SCIENTIFIC REPORTS

OPEN

Received: 27 April 2016 Accepted: 01 July 2016 Published: 26 July 2016

Hydrogen sulfide mediates the protection of dietary restriction against renal senescence in aged F344 rats

Wen-juan Wang^{1,2}, Guang-yan Cai¹, Yi-chun Ning^{1,2}, Jing Cui¹, Quan Hong¹, Xue-yuan Bai¹, Xiao-meng Xu^{1,2}, Ru Bu¹, Xue-feng Sun¹ & Xiang-mei Chen¹

Renal aging is always accompanied by increased oxidative stress. Hydrogen sulfide (H_2S) can be upregulated by 50% dietary restriction (DR) for 7-day and can block mitochondrial oxidative stress. H_2S production exerts a critical role in yeast, worm, and fruit fly models of DR-mediated longevity. In this study, we found that renal aging could be attenuated by 30% DR for 6-month (DR-6M) and life-long (DR-LL), but not for 6-week (DR-6W). The expressions of cystathionine- γ -lyase (CGL) and cystathionine- β - synthase (CBS) were improved by DR-6M and DR-LL. Endogenous H_2S production shared the same trend with CBS and CGL, while glutathione (GSH) didn't. When comparing efficiencies of DR for different durations, more evident production of H_2S was found in DR-6M and DR-LL than in DR-6W. Finally the level of oxidative stress was improved by DR-6M and DR-LL rather than by DR-6W. It concluded that aged rats had the ability to produce enough H_2S on 30% DR interventions protecting against renal aging, and the effect of DR for long-term were more significant than that of DR for short-term.

Increasing age is an independent risk factor for chronic kidney disease¹. For the kidney structure, aging is associated with decreased kidney weight, vascular sclerosis, tubular atrophy and interstitial fibrosis; For kidney function, aging not only aggravates the declining processes of the glomerular filtration rate, urinary sodium excretion and erythropoietin production, but also leads to increased glomerular capillary pressure and susceptibility to nephrotoxic injury, including drug-induced renal damage¹⁻³. All of these results suggest that delaying or reversing the process of renal aging is necessary to reduce the incidence of age-related kidney dysfunctions and pathological changes.

In 1957, the free radical theory of aging was first proposed. Free radical attack caused macromolecular impairments and accelerated the aging progression, laying the foundation for oxidative stress⁴. In 1990, Sohal *et al.* established that oxidative stress was a causal factor in differentiation and aging⁵. Oxidative stress refers to an imbalance in reduction-oxidation reactions. Once reactive oxygen species (ROS) can't be effectively cleared, they will produce various cascade reactions that lead to tissue damage and induce or accelerate physiological aging⁶⁷. NOX2/gp91, as the first identified factor of nicotinamide adenine dinucleotide phosphate oxidase (NOX) which transfers electrons across biological membranes, is responsible for the generation of ROS⁸. ROS avidly reacts with a large number of molecules, including small inorganic molecules as well as lipids, proteins, nucleic acids, and carbohydrates. As a result, it leads to significant increases in malondialdehyde (MDA), protein carbonyl (PC), 8-hydroxygluanine (8-OHdG), and some other oxidation products^{8,9}. What's more, oxidative stress also induces apoptosis increase, mitochondrial autophagy decrease, erythropoietin production restriction, and sodium homeostasis disorder in kidneys^{10,11}. All the data above show that oxidative stress participates in the process of renal aging and age-related alternations directly and indirectly.

Currently, it is widely accepted that DR, as a natural regimen of non-genetic transformation without malnutrition, brings numerous and beneficial effects, especially in extending the maximum and mean lifespan of a variety of organisms, from yeast to humans¹². DR encompasses various forms, including the reduction of 30% to

¹Department of Nephrology, Chinese PLA General Hospital, Chinese PLA Institute of Nephrology, State Key Laboratory of Kidney Diseases, National Clinical Research Center for Kidney Diseases, Beijing 100853, China. ²School of Medicine, Nankai University, TianJin 300071, China. Correspondence and requests for materials should be addressed to G.-Y.C. (email: caiguangyan@sina.com)



Figure 1. Pathway of the Krebs-Hanseleit cycle and TSP, which were responsible for productions of H₂S, GSH and some other substances mainly via CBS and CGL. When cystathionine is in the state of lack, it will cause a compensatory enhancement of CGL and CBS in TSP, as a result of increased expression of H₂S. SAM, S-adenosylmethionine; SAH, S-adenosylhomocysteine; CBS, cystathionine beta-synthase; CGL, cystathionine gamma-lyase; TSP, transsulfuration pathway; H₂S, hydrogen sulfide; GSH, glutathione.

50% of total calorie, protein and even essential amino acid intake for short-term or for life-long^{11,13}. Specifically restriction of essential amino acids (EAAs), especially Met, controls the benefits of longevity in diverse organisms, such as yeast, files, worms, elegans and rodents¹⁴⁻¹⁷. Lifespan is reduced by the addition back of some amino acids, in particular sulfur amino acid (SAA), indicating that restriction of SAA takes on a common role to mediate numerous benefits of DR¹⁵. Recently most of the biological mechanisms that underlie DR are focused on nutrient-sensing pathway activity, including the NAD⁺/sirtuins pathway¹⁸, adenosine monophosphate-activated protein kinase¹⁹, mammalian target of rapamycin²⁰, and insulin-like growth factor²¹. Similarly, SAA restriction extends lifespan in many species not only by altering insulin-like growth factor I, glucose, and insulin levels, but also by increasing the level of macrophage migration inhibition factor and the capacity of stress resistance¹⁶. Metformin, as an indispensable molecule regulating energy metabolism, retards aging in C. elegans mainly by altering methionine metabolism and microbial folate, indicating that transsulfuration pathway (TSP) has an essential role in the protection of metformin against aging¹⁷. TSP is responsible for methionine metabolism to produce endogenous H_2S . Zhang *et al.* reviewed that H_2S delayed the progression of aging mainly by inhibiting oxidative stress, activating silent information regulator of transcription 1 (SIRT1), and probably suppressing the expression of klotho²². Recently, Hine *et al.* found that TSP and H_2S mediated the beneficial effects of DR in the hepatic and renal ischemia-reperfusion models, and they also speculated that H₂S could participate in delaying aging²³. Based on these data, it comes to the conclusion that increased TSP activity is an evolutionarily conserved effector to multiple DR regimens, including protein restriction, caloric restriction, and SAA restriction, and that increased H₂S production under these conditions indicates a common molecular mechanism underlying multiple DR benefits.

 H_2S has been gaining increasing attention as a third signal gasotransmitter, following nitric oxide and carbon monoxide. It exerts beneficial effects mainly through antioxidant activity at physiological levels^{12,24}. It has been reported that H_2S and GSH are endogenously generated by CGL and CBS in TSP^{23,25,26}. The expressions of CBS and CGL increase when Cys is low to allow de novo synthesis from Met (Fig. 1)²⁷. H_2S influences the normal physiology of kidney as well as the pathogenesis of kidney diseases^{25,28}. It has been suggested that the elevated H_2S concentration in kidneys not only mediates the metabolism of homocysteine, but also regulates vascular and tubular functions, along with increased renal blood flow, glomerular filtration rate and urinary excretion²⁶. Because of its diverse properties and systemic effects, H_2S dysfunction has recently been identified as a key factor in the onset and progression of renal diseases, such as ureteral obstruction²⁹, chronic kidney disease³⁰, drug-induced nephrotoxicity³¹ and renal ischemia reperfusion injury³². Existing evidences show that H_2S is involved in delaying the progression of aging and age-associated diseases by inhibiting oxidative stress, activating SIRT1, and probably exerting indirect effects on anti-aging gene klotho²². Therefore, it would be interesting to investigate whether aged rats have the ability of producing sufficient H_2S on the intervention of DR for different durations to delay the process of aging.

This study, for the first time, observed whether aged rats had the ability to produce enough H_2S on the intervention of 30% DR in delaying kidney aging, and compared the efficiencies of DR for different durations (6-week, 6-month, and life-long) on the production of H_2S . Firstly, we observed that metabolic indexes, renal function, renal histology alternations and senescence markers on 30% DR for different durations. Secondly, we systematically detected the changes of both expressions and localizations of CBS and CGL in TSP and measured the levels of H_2S and GSH. Finally we observed the levels of ROS, oxidation products and anti-oxidative indicators. We found that aged rats had the ability to produce enough H_2S , but not GSH, to delay the process of aging, and it's more significant in DR-6M and DR-LL groups than in DR-6W group. DR-induced H_2S production would have an ROS-scavenging ability and antioxidant-improving properties to attenuate stress-induced senescence in the aged kidneys to some extent.

Results

Animal characteristics. Compared with young ad libitum (Young-AL), the body weight, serum urea nitrogen, the urine protein/urine creatinine ratio, triglycerides, and serum glucose in old ad libitum (Old-AL) were significantly increased (see Supplementary Table S1, p < 0.05), while the kidney weight/body weight ratio in

Old-AL was decreased (see Supplementary Table S1, p < 0.05). There was no change in the levels of serum creatinine, serum cholesterol, total protein and albumin in two groups. Compared with corresponding AL control group, significant improvements in the levels of body weight, kidney: body weight, serum urea nitrogen, urine protein/urine creatinine ratio, triglycerides, serum glucose were found in both DR-6M and DR-LL (see Supplementary Table S2, p < 0.05), but there was no obvious difference in other indexes (see Supplementary Table S2, p > 0.05). While it showed no obvious changes of all the indexes in DR-6W group (see Supplementary Table S2, p > 0.05). Compared with the effect of DR-6W in improving kidney weight/body weight and triglycerides, it was more prominent in DR-LL than in DR-6M (see Supplementary Table S2, p < 0.05).

Kidney structural alterations. Renal tissues were processed by routine Periodic Acid-Schiff (PAS) staining. Specific morphological changes and pathological grading were shown in Supplementary Figs S1 and S2. Glomerular lesions and interstitial renal tubular damage, such as glomerulosclerosis, fibrous, cell proliferation, renal tubule atrophy, renal tubular epithelial cell degeneration, renal tubular casts and inflammatory cell infiltrations, significantly increased in Old-AL (see Supplementary Fig. S1, p < 0.05). Compared with AL corresponding, renal tubular epithelial cell degeneration and glomerulosclerosis were improved in both DR-6M and DR-LL (see Supplementary Fig. S2, p < 0.05) rather than DR-6W (see Supplementary Fig. S2, p > 0.05). Some interstitial fibrosis, cell proliferation, loop necrosis and inflammatory cell infiltrations were largely blunted largely by DR-LL and partly by DR-6M (see Supplementary Fig. S2, p > 0.05).

Senescence markers in aged rat renal tissues. In this study, both the expression of p16, as a possible effector and a robust biomarker in mammalian aging, and the expression of p21, as a cell cycle inhibitor and the most extensive kinase inhibitor, were higher in Old-AL than in Young-AL (Fig. 2a–d, p < 0.05). To observe the effects of DR for different durations on aged kidney, the expressions of p16 and p21 in aged rats from different groups were further detected. As shown in Fig. 2, the expression of p16 protein could be reversed by both DR-6M and DR-LL, but not by DR-6W (Fig. 2e,f, p < 0.05). Similarly cell cycle inhibitor p21 showed the same trend (Fig. 2g,h, p < 0.05).

Alternations of capital enzymes, H_2S , and GSH in TSP. Firstly we observed the expressions of the capital enzymes in the TSP. There was little difference in the expressions of both CGL and CBS in DR-6W (Fig. 3a,b, p > 0.05). It also showed that there were increases in both CGL and CBS in DR-6M (Fig. 3c,d, p < 0.05) and DR-LL (Fig. 3e,f, p < 0.05). We further observed the trends in the endogenous H_2S -related enzymes in groups of DR for different durations together. Compared with the corresponding AL groups, the levels of H_2S -related enzymes were respectively improved by DR-6M and DR-LL rather than by DR-6W (Fig. 4a,b, p < 0.05); Compared with DR-6W, it took on a more significant improvement in DR-LL than DR-6M, which demonstrated that the expressions of the TSP enzymes were increased in a time-dependent mode to some extent (Fig. 4a,b, p < 0.05). We also determined the expressions of CGL and CBS in aged kidneys with an immunohistochemistry stain, which showed the remarkable alterations and locations of these two enzymes (Figs 5 and 6). Consistent with previous findings, both CGL and CBS were primarily expressed in renal tubular epithelial cells, whereas they were hardly expressed in the glomerulus.

We applied enzyme-linked immunosorbnent assay to quantify H_2S and GSH in aged kidney tissues and found that the expression of H_2S was improved by DR-6M and DR-LL (Fig. 7a, p < 0.05) while the expression of GSH was not (Fig. 7b, p > 0.05). Compared with DR-6W, the effect of DR-LL on H_2S was more substantial than that of DR-6M (Fig. 7a, p < 0.05).

Oxidant properties and antioxidant capacity. Western blot analysis showed that the expressions of NOX2/gp91 were significantly reduced in DR-6M and DR-LL (Fig. 8a,b, p < 0.05), and the production of ROS showed a similar trend as that of gp91/NOX2 (Fig. 8c, p < 0.05). Then we detected oxidant properties (MDA, and PC) and anti-oxidant indicators (catalase, CAT and total superoxide dismutase, T-SOD) in aged kidneys. It showed that the levels of MDA and PC had been reduced by DR-6M and DR-LL (Fig. 8d,e, p < 0.05) rather than by DR-6W (Fig. 8d,e, p > 0.05). Compared with DR-6W, it showed a significant reduction in expressions of both MDA and PC in DR-LL (Fig. 8d,e, p < 0.05), while it showed a vital decrease in expression of only MDA in DR-6M (Fig. 8d, p < 0.05). Finally we observed the expressions of T-SOD and CAT. Compared with the corresponding AL, the level of T-SOD was improved only by DR-LL (Fig. 8f, p < 0.05), it was enhanced in the expression of CAT in both DR-6M and DR-LL, but not in DR-6W (Fig. 8g, p < 0.05).

In addition, we also examined the effects of DR for different durations on the products of oxidative DNA damage in old renal tissues. According to the immunochemical staining results of 8-OHdG, we observed that it was mainly distributed in renal tubules (see Supplementary Fig. S3). Then we compared the effects of DR for different durations on renal 8-OHdG levels, and found that the immunoreactivities and the staining intensities of 8-OHdG in DR-6M and DR-LL groups were significantly lower than the corresponding AL groups, indicating that DR for long term could suppress the oxidative injury in old kidneys (see Supplementary Fig. S3, p < 0.05).

In short, DR-6M and DR-LL enhanced the expressions of CGL and CBS, contributing to elevating the levels of H_2S rather than GSH. Thus, our study demonstrated that aged rats had the ability of producing enough H_2S and that it was more significant in DR for long-term than in DR for short-term, which indicating that TSP and H_2S could mediate the DR effect in protecting against renal senescence partially by modulation of redox balance to some extent.

Discussion

Accumulated studies have shown that aging is an independent risk factor for the onset and development of renal diseases^{33,34}. DR can extend the lifespan or delay the aging process by decreasing oxidative stress level^{35–37}.



Figure 2. The expressions of senescent biomarker p16 and cell cycle inhibitor p21 in kidneys. Western blot results (**a**,**c**) and quantitative analysis of the band density (**b**,**d**) showed that the expressions of p16 and p21 were increased in kidneys of Old-AL vs. those of Young-AL. Western blot results (**e**,**g**) and quantitative analysis of the band density (**f**,**h**) showed that DR-6M and DR-LL decrease the expressions of p16 and p21 but DR-6W doesn't. The data are presented as the mean \pm SD (n = 5–8). *p < 0.05 vs. the corresponding AL. *p < 0.05 vs. DR-6W.

However, the specific mechanisms among DR, oxidative stress and aging remain unclear. Recently there was an study highlighted that 50% DR for 7 days could partly mediate the protection of DR and further increased expressions of CGL and CBS, resulting in the production of H_2S in young mice model²³. In this study, we attempted to









explore whether the aged rats had the potentiality to produce enough H_2S , and further to observe the efficiencies of DR for different durations in delaying renal aging.

To the best of our knowledge, this study was the first time to study protection against renal senescence by DR for different durations. We observed metabolic indexes, renal function, renal histology alternations and senescence markers, and found that DR-6M and DR-LL could significantly delay or reduce multiple abnormal or pathological manifestations of aged kidneys. We further respectively measured the levels of capital enzymes, H₂S, GSH, and oxidative stress. And we found that it was H₂S, not GSH, that worked in the protection against



AL-6W

DR-6W



AL-6M

DR-6M



AL-LL

DR-LL

Figure 5. Location of CGL in renal tissue by immunohistochemistry staining. Magnification, x400. It was mainly expressed in the renal tubular cytoplasm, but rarely in glomerulus.

stress-induced senescence in aged kidneys by reducing oxidative stress. Then we compared the efficiencies of H_2S induced by DR for different durations, and concluded that the effect of DR for long-term was better than that of DR for short-term.

It has been widely accepted that oxidative stress activity increases during organism aging and that the imbalance between oxidant substances and antioxidant products exerts a significant and causal factor in age-associated symptoms^{4,5,38}. Growing evidences indicate that a progressive accumulation of oxidative stress is involved in lipid, protein, and DNA damage, disturbing physiological homeostasis and contributing to aging-related kidney dysfunctions^{7,39}. A previous study in our laboratory confirmed that 40% DR for 8 weeks reduced the expression of 8-OHdG, a sensitive biomarker for mitochondrial DNA (mtDNA), and thus protected the kidney from oxidative damage in aged rats^{35,37}. The above results indicate that the increased oxidative stress in aged kidney can be decreased by DR.



AL-6W

DR-6W



AL-6M

DR-6M



Figure 6. Location of CBS in renal tissue by immunohistochemistry staining. Magnification, x400. It was mainly expressed in the renal tubular cytoplasm.

 $\rm H_2S$, which is previously regarded as a poisonous gas, has been gaining increasing recognition for its numerous beneficial effects. Recently, a study demonstrated that the expressions of both $\rm H_2S$ and $\rm H_2S$ -related enzymes increased with 50% DR for 7 days or methionine restriction, and further blocked mitochondrial oxidative stress²³. It is well known that TSP is responsible for the production of both $\rm H_2S$ and $\rm GSH^{23}$. In our study, TSP was evidently activated, and it was $\rm H_2S$, rather than GSH, that increased in DR-6M and DR-LL groups. Compared with DR-6W, DR-LL exerted larger roles in $\rm H_2S$ production than DR-6M, indicating it was necessary to maintain a long-term and regular rhythm of dietary restrictions.

Accumulated studies have shown that the role of H_2S has been widely considered as a direct and/or indirect mediator of renal oxidative stress response^{22,28}. H_2S exerts protection from stress in part by inner membrane component sulfide quinone oxidoreductase and the latter transfers electrons from H_2S to the electron transport



Figure 7. Effects of DR for different durations on H₂S and GSH level. It could be improved by DR-6M and DR-LL in the levels of H₂S, rather than GSH, in kidney tissues. The data are presented as the mean \pm SD (n = 5-8). *p < 0.05 vs. the corresponding AL. *p < 0.05 vs. DR-6W.

chain and coenzyme Q^{40,41}. Moreover, thiosulfate, as a product of H₂S oxidation via sulfide quinone oxidoreductase, can encounter further chemical modification in a thioredoxin-reductase-dependent reaction using glutathione to produce sulfate or sulfite, which serves as terminal electron acceptors for ATP production and thus results in H₂S generation in some unicellular organisms^{41,42}. Wu et al. noted that exogenous NaHS (100 µmol/L) treatment decreased ROS production and enhanced SOD, GSH and GST expression in H9c2 cardiomyocytes, while Ex 527 (10 µmol/L) reversed these protection effects significantly, which suggests that SIRT1 participated in anti-oxidative activities of H2S during cellular senescence^{43,44}. Consistent with previous research, our study found that H₂S was involved in inhibiting free-radical generation, such as MDA PC, and 8-OHdG. One study suggested that the anti-oxidant actions of H₂S were mediated by preventing p66Shc phosphorylation, thus inhibiting mitochondrial ROS production⁴⁵. Obviously, there are other mechanisms that involves in suppressing cellular senescence besides the sulfhydration of Kelch-like ECH-associated protein 1 and 8-nitroguanosine-3', 5'-cyclic monophosphate⁴⁶⁻⁴⁸. Many studies have concluded that H₂S protected kidney aging mainly by improving the level of oxidative stress, while specific molecular mechanisms remain to be validated by future investigations.

In summary, we demonstrated for the first time that aged rats had the ability to produce enough H₂S on the interventions of DR-6M and DR-LL, thus decreasing stress-induced senescence in aged kidneys. It was implied that H₂S could be a unified mediator of DR in reducing age-related oxidative stress. These novel results could be meaningful for identifying the increasingly important and complex roles of the H₂S system in stress-induced aging, which indicated that H₂S could be likely to be applied in improving the clinical outcomes of aging-related renal diseases in the future.

Methods

Animals. This study was approved by the Chinese PLA General Hospital and Military Medical Postgraduate College. It was performed in accordance with the National Institutes of Health guidelines for the use of experimental animals. The rats were approved in a 12:12 light/dark cycle at 22 ± 1 °C and $50 \pm 10\%$ relative humidity. They were fed with one male per cage and had free access to water under a specific pathogen-free condition. Young (3 months, n = 48) male Fischer 344 rats were divided into ad libitum dietary (AL) group (Young-AL, n = 40) and dietary restriction (DR) group (n = 8). The rats in the DR groups were fed with food that was approximately 70% of the food consumed by the AL group. The food consumption in AL group was measured every week, and then, the results obtained were used to calculate the daily food intake for the next week. When the rats were 24 months old (Old-AL), the AL groups were further divided into AL (n = 24) and DR (n = 16) groups, and then, 8 rats out of the AL and DR groups were sacrificed after 6-week DR (AL-6W and DR-6W, respectively). When they were 30 months old, all of the rats in each group (AL-6M, DR-6M, AL-LL and DR-LL) were sacrificed. Twenty-four hour urine samples were collected in metabolic cages individually and then were sent to detect the urinary protein/creatinine ratio by the coomassie brilliant blue method and the sarcosine oxidase method. Blood samples were collected from the inner canthus before the animals were sacrificed and analyzed. Specially, triglyceride and blood urea nitrogen were detected by colorimetric methods. The hexokinase method was used to detect blood sugar and enzymatic methods were used to detect total cholesterol and serum creatinine. The kidneys were stored at -80 °C for immunohistochemistry staining and western blot.

Renal histology and histological grading. Kidney specimens were fixed in a 10% formalin solution at 4°C overnight. After a series of graded alcohol dehydration, kidney pieces were embedded, sectioned and stained with PAS. The images were visualized and captured at a total magnification of ×200 with Nikon Element software (Nikon Instrument, Nikon Inc., Melville, NY, USA) and were analyzed by two investigators according to the standards of glomerular lesions and tubulointerstitial lesions previously described^{49,50}. Each specimen was measured from 20 random fields per rat using the National Institutes of Health (NIH, MD, USA) Semi-Quantitative Score. In particular, glomerular Lesions (inflammatory cell infiltration, glomerulosclerosis, fibrous and cellular crescents, loop necrosis/karyorrhexis) and renal interstitial lesion (protein casts) were graded by standard scores



Figure 8. DR for different durations improved the renal oxidative stress. (a) The expressions and (b) the quantitative analysis of the band density of gp91, and (c) the levels of ROS increased with aging, which could be reduced by DR-6M and DR-LL in the kidney tissues. The biochemical measurements with regard to the oxidative stress in aged kidney tissues, such as (d) CAT, (e) PC, (f) T-SOD, and (g) CAT, were improved with the interventions of DR-6M and DR-LL, rather than of DR-6W. The data are presented as the mean \pm SD (n = 5–8). *p < 0.05 vs. the corresponding AL. *p < 0.05 vs. DR-6W.

of 0 points (0%), 1 point (<25%), 2 points (25–50%), and 3 points (>50%). Furthermore, renal interstitial lesions (renal tubular epithelial cell degeneration, renal tubular atrophy, renal interstitial fibrosis and interstitial inflammatory cell infiltration) and glomerular lesions (cell proliferation) were measured by standard scores of 0 points (-), 1 point (mild), 2 points (medium), and 3 points (severe).

Western blot analysis. After the kidney tissues were lysed in radio immunoprecipitation assay buffer, the extracted protein concentration was measured with a BCA Protein Assay kit (Thermo Fisher Scientific, Rockford, IL, USA). The extracted proteins were mixed well with 5% sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SEMS-PAGE) sample buffer followed by heating at 95 °C for 10 min. A total of 50 to 80 µg protein was separated using 8–12% SEMS-polyacrylamide gel electrophoresis, transferred to a nitrocellulose (NC) membrane and then inoculated at 4 °C overnight with the primary antibodies: rabbit monoclonal anti-p16 antibody (1:500; Abcam, Cambridge MA, USA), rabbit polyclonal anti-p21 antibody (1:1000; Proteintech, Chicago IL, USA), rabbit polyclonal anti-CGL and anti-CBS antibody (1:1000; Abcam, Cambridge MA, USA), and rabbit monoclonal anti-gp91/NOX2 antibody (1:2000; Abcam, Cambridge MA, USA). The HRP-conjugated secondary antibodies (Santa cruz biotechnology, CA, USA) were incubated at room temperature for 2 h or at 4 °C for 4 h. Each membrane was detected by enhanced chemiluminescence, and densitometry was conducted using Quantity One software (Bio-Rad Laboratories, Hercules, CA, USA).

Oxidative stress-related indicator detection. The renal tissue was homogenized in a cold 0.9% sodium chloride solution to make a 10% homogenate. Then, the homogenate was centrifuged at 4000 rpm 4°C for 15 min. The tissue supernatant that was extracted was collected for further analysis. The protein concentration was determined with a BCA Protein Assay kit (Thermo Fisher Scientific, Rockford, IL, USA). The activities of ROS, MDA, PC, CAT, T-SOD and GSH, contents were assayed according to the recommended procedures provided by commercial reagent kits (Jiancheng Institute of Bioengineering, Nanjing, China). The level of H₂S in kidney was determined in tissue lysates using the Rat Hydrogen Sulfide Assay kit (Neobiolab, Cambridge MA, USA) according to the manufacturer's instructions.

Immunohistochemistry staining. The immunohistochemistry staining methods were performed as previously described³⁵. After dewaxing, antigen retrieval and blocking of endogenous peroxidase, the sections were fixed with goat serum and incubated with a 1:1000 dilution of rabbit monoclonal anti-CGL antibody (Abcam, Cambridge MA, USA), a 1:2000 dilution of rabbit monoclonal anti-CBS antibody (Abcam, Cambridge MA, USA), and a 1:20 dilution of mouse monoclonal anti-8-OHdG antibody (Santa cruz biotechnology, CA, USA) overnight at 4 °C, followed by incubation with biotin-conjugated goat anti-rabbit IgG (Invitrogen, Carlsbad, CA, USA) and streptavidin- conjugated peroxidase (Invitrogen, Carlsbad, CA, USA). Finally, sections were detected under a microscope after reaction using 3,3'-diaminobenzidine (DAB) (Invitrogen, Carlsbad, CA, USA). The intensity of immunohistochemical staining of 8-OHdG was analyzed in ten random fields (×400) of renal cortex per rat using Image J software (NIH, MD, USA).

Statistical analysis. All of the data analyses were performed using SPSS 17.0 (SPSS, Chicago, IL, USA) software. For the animal and kidney characteristics as well as the protein expressions, the data were provided as the mean \pm SD. Two group differences were determined by T test and multiple group differences were determined by ANOVA analysis.

References

- 1. Zhou, X. J. et al. The aging kidney. Kidney international 74, 710-720, doi: 10.1038/ki.2008.319 (2008).
- Musso, C. G. & Oreopoulos, D. G. Aging and physiological changes of the kidneys including changes in glomerular filtration rate. Nephron. Physiology 119 Suppl 1, p1–5, doi: 10.1159/000328010 (2011).
- Chung, H. Y. et al. Molecular inflammation: underpinnings of aging and age-related diseases. Ageing research reviews 8, 18–30, doi: 10.1016/j.arr.2008.07.002 (2009).
- 4. Harman, D. Aging: a theory based on free radical and radiation chemistry. Journal of gerontology 11, 298-300 (1956).
- 5. Sohal, R. S. & Allen, R. G. Oxidative stress as a causal factor in differentiation and aging: a unifying hypothesis. *Experimental gerontology* **25**, 499–522 (1990).
- Harman, D. The aging process. Proceedings of the National Academy of Sciences of the United States of America 78, 7124–7128 (1981).
 Liu, J. et al. Immobilization stress causes oxidative damage to lipid, protein, and DNA in the brain of rats. FASEB journal: official
- publication of the Federation of American Societies for Experimental Biology 10, 1532–1538 (1996).
 8. Bedard, K. & Krause, K. H. The NOX family of ROS-generating NADPH oxidases: physiology and pathophysiology. *Physiological reviews* 87, 245–313, doi: 10.1152/physrev.00044.2005 (2007).
- Qin, L., Liu, Y., Hong, J. S. & Crews, F. T. NADPH oxidase and aging drive microglial activation, oxidative stress, and dopaminergic neurodegeneration following systemic LPS administration. *Glia* 61, 855–868, doi: 10.1002/glia.22479 (2013).
- Hao, C. M. & Haase, V. H. Sirtuins and their relevance to the kidney. Journal of the American Society of Nephrology: JASN 21, 1620–1627, doi: 10.1681/asn.2010010046 (2010).
- 11. Kume, S. et al. Calorie restriction enhances cell adaptation to hypoxia through Sirt1-dependent mitochondrial autophagy in mouse aged kidney. The Journal of clinical investigation 120, 1043–1055, doi: 10.1172/jci41376 (2010).
- Hine, C. & Mitchell, J. R. Calorie restriction and methionine restriction in control of endogenous hydrogen sulfide production by the transsulfuration pathway. *Experimental gerontology* 68, 26–32, doi: 10.1016/j.exger.2014.12.010 (2015).
- Fontana, L., Partridge, L. & Longo, V. D. Extending healthy life span-from yeast to humans. Science 328, 321–326, doi: 10.1126/ science.1172539 (2010).
- 14. Ruckenstuhl, C. et al. Lifespan extension by methionine restriction requires autophagy-dependent vacuolar acidification. PLoS Genet 10, e1004347, doi: 10.1371/journal.pgen.1004347 (2014).
- Grandison, R. C., Piper, M. D. & Partridge, L. Amino-acid imbalance explains extension of lifespan by dietary restriction in Drosophila. Nature 462, 1061–1064, doi: 10.1038/nature08619 (2009).

- Miller, R. A. *et al.* Methionine-deficient diet extends mouse lifespan, slows immune and lens aging, alters glucose, T4, IGF-I and insulin levels, and increases hepatocyte MIF levels and stress resistance. *Aging cell* 4, 119–125, doi: 10.1111/j.1474-9726.2005.00152.x (2005).
- 17. Cabreiro, F. *et al.* Metformin retards aging in C. elegans by altering microbial folate and methionine metabolism. *Cell* **153**, 228–239, doi: 10.1016/j.cell.2013.02.035 (2013).
- 18. Ledford, H. Mice share yeast's ageing system. Nature 456, 433, doi: 10.1038/456433a (2008).
- Mair, W. et al. Lifespan extension induced by AMPK and calcineurin is mediated by CRTC-1 and CREB. Nature 470, 404–408, doi: 10.1038/nature09706 (2011).
- 20. Harrison, D. E. *et al.* Rapamycin fed late in life extends lifespan in genetically heterogeneous mice. *Nature* **460**, 392–395, doi: 10.1038/nature08221 (2009).
- Sharples, A. P. *et al.* Longevity and skeletal muscle mass: the role of IGF signalling, the sirtuins, dietary restriction and protein intake. *Aging cell* 14, 511–523, doi: 10.1111/acel.12342 (2015).
- Zhang, Y. et al. Hydrogen sulfide, the next potent preventive and therapeutic agent in aging and age-associated diseases. Molecular and cellular biology 33, 1104–1113, doi: 10.1128/mcb.01215-12 (2013).
- Hine, C. et al. Endogenous hydrogen sulfide production is essential for dietary restriction benefits. Cell 160, 132–144, doi: 10.1016/j. cell.2014.11.048 (2015).
- Zhang, X. & Bian, J. S. Hydrogen sulfide: a neuromodulator and neuroprotectant in the central nervous system. ACS chemical neuroscience 5, 876–883, doi: 10.1021/cn500185g (2014).
- Lobb, I., Sonke, E., Aboalsamh, G. & Sener, A. Hydrogen sulphide and the kidney: important roles in renal physiology and pathogenesis and treatment of kidney injury and disease. *Nitric oxide: biology and chemistry / official journal of the Nitric Oxide* Society 46, 55–65, doi: 10.1016/j.niox.2014.10.004 (2015).
- Xia, M., Chen, L., Muh, R. W., Li, P. L. & Li, N. Production and actions of hydrogen sulfide, a novel gaseous bioactive substance, in the kidneys. *The Journal of pharmacology and experimental therapeutics* 329, 1056–1062, doi: 10.1124/jpet.108.149963 (2009).
- Stipanuk, M. H. & Ueki, I. Dealing with methionine/homocysteine sulfur: cysteine metabolism to taurine and inorganic sulfur. Journal of inherited metabolic disease 34, 17–32, doi: 10.1007/s10545-009-9006-9 (2011).
- Koning, A. M., Frenay, A. R., Leuvenink, H. G. & van Goor, H. Hydrogen sulfide in renal physiology, disease and transplantation-the smell of renal protection. Nitric oxide: biology and chemistry/official journal of the Nitric Oxide Society 46, 37–49, doi: 10.1016/j. niox.2015.01.005 (2015).
- 29. Song, K. *et al.* Hydrogen sulfide inhibits the renal fibrosis of obstructive nephropathy. *Kidney international* **85**, 1318–1329, doi: 10.1038/ki.2013.449 (2014).
- Aminzadeh, M. A. & Vaziri, N. D. Downregulation of the renal and hepatic hydrogen sulfide (H2S)-producing enzymes and capacity in chronic kidney disease. Nephrology, dialysis, transplantation: official publication of the European Dialysis and Transplant Association - European Renal Association 27, 498–504, doi: 10.1093/ndt/gfr560 (2012).
- Della Coletta Francescato, H. et al. Inhibition of hydrogen sulphide formation reduces cisplatin-induced renal damage. Nephrology, dialysis, transplantation: official publication of the European Dialysis and Transplant Association - European Renal Association 26, 479–488, doi: 10.1093/ndt/gfq447 (2011).
- Bos, E. M. et al. Cystathionine gamma-lyase protects against renal ischemia/reperfusion by modulating oxidative stress. Journal of the American Society of Nephrology: JASN 24, 759–770, doi: 10.1681/asn.2012030268 (2013).
- Striker, G. E. The aging kidney phenotype and systemically derived stem cells. Journal of the American Society of Nephrology: JASN 22, 1958–1960, doi: 10.1681/asn.2011090946 (2011).
- Wulfert, F. M. et al. Age-dependent role of microvascular endothelial and polymorphonuclear cells in lipopolysaccharide-induced acute kidney injury. Anesthesiology 117, 126–136, doi: 10.1097/ALN.0b013e31825b57c9 (2012).
- 35. Cui, J. et al. Age-related changes in the function of autophagy in rat kidneys. Age 34, 329–339, doi: 10.1007/s11357-011-9237-1 (2012).
- Ning, Y. C. et al. Short-term calorie restriction protects against renal senescence of aged rats by increasing autophagic activity and reducing oxidative damage. Mechanisms of ageing and development 134, 570–579, doi: 10.1016/j.mad.2013.11.006 (2013).
- Ning, Y. C. et al. Beneficial effects of short-term calorie restriction against cisplatin-induced acute renal injury in aged rats. Nephron. Experimental nephrology 124, 19–27, doi: 10.1159/000357380 (2013).
- Pincemail, J., Ricour, C., Defraigne, J. O. & Petermans, J. [Oxidative stress, antioxydants and the ageing process]. Revue medicale de Liege 69, 270–275 (2014).
- Aydin, S. et al. Comparison of oxidative stress biomarkers in renal tissues of D-galactose induced, naturally aged and young rats. Biogerontology 13, 251–260, doi: 10.1007/s10522-011-9370-3 (2012).
- Modis, K., Coletta, C., Erdelyi, K., Papapetropoulos, A. & Szabo, C. Intramitochondrial hydrogen sulfide production by 3-mercaptopyruvate sulfurtransferase maintains mitochondrial electron flow and supports cellular bioenergetics. FASEB journal: official publication of the Federation of American Societies for Experimental Biology 27, 601–611, doi: 10.1096/fj.12-216507 (2013).
- Ölson, K. R. et al. Thiosulfate: a readily accessible source of hydrogen sulfide in oxygen sensing. American journal of physiology. Regulatory, integrative and comparative physiology 305, R592–603, doi: 10.1152/ajpregu.00421.2012 (2013).
- 42. Muyzer, G. & Stams, A. J. The ecology and biotechnology of sulphate-reducing bacteria. *Nature reviews. Microbiology* **6**, 441–454, doi: 10.1038/nrmicro1892 (2008).
- 43. Yu, L. *et al.* Melatonin receptor-mediated protection against myocardial ischemia/reperfusion injury: role of SIRT1. *Journal of pineal research* 57, 228–238, doi: 10.1111/jpi.12161 (2014).
- Wu, D. et al. Hydrogen sulfide protects against apoptosis under oxidative stress through SIRT1 pathway in H9c2 cardiomyocytes. Nitric oxide: biology and chemistry / official journal of the Nitric Oxide Society 46, 204–212, doi: 10.1016/j.niox.2014.11.006 (2015).
- Xie, Z. Z. et al. Sulfhydration of p66Shc at cysteine59 mediates the antioxidant effect of hydrogen sulfide. Antioxidants & redox signaling 21, 2531–2542, doi: 10.1089/ars.2013.5604 (2014).
- Yang, G. et al. Hydrogen sulfide protects against cellular senescence via S-sulfhydration of Keap1 and activation of Nrf2. Antioxidants & redox signaling 18, 1906–1919, doi: 10.1089/ars.2012.4645 (2013).
- 47. Nishida, M. *et al.* Hydrogen sulfide anion regulates redox signaling via electrophile sulfhydration. *Nature chemical biology* **8**, 714–724, doi: 10.1038/nchembio.1018 (2012).
- Mustafa, A. K. *et al.* Hydrogen sulfide as endothelium-derived hyperpolarizing factor sulfhydrates potassium channels. *Circulation research* 109, 1259–1268, doi: 10.1161/circresaha.111.240242 (2011).
- Bazzi, C., Petrini, C., Rizza, V., Arrigo, G. & D'Amico, G. A modern approach to selectivity of proteinuria and tubulointerstitial damage in nephrotic syndrome. *Kidney international* 58, 1732–1741, doi: 10.1046/j.1523-1755.2000.00334.x (2000).
- 50. Debelle, F. D. *et al.* Aristolochic acids induce chronic renal failure with interstitial fibrosis in salt-depleted rats. *Journal of the American Society of Nephrology: JASN* **13**, 431–436 (2002).

Acknowledgements

This work was supported by the National Key Basic Research Program of China (973 Program, 2013CB530800), National High Technology Research and Development Program of China (863 program, 2012AA02A512), National Key Technology R&D program (2015BAI12B06), and the NSFC (81070267, 81171645).

Author Contributions

G.-Y.C. is the corresponding author designed and supervised the research, amended all drafts and provided the final approval of the version to be published. Y.-C.N., J.C., X.-M.X. and R.B. contributed to breeding the rats. W.-J.W. performed all the experiments, interpreted the data and wrote the manuscript. Q.H., X.-Y.B., X.-F.S. and X.-M.C put forward both very valuable comments and important insights and contributed to the collection, interpretation and analysis of data throughout the experiment.

Additional Information

Supplementary information accompanies this paper at http://www.nature.com/srep

Competing financial interests: The authors declare no competing financial interests.

How to cite this article: Wang, W.-J. *et al.* Hydrogen sulfide mediates the protection of dietary restriction against renal senescence in aged F344 rats. *Sci. Rep.* **6**, 30292; doi: 10.1038/srep30292 (2016).

This work is licensed under a Creative Commons Attribution 4.0 International License. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in the credit line; if the material is not included under the Creative Commons license, users will need to obtain permission from the license holder to reproduce the material. To view a copy of this license, visit http://creativecommons.org/licenses/by/4.0/

© The Author(s) 2016