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## Ramatroban as a Novel Immunotherapy for COVID-19

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### Abstract

SARS-CoV-2 virus suppresses host innate and adaptive immune responses, thereby allowing the virus to proliferate, and cause multiorgan failure, especially in the elderly. Respiratory viruses stimulate cyclooxygenase-2 (COX-2) to generate prostanoids including Prostaglandin D<sub>2</sub> (PGD<sub>2</sub>) and thromboxane A<sub>2</sub>. Furthermore, PGD<sub>2</sub> concentrations in the airways increase with aging. PGD<sub>2</sub> action mediated via DP<sub>2</sub> receptors suppresses both innate and adaptive immune responses, by inhibiting interferon-λ and stimulation of myeloid monocyte-derived suppressor cells respectively. PGD<sub>2</sub> and thromboxane A<sub>2</sub> actions via the TP receptors activate platelets leading to a prothrombotic state. Ramatroban, a small-molecule antagonist of DP<sub>2</sub> and TP receptors, reverses viremia-associated proinflammatory, immunosuppressive<sup>5</sup> and prothrombotic processes which are similar to those induced by SARS-Cov-2. Ramatroban, used for the treatment of allergic rhinitis in Japan for the past 20 years has an excellent safety profile. Therefore, Ramatroban merits investigation as a novel immunotherapy for the treatment of COVID-19 disease.

### Keywords

SARS-CoV-2 Virus; Ramatroban; COVID-19; Immune Function

### Introduction

Novel coronavirus disease 2019, also known as COVID-19, is a highly infectious, rapidly spreading viral disease with very high morbidity and mortality [1]. The death rate with COVID-19 increases with advancing age, which could be due to several factors including

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Author Contributions

A. Gupta conceptualized, created the inventive concept and the framework for the manuscript; K. Kalantar-Zadeh and A. Gupta wrote the original draft; and all authors reviewed and edited.

Disclosure of Potential Sources of Conflict of Interest

A. Gupta has filed three provisional patent applications for use of PGD<sub>2</sub> and thromboxane A<sub>2</sub> antagonists, including ramatroban, as a treatment for COVID-19 (Application numbers: 63/003,286 filed on March 31, 2020; 63/005,205 filed on April 3, 2020; and 63/027,751 filed on May 2, 2020). Other authors have not declared conflict of interest. Ramatroban (Baynas®) was approved in Japan for allergic rhinitis in 2000.

progressive decline in immune function with aging [2]. Although SARS-CoV-2 and the 2003 SARS-CoV infections share a number of common clinical manifestations, SARS-CoV-2 virus appears to be highly efficient in person-to-person transmission and frequently cause asymptomatic infections, but the underlying mechanisms that confer the above characteristics of COVID-19 disease remain incompletely understood [3].

## Literature Review

SARS-CoV-2 and SARS-CoV are similar in cell tropism, with both targeting types I and II pneumocytes, and alveolar macrophages [3]. In mouse models of viral respiratory infections, airway epithelial cells produce prostaglandin D<sub>2</sub> (PGD<sub>2</sub>) in an age dependent manner, with higher levels in older mice [4]. There is increasing evidence that following respiratory virus infection, PGD<sub>2</sub> mediates airway inflammation while suppressing the host immune response to the virus [5]. We propose the hypotheses that higher basal production of PGD<sub>2</sub> in the airways of the elderly is causally linked to more severe disease with COVID-19; and that PGD<sub>2</sub> suppresses the innate and adaptive immune responses to SARS-CoV-2 allowing robust unchecked viral replication and virulence. We present evidence that Ramatroban, a small molecule antagonist of PGD<sub>2</sub> and thromboxane A<sub>2</sub>, currently approved and used in Japan for the treatment of allergic rhinitis, merits immediate investigation as a potential therapeutic agent against SARS-CoV-2.

### Respiratory viruses' upregulate PGD<sub>2</sub> production in lungs and airways, and PGD<sub>2</sub> regulates host immune responses to the virus

SARS-CoV-2 was found to be capable of infecting and replicating about 3 times more robustly than SARS-CoV in ex-vivo studies of human lung tissues [3]. The first line of defense against respiratory viruses, including influenza, rhino, respiratory syncytial, SARS-CoV and SARS-CoV-2 viruses are Type III interferons, including interferon- $\lambda$  (IFN- $\lambda$ ) [interleukin-28A/B (IL-28A/B)] [6]. In the respiratory tract, IFN- $\lambda$  expression is selective to mostly respiratory epithelial cells and dendritic cells. Both in vitro and in vivo studies have demonstrated that IFN- $\lambda$  is as effective as Type I interferons in anti-viral activity [7]. The role of interferons in differential replication rates has been studied. The 2003 SARS-CoV infection resulted in significant upregulation of types I (IFN $\beta$ ), II (IFN $\gamma$ ), and III (IFN $\lambda$ 1, IFN $\lambda$ 2, and IFN $\lambda$ 3) IFNs in human lung tissues, but SARS-CoV-2 infection did not significantly trigger the expression of any IFN [3,8]. Recent studies have demonstrated that interferon response to viral infections is regulated by local production of prostaglandins, especially prostaglandin D<sub>2</sub> [9].

Prostaglandins are known to play a key role in immune and inflammatory responses [10]. Respiratory viruses, including respiratory syncytial and SARS-CoV viruses, increase prostaglandin D<sub>2</sub> (PGD<sub>2</sub>) in the airways in pathogen dependent manner [4,9]. Infection with respiratory syncytial virus increased PGD<sub>2</sub> release by cultured human primary airway epithelial cells [9]. Intranasal administration of SARS-CoV virus to mice resulted in marked increase in PGD<sub>2</sub> levels in the lungs [11]. Moreover, PGD<sub>2</sub> production was elevated in nasopharyngeal samples from young infants hospitalized with RSV bronchiolitis, compared to healthy controls [9]. To the best of our knowledge PGD<sub>2</sub> levels have not been examined in

lung, airways or hematopoietic cells from patients with SARS-CoV or SARS-CoV-2 infections.

The increased generation of PGD<sub>2</sub> in response to viral infections is primarily mediated by upregulation of COX-2, phospholipase A<sub>2</sub> and PGD<sub>2</sub> synthase as reviewed here. Transcriptional activation of COX-2 is mediated by multiple mechanisms. Intratracheal administration of a synthetic double stranded viral RNA (dsRNA) to mimic viral infection induced gene expression of cyclooxygenase-2 (COX-2) by direct binding to the COX-2 promoter [5]. Additionally, the nucleocapsid protein of SARS-CoV activates the expression of COX-2 by binding directly to the regulatory elements for nuclear factor-kappa B and CCAAT/enhancer binding protein [12]. HIV-2 efficiently induces COX-2 transcription in human astrocytes through regulation of NF- $\kappa$ B p65/relA phosphorylation and transactivation [13]. COX-2 transcription is also induced directly or indirectly by inflammatory molecules released either as a result of viral infection per se or the host response to the virus [12]. Intranasal administration of SARS-CoV virus to mice resulted in marked increase in phospholipase A<sub>2</sub> expression and PGD<sub>2</sub> levels in the lungs.<sup>11</sup> Infection with respiratory syncytial virus up-regulated hematopoietic prostaglandin D synthase expression in cultured human primary airway epithelial cells [9]. Therefore, as reviewed here, it has been conclusively demonstrated that respiratory viruses upregulate PGD<sub>2</sub> production in the airways and the lungs.

Studies over the past decade have investigated the role of PGD<sub>2</sub> in regulating the innate and adaptive immune responses to the SARS-CoV viruses. PGD<sub>2</sub> signals primarily through three G-protein coupled receptors, first the D-prostanoid receptor 1 (DP<sub>1</sub>); second, prostanoid receptor 2 (DP<sub>2</sub>) which was identified previously as the “chemoattractant receptor-homologous molecule expressed on Th2 cells” (CRTH2); and third, the thromboxane receptor (TP) [14,15]. PGD<sub>2</sub> effects on cellular elements in the respiratory tree including pulmonary capillary endothelial cells, airway epithelial cells and cells of the innate and adaptive immune system are diverse and cell specific [4].

Werder RB, et al. have demonstrated in a neonatal mouse model of severe viral bronchiolitis that production of IFN- $\lambda$  is dependent on PGD<sub>2</sub>/DP<sub>2</sub> signaling; PGD<sub>2</sub>/DP<sub>2</sub> antagonism decreases viral load, immunopathology, and morbidity [9]. The beneficial effects of DP<sub>2</sub> blockade were associated with increased IFN- $\lambda$  (IL-28A/B) expression and were lost upon IFN- $\lambda$  neutralization [9]. This suggests that PGD<sub>2</sub>/DP<sub>2</sub> antagonists may be useful antivirals for the treatment of respiratory infections including SARS-CoV-2.

T cells are necessary for viral clearance, and, the development of robust T cell responses in the lung requires well-functioning respiratory dendritic cells (rDC) to process and present antigens, migrate to draining lymph nodes and stimulate adaptive immunity against the virus, including cell and humoral mediated immune responses. Experimental data suggests that increase in PGD<sub>2</sub> expression in mouse lungs following viral infection leads to impairment in rDC migration to mediastinal lymph nodes [4]. The production of PGD<sub>2</sub> increases with aging, which results in diminished T cell responses and a more severe clinical disease in older mice infected with respiratory viruses [4]. Furthermore, PGD<sub>2</sub> drives ‘group 2 innate lymphoid cells’ (ILC2) to secrete interleukin-13 (IL-13), which activates

“monocytic myeloid-derived suppressor cells (M-MDSCs)” to suppress downstream immune responses [16]. Blocking the PGD<sub>2</sub> pathway by a specific antagonist of the DP<sub>2</sub> receptor led to a decrease in ILC2 and M-MDSC cells, demonstrating that the ILC2/M-MDSC immunosuppressive axis is partly driven by high PGD<sub>2</sub> concentrations acting upon the DP<sub>2</sub> receptor on ILC2 cells (Figure 1).

It is important to note that production of PGD<sub>2</sub> by airway epithelial cells upon infection with respiratory viruses has salutary effects through the other PGD<sub>2</sub> receptor, D-prostanoid receptor (DP<sub>1</sub>), in that PGD<sub>2</sub>/DP<sub>1</sub> signaling upregulates interferon and accelerates viral clearance. There may be beneficial effects of PGD<sub>2</sub>/DP<sub>1</sub> system in acute lung injury. PGD<sub>2</sub>/DP<sub>1</sub> signaling tightens endothelial barrier function in lipopolysaccharide (LPS) induced acute lung injury, while DP<sub>2</sub> antagonism was not harmful [17]. The biological effects of PGD<sub>2</sub>/DP<sub>1</sub> axis in the respiratory system seem to be largely anti-inflammatory and opposed to the effects of PGD<sub>2</sub>/DP<sub>2</sub> signaling which has deleterious effects, as stated above [9,16]. Ramatroban is a selective blocker of DP<sub>2</sub> and TP receptors, and does not affect PGD<sub>2</sub> signaling through DP<sub>1</sub> receptor.

### **PGD<sub>2</sub>, a potential mediator of increased morbidity and mortality in COVID-19 disease.**

It is well known that during the aging process, immune functions decline, rendering the host more vulnerable to certain viruses. The morbidity and mortality from SARS-CoV-2 also increases with both aging and lymphopenia [18,19]. The mechanisms underlying this age-dependent susceptibility to viral infections are an active area of research. Stanley Perlman and colleagues have demonstrated that with aging, mice exhibit a higher basal PGD<sub>2</sub> levels in the airways and lungs, with 22-months old mice exhibiting PGD<sub>2</sub> levels that are 4–5 fold higher, compared to the 6-weeks old mice [4]. They further infected mice of various ages with SARS-CoV intranasally. While all 8 of the 22-month old mice died, all 14 of the 6-week old mice survived. Furthermore, increase in PGD<sub>2</sub> expression in mouse lungs upon aging correlated with a progressive impairment in respiratory dendritic cell (rDC) migration to mediastinal lymph nodes resulting in diminished T cell responses and more severe clinical disease in older mice [4]. Vijay et al have subsequently demonstrated that secreted phospholipase A<sub>2</sub> (PLA<sub>2</sub>) group IID (PLA<sub>2</sub>G2D) is critical in determining the impact of age on host susceptibility to SARS-CoV [11].

Furthermore, PGD<sub>2</sub> via DP<sub>2</sub> signaling may have a pro-inflammatory effect by increasing the production of monocyte chemoattractant protein 1 (MCP-1) and interleukin-6 (IL-6), while IL-6 may promote virus survival and/or exacerbation of clinical disease [20,21].

### **Ramatroban, an antagonist of PGD<sub>2</sub>/DP<sub>2</sub> and Thromboxane/TP axis, as a novel immunotherapeutic drug for COVID-19.**

Downregulation of the innate and adaptive immune responses to respiratory viruses is mediated by PGD<sub>2</sub>/DP<sub>2</sub> signaling, as discussed above. This suggests that a selective blockade of PGD<sub>2</sub>/DP<sub>2</sub> signaling without blocking DP<sub>1</sub>, if effectively operational, can favorably accelerate viral clearance and reduce immunopathology and morbidity. Ramatroban is a potent but reversible antagonist of PGD<sub>2</sub>/DP<sub>2</sub> receptors, while sparing the DP<sub>1</sub> receptors [14,22,23]. Ramatroban is also a potent antagonist of thromboxane receptors

(TP), and inhibits tumor necrosis factor and platelet activating factor induced expression and production of monocyte chemoattractant protein-1 (MCP-1), and expression of adhesion molecules in human endothelial cells, while reducing inflammation.<sup>22</sup> Ramatroban enhances vascular response to acetylcholine, and has an inhibitory effect on vascular smooth muscle contraction and platelet aggregation [22]. Ramatroban reduces myocardial infarct size, and prevents neointimal formation after balloon arterial injury in hypercholesterolemic rabbits [22,24].

Ramatroban exhibits a large safety factor. The usual dose of ramatroban is 50 to 150 mg orally, twice a day. The intravenous LD<sub>50</sub> values in mice and rabbits were > 600 and > 100 mg/kg respectively, while no dogs died with an intravenous dose of 250 mg/kg [22]. In the 12-months toxicity study in dogs, no toxicologically important changes were observed in any dog given up to 30 mg/kg/day of ramatroban. In this study plasma concentration of ramatroban in animals at 2-hours after oral administration of 30 mg/kg of the drug was between 11.9 to 32.7 mg/mL, while C<sub>max</sub> in healthy adult male volunteers given 75 mg of ramatroban twice daily (usual clinical dose) was about 0.4 mg/mL. Accordingly, the doses tested were judged to be sufficiently high to indicate clinical safety of ramatroban in humans. As a thromboxane A<sub>2</sub> antagonist, ramatroban has been used for the treatment of allergic diseases. The clinical efficacy and safety of ramatroban have been demonstrated in clinical studies as treatment of allergic rhinitis, and for the past 2 decades ramatroban (Baynas®) has been marketed in Japan for this indication. COVID-19 is characterized by a prothrombotic state including disseminated intravascular coagulopathy (DIC), thrombosis and infarctions which are associated with poor outcomes and higher mortality [25,26]. The anti-platelet action of ramatroban, as a thromboxane A<sub>2</sub> antagonist, could potentially reduce thrombotic events in patients with COVID-19 disease. Therefore, ramatroban by its DP<sub>2</sub> antagonism can potentially help restore the IFN-λ, T and B cell responses that have been suppressed by the SARS-Co-V2 virus in COVID-19 disease, especially in the elderly; and help control inflammation by inhibiting production of IL-6. Furthermore, ramatroban, as a potent thromboxane A<sub>2</sub> antagonist can potentially reduce the severity of coagulopathy, a sequela of severe COVID-19 disease.

## Discussion and Conclusion

COVID-19 disease in more severe cases is characterized by immune suppression, inflammation and a prothrombotic state (Figure 1). Prostaglandin D<sub>2</sub> has been found to be a key mediator of immunosuppressive effects in animal models of SARS-CoV infection. Ramatroban selectively antagonizes the actions of thromboxane A<sub>2</sub> via the TP receptors and PGD<sub>2</sub> via the DP<sub>2</sub> and TP receptors while sparing PGD<sub>2</sub>/DP<sub>1</sub> signaling (Table 1). Hence, Ramatroban, holds great potential for restoring or enhancing immune function in patients with COVID-19 disease, especially in the elderly patients. Ramatroban is expected to decrease inflammation, improve endothelial function, inhibit thrombosis and improve outcomes. Ramatroban has excellent safety profile. Therefore, Ramatroban is a highly promising therapy for patients with COVID-19 infection. Urgent and fast-track clinical trials are needed to investigate the efficacy and safety of Ramatroban across different levels of severity of COVID-19 disease. This requires cooperation between scientists, industry and governmental agencies globally.

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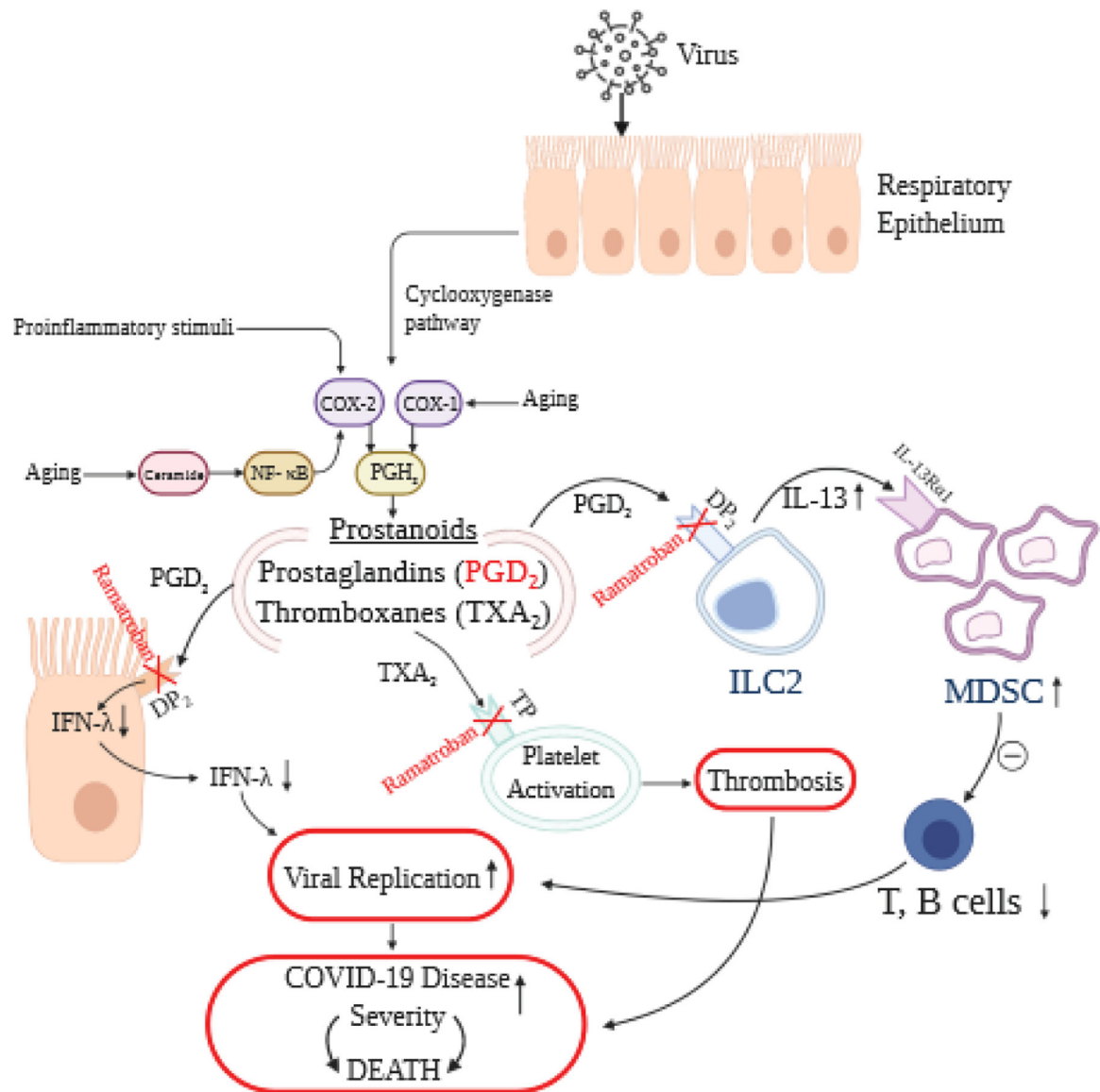
## Abbreviations:

<b>PGD<sub>2</sub></b>	Prostaglandin D <sub>2</sub>
<b>DP<sub>1</sub></b>	D-Prostanoid Receptor 1
<b>DP<sub>2</sub></b>	D-Prostanoid Receptor 2
<b>TP Receptor</b>	Thromboxane Receptor
<b>IFN-λ</b>	Interferon-λ [interleukin-28A/B (IL-28A/B)]
<b>IP3</b>	Inositol Trisphosphate ILC2, Group 2 Innate Lymphoid Cells
<b>M-MDSC</b>	Monocytic Myeloid-Derived Suppressor Cells
<b>COX</b>	cyclooxygenase
<b>Phospholipase A<sub>2</sub></b>	(PLA <sub>2</sub> ) Group IID (PLA <sub>2</sub> G2D)
<b>rDC</b>	Respiratory Dendritic Cell

## References

1. Grasselli Giacomo, Zangrillo Alberto, Zanella Alberto and Antonelli Massimo, et al. "Baseline Characteristics and Outcomes of 1591 Patients Infected with SARS-CoV-2 Admitted to ICUs of the Lombardy Region, Italy". *JAMA* 323(2020): 1574–1581.
2. Montecino-Rodriguez Encarnacion, Berent-Maoz Beata, and Kenneth Dorshkind. "Causes, Consequences, and Reversal of Immune System Aging". *J Clin Invest* 123(2013): 958–965. [PubMed: 23454758]
3. Chu Hin, Jasper Fuk-Woo Chan, Yixin Wang and Terrence Tsz-Tai Yuen, et al. "Comparative Replication and Immune Activation Profiles of SARSCoV-2 and SARS-CoV in Human Lungs: An *ex vivo* Study with Implications for the Pathogenesis of COVID-19". *Clin Infect Dis* 2(2020): 2–4.
4. Zhao Jincun, Zhao Jingxian, Legge Kevin and Perlman Stanley. "Age-Related Increases in PGD(2) Expression Impair Respiratory DC Migration, Resulting in Diminished T Cell Responses upon Respiratory Virus Infection in Mice". *J Clin Invest* 121(2011): 4921–4930. [PubMed: 22105170]
5. Shiraiishi Yoshiki, Asano Koichiro, Niimi Kyoko and Fukunaga Koichi, et al. "Cyclooxygenase-2/ Prostaglandin D2/CRTH2 Pathway Mediates Double-Stranded RNA-Induced Enhancement of Allergic Airway Inflammation". *J Immunol* 180(2008): 541–549. [PubMed: 18097056]
6. Andreakos Evangelos, Salagianni Maria, Galani Ioanna E and Ourania Koltsida. "Interferon-Lambdas: Front-Line Guardians of Immunity and Homeostasis in the Respiratory Tract". *Front Immunol* 8(2017): 1232. [PubMed: 29033947]
7. Lazear Eric, Ghasemi Reza, Hein Sarah M and Westwick John, et al. "Targeting of IL-2 to Cytotoxic Lymphocytes as an Improved Method of Cytokine-Driven Immunotherapy". *Oncoimmunol* 6(2017): e1265721.
8. O'Brien TR, Thomas DL, Jackson SS and Prokunina-Olsson L, et al. "Weak Induction of Interferon Expression by SARS-CoV-2 Supports Clinical Trials of Interferon Lambda to Treat Early COVID-19". *Clin Infect Dis* 2(2020): 2–4.

9. Werder Rhiannon B, Lynch Jason P, Simpson Jennifer Cand Zhang Vivian, et al. "PGD2/DP2 Receptor Activation Promotes Severe Viral Bronchiolitis by Suppressing IFN-Lambda Production". *Sci Transl Med* 10(2018): 2–4.
10. Ricciotti Emanuela and FitzGerald Garret A. "Prostaglandins and Inflammation". *Arterioscler Thromb Vasc Biol* 31(2011): 986–1000. [PubMed: 21508345]
11. Vijay Rahul, Hua Xiaoyang, Meyerholz David K and Miki Yoshimi, et al. "Critical Role of Phospholipase A2 Group IID in Age-Related Susceptibility to Severe Acute Respiratory Syndrome-CoV Infection". *J Exp Med* 212(2015): 1851–1868. [PubMed: 26392224]
12. Yan Xiaohong, Hao Qian, Mu Yongxin and Timani Khalid Amine, et al. "Nucleocapsid Protein of SARS-CoV Activates the Expression of Cyclooxygenase-2 by Binding Directly to Regulatory Elements for Nuclear Factor-Kappa B and CCAAT/Enhancer Binding Protein". *Int J Biochem Cell Biol* 38(2006): 1417–1428. [PubMed: 16546436]
13. Álvarez Susana, Blanco Almudena, Kern Florian and Fresno Manuel, et al. "HIV-2 Induces NF- $\kappa$ B Activation and Cyclooxygenase-2 Expression In Human Astroglial Cells". *Virol* 380(2008): 144–151.
14. Kupczyk Maciej and Kuna Piotr. "Targeting the PGD2/CRTH2/DP1 Signaling Pathway in Asthma and Allergic Disease: Current Status and Future Perspectives". *Drugs* 77(2017): 1281–1294. [PubMed: 28612233]
15. Brightling Christopher E, Guy Brusselleand Pablo Altman. "The impact of the Prostaglandin D 2 receptor 2 and its Downstream Effects on the Pathophysiology of Asthma". *Allergy* 75(2020): 761–768. [PubMed: 31355946]
16. TrabANELLI Sara, Chevalier Mathieu F, Amaia Martinez-Usatorre and Alejandra Gomez-Cadena, et al. "Tumour-Derived PGD2 and NKp30-B7H6 Engagement Drives an Immunosuppressive ILC2-MDSC Axis". *Nat Commun* 8(2017): 593. [PubMed: 28928446]
17. Murata Takahisa, Aritake Kosuke, Tsubosaka Yoshiki and Maruyama Toshihiko, et al. "Anti-inflammatory Role of PGD2 in Acute Lung Inflammation and Therapeutic Application of its Signal Enhancement". *Proc Natl Acad Sci USA* 110(2013): 5205–5210. [PubMed: 23479612]
18. Mahase Elisabeth. "Covid-19: Death Rate is 0.66% and Increases with Age, Study Estimates". *BMJ* 2(2020): m1327.
19. Tan Li, Wang Qi, Zhang Duanyang and Ding Jinya, et al. "Lymphopenia Predicts Disease Severity of COVID-19: A Descriptive and Predictive Study". *Sig Transd Targeted Ther* 5(2020): 1–3.
20. Peeraully Muhammad R, Henning Sievert, Mònica Bulló and Bohan Wang, et al. "Prostaglandin D2 and J2-series (PGJ2, 12-PGJ2) Prostaglandins Stimulate IL-6 and MCP-1, but Inhibit Leptin, Expression and Secretion by 3T3-L1 Adipocytes". *Pflügers Archiv* (2006): 177–187. [PubMed: 16924534]
21. Velazquez-Salinas Lauro, Antonio Verdugo-Rodriguez, Rodriguez Luis Land Borca Manuel V. "The Role of Interleukin 6 during Viral Infections". *Front Microbiol* 10(2019): 1057. [PubMed: 31134045]
22. Ishizuka Toshiaki, Matsui Takemi, Okamoto Yasuhiro and Ohta Atsuko, et al. "(BAY u 3405): A Novel Dual Antagonist of TXA2 Receptor and CRTh2, A Newly Identified Prostaglandin D2 Receptor". *Cardiovasc Drug Rev* 22(2004): 71–90. [PubMed: 15179446]
23. Ulven Trond and Kostenis Evi. "Minor Structural Modifications Convert the Dual TP/CRTH2 Antagonist Ramatroban into a Highly Selective and Potent CRTH2 Antagonist". *J Med Chem* 48(2005): 897–900. [PubMed: 15715457]
24. Fiedler Volker B, Elisabeth Perzbornand Friedel Seuter. "Protective Effect of a Novel Thromboxane Antagonist, BAY-U3405, on Canine Myocardial Damage after Coronary Artery Occlusion and Reperfusion". *Pharmacother* 11(1991): 77–84.
25. Wang Janice, Hajizadeh Negin, Moore Ernest E and McIntyre Robert C, et al. "Tissue Plasminogen Activator (tPA) Treatment for COVID-19 Associated Acute Respiratory Distress Syndrome (ARDS): A Case Series". *J Thromb Haemost* 2(2020): 2–4.
26. Zhang Yan, Xiao Meng, Zhang Shulan and Xia Peng, et al. "Coagulopathy and Antiphospholipid Antibodies in Patients with Covid-19". *N Engl J Med* 382(2020): e38. [PubMed: 32268022]



**Figure 1.** COVID-19 disease characterized by immune suppression, inflammation and a prothrombotic state.



**Table 1.**

Potential effects of Ramatroban on prostanoid signaling and anticipated biological effects.

Prostanoid / Receptor [14,15]	Signal Transduction [14]	Effect of Prostanoid on Effector Cells and Cytokines [4,9,10,16]	Effect on Airway Inflammation [5,16]	Effect of Ramatroban [14,22]
PGD <sub>2</sub> /DP <sub>2</sub>	•	IFN- $\lambda$ ↓		
	•	ILC2 ↑		
	•	MDSC ↑		
	•	cAMP ↓	Airway inflammation to virus ↑ (chemotaxis, WBC infiltration)	Inhibits
	•	Ca ↑	Dendritic cell function ↓	
	•		T & B cell function ↓ Lymphocyte count ↓	
Thromboxane A <sub>2</sub> /TP	•	IP3 ↑	Activation and aggregation of platelets	
	•	Ca ↑	Arterial and venous thrombosis	Inhibits
	•		DIC	
PGD <sub>2</sub> /DP <sub>1</sub>	•	cAMP ↑	Thymic function ↑	
	•		IFN- $\lambda$ ↑	No effect
			Airway inflammation ↓	