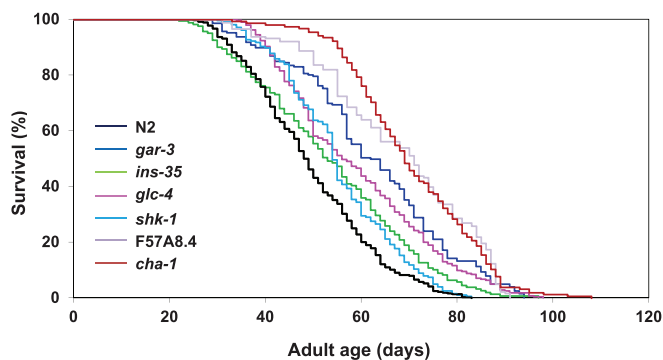
**PolyQ aggregate measurement.** To measure polyQ aggregates, a *C. elegans* transgenic line expressing a chimeric fusion protein of a 35-glutamine repeat (Q35) and yellow fluorescent protein (YFP) in the body wall muscle, as described previously<sup>8</sup>, was used. The Q35 aggregates of worms were counted with a fluorescence microscope system (VB-G25/S20/L10/7010; Keyence CO., Osaka, Japan). Aggregates that were more than 1  $\mu$ m long were defined as discrete structures.

**Real-time RT-PCR.** To confirm the gene expression differences between ground control worms and space-flown worms, real-time PCR was performed. Total RNA of a mixed population of space-flown and ground control N2 worms was reverse-transcribed to cDNA using ExScript RT reagent kit (TaKaRa Bio Inc., Shiga, Japan) and iCycler thermal cycler (Bio-Rad Laboratories Inc., Hercules, CA, USA). The reaction mixture for measuring the cDNA quantity was prepared using Premix EX Taq (TaKaRa Bio) according to the manufacturer's instructions. Reaction and fluorescence monitoring were done using a Thermal Cycler Dice Real Time System (Takara Bio). The following primers were used:

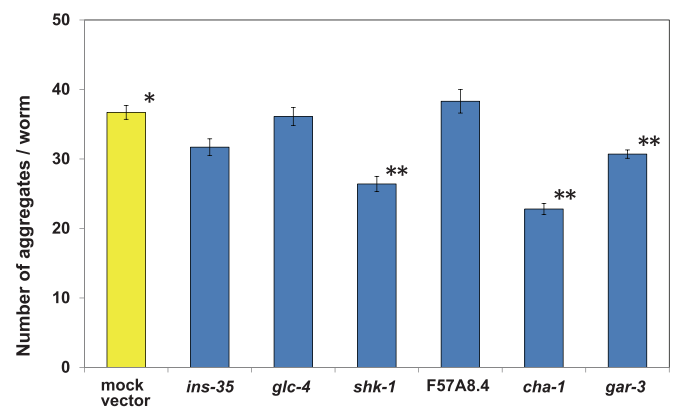
*gpd-2* forward, 5'-accggagtctccaccaccatc-3'; *gpd-2* reverse, 5'-acgactcagaggtta-caagca-3';  
*gar-3* forward, 5'-tgacaccgatgccgacta-3'; *gar-3* reverse, 5'-gcgacttcgaaaccttgctga-3';  
*ins-35* forward, 5'-actgccaacgcgactaaag-3'; *ins-35* reverse, 5'-tggctctgggcagcagataag-3';  
*glc-4* forward, 5'-ccgttggtgcagttctgt-3'; *glc-4* reverse, 5'-catgactcgtccggatcctc-3';  
*unc-17* forward, 5'-ccacggctatattgtcttggga-3'; *unc-17* reverse, 5'-ctaggcaccggaac-gagatga-3';  
*shk-1* forward, 5'-tgcgcaatgattggtgtctt-3'; *shk-1* reverse, 5'-atgtctcgtctgcatcgtc-3';  
*F57A8.4* forward, 5'-tcgcatcagacgctgtcagta-3'; *F57A8.4* reverse, 5'-tcctgcactcca-caggtgag-3';  
*cha-1* forward, 5'-cgttggtgtcttgcacatgga-3'; *cha-1* reverse, 5'-cgattgagcccaacttga-3';  
*lgc-54* forward, 5'-ccaatgcaatgcagcagag-3'; *lgc-54* reverse, 5'-tgtgcagagcgtg-caaag-3';



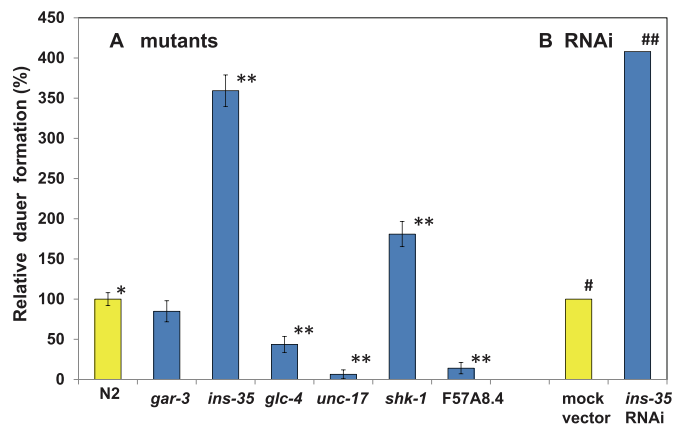
**Figure 4 | Lifespan of the mutants in the genes down-regulated by spaceflight.** (A) The survival curves of N2 and the mutants in the genes that were down-regulated by spaceflight, on NGM with UV-killed OP50 are shown. The percentage of live worms is plotted against adult age. Lifespans of the mutants in the seven genes were longer than those of the wild type. Data are one of two experiments, each of which gave similar results, detailed parameters of which are indicated in Supplementary Table S1. The survival curves of N2 and the mutants treated with *daf-16* RNAi (B), *skn-1* RNAi (C) and *eat-2* RNAi (D) from the L1 stage until death. Data are one of two experiments detailed parameters of which are indicated in Supplementary Table S2 with the data of worms treated with mock-vector RNAi bacteria as control experiments.



**Figure 5 | The survival curves of N2 and the mutants in the genes that were down-regulated by spaceflight, in liquid CeMM.** The percentage of live worms is plotted against adult age. Day 0 corresponds to the L4 molt. Mean adult lifespan  $\pm$  S.E. (day), number of assayed worms and statistical significance with N2 wild type are: N2:  $49.5 \pm 0.7$ ,  $n=364$ ; *gar-3*(*gk337*):  $62.3 \pm 1.1$ ,  $n=205$ ,  $p<0.001$ ; *ins-35*(*ok3297*):  $53.5 \pm 1.3$ ,  $n=159$ ,  $p<0.01$ ; *glc-4*(*ok212*):  $59.0 \pm 0.9$ ,  $n=286$ ,  $p<0.001$ ; *shk-1*(*ok1581*):  $55.2 \pm 0.8$ ,  $n=204$ ,  $p<0.001$ ; F57A8.4 (*tm4341*):  $68.1 \pm 1.2$ ,  $n=179$ ,  $p<0.001$  and *cha-1*(*p1152*):  $70.5 \pm 1.1$ ,  $n=155$ ,  $p<0.001$ .



**Figure 6 | The effect of RNAi of the genes on Q35 aggregate formation.** The feeding RNAi of the designated genes against Q35-YFP transgenic worms was conducted as described in Methods for RNAi. L3 larval worms were placed on RNAi-expressing or mock-vector control bacteria. Worms were incubated for 2 days at  $20^{\circ}\text{C}$  until they became gravid adults. The gravid adults were transferred to fresh RNAi-expressing bacteria lawns and allowed to lay eggs for 2 hours and removed and eggs were allowed to grow for subsequent assays. Aggregates of Q35-YFP were counted at 7 days after the treatment was started. Numbers of Q35 aggregates per worm are shown. Data are expressed as the mean  $\pm$  S.E. \* vs \*\*:  $p<0.001$  (Student's *t* test).



**Figure 7 | Pheromone-induced dauer formation in the mutants.** Dauer formation of N2 and mutant worms (A) and RNAi fed *rrf-3* worms (B) was scored after 3 days incubation with 1% crude pheromone extract at 25°C from the egg stage. The relative percentages of dauer arrest in the mutants compared with N2 (A) and in the RNAi fed *rrf-3* worms compared with vector RNAi fed *rrf-3* worms (B) are shown. Data are expressed as mean  $\pm$  S.E. \* vs \*\* and # vs ##:  $p < 0.001$  (Student's *t* test). Percentages of dauer larvae (mean  $\pm$  S.E.  $n$ =number of worms examined): N2 ( $19.9 \pm 1.6\%$ ,  $n=367$ ), *gar-3*(*gk337*) ( $16.9 \pm 2.6\%$ ,  $n=418$ ), *ins-35*(*ok3297*) ( $71.5 \pm 3.9\%$ ,  $n=216$ ), *glc-4*(*ok212*) ( $8.7 \pm 2.0\%$ ,  $n=102$ ), *unc-17*(*e245*) ( $1.3 \pm 1.1\%$ ,  $n=171$ ), *shk-1*(*ok1581*) ( $36.0 \pm 3.1\%$ ,  $n=306$ ), F57A8.4(*tm4341*) ( $2.8 \pm 1.4\%$ ,  $n=170$ ), vector RNAi ( $16.2 \pm 2.3\%$ ,  $n=215$ ), and *ins-35* RNAi ( $66.1 \pm 3.1\%$ ,  $n=176$ ). These data are from one of two or three trials, each of which gave similar results. *cha-1*(*pr1152*) worms did not grow beyond the L2 stage under this condition.

*glr-1* forward, 5'-aggtgcagttgcacaaccag-3'; *glr-1* reverse, 5'-gtttgcaggtgatccacatta-3';

*sym-5* forward, 5'-aagccggagccagtgctagt-3'; *sym-5* reverse, 5'-cctgtcgttatggagcttc-3';

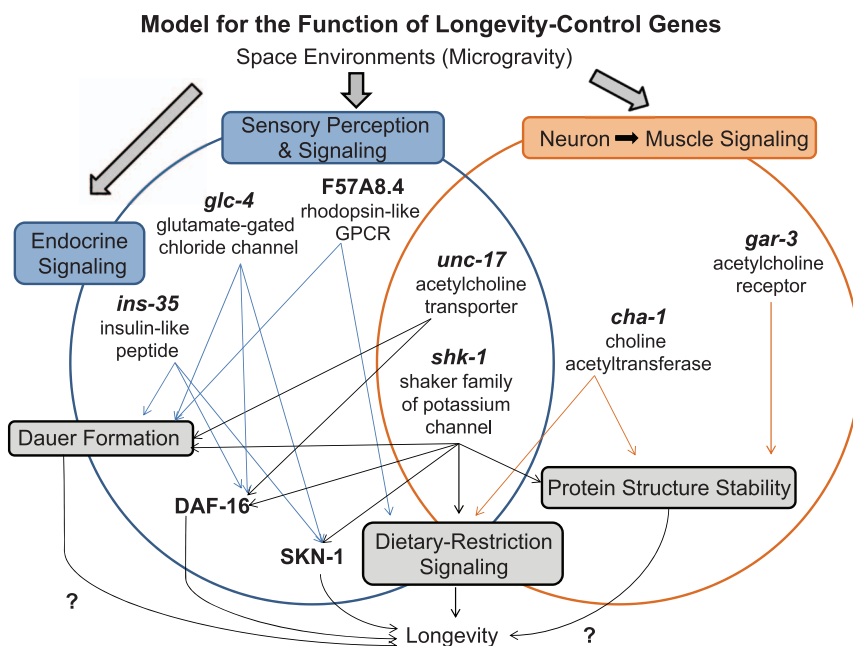
*sra-12* forward, 5'-gacgtcaccagcgatatttg-3'; *sra-12* reverse, 5'-gaaacacagccatattggagag-3'.

Real-time RT-PCR was performed in triplicate for one sample each from the spaceflight experiment and ground control. The mRNA levels of each gene were adjusted to that of *gpd-2* (glyceraldehyde-3-phosphate dehydrogenase), which was used as the internal standard. Flight/ground ratios of these were statistically analyzed by Student's *t*-test using PRISM software (GraphPad Software Inc., San Diego, CA, USA). Significance was accepted at  $p < 0.05$ .

**RNAi.** RNAi experiments were performed according to the RNAi feeding method described previously<sup>38</sup>. *E. coli* HT115 strains expressing double-stranded RNA that inactivated the target genes were obtained from Open Biosystems (Huntsville, AL, USA) and Source BioScience (Nottingham, UK).

**Determination of lifespan.** Eggs isolated after treatment of gravid adult worms with hypochlorite were incubated in S basal at 20°C for 1 day. Lifespan was determined in three ways. First, the hatched L1 larvae were placed on NGM agar covered with UV-killed OP50. UV killing was used to avoid any toxic effects of live *E. coli* on *C. elegans* lifespan<sup>39</sup>. Second, the hatched L1 worms were placed on NGM agar covered with RNAi-expressing or vector control bacteria. In these two assay systems, worms were transferred to fresh plates every other day. Third, the hatched L1 worms were inoculated into CeMM. In every case, synchronous L4/young adult stage worms estimated as 0-day adults in the lifespan assay were treated with 40  $\mu$ M FUdU to prevent self-fertilization. The number of surviving worms was monitored until death as judged by nonresponse to a mechanical stimulus. To focus on aging, we excluded worms that had become desiccated on the side of the plate after crawling off in the former two assay systems, and that displayed extruded internal organs, and those that died because of progeny hatching inside the uterus (matricidal death). The results of the survival assays were analyzed using the Kaplan-Meier method, and significance was measured with the log-rank test using the statistical analysis package StatMate III (ATMS, Tokyo, Japan).

**Dauer formation.** Dauer forming ability was tested in the presence of crude pheromone extract<sup>3</sup>. Crude pheromone extract was added to a final concentration of 1% to liquid NGM that had been autoclaved and cooled to 50°C. The media were immediately dispensed into petri plates. Gravid adult hermaphrodites were allowed to lay eggs on the plates covered with UV-killed OP50 for 6 h. After removal of the adult hermaphrodites, the plates were incubated at 25°C for 3 days and scored for



**Figure 8 | Model for the action of longevity-control genes whose expressions are suppressed by spaceflight.** In response to space-environmental stimuli including microgravity, sensory perception, neuronal and endocrine signaling, and motorneuron-muscle signaling work through changes in these gene activities. These changes, some of which activate DAF-16 and/or SKN-1 transcription factors, induce “longevity-promoting” processes including dietary-restriction signaling, stabilization of protein structure or dauer-related efficient life maintenance during harsh conditions.



dauer arrest. The dauer stage was confirmed by visual inspection of the animals as the presence of dark pigment granules and constriction of the body and pharynx and the loss of pharyngeal pumping.

- Klass, M. R. Aging in the nematode *Caenorhabditis elegans*: major biological and environmental factors influencing life span. *Mech. Ageing Dev.* **6**, 413–429 (1977).
- Honda, S., Ishii, N., Suzuki, K. & Matsuo, M. Oxygen-dependent perturbation of life span and aging rate in the nematode. *J. Gerontol.* **48**, B57–61 (1993).
- Kawano, K. *et al.* Lifespan extending activity of substances secreted by the nematode *Caenorhabditis elegans* that include the dauer-inducing pheromone. *Biosci. Biotechnol. Biochem.* **69**, 2479–2481 (2005).
- Masoro, E. J. Overview of caloric restriction and ageing. *Mech. Ageing Dev.* **126**, 913–922 (2005).
- Apfeld, J. & Kenyon, C. Regulation of lifespan by sensory perception in *Caenorhabditis elegans*. *Nature* **402**, 804–809 (1999).
- Vernikos, J. Human physiology in space. *Bioessays* **18**, 1029–1037 (1996).
- Szewczyk, N. J. *et al.* Description of International *Caenorhabditis elegans* Experiment first flight (ICE-FIRST). *Adv. Space Res.* **42**, 1072–1079 (2008).
- Morley, J. F., Brignull, H. R., Weyers, J. J. & Morimoto, R. I. The threshold for polyglutamine-expansion protein aggregation and cellular toxicity is dynamic and influenced by aging in *Caenorhabditis elegans*. *Proc. Natl. Acad. Sci. USA* **99**, 10417–10422 (2002).
- Oczypok, E. A. *et al.* Remote automated multi-generational growth and observation of an animal in low Earth orbit. *J. R. Soc. Interface* **9**, 596–599 (2012).
- Higashibata, A. *et al.* Decreased expression of myogenic transcription factors and myosin heavy chains in *Caenorhabditis elegans* muscles developed during spaceflight. *J. Exp. Biol.* **209**, 3209–3218 (2006).
- Higashitani, A. *et al.* Checkpoint and physiological apoptosis in germ cells proceeds normally in spaceflown *Caenorhabditis elegans*. *Apoptosis* **10**, 949–954 (2005).
- Kenyon, C. J. The genetics of ageing. *Nature* **464**, 504–512 (2010).
- Tullet, J. M. A. *et al.* Direct inhibition of the longevity promoting factor SKN-1 by Insulin-like signaling in *C. elegans*. *Cell* **132**, 1025–1038 (2008).
- Bishop, N. A. & Guarente, L. Two neurons mediate diet-restriction-induced longevity in *C. elegans*. *Nature* **447**, 545–549 (2007).
- Lakowski, B. & Hekimi, S. The genetics of caloric restriction in *Caenorhabditis elegans*. *Proc. Natl. Acad. Sci. USA* **95**, 13091–13096 (1998).
- Fawcett, G. L. *et al.* Mutant analysis of the Shal (Kv4) voltage-gated fast transient K<sup>+</sup> channel in *Caenorhabditis elegans*. *J. Biol. Chem.* **281**, 30725–30735 (2006).
- Hunt-Newbury, R. *et al.* High-throughput in vivo analysis of gene expression in *Caenorhabditis elegans*. *PLoS Biol.* **5**, e237 (2007).
- Altun-Gultekin, Z. *et al.* A regulatory cascade of three homeobox genes, *ceh-10*, *ttx-3* and *ceh-23*, controls cell fate specification of a defined interneuron class in *C. elegans*. *Development* **128**, 1951–1969 (2001).
- Palczewski, K. G protein-coupled receptor rhodopsin. *Annu. Rev. Biochem.* **75**, 743–767 (2006).
- Kenyon, C., Chang, J., Gensch, E., Rudner, A. & Tabtiang, R. A *C. elegans* mutant that lives twice as long as wild type. *Nature* **366**, 461–464 (1993).
- Tatar, M., Bartke, A. & Antebi, A. The endocrine regulation of aging by insulin-like signals. *Science* **299**, 1346–1351 (2003).
- Selch, F. *et al.* Genomic response of the nematode *Caenorhabditis elegans* to spaceflight. *Adv. Space Res.* **41**, 807–815 (2008).
- Kim, N. *et al.* Gravity Force Transduced by the MEC-4/MEC-10 DEG/ENaC Channel Modulates DAF-16/FoxO Activity in *Caenorhabditis elegans*. *Genetics* **177**, 835–845 (2007).
- Liu, P. *et al.* Genetic dissection of ion currents underlying all-or-none action potentials in *C. elegans* body-wall muscle cells. *J. Physiol.* **589**, 101–117 (2011).
- Duerr, J. S., Han, H. P., Fields, S. D. & Rand, J. B. Identification of major classes of cholinergic neurons in the nematode *Caenorhabditis elegans*. *J. Comp. Neurol.* **506**, 398–408 (2008).
- Liu, Y., LeBoeuf, B. & Garcia, L. R.  $G\alpha_q$ -coupled muscarinic acetylcholine receptors enhance nicotinic acetylcholine receptor signaling in *Caenorhabditis elegans* mating behavior. *J. Neurosci.* **27**, 1411–1421 (2007).
- Alfonso, A., Grundahl, K., McManus, J. R., Asbury, J. M. & Rand, J. B. Alternative splicing leads to two cholinergic proteins in *Caenorhabditis elegans*. *J. Mol. Biol.* **241**, 627–630 (1994)
- García, S. M., Casanueva, M. O., Silva, M. C., Amaral, M. D. & Morimoto, R. I. Neuronal signaling modulates protein homeostasis in *Caenorhabditis elegans* post-synaptic muscle cells. *Genes Dev.* **21**, 3006–3016 (2007).
- Taylor, R. C. & Dillin, A. Aging as an event of proteostasis collapse. *Cold Spring Harb. Perspect. Biol.* **3**, a004440 (2011).
- Shephard, F., Adenle, A. A., Jacobson, L. A. & Szewczyk, N. J. Identification and functional clustering of genes regulating muscle protein degradation from amongst the known *C. elegans* muscle mutants. *PLoS ONE* **6**, e24686 (2011)
- Szewczyk, N. J., Hartman, J. J., Barmada, S. J. & Jacobson, L. A. Genetic defects in acetylcholine signalling promote protein degradation in muscle cells of *Caenorhabditis elegans*. *J. Cell Sci.* **113**, 2003–2010 (2000).
- Nollen, E. A. A. *et al.* Genome-wide RNA interference screen identifies previously undescribed regulators of polyglutamine aggregation. *Proc. Natl. Acad. Sci. USA* **101**, 6403–6408 (2004).
- Prahlad, V. & Morimoto, R. I. Neuronal circuitry regulates the response of *Caenorhabditis elegans* to misfolded proteins. *Proc. Natl. Acad. Sci. USA* **108**, 14204–14209 (2011).
- Rand, J. B. & Russell, R. L. Choline acetyltransferase-deficient mutants of the nematode *Caenorhabditis elegans*. *Genetics* **106**, 227–248 (1984).
- Bengur, A. *et al.* Microgravity effects on *Drosophila melanogaster* behavior and aging. Implications of the IML-2 experiment. *J. Biotech.* **47**, 191–201 (1996).
- Brenner, S. The genetics of *Caenorhabditis elegans*. *Genetics* **77**, 71–94 (1974).
- Szewczyk, N. J., Kozak, E. & Conley, C. A. Chemically defined medium and *Caenorhabditis elegans*. *BMC Biotechnol.* **3**, 19 (2003).
- Timmons, L., Court, D. L. & Fire, A. Ingestion of bacterially expressed dsRNAs can produce specific and potent genetic interference in *Caenorhabditis elegans*. *Gene* **263**, 103–112 (2001).
- Garigan, D. *et al.* Genetic analysis of tissue aging in *Caenorhabditis elegans*: a role for heat-shock factor and bacterial proliferation. *Genetics* **161**, 1101–1112 (2002).

## Acknowledgements

We are grateful to the entire crew of the ICE 1st for operation in orbit. The ICE 1st was well organized with the support of the European Space Agency, the Space Research Organization of the Netherlands, the Japan Aerospace Exploration Agency, National Aeronautics and Space Administration (NASA), the Canadian Space Agency and the French Space Agency. We thank Drs. M. Viso, G. Gasset, B. Eche, and K. Fukui for their kind support in the space experiment. We acknowledge NASA Ames Research Center for preparation of CeMM. We also thank Dr. R. Morimoto for kindly supplying the Q35-YFP strain. We are grateful to N. Endo for his support with respect to fluorescence microscope observation. Some strains were provided by the *Caenorhabditis* Genetics Center funded by the National Institutes of Health National Center for Research Resources (NCRR) and the International *C. elegans* Gene Knockout Consortium and National Bioresource Project for the nematode. YH was supported by a grant-in-aid from the Ministry of Education, Science, and Culture of Japan and by the program “Grand-based Research Announcement for Space Utilization” promoted by the Japan Space Forum. NJS was supported by NIAMS AR054342.

## Author contributions

YH and SH conceived and designed the experiments. YH, SH, A. Higashibata, YM, YY, TK, A. Higashitani, and NI carried out the experiments and analysed the data. KK and TS prepared spaceflight experiments. NJS contributed materials. MT and NJS discussed results. YH and SH wrote the main manuscript text. NJS edited and all authors reviewed the manuscript.

## Additional information

Supplementary information accompanies this paper at <http://www.nature.com/scientificreports>

**Competing financial interests:** The authors declare no competing financial interests.

**License:** This work is licensed under a Creative Commons Attribution-NonCommercial-ShareAlike 3.0 Unported License. To view a copy of this license, visit <http://creativecommons.org/licenses/by-nc-sa/3.0/>

**How to cite this article:** Honda, Y. *et al.* Genes down-regulated in spaceflight are involved in the control of longevity in *Caenorhabditis elegans*. *Sci. Rep.* **2**, 487; DOI:10.1038/srep00487 (2012).