## **Short Communication**

# Prevalence of *Bartonella henselae* and *Bartonella clarridgeiae* in cats and dogs in Korea

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Blood, saliva, and nail samples were collected from 54 dogs and 151 cats and analyzed for the presence of *Bartonella henselae* with a novel nested polymerase chain reaction (PCR) method. *Bartonella* (*B.*) *henselae* was detected in feral cat blood (41.8%), saliva (44.1%), and nail (42.7%) samples. *B. henselae* was also detected in pet cat blood (33.3%), saliva (43.5%), and nail (29.5%) samples and in pet dog blood (16.6%), saliva (18.5%), and nail (29.6%) samples. Nine samples were infected with *B. clarridgeiae* and 2 were co-infected with *B. henselae* and *B. clarridgeiae* of blood samples of dogs. This report is the first to investigate the prevalence of *B. henselae* and *B. clarridgeiae* in dogs and cats in Korea, and suggests that dogs and cats may serve as potential Bartonella reservoirs.

Keywords: Bartonella, cats, cat-scratch disease, dogs, Korea

# Introduction

The genus *Bartonella* (*B.*) includes at least 20 species and subspecies, and several of these are human pathogens [22]. Clinical manifestations of Bartonella infection include Carrion's disease, trench fever, cat scratch disease, bacillary angiomatosis, endocarditis, chronic bacteremia, neuroreti nitis, and osteomyelitis [13]. Cat scratch disease is zoonotic and primarily caused by *B. henselae* [15]. *B. clarridgeiae* can also cause cat scratch fever. *B. henselae* and *B. clarridgeiae* can also infect dogs [4,7], and both species can function as bacterial reservoirs for infection [5,9,10, 18,19]. Cat scratch disease was recently reported in a woman with a pet dog in Korea [5]. However, the prevalence of *Bartonella* spp. from companion animals in Korea has

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not been previously investigated. We examined the prevalence of *B. henselae* and *B. clarridgeiae* in dogs and cats in the present study using a recently developed nested PCR method.

Blood, saliva, and nail samples were collected from healthy pet dogs (n = 54) and cats (n = 48) at the Veterinary Medical Teaching Hospital of Seoul National University, Korea. All samples were collected from November 2005 to July 2006. Feral cats (n = 103) were captured in neighborhoods throughout Seoul and were isolated in an animal shelter. B. henselae strain Houston-1 (ATCC 49882) and B. clarridgeiae strain (ATCC 51734) were obtained from the American Type Culture Collection (USA) and used for positive control samples. Genomic DNA was extracted using Genomic Blood DNA and Genomic Cell/ Tissue DNA Extraction Kits (iNtRoN Biotechnology, Korea), per the manufacturer's instructions. Primary PCR was performed with the P-bhenfa (5'-TCTTCGTTTCTCT TTCTTCA-3') and P-benr1 (5'-CAAGCGCGCGCTCTA ACC-3') primers which amplified B. henselae (186 bp) and B. clarridgeiae (168 bp) fragments. Nested PCR amplified B. henselae (152 bp) and B. clarridgeiae (134 bp) fragments with the N-bhenf1a (5'-GATGATCCCAAG CCTTCTGGC-3') and N-bhenr (5'-AACCAACTGAGC TACAAGCC-3') primers [15]. Primary and nested PCR reactions were performed as previously described [15].

All PCR products were analyzed by sequencing with an automated sequencer ABI 3100 Genetic Analyzer (Bionics, Korea) and results were confirmed to be from *B. henselae* (GeneBank access number DQ000494) and from *B. clarridgeiae* (GeneBank access number: DQ003029).

*B. henselae* was detected in 14.2% of blood samples (14/98), 3.9% of saliva samples (4/102), and 4.8% of nail samples (5/103) from feral cats. In contrast, only 6.3% (3/48) of blood samples from pet cats were positive for *B. henselae*. *B. henselae* was not detected in pet cat saliva samples (n = 46), pet cat nail samples (n = 44), or in any pet

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#### dog samples (n = 54).

*B. henselae* was detected in 41.8% (41/98) of blood samples, 44.1% (45/102) of saliva samples, and 42.7% (44/103) of nail samples from feral cats by nested PCR. In addition, 33.3% of blood samples (16/48), 43.5% of saliva samples (20/46), and 29.5% of nail samples (13/44) from pet cats were *B. henselae* positive. *B. henselae* DNA was also detected in 16.6% (9/54) blood samples, 18.5% (10/54) of saliva samples and 29.6% (16/54) of nail samples from dogs (Table 1).

*B. clarridgeiae* was detected in 2 feral cat blood samples, a feral cat saliva sample, 3 dog blood samples, a dog saliva sample, and 2 dog nail samples. Additionally, 2 samples (1 dog blood and 1 dog nail) were co-infected with *B. henselae* and *B. clarridgeiae* (Table 2). PCR product and DNA sequencing data are shown in Fig. 1.

Cats are usually the main zoonotic reservoir for *Bartonella* infection [14], although dogs may also serve as zoonotic reservoirs secondary to *B. henselae* and *B. clarridgeiae* infection [4,7]. Cat scratch disease was identified in a case with suspected human:canine transmission in Korea [5,18]. However, there are no current surveys evaluating *Bartonella* spp. prevalence in cats and dogs.

A previous study reported that 39% of cats were *B. henselae* positive among a population of 146 cats in Japan [11], and this result was significantly higher than the previous 7.2% prevalence among cats in Japan. Previous studies conducted in various countries identified higher Bartonella bacteremia prevalence in shelter cats than in pet cats [2,3,8,10]. The Bartonella prevalence in pet cats in the prevalence in previous studies, including Germany (13%), France (11%), and the Netherlands (22%) [1,5,17].

Conversely, the prevalence of *B. henselae* in sheltered cats (41.8%) was similar to the prevalence identified in other studies. These findings suggest that pet cats may serve as a reservoir for *B. henselae* infection to their owners. This is particularly relevant to immunocompromised pet owners.

*B. henselae* prevalence in cats is higher than *B. clarridgeiae* prevalence [16], but this may be dependent on age, sex, and type of breeding [6]. The *B. henselae* prevalence in cats and dogs was greater than *B. clarridgeiae* and was higher in cats than in dogs. These results supported previous studies which suggested that *B. henselae* was the major zoonotic pathogen. A recent survey of Bartonella seropositive healthy blood donor in Sweden demonstrated a similar prevalence to



**Fig. 1.** *Bartonella* (*B*.) *henselae* (S2) and *B. clarridgeiae* (S1) nested PCR amplification bands from 2 cats. The negative control band (S3) is visualized on the right side. M: standard maker.

Table 1. Prevalence of Bartonella henselae infection in cat and dog blood, saliva, and nail samples detected by nested PCR

Samples	Number of positive samples / Number of tested samples (%)			
	Feral cats	Pet cats	Subtotal (Feral + Pet)	Dogs
Blood	41/98 (41.8)	16/48 (33.3)	57/146 (39.0)	9/54 (16.6)
Saliva	45/102 (44.1)	20/46 (43.5)	65/148 (43.9)	10/54 (18.5)
Nails	44/103 (42.7)	13/44 (29.5)	57/147 (38.8)	16/54 (29.6)

Table 2. Prevalence of Bartonella clarridgeiae infection in cat and dog blood, saliva, and nail samples detected by nested PCR

Samples	Number of positive samples / Number of tested samples (%)			
	Feral cats	Pet cats	Subtotal (Feral + Pet)	Dogs
Blood	2/98 (2.04)	0/48 (0.00)	2/146 (1.37)	3/54 (5.56)
Saliva	2/102 (1.96)	0/46 (0.00)	2/148 (1.35)	1/54 (1.85)
Nails	0/103 (0.00)	0/44 (0.00)	0/147 (0.00)	1/54 (1.85)

dogs in the present study [12]. Zoonotic diseases have become an increasingly important public health concern [5]. Our results suggest that *B. henselae* and *B. clarridgeiae* are highly prevalent in Korean cats and dogs. Further, cats and dogs may serve as reservoirs for human Bartonella infection.

In conclusion, data from the present study suggests that Bartonella infection prevalence in Korean shelter cats is similar to those of previously described countries. However, the prevalence of *B. henselae* in Korean pet cats was higher than reported prevalence in other countries. This is the first report examining the prevalence of *B. henselae* and *B. clarridgeiae* infection in domestic cats and dogs in Korea.

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