


 Cite this: *RSC Adv.*, 2020, **10**, 23270

A porous nano-adsorbent with dual functional groups for selective binding proteins with a low detection limit

 Xueyan Zou,^{aef} Yu Zhang,^b Jinqiu Yuan,^c Zhibo Wang,^b Rui Zeng,^c Kun Li,^d Yanbao Zhao^{*aef} and Zhijun Zhang^{id}^{*aef}

In this study, porous silica nanoparticles functionalized with a thiol group (SiO₂-SH NPs) were synthesized via a one-pot method. Subsequently, iminodiacetic acid was modified, and further adsorption of Ni²⁺ ions was conducted to obtain a SiO₂-S/NH-Ni nano-adsorbent. Then, transmission electron microscopy (TEM), scanning electron microscopy (SEM), Fourier transform infrared (FT-IR) spectroscopy, thermogravimetric analysis (TG) and X-ray diffraction (XRD) were employed to characterize its morphology and composition. The results indicate that the SiO₂-S/NH-Ni nano-adsorbent is porous, has an average diameter of 77.1 nm and has a small porous structure of about 3.7 nm in the silica skeleton. The Brunauer-Emmett-Teller (BET) surface area and total pore volume were 537.2 m² g⁻¹ and 3.3 cm³ g⁻¹, respectively, indicating a large BET surface area. The results indicate that the as-prepared SiO₂-S/NH-Ni nano-adsorbent would be suitable to selectively and efficiently bind His-tagged proteins from an *E. coli* cell lysate. The SDS-PAGE results show that the as-prepared nano-adsorbent presents specifically to both His-tagged CPK4 and His-tagged TRX proteins, indicating the nano-adsorbent can be used to effectively separate His-tagged proteins and is universal to all His-tagged fusion proteins. We also found that the as-prepared nano-adsorbent exhibits a low detection limit (1.0 × 10⁻⁷ mol L⁻¹) and a strong regeneration ability based on four regeneration experiments that were particularly suited to the separation of His-tagged proteins.

Received 7th February 2020

Accepted 14th May 2020

DOI: 10.1039/d0ra01193b

rsc.li/rsc-advances

Introduction

Proteins play a variety of crucial roles in organisms, and the enrichment and purification of proteins, particularly low-abundance proteins, are currently hot topics.¹⁻⁶ Histidine is a semi essential amino acid for humans that constitutes the proteins of the body. It has a very wide range of applications, such as in food, medicine and other fields.⁷⁻¹⁰ Furthermore, biotechnologies nowadays have enabled proteins to easily express with a tag, and numerous protein-purification methods are based on the specific interactions between immobilized ligands and affinity tags.¹¹⁻¹⁶ Among affinity tags, histidine tags are particularly popular as they can be readily incorporated into

desired proteins by the expression of the target gene in micro-organism cells using commercial expression vectors.^{17,18} Therefore, the easy separation and purification of His-tagged proteins are very important in biology and medicine.

Recently, metal-chelate affinity precipitation has been employed to separate His-tagged recombinant proteins based on the interactions between metal ions (Ni²⁺, Cu²⁺, Zn²⁺ and so on) and the histidine residues exposed on the surface of the proteins.¹⁹⁻²² Although metal-chelate affinity precipitation is easily adaptable to any protein expression system, there are some limits to using this method, such as a requirement for pretreatment, solvent consumption, and protein solubility.²³⁻²⁵

With the development of nanotechnology, nanomaterials are used in many fields, such as protein purification, environmental treatment, targeted drugs and so on.²⁶⁻²⁸ However, the major limitations of current nanomaterial systems are the detection limit and poor regeneration ability, which leads to low purification efficiency. In this study, porous silica with dual functional groups (-SH and -NH₂ groups) was employed as an adsorbent to separate His-tagged proteins. This material can provide a larger surface area, comparable to solid particles, and improve the separation capacity. The as-prepared nano-adsorbent can be used to directly separate His-tagged proteins from the *E. coli* lysate.

^aEngineering Research Center for Nanomaterials, Henan University, Kaifeng 475004, China. E-mail: zhangzj09@126.com; zhaoyb902@henu.edu.cn

^bCollege of Chemistry and Chemical Engineering, Henan University, Kaifeng 475004, China

^cInstitute of Technology, Henan University Minsheng College, Kaifeng 459000, China

^dState Key Laboratory of Crop Stress Adaptation and Improvement, Kaifeng 459000, China

^eNational & Local Joint Engineering Research Center for Applied Technology of Hybrid Nanomaterials, Kaifeng 459000, China

^fKey Laboratory for Monitor and Remediation of Heavy Metal Polluted Soils of Henan Province, Jiyuan 459000, China



Results and discussion

Fig. 1 shows the SEM (a) and TEM (b) images of the as-synthesized SiO₂-S/NH-Ni sample. It can be seen in Fig. 1a that the main products were spherical in shape and presented uniform spheres. Moreover, we could see that the as-prepared SiO₂-S/NH-Ni sample displayed a narrow size distribution and had an average diameter of 77.1 nm (Fig. 1c). Evidently, the surfaces of these nanospheres were rough, which may be due to the oriented growth and crystallinity.²⁷ As shown in Fig. 1b, the obtained SiO₂-S/NH-Ni sample displays a small porous structure, with about 3.7 nm, in the silica skeleton.

Fig. 2a shows the FT-IR absorption spectra of the as-prepared SiO₂-SH NPs (curve 1) and SiO₂-SH/NH₂ NPs (curve 2). In the FT-IR spectra, the peak at 3430 cm⁻¹ is attributed to the -OH stretching vibration of porous silica spheres. The peaks at around 1105, 807 and 471 cm⁻¹ were due to Si-O-Si stretching vibrations. The strong peaks at 2939 and 2836 cm⁻¹ belong to the stretching vibrations of the -CH₂- bonds in MPS, which indicates the presence of -SH in the SiO₂-SH NPs (Fig. 2a curve 1). It is clear that the curve of the SiO₂-SH/NH₂ NPs is very similar to that of the SiO₂-SH NPs, which indicates that the main component of SiO₂-SH/NH₂ NPs is SiO₂. The presence of the N-H bending vibration at around 695 cm⁻¹ confirms the incorporation of the imino group of IDA.²⁹

Fig. 2b shows the TG curve of the SiO₂-SH/NH₂ NPs. It can be seen that the sample displays successive weight loss from room temperature to 850 °C. The initial mass loss below 250 °C can be attributed to the desorption of adsorbed water from the surface of the sample powder, and the following mass loss was related to the further release of the inner adsorbed or crystal water (total mass loss 25%). The major mass loss (about 15.5%) took place in the range of 250–600 °C, which might be attributed to the thermal decomposition of the -SH and -NH₂ groups of the as-prepared SiO₂-SH/NH₂ NPs. This indicates that there are two

groups (-SH and -NH₂) on the surface of the as-synthesized NPs. Fig. 2c shows the XRD pattern of the as-prepared SiO₂-SH/NH₂ NPs. The XRD pattern exhibits an obvious broad diffraction peak at around 23°, which was indexed to the scattering of amorphous SiO₂ (JCPDS 76-0933).³⁰

The surface and pore properties of the as-prepared SiO₂-SH/NH₂ NPs was examined *via* N₂ sorption, as shown in Fig. 3. It can be seen in Fig. 3a that the N₂ adsorption-desorption isotherms are characteristic of type IV curves with the H4 hysteresis loop in the relative-pressure range of 0.04–0.3, which indicates the presence of well-defined mesoporous structures with narrow slit-like pores.^{31,32} It can be seen that the pore size distributions calculated using the BJH method were 3.3 and 3.7 nm (Fig. 3b). Moreover, the Brunauer-Emmett-Teller (BET) surface area and total pore volume were 537.2 m² g⁻¹ and 3.3 cm³ g⁻¹, respectively, which represents a large BET surface area that is suitable for separating proteins.

Fig. 4 shows a scheme of the preparation of the SiO₂-S/NH-Ni nano-adsorbent and their separation of His-tagged proteins. It can be seen that there are three steps required to synthesize the SiO₂-S/NH-Ni nano-adsorbent. First, SiO₂-SH was prepared by the hydrolysis of TEOS and MPS *via* a one-pot method. Second, the IDA molecules were modified on the surfaces of the SiO₂-SH NPs. Third, the SiO₂-S/NH-Ni nano-adsorbent was obtained by chelating Ni²⁺. Then, the nano-adsorbent can be used to separate His-tagged proteins from the *E. coli* lysate, and the obtained SiO₂-S/NH-Ni/His-tagged proteins can be eluted by imidazole. Subsequently, the His-tagged proteins can be obtained, and the remaining SiO₂-S/NH-Ni nano-adsorbent could be reused by regeneration with EDTA and NiCl₂ solutions. The results show that the as-prepared SiO₂-S/NH-Ni nano-adsorbent has a strong regeneration ability, particularly suited to the separation of the His-tagged fusion proteins.

To estimate the separation effect of the as-prepared samples, SDS-PAGE was employed as a general method for assessing the target proteins. Fig. 5 shows the SDS-PAGE analysis of His-tagged proteins that were purified by the SiO₂-S/NH-Ni nano-adsorbent. Lanes 1–4 contain the fractions washed off from the SiO₂-S/NH-Ni nano-adsorbent with different amounts of imidazole (lane 1, 0.5 mol L⁻¹; lane 2, 1 mol L⁻¹; lane 3, 2 mol L⁻¹; lane 4, 3 mol L⁻¹). Lane 5 contains the marker, and lane 6 contains the *E. coli* lysate. It was found that the SiO₂-S/NH-Ni nano-adsorbent could efficiently enrich target proteins from cell lysate. By changing the concentration of imidazole (0.5–3 mol L⁻¹) and keeping the others unchanged, the quantity of disassociated proteins remained basically the same with increment in the concentration of imidazole (Fig. 5 lane 1–4), so 0.5 mol L⁻¹ imidazole was used in subsequent experiments.

Keeping the imidazole concentration constant (0.5 mol L⁻¹), the effect of the concentration of the target proteins was investigated, as shown in Fig. 5. The concentrations in the lanes 7–10 are as follows: lane 7, 1.0 × 10⁻⁴ mol L⁻¹; lane 8, 1.0 × 10⁻⁵ mol L⁻¹; lane 9, 1.0 × 10⁻⁶ mol L⁻¹; lane 10, 1.0 × 10⁻⁷ mol L⁻¹; lane 11, 1.0 × 10⁻⁸ mol L⁻¹. When the concentrations of the TRX proteins were 1.0 × 10⁻⁴, 1.0 × 10⁻⁵, 1.0 × 10⁻⁶, 1.0 × 10⁻⁷ and 1.0 × 10⁻⁸ mol L⁻¹, the target proteins

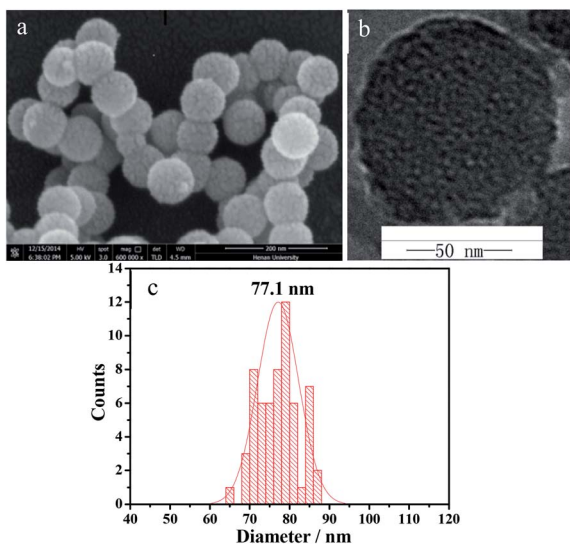


Fig. 1 SEM (a), TEM (b) images and distribution histogram (c) of the as-prepared SiO₂-S/NH-Ni NPs.

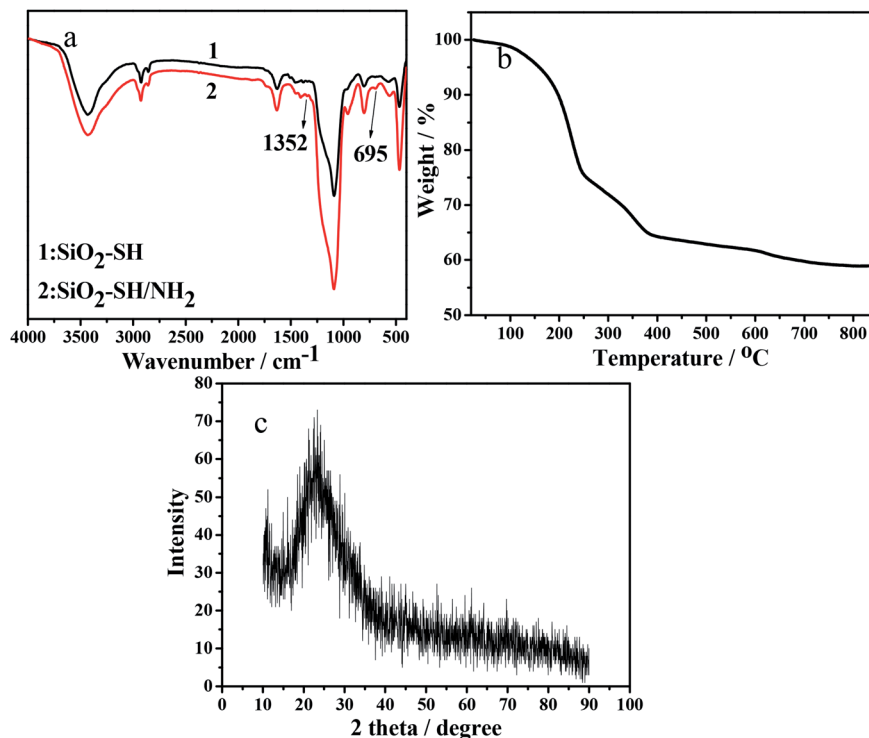


Fig. 2 FT-IR spectrum (a), TG curve (b) and XRD pattern (c) of the SiO₂-SH/NH₂ NPs.

could be detected effectively by the as-prepared SiO₂-S/NH-Ni nano-adsorbent, which shows that the samples can be used to detect target proteins with a detection limit lower than $1.0 \times 10^{-7} \text{ mol L}^{-1}$.

In order to investigate the reusability properties of the as-prepared SiO₂-S/NH-Ni nano-adsorbent, we separated the target proteins four times. Fig. 6 shows the SDS-PAGE for the four recycling times using lane 1, Ni-NTA agarose; lane 2, marker; lane 3, His-tagged TRX *E. coli* lysate; lane 5, 1st; lane 6, 2nd; lane 7, 3rd; lane 8, 4th. The adsorbent could be repeatedly used in the experiment if it was treated with EDTA and NiCl₂ as needed. It can be seen that the specificity and affinity of the SiO₂-S/NH-Ni nano-adsorbent remained unaffected after four recycling times. The binding capacities of the target protein were 9.8, 9.8, 9.6 and 9.2 mmol g⁻¹.

In order to verify the universality of the SiO₂-S/NH-Ni nano-adsorbent, we used the adsorbent to separate His-tagged Ca²⁺-dependent protein kinase 4 (His-tagged CPK4) proteins from the *E. coli* lysate, as shown in Fig. 6. The lanes are as follow: lane 3, His-tagged TRX *E. coli* lysate; lane 4, His-tagged CPK4 *E. coli* lysate; lane 9, His-tagged TRX separated by the SiO₂-S/NH-Ni nano-adsorbent; lane 10, His-tagged CPK4 separated by the SiO₂-S/NH-Ni nano-adsorbent. It is clear that the His-tagged CPK4 proteins could be separated specifically from the *E. coli* lysate, and there was no nonspecific binding. These results indicate that the as-prepared SiO₂-S/NH-Ni nano-adsorbent could be used effectively in His-tagged protein affinity separation or purification, and it is universally applicable to all His-tagged fusion proteins. These results are due to the Ni of the as-prepared SiO₂-S/NH-Ni nano-adsorbent being specific to

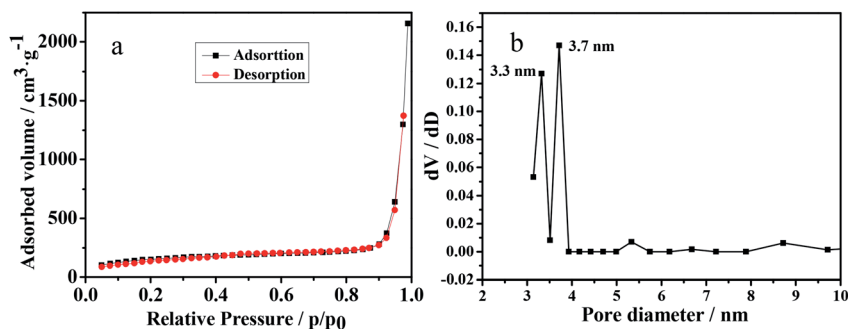


Fig. 3 N₂ adsorption-desorption isotherm (a) and the pore size distribution (b) of the SiO₂-S/NH-Ni NPs.

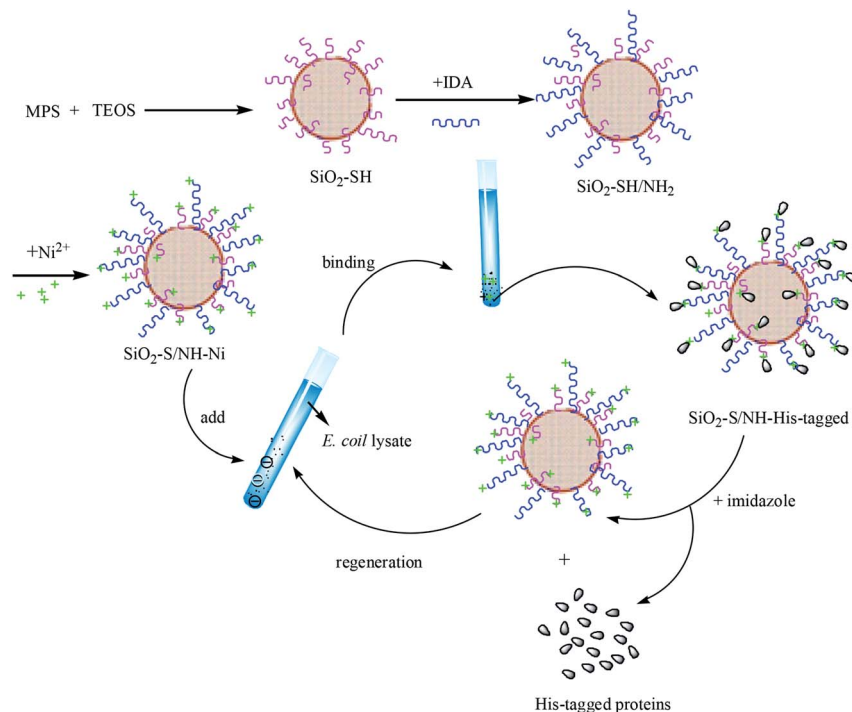


Fig. 4 Scheme for the preparation of the SiO₂-S/NH-Ni samples and their separation of His-tagged proteins.

histidine and the specific binding being independent of the type of fusion proteins.

Experimental

Materials

Ni-NTA agarose was purchased from QIAGEN. Phosphate buffer saline solutions (abridged as PBS; concentration 0.1 mol L⁻¹, pH = 8.0; concentration 0.01 mol L⁻¹, pH = 7.4) were purchased from Sigma-Aldrich, America. 3-Glycidyloxy-propyltrimethoxysilane (GPTMS) and iminodiacetic acid (IDA) were purchased from Aladdin. Tetraethyl orthosilicate (TEOS) was from Tianjin Fuchen Chemicals. 3-Mercaptopropyltrimethoxysilane (MPS) was supplied

from Alfa-Aesar. Hexadecyl-trimethylammonium bromide (CTAB) was from Sinopharm Chemicals. Triethanolamine (TEA), ethylene diamine tetraacetic acid (denoted as EDTA), absolute alcohol and aqueous ammonia (28 wt%) were purchased from Tianjin Kernel Chemicals.

Synthesis of the SiO₂-S/NH-Ni nano-adsorbent

In a typical synthesis, 93.75 μmol of CTAB was added into 19 mL of an ethanol solution (v(CH₃CH₂OH)/v(H₂O) = 6 : 1) and stirred for a few minutes. Then, 1.1 mL of TEA was also added into

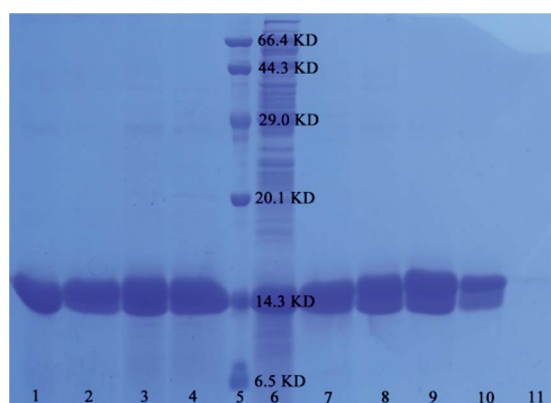


Fig. 5 SDS-PAGE analysis of His-tagged proteins separated by the SiO₂-S/NH-Ni nano-adsorbent.

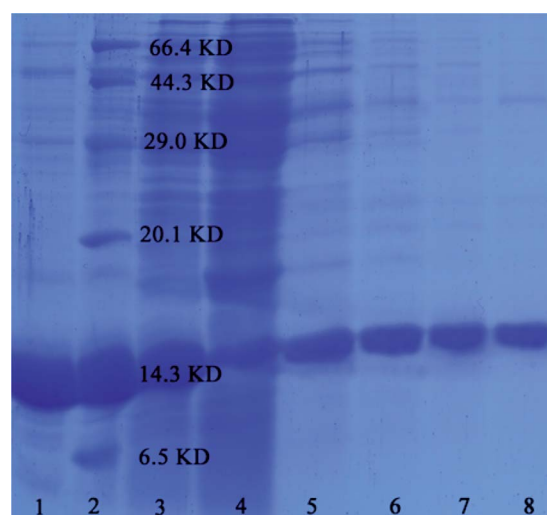


Fig. 6 SDS-PAGE analysis of His-tagged proteins separated by the as-prepared SiO₂-S/NH-Ni nano-adsorbent.

the above solution. Subsequently, 1.54 mL of the mixed solution of TEOS and MPS ($v/v = 10 : 1$) was dropped slowly into the above solution at $60\text{ }^{\circ}\text{C}$ ($28\ \mu\text{L s}^{-1}$) and reacted for 3 h. Then, the precipitate was treated with an ethanol-HCl solution for 12 h to remove the CTAB template and was then washed with deionized water three times and dried for 48 h at $60\text{ }^{\circ}\text{C}$ to obtain $\text{SiO}_2\text{-SH}$ NPs.

Then, 20 mL of an IDA solution ($0.085\ \text{g mL}^{-1}$, $\text{pH} = 11$) was added into a flask, and the solution was kept at $0\text{ }^{\circ}\text{C}$ in an ice-bath. Then, 2 mL of GPTMS was added under stirring conditions, and the mixture was reacted at $65\text{ }^{\circ}\text{C}$ for 12 h. Subsequently, 0.2 g of $\text{SiO}_2\text{-SH}$ NPs was dispersed in 6 mL of the GPTMS-IDA solution ($\text{pH} = 2$), and the suspension was sustained at $90\text{ }^{\circ}\text{C}$ for 3 h. Finally, the suspension was centrifuged, washed and dried to get $\text{SiO}_2\text{-SH/NH}_2$ NPs. For chelating the Ni^{2+} ions, 5 mg of $\text{SiO}_2\text{-SH/NH}_2$ NPs was dispersed in 50 mL of a $0.2\ \text{mol L}^{-1}$ NiCl_2 solution and reacted at $25\text{ }^{\circ}\text{C}$ for 24 h. Then, the $\text{SiO}_2\text{-S/NH-Ni}$ nano-adsorbent was washed by deionized water for several times and stored at $4\text{ }^{\circ}\text{C}$.

Affinity separation of His-tagged proteins

After being washed with a $0.02\ \text{mol L}^{-1}$ Tris-HCl binding buffer three times, the $\text{SiO}_2\text{-S/NH-Ni}$ nano-adsorbent was dispersed directly into 1 mL of *E. coli* lysate and shaken for 2 h at a rotation speed of 70 rpm at $4\text{ }^{\circ}\text{C}$. Then, the $\text{SiO}_2\text{-S/NH-Ni}$ nano-adsorbent with the captured His-tagged proteins was washed with Tris-HCl buffer three times to remove any residual uncaptured proteins. Finally, the $\text{SiO}_2\text{-S/NH-Ni}$ nano-adsorbent with the captured His-tagged proteins was eluted with 0.3 mL of $0.05\ \text{mol L}^{-1}$ imidazole solution. The $\text{SiO}_2\text{-S/NH-Ni}$ nano-adsorbent can be recovered and reused by washing sequentially with $0.1\ \text{mol L}^{-1}$ EDTA and $0.2\ \text{mol L}^{-1}$ NiCl_2 solutions.

Characterization

Transmission electron microscopy (TEM, JEM-2010), scanning electron microscopy (SEM, JSM 5600LV), Fourier transform infrared spectroscopy (FT-IR, AVATAR360) and thermogravimetric analysis (TG, EXSTAR 6000) were employed to detect the morphology and composition of the as-prepared nano-materials. The surface area was measured *via* the Brunauer-Emmett-Teller method (BET, QUADRASORB). The separated His-tagged proteins were detected with sodium dodecylsulfate polyacrylamide gel electrophoresis (SDS-PAGE, Power PAC 300). The concentration of the target proteins was analyzed at 280 nm on a UV-Vis spectrophotometer (UV-Vis, nanodrop 2000c).

Conclusions

In this study, thiol-functionalized porous silica nanoparticles were synthesized *via* a one-pot method. Then, the $\text{SiO}_2\text{-SH}$ NPs were further modified by iminodiacetic acid and chelating Ni^{2+} ions to get the porous affinity adsorbent, which could then be applied to separate His-tagged proteins. The results showed that the as-prepared adsorbent had good specificity and regeneration ability for use with His-tagged fusion proteins.

Moreover, the $\text{SiO}_2\text{-SH}$ NP adsorbent had high sensitivity to His-tagged proteins, and the detection limit for the target proteins was lower than $1.0 \times 10^{-7}\ \text{mol L}^{-1}$.

Conflicts of interest

There are no conflicts to declare.

Acknowledgements

The authors acknowledge the financial support provided by the National Natural Science Foundation of China (grant no. 21571051), the Science and Technology Department of Henan Province (in the name of "Major Science and Technology Project", grant no. 181100310600), Henan University (in the name of "Interdisciplinary Research for First-Class Discipline Construction Project", grant no. 2019YLXKJC04), Henan University Minsheng College (in the name of "Innovative Experimental and Practical Projects for College Students", grant no. MSCXSY2019007 and MSCXSY2019016).

Notes and references

- 1 W. Li, F. A. Zhao, W. P. Fang, D. Y. Xie and J. N. Hou, *Front. Plant Sci.*, 2015, **6**, 732–745.
- 2 L. Q. Chai, J. H. Meng, J. Gao, Y. H. Xu and X. W. Wang, *Fish Shellfish Immunol.*, 2018, **80**, 155–164.
- 3 J. F. Hua, S. Zhang, J. J. Cui, D. J. Wang and C. Y. Wang, *J. Insect Physiol.*, 2013, **59**, 690–696.
- 4 M. Y. Chen, K. Li, H. P. Li, C. P. Song and Y. C. Miao, *Sci. Rep.*, 2017, **7**, 44743–44757.
- 5 S. S. Liang, L. Xiao, P. H. Huang, Z. Y. Cheng, F. L. Chen, Y. C. Miao, Y. F. Fu and Q. S. Chen, *Plant Cell Rep.*, 2019, **38**, 1263–1271.
- 6 L. Li, M. J. Hou, L. Cao, Y. Xia and Z. G. Shen, *Environ. Exp. Bot.*, 2018, **155**, 313–320.
- 7 B. K. Oh, S. Park, J. E. Millstone, S. W. Lee, K. B. Lee and C. A. Mirkin, *J. Am. Chem. Soc.*, 2006, **128**, 11825.
- 8 J. H. Lee, Y. M. Huh, Y. W. Jun, J. W. Seo, J. T. Jang, H. T. Song, S. Kim, E. J. Cho, H. G. Yoon, J. S. Suh and J. Cheon, *Nat. Med.*, 2007, **13**, 95–99.
- 9 D. Driss, F. Bhiri, R. Ghorbel and S. E. Chaabouni, *Protein Expression Purif.*, 2012, **83**, 8–14.
- 10 M. Mielecki, J. Wojtasikb, M. Zborowskab, K. Kurzatowska, K. Grzelak, W. Dehaen, J. Radecki and H. Radecka, *Electrochim. Acta*, 2013, **96**, 147–154.
- 11 K. J. Lei, Y. Lin, J. Ren, L. Bai and Y. C. Miao, *Plant Cell Physiol.*, 2016, **57**, 192–203.
- 12 M. Perrier, M. Gary-Bobo, L. Lartigue, D. Brevet, A. Alain Morère, M. Garcia, P. Maillard, L. Raehm, Y. Guari, J. Larionova, J. O. Durand, O. Mongin and M. J. Blanchard-Desce, *J. Nanopart. Res.*, 2013, **15**, 1602–1618.
- 13 P. C. Wang, Y. Y. Du, Y. J. Hou, Y. Zhao and C. C. P. Hsu, *Proc. Natl. Acad. Sci. U. S. A.*, 2015, **112**, 613–618.
- 14 D. R. Bae, S. J. Lee, S. W. Han, J. M. Lim, D. M. Kang and J. H. Jung, *Chem. Mater.*, 2008, **20**, 3809–3813.

- 15 X. Zhao, J. Wang, J. Yuan, X. L. Wang and Q. P. Zhao, *New Phytol.*, 2015, **207**, 211–224.
- 16 N. Li, X. Sun and M. Q. J. Wang, *J. Appl. Entomol.*, 2017, **141**, 751–757.
- 17 Y. B. Yin, G. M. Wei, X. Y. Zou and Y. B. Zhao, *Sens. Actuators, B*, 2015, **209**, 5108–5511.
- 18 X. Y. Zou, K. Li, Y. B. Yin, Y. B. Zhao, Y. Zhang, B. J. Li, S. S. Yao and C. P. Song, *Mater. Sci. Eng., C*, 2014, **34**, 468–473.
- 19 F. Xu, J. H. Geiger, G. L. Baker and M. L. Bruening, *Langmuir*, 2011, **27**, 3106–3112.
- 20 X. Y. Zou, K. Li, Y. B. Yin, Y. B. Zhao, Y. Zhang, B. J. Li, S. S. Yao and C. P. Song, *Mater. Sci. Eng., C*, 2014, **34**, 468–473.
- 21 W. J. Fang, X. L. Chen and N. F. Zheng, *J. Mater. Chem.*, 2010, **20**, 8624–8630.
- 22 W. B. Yap, B. T. Tey, N. B. M. Alitheen and W. S. Tan, *J. Chromatogr. A*, 2010, **1217**, 3473–3480.
- 23 C. J. Xu, K. M. Xu, H. W. Gu, X. F. Zhong, Z. H. Guo, R. K. Zheng, X. X. Zhang and B. Xu, *J. Am. Chem. Soc.*, 2004, **126**, 3392–3393.
- 24 K. S. Lee and I. S. Lee, *Chem. Commun.*, 2008, **14**, 709–711.
- 25 B. Mizrahi, S. Irusta, M. Mckenna, C. Stefanescu, L. Yedidsion, M. Myint, R. Langer and D. S. Kohane, *Adv. Mater.*, 2011, **23**, H258–H262.
- 26 Y. Y. Wang, Y. D. Liu, W. H. Zhan, K. X. Zheng, J. N. Wang, C. S. Zhang and R. H. Chen, *Sci. Total. Environ.*, 2020, **729**, 139060.
- 27 X. Y. Zou, K. Li, Y. B. Zhao, Y. Zhang, B. J. Li and C. P. Song, *J. Mater. Chem. B*, 2013, **1**, 5108–5113.
- 28 Y. Y. Wang, Y. D. Liu, W. H. Zhan, K. X. Zheng, M. M. Lian, C. S. Zhang, X. L. Ruan and T. Li, *Ecotoxicol. Environ. Saf.*, 2020, **197**, 110600.
- 29 M. F. Jeng, A. P. Campbell, T. Begley, A. Holmgren, D. A. Case, P. E. Wright and H. J. Dyson, *Structure*, 1994, **2**, 853–868.
- 30 H. Zhu, B. Lee, S. Dai and S. H. Overbury, *Langmuir*, 2003, **19**, 3974–3980.
- 31 S. S. Wu, X. Huang and X. Z. Du, *J. Mater. Chem. B*, 2015, **3**, 1426–1432.
- 32 S. Q. Liu, M. Y. Wei, J. C. Rao, H. X. Wang and H. Zhao, *Mater. Lett.*, 2011, **65**, 2083–2085.