Household Risk Factors for Colonization with Multidrug-Resistant *Staphylococcus aureus* Isolates

Meghan F. Davis¹*, Amy E. Peterson², Kathleen G. Julian³, Wallace H. Greene³, Lance B. Price⁴, Kenrad Nelson², Cynthia J. Whitener³, Ellen K. Silbergeld¹

1 Department of Environmental Health Sciences, Johns Hopkins Bloomberg School of Public Health, Baltimore, Maryland, United States of America, 2 Department of Epidemiology, Johns Hopkins Bloomberg School of Public Health, Baltimore, Maryland, United States of America, 3 Penn State Hershey Medical Center, Hershey, Pennsylvania, United States of America, 4 Division of Pathogen Genomics, The Translational Genomics Research Institute (TGen North), Phoenix, Arizona, United States of America

Abstract

Antimicrobial resistance, particularly in pathogens such as methicillin-resistant Staphylococcus aureus (MRSA), limits treatment options and increases healthcare costs. To understand patient risk factors, including household and animal contact, potentially associated with colonization with multidrug-resistant MRSA isolates, we performed a prospective study of case patients colonized with MRSA on admission to a rural tertiary care hospital. Patients were interviewed and antimicrobial resistance patterns were tested among isolates from admitted patients colonized with MRSA in 2009–10. Prevalence of resistance was compared by case-patient risk factors and length-of-stay outcome among 88 MRSA case patients. Results were compared to NHANES 2003-04. Overall prevalence of multidrug resistance (non-susceptibility to ≥four antimicrobial classes) in MRSA nasal isolates was high (73%) and was associated with a 1.5-day increase in subsequent length of stay (p = 0.008). History of hospitalization within the past six months, but not antimicrobial use in the same time period, was associated with resistance patterns. Within a subset of working-age case patients without recent history of hospitalization, animal contact was potentially associated with multidrug resistance. History of hospitalization, older age, and small household size were associated with multidrug resistance in NHANES data. In conclusion, recent hospitalization of case patients was predictive of antimicrobial resistance in MRSA isolates, but novel risk factors associated with the household may be emerging in CA-MRSA case patients. Understanding drivers of antimicrobial resistance in MRSA isolates is important to hospital infection control efforts, relevant to patient outcomes and to indicators of the economic burden of antimicrobial resistance.

Citation: Davis MF, Peterson AE, Julian KG, Greene WH, Price LB, et al. (2013) Household Risk Factors for Colonization with Multidrug-Resistant Staphylococcus aureus Isolates. PLoS ONE 8(1): e54733. doi:10.1371/journal.pone.0054733

Editor: Herminia de Lencastre, Rockefeller University, United States of America

Received August 29, 2012; Accepted December 14, 2012; Published January 24, 2013

Copyright: © 2013 Davis et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: Research was supported by grants from the Johns Hopkins Center for a Livable Future (http://www.jhsph.edu/clf/) and the Heinz Foundation (http:// www.heinzfamily.org/aboutus/philanthropies_02.html). AEP and MFD were supported by Johns Hopkins Center for a Livable Future Predoctoral Fellowships. Additionally, MFD was supported by a Sommer Scholarship. The funders had no role in the study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing Interests: The authors have declared that no competing interests exist.

* E-mail: mdavis@jhsph.edu

Introduction

Methicillin-resistant *Staphylococcus aureus* (MRSA) is a pandemic antimicrobial-resistant pathogen [1]. In 2004, an estimated 1.5% of the United States population, or approximately 4 million people, were nasally colonized with MRSA [2]. Nasal colonization increases risk for development of clinical infection [3]. Antimicrobial-resistant pathogens, which include MRSA, have human costs in morbidity and mortality, and they have been estimated to have healthcare costs in excess of \$4 billion annually in the U.S. [4]. As a result, understanding the epidemiology of multidrug-resistant MRSA case-patients is both clinically and economically relevant to healthcare surveillance and control efforts.

MRSA epidemiology in the United States is shifting rapidly, as strains historically considered community-associated enter hospitals, and hospital strains disseminate into the community [5–8]. However, some authors have suggested that isolate antimicrobial susceptibility may continue to distinguish community-acquired (CA-)MRSA isolates from those acquired in the hospital, and that isolate resistance to certain antimicrobials (ciprofloxacin, clindamycin, and aminoglycosides) may typify hospital-acquired (HA-)MRSA isolates [9–12]. In addition, new risk factors for acquisition of MRSA may be emerging. Human households [13–15] and animals [16,17] recently have been described as potential community reservoirs for MRSA.

To describe case-patient epidemiology and evaluate novel household and animal risk factors as potential drivers of antimicrobial resistance, we interviewed MRSA positive case patients identified from nasal colonization surveillance at a tertiary care center serving largely rural and suburban communities. MRSA isolates were tested for antimicrobial susceptibility. We also evaluated associations between isolate drug resistance and subsequent length-of-stay (LOS) among case-patients, using LOS as an economic and human cost marker for potential associations between drug resistance and factors related to hospitalization. Risk factor results were compared to data from MRSA-colonized participants in the National Health and Nutrition Examination Survey (NHANES) 2003–04.



Figure 1. Study design for analysis of risk factors from casepatients interviewed at Penn State Hershey Medical Center. doi:10.1371/journal.pone.0054733.g001

Methods

Research Design

We enrolled patients over the age of 18 years at Penn State Hershey Medical Center (PSHMC) between August 2009 and March 2010 as previously described [18]. As part of a larger casecontrol study, MRSA case patients identified on admission via screening nasal swabs were interviewed as a prospective case cohort to characterize MRSA isolates by multi-locus sequence typing (MLST) and antimicrobial susceptibility patterns from a hospital source population that included predominantly rural and suburban communities [18]. This manuscript is limited to analysis of the case-patients from whom a MRSA isolate was available for antimicrobial susceptibility testing and who had complete data for all risk factors.

Survey

Patients were interviewed for self-reported risk factors that included demographic information; hospitalization within the past month, six months, or year prior to admission for themselves and for family members; antimicrobial drug use within the past month, six months or year prior to admission for themselves and for family members; animal contact, including livestock (cows, pigs, and poultry); household pet ownership (dogs and cats only); and number of people living in the household. Subsequent length of stay was determined through record review.

Sample Collection

Swabs of the anterior nares of patients were collected within 48 hours of admission and these swabs were processed at the PSHMC virology laboratory using the BD GeneOhmTM MRSA Assay (Becton Dickinson Diagnostics, San Diego, CA). MRSA-positive swabs by this PCR method were archived and subsequently cultured for viable MRSA isolates using commercial MRSA

SelectTM agar plates (Bio-Rad Laboratories, Hercules, CA). Isolates were confirmed as MRSA using a real-time PCR assay by detection of *mecA* and *femA* genes (Pathogene, LLC). Due to potential presence of variant *mecA* genes, MSSA isolates found to be beta-lactam resistant were tested for presence of *mecC* using a newly designed universal primer as previously described [19,20].

Antimicrobial Susceptibility Testing

MRSA and MSSA isolates were tested for antimicrobial susceptibility using disc diffusion methods [21,22], including ervthromycin-induced resistance to clindamycin (D-test), following CLSI guidelines [23] to nine antimicrobials: chloramphenicol, quinupristin/dalfopristin (Svnercid), tetracycline, gentamicin, amikacin, trimethoprim/sulfamethoxazole, clindamycin, ciprofloxacin, and erythromycin. Multi-drug resistance (MDR4) was defined as beta-lactam resistance by mecA gene presence plus nonsusceptibility (inducible, intermediate or high-level resistance) to three additional classes of antimicrobial drugs by disc diffusion methods, based on a definition reported by SENTRY [24]. An additional category of high multidrug resistance (MDR5) was included to evaluate whether risk factors differed for isolates more difficult to treat, and this was defined as beta-lactam resistance (mecA gene presence, all isolates by definition) plus high-level (complete) resistance to four additional classes of antimicrobial drugs (*i.e.*, resistance to \geq five antimicrobials). For MDR5 and for individual antimicrobial drug evaluation, risk factors were compared to highlevel (complete) resistance only, including intermediate with susceptible isolates in models, because high-level resistance may be associated with a higher probability of acquired resistance [25], as opposed to resistance based on other mechanisms, e.g. via multiple mutations in cell wall biosynthesis. For clindamycin, inducible resistance was included with high-level resistance phenotypes [26].

All isolates were screened for vancomycin resistance by realtime PCR assay for the vanA gene (Pathogene, LLC) and disc diffusion testing. Because of previous findings of hVISA isolates in this patient population [27], 33 of the isolates were selected for further vancomycin susceptibility testing on the basis of their susceptibility profiles (e.g. quinupristin/dalfopristin non-susceptibility or MDR4). These isolates were screened using a standard VA E-test and also a GRD E-test (vancomycin and teicoplanin) for potential hGISA phenotype as previously described [27]. Isolates with positive GRD E-tests [28] were tested using a population analysis as previously described [27]. Briefly, 10^7 and 10^6 inoculations were placed on agar plates containing 1, 2, 4, 7, and 8 ug/ml vancomycin. Growth at the level of 4 ug/ml with a 10⁶ inoculation was considered indicative of hGISA positivity. Results were validated against quality control strains ATCC 29213, ATCC Mu3 (hGISA) and ATCC Mu50 (GISA).

Statistical Analysis

We estimated unadjusted and adjusted associations for antimicrobial resistance by risk factor using prevalence ratios (PRs). We calculated PRs using Poisson models with robust estimation of standard errors as described previously [29,30] using Stata 11 (College Station, TX). P-values ≤ 0.05 were considered statistically significant, and p-values ≤ 0.10 were considered to approach statistical significance.

A priori, covariates included self-reported age, gender, race, history of hospitalization, prior use of antimicrobials, exposure to animals, and household size. Categorical dummy variables for hospitalization or antimicrobial use within one month compared to within six months were created, assigning 0 if patients selfreported no hospital contact or antimicrobial use within six
 Table 1. Prevalences of antimicrobial resistance by risk factor among 88 MRSA isolates from Penn State Hershey Medical Center

 admitted patients, August 2009 to February 2010.

	Multidrug Resistance (MDR4)	High Multidrug Resistance (MDR5)	Ciprofloxacin Resistant (CIPR)	Clindamycin Resistant (CLIR)	Amikacin Resistant (AMKR) - high-level resistance	
	4+ classes of non- susceptibility	5+ classes of high-level resistance	high-level resistance	high-level resistance		
Overall, N (%)	64 (73%)	20 (23%)	72 (82%)	51 (58%)	21 (24%)	
Gender						
Female, <i>n</i> = 35	25 (71%)	11 (31%)	29 (83%)	20 (57%)	10 (29%)	
Male (<i>ref</i>), <i>n</i> = 53	39 (74%)	9 (17'%)	43 (81%)	31 (58%)	11 (21%)	
Age						
65 years or older, $n = 38$	30 (80%)	10 (26%)	35 (92%)	26 (68%)	7 (19%)	
Under 65 (<i>ref</i>), <i>n</i> = 50	34 (68%)	10 (20%)	37 (74%)	25 (50%)	14 (28%)	
Hospitalization						
Within 1 month, $n = 24$	20 (83%)	9 (38%)	22 (92%)	17 (71%)	8 (33%)	
1–6 mo, <i>n</i> = 18	17 (94%)	5 (28%)	17 (94%)	13 (72%)	2 (11%)	
Over 6 mo (<i>ref</i>), <i>n</i> = 46	27 (59%)	6 (13%)	33 (72%)	21 (46%)	11 (24%)	
Antimicrobial use						
Within 1 month, $n = 36$	31 (86%)	11 (31%)	33 (92%)	24 (67%)	10 (28%)	
1–6 mo, <i>n</i> = 24	18 (75%)	4 (17%)	20 (83%)	13 (54%)	6 (25%)	
Over 6 mo (<i>ref</i>), <i>n</i> = 28	15 (54%)	5 (18%)	19 (68%)	14 (50%)	5 (18%)	
Livestock Exposure						
Direct contact, $n = 12^{\ddagger}$	7 (58%)	1 (8%)	8 (67%)	7 (58%)	3 (25%)	
No direct contact (<i>ref</i>), $n = 76$	57 (75%)	19 (25%)	64 (84%)	44 (58%)	18 (24%)	
Household pets						
Have pets, $n = 49^{\ddagger\ddagger}$	39 (80%)	12 (24%)	40 (82%)	30 (61%)	12 (24%)	
Don't have pets (<i>ref</i>), $n = 39$	25 (64%)	8 (21%)	32 (82%)	21 (54%)	9 (23%)	
Household Size*						
Over 2, <i>n</i> = 32	24 (75%)	8 (25%)	23 (72%)	18 (56%)	9 (28%)	
2 or fewer (<i>ref</i>), <i>n</i> = 56	40 (71%)	12 (21%)	49 (88%)	33 (59%)	12 (21%)	

N (%) shown are for the resistant population compared to the susceptible population. Intermediates are included with resistant isolates for the SENTRY MDR definition, but are included with the susceptible population for the remainder of the categories. Race was not included due to small numbers of non-white participants (N = 6). [‡]Six case patients reported living on a farm; six reported farm occupation; two reported chicken contact, six reported cow contact; and one reported pig contact (categories non-exclusive).

^{‡‡}34 dogs and 22 cats.

*Household size includes index patient.

doi:10.1371/journal.pone.0054733.t001

months, assigning 1 if patients self-reported hospitalization or antimicrobial use within six months prior to admission, and 2 if patients self-reported hospitalization or antimicrobial use within a month of admission. The six-month cut-off and definitions for HA- versus CA-MRSA assignment were selected based on prior work in this study population [18]. Self-reported contact with dogs and cats was colinear with self-reported household pet ownership; hence these variables were aggregated. Due to small numbers, livestock contact was aggregated from individual reporting of pig, poultry, or cow contact. Age, household size, and animal exposure variables were dichotomized. Because of the small numbers with non-white race (n = 6), this risk factor was not examined further.

Linear regression models were run with log-transformed length of stay (LOS) as an outcome, evaluating potential association with antimicrobial resistance patterns. Beta coefficients from logtransformed models were exponentiated to return a point estimate, in days, for average LOS increase in patients colonized with MDR isolates. The Penn State Hershey Medical Center and Johns Hopkins Bloomberg School of Public Health Institutional Review Boards reviewed and approved this study. Patients provided written informed consent to participate in the study.

NHANES Analysis

To provide a descriptive comparison between this geographically-limited study of hospital inpatients and data from a wider U.S. population, statistical analysis was run on a subset of all MRSA-colonized participants in NHANES 2003–04, which represented the most recent national data available to the public on MRSA colonization [31]. Risk factor data available in NHANES 2003–04 included gender, age, self-reported history hospitalization within the past year, and household size [31]. Antimicrobial use data was available only for the prior one month; this variable was not included in analysis due to the inconsistent time frame with the hospitalization variable. Data on animal contact or pet ownership was not available. Methods for **Table 2.** Unadjusted and adjusted prevalence ratios for antimicrobial resistance by risk factor among 88 MRSA isolates from Penn State Hershey Medical Center admitted patients, August 2009 to February 2010.

				High M	ultidrug Resist	ance				Clindar	nycin Resist	ant	:		
	Multid	rug Resistance (MDR4)	(MDR5)			Ciprofl	oxacin Resista	ant (CIPR)	(CLIR)			Amika	cin Resistan	t (AMKR)
	4+ clas: non-su	ses of sceptibility		5+ class high-lev	es of el resistance		high-lev	vel resistance		high-lev	el resistance		high-le	vel resistance	
	æ	95% CI	p- value	R	95% CI	p-value	Æ	95% CI	p-value	R	95% CI	p-value	æ	95% CI	p-value
Gender (male is ref)															
Unadjusted	0.97	0.74-1.27	0.83	1.85	0.85-4.02	0.12	1.02	0.84-1.25	0.84	0.98	0.68-1.41	06.0	1.38	0.65-2.90	0.40
Adjusted	0.96	0.73-1.26	0.78	1.70	0.79–3.64	0.18	1.00	0.82-1.21	0.97	0.91	0.63-1.31	0.61	1.58	0.74-3.38	0.24
Age (under 65 is ref)															
Unadjusted	1.16	0.90-1.49	0.25	1.32	0.61–2.85	0.49	1.24	1.03-1.50	0.02	1.37	0.96-1.95	0.08	0.66	0.29-1.48	0.31
Adjusted	1.23	0.93–1.61	0.15	1.12	0.51–2.47	0.78	1.24	1.02-1.51	0.03	1.42	0.98–2.06	0.07	0.58	0.26-1.27	0.17
Hospitalization (>6 mo is ref)															
Within 1 month, unadjusted	1.42	1.05-1.92	0.02	2.88	1.15-7.16	0.02	1.28	1.03-1.59	0.03	1.55	1.03-2.34	0.04	1.39	0.65-3.01	0.40
Within 1 month, adjusted	1.14	0.79–1.65	0.49	3.15	1.16-8.57	0.03	1.12	0.88-1.44	0.36	1.42	0.82-2.45	0.22	1.28	0.45-3.62	0.64
1–6 months, unadjusted	1.61	1.23-2.10	0.001	2.13	0.74-6.15	0.16	1.32	1.06-1.63	0.01	1.58	1.03-2.43	0.04	0.46	0.11-1.91	0.29
1–6 months, adjusted	1.46	1.09-1.96	0.01	2.38	0.85-6.67	0.10	1.28	1.02-1.62	0.04	1.72	1.06-2.79	0.03	0.31	0.08-1.19	0.09
Antimicrobial use (>6 mo is ref)															
Within 1 month, unadjusted	1.61	1.11-2.33	0.01	1.71	0.67-4.38	0.26	1.35	1.03-1.78	0.03	1.33	0.86-2.07	0.20	1.56	0.60-4.06	0.37
Within 1 month, adjusted	1.42	0.93–2.15	0.10	0.84	0.29–2.46	0.75	1.31	0.98–1.75	0.06	1.10	0.61-1.97	0.75	1.51	0.46–4.99	0.50
1–6 months, unadjusted	1.40	0.92-2.13	0.11	0.93	0.28–3.11	0.91	1.23	0.90-1.68	0.20	1.08	0.64-1.83	0.77	1.40	0.48-4.04	0.53
1–6 months, adjusted	1.16	0.74-1.82	0.51	0.57	0.18-1.84	0.35	1.15	0.83-1.59	0.42	0.81	0.45-1.45	0.47	1.74	0.57-5.34	0.33
Livestock Exposure (no contact is re	ef)														
Bivarite	0.78	0.47-1.28	0.32	0.33	0.04-2.29	0.26	0.79	0.52-1.20	0.27	1.01	0.60-1.69	0.98	1.06	0.36-3.07	0.92
Adjusted	0.79	0.53-1.17	0.24	0.31	0.04-2.39	0.26	0.81	0.57-1.15	0.24	1.03	0.63-1.67	0.92	0.93	0.31-2.79	0.89
Household pets (no pets is ref)															
Unadjusted	1.24	0.94-1.64	0.12	1.19	0.54-2.64	0.66	0.99	0.82-1.21	0.96	1.14	0.79-1.64	0.49	1.06	0.50-2.27	0.88
Adjusted	1.23	0.93–1.61	0.14	1.12	0.49–2.55	0.79	1.03	0.84-1.24	0.80	1.07	0.73-1.57	0.71	0.83	0.37-1.88	0.66
Household Size* (2 or fewer is ref)															
Unadjusted	1.05	0.81–1.36	0.71	1.17	0.53–2.56	0.70	0.82	0.65-1.04	0.11	0.95	0.65–1.39	0.81	1.31	0.62–2.78	0.48
Adjusted	0.93	0.73-1.20	0.59	1.05	0.49–2.27	0.90	0.79	0.63-0.99	0.04	0.94	0.64–1.38	0.51	1.42	0.68-2.96	0.35
Intermediates are included with resistant individuals for whom complete data on al Prevalence ratios (PRs) shown are estimat non-significant but approach significance *Household size includes index patient.	t isolates f ill potentia ted from J e (two-side	for the SENTRY MI al covariates is ava poisson regression ed p<0.10) are ita	DR definition ilable. Adjust models (cat licized. Race	, but are ir ed models egorical m was not in	icluded with the control for gend odels used for hic cluded due to si cluded due to si	susceptible er, age, hist sspitalizatio mall numbe	populatic ory of hos _i n and anti rs of non-	on for the remal pitalization, hist ibiotic use). Sigr white participar	inder of the ory of antin nificant asso nts (<i>n</i> = 6).	e categori nicrobial 1 nciations (es. Unadjuste Jse, livestock two-sided p<	:d and adju exposure, ŀ <0.05) are s.	isted resu nouseholc hown in	ults are limited d pets, and hc bold. Associat	l to the 88 usehold size. ions that are



Figure 2. Risks for antimicrobial resistance in CA-MRSA case-patients of working age (18–65), n=27. *p<0.10. No estimates were statistically significant at the p<0.05 level. ‡ A PR could not be estimated for MDR5 (high multi-drug resistance, 5+ classes of antimicrobial) for household pet presence due to a 0 stratum. Antimicrobial resistance patterns: MDR4: nonsusceptibility to four or more classes of antimicrobial drug; MDR5 ("high multidrug resistance"): high-level (complete) resistance to five or more classes of antimicrobial drug; CIPR: high-level resistance to clindamycin, including inducible resistance; AMKR: high-level (complete) resistance to amikacin.

doi:10.1371/journal.pone.0054733.g002

antimicrobial susceptibility testing in NHANES previously have been described [32]. Identical analysis was run on the NHANES data and the PSHMC datasets for descriptive comparison. Trends also were evaluated for *S. aureus* nasal colonization and identified risk factors in NHANES. Survey weighting was not used for NHANES models limited to MRSA-positive individuals due to the small sample size.

Results

Case-patient Selection

Figure 1 presents the selection process for case-patient inclusion in these analyses. Analysis was restricted to 88 individuals for whom data was complete for risk factors and from whom isolates were available for antimicrobial susceptibility testing. Epidemiologic comparison of the 63 case patients not included in this analysis demonstrated that these patients did not differ significantly in demographic characteristics, rates of prior hospitalization, or self-reported antimicrobial use as the included 88 case patients.

Prevalence of Antimicrobial Resistance

Overall, the prevalence of inducible and high-level resistance to individual antimicrobial drugs among the 88 isolates was: erythromycin, 90%; ciprofloxacin, 82%; clindamycin, 58%; amikacin, 24%; trimethoprim-sulfamethoxazole, 15%; gentamicin, 10%; tetracycine, 7%; quinupristin/dalfopristin (Synercid), 1%; and chloramphenicol, 1%.

All isolates were negative for *vanA* by real-time PCR and were susceptible on vancomycin disc diffusion testing. Based on

antimicrobial susceptibility profile screening, we selected a subset of 33 isolates to evaluate further. All were classified as vancomycin-susceptible based on a MIC of 2 or less by standard vancomycin E-test analysis. However, 16 (48%) were considered suspect for heterogeneous glycopeptide intermediate resistant *S. aureus* (hGISA) phenotype by vancomycin-teicoplanin (GRD) Etest. One (6%) of these 16 isolates was confirmed as a heterogeneous glycopeptide intermediate resistant *S. aureus* (hGISA) on the basis of population analysis, and details on this case are described below. Due to the low prevalence in this cohort (n = 1), vancomycin, quinupristin-dalfopristin, and chloramphenicol resistance were excluded from further analysis.

Case Report

This paper reports on the finding of a methicillin-resistant and heterogeneous glycopeptide-intermediate S. aureus (hGISA) isolate, typed as a ST5 (CC5) on the basis of multi-locus sequence testing [33], from a nasally colonized 76-year old Caucasian female casepatient who reported no personal or household member (n=1)history of hospitalization, antimicrobial drug use, or healthcare occupational contact in the past year and who did not currently reside in a nursing home or have home nursing care on the basis of record review. She did have routine outpatient contact for follow up of chronic medical conditions. Her isolate was highly resistant to all nine antimicrobials tested and represented the only strain we identified with high-level resistance to quinupristin-dalfopristin and chloramphenicol. Although CC5 isolates historically have been associated with healthcare acquisition, recent reports have demonstrated that these strains may be establishing circulation patterns in the community [7], and the finding of such a highly
 Table 3.
 Antimicrobial resistance by risk factor among MRSA isolates comparing data from NHANES 2003–04 and data from Penn

 State Hershey Medical Center 2009–10.
 Pennesistance

	NHANE	S 2003–04					PSHM	C 2009-10				
	n=133						n=88					
	Multidr (MDR4)	ug Resistan	ce	Clinda (CLIR)	mycin Resis	tant	Multidrug Resistance (MDR4)			Clinda (CLIR)	mycin Resi	stant
	4+ class non-sus	es of ceptibility		high-le	evel resistanc	e	4+ cla non-su	sses of usceptibility		high-le	evel resistanc	e
Prevalence	73%			63%			73%			58%		
	PR	95% CI	p-value	PR	95% CI	p-value	PR	95% CI	p-value	PR	95% CI	p-value
Gender (male is ref)												
Unadjusted	1.00	0.81-1.23	0.98	1.04	0.80-1.35	0.78	0.97	0.74-1.27	0.83	0.98	0.68-1.41	0.90
Adjusted	0.98	0.81-1.20	0.87	1.01	0.79–1.30	0.91	0.93	0.71-1.22	0.60	0.91	0.64–1.32	0.63
Age (18–65 is ref)												
Over 65, Unadjusted	1.61	1.24-2.09	0 <0.001	1.82	1.34-2.48	<0.001	1.16	0.90-1.49	0.25	1.37	0.96–1.95	0.08
Over 65, Adjusted	1.36	1.03-1.78	3 0.03	1.46	1.03-2.04	0.03	1.20	0.93–1.55	0.17	1.40	0.97–2.02	0.07
Under 18, Unadjusted	1.30	0.97–1.75	0.08	1.17	0.80-1.71	0.41	-			-		
Under 18, Adjusted	1.44	1.05-1.97	0.02	1.31	0.89–1.93	0.17	-			-		
Hospitalization (>1 year is r	ef)											
Within 1 year, unadjusted	1.30	1.08-1.57	0.005	1.53	1.22-1.92	<0.001	1.22	0.90-1.67	0.20	1.13	0.76–1.68	0.54
Within 1 month, adjusted	1.22	1.00-1.48	8 0.05	1.36	1.07-2.04	0.01	1.23	0.91-1.66	0.19	1.13	0.77-1.67	0.53
Household Size* (under 2 is	ref)											
Unadjusted	0.79	0.65-0.96	5 0.02	0.68	0.53-0.88	0.003	1.05	0.81-1.36	0.71	0.95	0.65–1.39	0.81
Adjusted	0.93	0.73-1.20	0.59	0.76	0.55-1.06	0.10	1.10	0.85-1.41	0.47	1.04	0.70-1.52	0.86

Adjusted models include gender, age, hospitalization, and household size.

doi:10.1371/journal.pone.0054733.t003

drug resistant isolate in a patient with CA-MRSA epidemiology is notable.

Prevalence of Antimicrobial Resistance by Risk Factor

Table 1 presents prevalence for individual resistance patterns and for multidrug resistance (MDR4 and MDR5) according to self-reported variables for gender, age, history of hospitalization and antimicrobial use, animal contact (livestock and household pets separately), and human household size for the 88 casepatients. The rate of multidrug resistance (MDR4) was high overall (73%). Highly multidrug-resistant isolates (MDR5) comprised almost a quarter of isolates. Almost half (48%) of casepatients reported a history of hospitalization within the prior six months, and antimicrobial use in the same period (68%) frequently was found.

Three antimicrobials-clindamycin, ciprofloxacin, and amikacin-were selected to demonstrate trends according to risk factor in part based on the potential utility of these resistance patterns to differentiate isolates as HA-MRSA or CA-MRSA [9–11]. Erythromycin was excluded from analysis due to extremely high resistance prevalence, and others were excluded due to low prevalence, which led to unstable model estimates (zero strata).

Unadjusted and Adjusted Models

Table 2 presents unadjusted and adjusted model estimates of prevalence ratios for each resistance pattern by risk factor. In adjusted models, history of hospitalization was associated with multidrug resistance, clindamycin resistance, and ciprofloxacin resistance, controlling for the effects of other covariates. Such associations with hospitalization were not present for amikacin resistance; instead, history of hospitalization in the one to six months prior to admission had a prevalence ratio (PR) of 0.31, although this association was not significant (p = 0.09). A sensitivity analysis including self-reported information on healthcare occupation reported by the case patient or a household member as recent (≤ 1 mo) hospital contact produced no significant changes in inference, although estimates of association strengthened slightly in all cases.

Estimates of association between resistance patterns and antimicrobial use were weaker, and attenuated to non-significance in adjusted models in all cases. When sensitivity analysis was performed including intermediate with high-level (complete) resistance, antimicrobial use in the previous month was significantly associated with risk for resistance to ciprofloxacin or to multidrug resistance (MDR4), and antimicrobial use remained a significant predictor in adjusted models. In these adjusted models, estimates of association with hospitalization were attenuated and generally non-significant.

Most animal contact or household variables did not show strong trends for association with high-level resistance patterns, except that greater household size was negatively associated with ciprofloxacin resistance (PR 0.79 for household sizes over two, p = 0.04). However, 29 (76%) of the older case-patients lived in households with, at most, one other person. No significant trends in multidrug resistance over time (August 2009 to March 2010) were found (results not shown).

We also performed a sensitivity analysis examining inclusion of indirect effects from household members. We created a dummy variable in which we assigned: (2) the patient reported hospitalization and/or antimicrobial use within the past six months, (1) the patient reported that a he or she had not been hospitalized or taken antimicrobial drugs, but a household member had done so in the last six months, or (0) neither patient nor household member reported a history of hospitalization or antimicrobial use in the past six months. Patient history of hospitalization or antimicrobial use was associated with a 1.51 [95% CI: 1.01–2.25] fold higher risk of multidrug resistance (MDR4) in the patient's isolate, and this was statistically significant (p = 0.05). Household member risk in the absence of patient risk was not associated with multidrug resistance (PR 0.46 [95% CI: 0.08–2.64], p = 0.38), but only four patients reported household member risk in the absence of patient risk.

Associations in CA-MRSA Case-patients

Because of interest in evaluating risks for colonization with multidrug-resistant MRSA among community associated (CA-)MRSA case-patients, we also performed analysis restricted to PSHMC case-patients who had reported no history of hospitalization in the past six months. We stratified this analysis by age due to a priori concerns with potential differences in risk factors for community acquisition of MRSA by age, and also based on evidence from the data that age independently was associated with increases in risk for antimicrobial resistance. We evaluated 27 CA-MRSA case-patients of working age (18-65) and 18 CA-MRSA case-patients over the age of 65. Due to statistical properties (strata with zero observations) in the older sub-cohort, few associations could be estimated for case-patients over the age of 65, but estimates that could be made sometimes were associated in the opposite direction from those of working-age CA-MRSA casepatients, supporting our decision to stratify by age. For workingage CA-MRSA case-patients, Figure 2 presents the predicted PR estimates according to gender, use of antimicrobials in the last month, contact with livestock, household pet contact, and human household size. Household pet contact was associated with a 2.35fold higher prevalence of multidrug resistance (MDR4), but this association was not significant (p = 0.09). Although a PR could not be estimated for associations between MDR5 and household pet contact due to a zero stratum, data from other strata suggested that this was a high-risk category. Estimates of risk with livestock were heavily influenced by a single case patient reporting direct contact with livestock who had an isolate resistant to beta-lactam, erythromycin, ciprofloxacin, clindamycin, and amikacin antimicrobials.

Length of Stay Outcome

Geometric mean length of stay (LOS) was 5.73 days, ranging from 1.25 to over 45 days. On average, among PSHMC casepatients, having a multidrug-resistant isolate (MDR4) on admission was associated with a 1.5-day [95% CI: 1.11–1.97] increase in subsequent length of stay (LOS) in unadjusted models, and this association was statistically significant (p = 0.008). LOS estimates were similar for having a clindamycin-resistant or ciprofloxacinresistant isolate. When history of hospitalization within the prior six months was included in LOS models, the estimate attenuated to a 1.3-day [95% CI: 1.01–1.80] increase (p = 0.05) and for having a clindamycin resistant isolate attenuated to a 1.4-day [95% CI: 1.06–1.80] increase (p = 0.02); the association between ciprofloxacin resistance and LOS lost significance.

NHANES Analysis

In the 2003–04 NHANES population, prevalence of multidrug resistance (MDR4) was 73%, but prevalence of high multidrug

resistance (MDR5) was only 4%, precluding further analysis. Prevalence of clindamycin resistance (inducible or high-level resistance) was 63%. Susceptibility to amikacin and ciprofloxacin were not tested; prevalence of levofloxacin resistance was 46%. Age distribution was: n = 47 for under 18, n = 49 for 18–65, and n = 37 for over 65 years of age. **Table 3** provides results of unadjusted and adjusted models from 133 NHANES MRSAcolonized individuals (2003–04) and 88 PSHMC MRSA-colonized inpatients (2009–10). In both the NHANES 2003–04 and PSHMC 2009–10 populations, older age and history of hospitalization were positively associated with multidrug resistance. In the NHANES 2003–04 population, greater household size was negatively associated with multidrug resistance in MRSA isolates.

Conversely, among the 9004 NHANES participants tested in 2003–04 for any nasal colonization with *S. aureus* (MRSA and MSSA combined), odds were 1.32 fold higher [95% CI: 1.10–1.57] for colonization among participants who lived in larger households (p = 0.005). This association remained significant in survey-weighted, adjusted models controlling for gender, age, and hospitalization within the prior year (OR 1.37 [95% CI: 1.15–1.62], p = 0.001).

Discussion

In this cohort of MRSA case-patients from primarily rural and suburban Pennsylvania, most isolates were susceptible to quinupristin-dalfopristin (Synercid); chloramphenicol; tetracycline; gentamicin; and trimethoprim/sufamethoxazole. Low rates of resistance to tetracycline and trimethoprim/sulfamethoxazole have been found in clinical MRSA isolates in other studies [34,35]. Overall, most isolates were resistant to erythromycin and ciprofloxacin. Other studies have shown high levels of erythromycin resistance in MRSA isolates [9,35-37] and increasing resistance to fluoroquinolones [9,37], including in NHANES [32]. However, the 82% rate of resistance to ciprofloxacin in this cohort was much higher than rates of 40-45% reported in a North American hospital-based prevalence study in 2001-02 [35] or 50-55% rates reported through NHANES for 2001-2004 [32]. The 58% overall rate of resistance to clindamycin in this study was higher than the 30-35% reported in the 2001-02 North American hospital data [35], but was comparable with national trends of inducible or constitutive resistance in isolates collected between 2001-2004 [32].

In this study, recent hospitalization was associated with patient risk for carrying a multidrug-resistant strain of MRSA, and carriage of such strains was associated with increases in length of stay among case-patients admitted to the hospital for medical reasons unrelated to colonization status. Patients with prior hospital contact were more likely to be colonized with multidrug resistant isolates, and patients with prior hospital contact might have been a biased group with more severe medical conditions. However, inclusion of history of hospitalization in models did not negate the association.

When patients with recent hospital contact were excluded from PSHMC analysis and models were limited to working-age casepatients, potential but non-significant associations with animal contact emerged, particularly with dogs and cats. We did not collect information on antimicrobial use in pets, which might be a source of selective pressure for antimicrobial resistance in households. A prior study by Lin and colleagues described five MRSA isolates in animals from PA, with four of canine origin, from a study of clinical isolates collected during 2006–08 [38]. These isolates were of human HA-MRSA MLST types (primarily CC5) and displayed high levels of antimicrobial non-susceptibility to erythromycin, and additionally to veterinary fluoroquinolones, e.g., enrofloxacin, (clindamycin was not tested) [38]. Clinical veterinary data analyzed by Rankin, Morris and colleagues from the Matthew J. Ryan veterinary hospital in Philadelphia demonstrated 100% resistance to clindamycin, ciprofloxacin, and erythromycin among nine PVL-positive clinical MRSA isolates from tested dogs and cats [39], and high clindamycin (72%), erythromycin (85%) and fluoroquinolone (90%) resistance among all 39 MRSA companion animal isolates submitted for 2003-04 [40]. The epidemic of MRSA in humans has been speculated to drive the parallel epidemic in companion animals, in part because companion animals tend to carry strains of MRSA typically associated with human transmission, and in part because households may be points of transmission between humans and companion animals [41-44]. The pilot data we report here may suggest that companion animals can serve as sources of antimicrobial resistance (potentially by harboring drug-resistant S. aureus, veterinary pathogens methicillin-resistant S. pseudintermedius and S. schleiferi, and other staphylococci [15]), but conclusions are limited by the small sample size and cross-sectional nature of the study. Longitudinal studies are needed to evaluate the potential association between animal contact and antimicrobial resistance in MRSA strains found in humans.

This study evaluated isolates from nasal colonization of newlyadmitted hospital inpatients, a group likely to have greater prior exposure to healthcare settings and antimicrobial use than an outpatient population. Due to this potential bias, we also evaluated data from NHANES 2003-04 for similar risk factors. Typically, associations with antimicrobial resistance were in the same direction, although somewhat stronger in the NHANES data, for older age, hospitalization history, and smaller household size risk factors among individuals with MRSA colonization. However, larger household size was a risk factor for S. aureus nasal colonization in the U.S. population generally, indicating that household size may play a complicated role in S. aureus and MRSA epidemiology. Evaluation of MRSA-colonized individuals demonstrated higher risk for isolate antimicrobial resistance in smaller households, although this association was attenuated when models accounted for gender, age, and history of hospitalization. Comparisons between PSHMC and NHANES data are limited

References

- Monecke S, Coombs G, Shore AC, Coleman DC, Akpaka P, et al (2011) A field guide to pandemic, epidemic and sporadic clones of methicillin-resistant *Staphylococcus aureus*. PloS One 6(4): e17936.
- Gorwitz RJ, Kruszon-Moran D, McAllister SK, Mcquillan G, McDougal LK, et al (2008) Changes in the prevalence of nasal colonization with *Staphylococcus aureus* in the United States, 2001–2004. The Journal of Infectious Diseases 197(9): 1226–1234.
- Huang SS, Platt R (2003) Risk of methicillin-resistant *Staphylococcus aureus* infection after previous infection or colonization. Clinical Infectious Diseases 36(3): 281–285.
- 4. American Society for Microbiology (1995) Report of the ASM task force on antibiotic resistance.
- Freitas EAF, Harris RM, Blake RK, Salgado CD (2010) Prevalence of USA300 strain type of methicillin-resistant *Staphylococcus aureus* among patients with nasal colonization identified with active surveillance. Infection Control and Hospital Epidemiology 31(5): 469–475.
- Milstone AM, Carroll KC, Ross T, Shangraw KA, Perl TM (2010) Communityassociated methicillin-resistant *Staphylococcus aureus* strains in pediatric intensive care unit. Emerging Infectious Diseases 16(4): 647–655.
- Miller R, Walker AS, Knox K, Wyllie D, Paul J, et al (2010) 'Feral' and 'wild'type methicillin-resistant *Staphylococcus aureus* in the United Kingdom. Epidemiology and Infection 138(5): 655–665.
- Alvarez CA, Yomayusa N, Leal AL, Moreno J, Mendez-Alvarez S, et al (2010) Nosocomial infections caused by community-associated methicillin-resistant *Staphylococcus aureus* in Colombia. American Journal of Infection Control 38(4): 315–318.

by differences in methodology and potential temporal trends in MRSA epidemiology between 2003–04 (NHANES) and 2009–10 (PSHMC). Future NHANES surveys of national *S. aureus* colonization should consider including survey questions on pet ownership and animal contact.

In conclusion, carriage of ciprofloxacin-resistant, clindamycinresistant, MDR4 and MDR5 MRSA was associated with prior history of hospitalization, but not with history of antimicrobial use, in MRSA case-patients colonized at admission to a tertiary care center. Similarly, prior history of hospitalization, older age (over 65), younger age (under 18), and small household size were risk factors for multidrug resistance and resistance to clindamycin in MRSA-colonized individuals who participated in NHANES 2003–04. Colonization with multidrug-resistant isolates was associated with increases in subsequent length-of-stay for patients in the hospital, with economic and clinical implications. Animal contact, particularly with household pets, may be an emerging risk factor for isolate antimicrobial resistance in case-patients lacking recent history of hospitalization, but this potential association should be confirmed with larger studies.

Acknowledgments

The authors are grateful to Dr. Avanthi Doppalapudi, Nancy L. Sperry and Dr. Michael Spitzer from PSHMC, and Dr. Beth Feingold and Grace Awantang from JHSPH for their invaluable assistance in sample collection. The authors would like to thank Drs. Peter Appelbaum and Klaudia Kosowska-Shick for their perspectives on MRSA in this PSHMC cohort; the PSHMC Diagnostic Virology Medical Technologists for their assistance in isolate collection; and Tania Contente, Jordan Buchhagen, Lindsey Watson, and Lauren Buck for their assistance in MLST preparation and analysis. AEP and MFD appreciate the assistance of thesis mentors, particularly Drs. Patti Gravitt, Jackie Agnew, and Bill Moss from JHSPH and Dr. Karen Carroll from JHSOM.

Author Contributions

Conceived and designed the experiments: MFD AEP KGJ KN CJW EKS. Performed the experiments: MFD AEP KGJ WHG LBP CJW. Analyzed the data: MFD AEP KJG CJW EKS. Contributed reagents/materials/ analysis tools: MFD AEP WHG LBP EKS. Wrote the paper: MFD AEP KGJ WHG LBP KN CJW EKS.

- David MZ, Glikman D, Crawford SE, Peng J, King KJ, et al (2008) What is community-associated methicillin-resistant *Staphylococcus aureus*? The Journal of Infectious Diseases 197(9): 1235–1243.
- Popovich K, Hota B, Rice T, Aroutcheva A, Weinstein RA (2007) Phenotypic prediction rule for community-associated methicillin-resistant *Staphylococcus* aureus. Journal of Clinical Microbiology 45(7): 2293–2295.
- Popovich KJ, Weinstein RA, Hota B (2008) Are Community-Associated Methicillin-Resistant *Staphylococcus aureus* (MRSA) strains replacing traditional nosocomial MRSA strains? Clinical Infectious Diseases 46(6): 787–794.
- Millar BC, Loughrey A, Elborn JS, Moore JE (2007) Proposed definitions of community-associated meticillin-resistant *Staphylococcus aureus* (CA-MRSA). Journal of Hospital Infection 67(2): 109–113.
- Lautenbach E, Tolomeo P, Nachamkin I, Hu B, Zaoutis TE (2010) The impact of household transmission on duration of outpatient colonization with methicillin-resistant *Staphylococcus aureus*. Epidemiology and Infection 138(5): 683–685.
- Lucet J, Paoletti X, Demontpion C, Degrave M, Vanjak D, et al (2009) Carriage of methicillin-resistant *Staphylococcus aureus* in home care settings: Prevalence, duration, and transmission to household members. Archives of Internal Medicine 169(15): 1372–1378.
- Davis MF, Iverson SA, Baron P, Vasse A, Silbergeld EK, et al (2012) Household transmission of meticillin-resistant *Staphylococcus aureus* and other staphylococci. The Lancet Infectious Diseases 12(9): 703–716.
- Smith TC, Moritz ED, Leedom Larson K,R., Ferguson DD (2010) The environment as a factor in methicillin-resistant *Staphylococcus aureus* transmission. Reviews on Environmental Health 25(2): 121–134.

- Cuny C, Friedrich A, Kozytska S, Layer F, Nubel U, et al (2010) Emergence of methicillin-resistant *Staphylococcus aureus* (MRSA) in different animal species. International Journal of Medical Microbiology 300(2–3): 109–117.
- Peterson AE, Davis MF, Julian KG, Awantang G, Greene WH, et al (2012) Molecular and phenotypic characteristics of healthcare- and communityassociated methicillin-resistant *Staphylococcus aureus* at a rural hospital. PloS One 7(6): e38354.
- Garcia Alvarez L, Holden M, Lindsay H, Webb CR (2011) Meticillin-resistant Staphylococcus aureus with a novel mecA homologue in human and bovine populations in the UK and Denmark: a descriptive study. The Lancet Infectious Diseases 11(8): 595–603.
- Ito T, Hiramatsu K, Tomasz A, de Lencastre H, Perreten V, et al (2012) Guidelines for reporting novel *mecA* gene homologues. Antimicrobial Agents and Chemotherapy 56(10): 4997–4999.
- Barry AL, Garcia F, Thrupp LD (1970) An improved single-disk method for testing the antibiotic susceptibility of rapidly-growing pathogens. American Journal of Clinical Pathology 53(2): 149–158.
- Bauer AW, Kirby WM, Sherris JC, Turck M (1966) Antibiotic susceptibility testing by a standardized single disk method. American Journal of Clinical Pathology 45(4): 493–496.
- 23. Clinical Laboratory Standards Institute (2010) 2009 S. aureus CLSI breakpoint guide.
- 24. Moet G, Jones R, Biedenbach D, Stilwell M, Fritsche T (2007) Contemporary causes of skin and soft tissue infections in North America, Latin America, and Europe: Report from the SENTRY antimicrobial surveillance program (1998– 2004). Diagnostic Microbiology and Infectious Disease 57(1): 7–13.
- Turnidge J, Kahlmeter G, Kronvall G (2006) Statistical characterisation of bacterial wild-type MIC value distributions and the determination of epidemiological cut-off values. Clinical Microbiology and Infection 12(5): 418– 425.
- Fiebelkorn KR, Crawford SA, McElmeel ML, Jorgensen JH (2003) Practical disk diffusion method for detection of inducible clindamycin resistance in *Staphylococcus aureus* and coagulase-negative staphylococci. Journal of Clinical Microbiology 41(10): 4740–4744.
- Kosowska-Shick K, Ednie LM, McGhee P, Smith K, Todd CD, et al (2008) Incidence and characteristics of vancomycin nonsusceptible strains of methicillin-resistant *Staphylococcus aureus* at Hershey Medical Center. Antimicrobial Agents and Chemotherapy 52(12): 4510–4513.
- Appelbaum PC (2007) Reduced glycopeptide susceptibility in methicillinresistant *Staphylococcus aureus* (MRSA). International Journal of Antimicrobial Agents 30(5): 398–408.
- Deddens JA, Petersen MR (2008) Approaches for estimating prevalence ratios. Occupational and Environmental Medicine 65(7): 501–506.
- Barros AJD, Hirakata V (2003) Alternatives for logistic regression in crosssectional studies: An empirical comparison of models that directly estimate the prevalence ratio. BMC Medical Research Methodology 3(1): 21.
- Centers for Disease Control and Prevention (2003–2004) National Center for Health Statistics (NCHS). National Health and Nutrition Examination Survey

data. Hyattsville, MD: U.S. Department of Health and Human Services, Centers for Disease Control and Prevention.

- Tenover FC, McAllister S, Fosheim G, McDougal LK, Carey RB, et al (2008) Characterization of *Staphylococcus aureus* isolates from nasal cultures collected from individuals in the United States in 2001 to 2004. Journal of Clinical Microbiology 46(9): 2837–2841.
- Enright MC, Day NP, Davies CE, Peacock SJ, Spratt BG (2000) Multilocus sequence typing for characterization of methicillin-resistant and methicillinsusceptible clones of *Staphylococcus aureus*. Journal of Clinical Microbiology 38(3): 1008–1015.
- Bordon J, Master RN, Clark RB, Duvvuri P, Karlowsky JA, et al (2010) Methicillin-resistant *Staphylococcus aureus* resistance to non beta-lactam antimicrobials in the United States from 1996 to 2008. Diagnostic Microbiology and Infectious Disease 67(4): 395–398.
- Winston LG (2009) Antimicrobial resistance in staphylococci: Mechanisms of resistance and clinical implications. Chapter in: Antimicrobial Drug Resistance.
- Limbago B, Fosheim GE, Schoonover V, Crane CE, Nadle J, et al (2009) Characterization of methicillin-resistant staphylococcus aureus isolates collected in 2005 and 2006 from patients with invasive disease: A population-based analysis. Journal of Clinical Microbiology 47: 1344–1351.
- Diep BA, Chambers HF, Graber CJ, Szumowski JD, Miller LG, et al (2008) Emergence of multidrug-resistant, community-associated, methicillin-resistant *Staphylococcus aureus* clone USA300 in men who have sex with men. Annals of Internal Medicine 148: 249–257.
- Lin Y, Barker E, Kislow J, Kaldhone P, Stemper ME, et al (2011) Evidence of multiple virulence subtypes in nosocomial and community-associated MRSA genotypes in companion animals from the upper midwestern and northeastern United States. Clinical Medicine & Research 9(1): 7–16.
- Rankin S, Roberts S, O'Shea K, Maloney D, Lorenzo M, et al (2005) Panton Valentine Leukocidin (PVL) toxin positive MRSA strains isolated from companion animals. Veterinary Microbiology 108(1–2): 145–148.
- Morris DO, Rook KA, Shofer FS, Rankin SC (2006) Screening of Staphylococcus aureus, Staphylococcus intermedius, and Staphylococcus schleiferi isolates obtained from small companion animals for antimicrobial resistance: A retrospective review of 749 isolates (2003–04). Veterinary Dermatology 17(5): 332–337.
- Weese JS, van Duijkeren E (2010) Methicillin-resistant Staphylococcus aureus and Staphylococcus pseudintermedius in veterinary medicine. Veterinary Microbiology 140(3–4): 418–429.
- Loeffler A, Lloyd DH (2010) Companion animals: A reservoir for methicillinresistant *Staphylococcus aureus* in the community? Epidemiology and Infection 138(5): 595–605.
- Bramble M, Morris D, Tolomeo P, Lautenbach E (2011) Potential role of pet animals in household transmission of methicillin-resistant *Staphylococcus aureus*: A narrative review. Vector Borne and Zoonotic Diseases 11(6): 617–620.
- Baptiste KE, Williams K, Willams NJ, Wattret A, Clegg PD, et al (2005) Methicillin-resistant staphylococci in companion animals. Emerging Infectious Diseases 11(12): 1942–1944.