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Anti-SARS-CoV-2 and cytotoxic activity of two marine alkaloids from green alga *Caulerpa cylindracea* Sonder in the Dardanelles†

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Caulerpa cylindracea Sonder is a green alga belonging to the Caulerpaceae family. This is the first chemical investigation of *C. cylindracea* in the Dardanelles which resulted in the isolation of four compounds, caulerpin (1), monomethyl caulerpinate (2), beta-sitosterol (3), and palmitic acid (4). Their structures were elucidated by spectroscopic analyses including 1D- and 2D NMR and mass. The isolated compounds 1 and 2 were tested against the SARS-CoV-2 viral targets spike protein and main protease (3CL) enzyme, and both compounds significantly inhibit the interaction of spike protein and ACE2, while the main protease activity was not significantly reduced. Docking studies suggested that compounds 1 and 2 may bind to the ACE2 binding pocket on spike, and compound 2 may also bind to an allosteric site on spike. As such, these compounds may inhibit the spike–ACE2 complex formation competitively and/or allosterically and have the potential to be used against SARS-CoV-2 virus infection. In addition, compounds 1 and 2 showed at least two-fold higher cytotoxicity against breast cancer cell lines MCF7 and MDA-MB-231 compared to the CCD fibroblast control cell line.

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1 Introduction

Severe Acute Respiratory Syndrome Coronavirus 2 (SARS-CoV-2) emerged in China at the end of 2019 and has spread all over the world, causing serious health risks to both humans and animals. Up to now, the total number of cases in the world is over 595 million, while the number of deaths is 6.4 million according to Worldometer Online, <https://www.worldometers.info/coronavirus/>, (accessed August 2022). The U.S. Food and Drug Administration (FDA) approved the first mRNA-based vaccine (Pfizer-BioNTech Covid-19) on August 23, 2021. This vaccine and other vaccines have emergency use authorization in the world, and are rejected by a certain majority due to short-term side effects such as low-

grade fever or pain or redness at the injection site or uncertainty of long-term side effects. In addition, no specific approved anti-viral drug is yet available for the treatment of the disease except FDA-approved drug remdesivir (Veklury). Considering all of this, drug discovery studies, particularly for antiviral agents, have become even more important, especially in natural products, which have low toxicity and fewer adverse effects. The entry of SARS-CoV-2 into host cells depends on the combination of its spike glycoprotein (S protein) and angiotensin-converting enzyme II (ACE2) on the human cell membranes.¹ Interfering or blocking the binding of S protein and ACE2 in cells will be beneficial in drug discovery against COVID-19. In recent years, many research groups have worked on this target to discover lead compounds. Therefore, several publications of new anti-viral agents derived from synthetic and natural sources including repurposing studies have been made and are still going on to be published.^{2–4}

Seaweeds have been consumed as food by humans for centuries, and the present worldwide market is worth more than \$6 billion per year, with an annual volume of around 12 million tonnes in 2018. Macroalgae are prevalent in coastal ecosystems and provide a source of bioactive metabolites with a variety of potential and noteworthy biological activities, capable of impacting the abundance, distribution, and survival of marine organisms.⁵

Seaweeds are classified into three major groups, including Chlorophyta (green algae), Rhodophyta (red algae), and Ochrophyta (brown algae) based on their color. It is estimated

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that 1800 green, and about 1800 brown macroalgae, and 6200 red macroalgae are found in the marine environment.⁶ Caulerpaceae is one of the green alga families, and represented by a unique genus "*Caulerpa*". The genus *Caulerpa* has 101 species, with 40 varieties and 67 forms in southern Australia.⁷ They are typically grown in shallow-water tropical and subtropical marine habitats. Seaweeds are well-known for their polysaccharides, minerals, and vitamins.⁸ Marine algae have also been identified as a possible source of antioxidants^{9,10} as well as readily available food, particularly in coastal populations.^{11–13} The fatty acids and enzymatic and non-enzymatic antioxidant capabilities of *Caulerpa* species have been studied by some researchers.^{14–16}

Caulerpa cylindracea Sonder is a species of seaweed that is native to the Australian coast in a large area extending from around Perth North into the Mid-West and Pilbara to the Kimberley region of the Western Australia. However, *C. cylindracea* was first introduced to the Mediterranean Sea in the early 1990s, where it has spread extensively and is considered an invasive species. Although this species has been explored in terms of taxonomy, invasiveness, and ecological features, as well as bioactivities and applications,¹⁷ it has not been studied in terms of isolation of its secondary metabolites.

In the present study, the green alga *Caulerpa cylindracea* Sonder was collected in the North Aegean Sea (Dardanelles), and investigated for secondary metabolites for the first time. The four pure compounds caulerpin (1), monomethyl caulerpinate (2), beta-sitosterol (3) and palmitic acid (4) were isolated and their structures were elucidated by 1D- and 2D-NMR and HRMS techniques.

Caulerpin is a red pigment, which was first isolated from a green alga *Caulerpa* spp., and characterized as a phenazine derivative in 1970,¹⁸ the chemical structure was later reasigned.¹⁹ It was then isolated from several *Caulerpa* species, such as *Caulerpa peltata*,²⁰ *C. ashmeadii*,²¹ *C. racemosa*,²² *C. serrulata*,²³ *C. prolifera*,²⁴ *C. lentillifera*,²⁵ *C. microphysa*,²⁵ *C. sertularioides*,²⁵ *C. taxifolia*,²⁶ and *C. lamourouxii*.²⁷ Caulerpin was also isolated from other alga genera: *Halimeda incrossate*, *Hypnea concornis*, *Caloglossa lepriouri*,²⁸ *Chondria armata*,²⁹ *Laurencia majuscula*,²⁸ and from a few brown alga *Spatoglossum asperum*,³⁰ and *Sargassum platycarpum*.³¹ However, we couldn't isolate caulerpin or derivatives in *Laurencia obtusa* collected from North Aegean Sea in our previous study.³² A number of bioactivity studies have been carried out with caulerpin, including cytotoxic, antitumor, anticancer, anti-larvicidal, anticorrosion, antitubercular, antimicrobial, antiviral, antiherpes, antidiabetic, anti-inflammatory, spasmolytic, antinociceptive, and plant growth regulatory activities.^{28,33–35}

Searches for combination therapies were carried out with a molecular docking approach, and many potential drugs were investigated.¹ There are several antiviral studies on caulerpin. In an earlier investigation, caulerpin was found to be an alternative agent against herpes simplex virus type-1.³⁶ In another study, caulerpin exhibited strong anti-viral activity ($EC_{50} = 0.8 \mu\text{M}$) against Chikungunya virus.³⁷ Caulerpin is also being studied as a possible SARS-CoV-2 protein

inhibitor. Results of a recent study based on *in silico* techniques, caulerpin has the highest binding affinity among all examined receptors when compared to other bioactive compounds.^{31,38}

Thus, we have investigated isolated two compounds from *C. cylindracea* against the SARS-CoV-2 viral targets spike protein and main protease (3CL) enzyme along with their docking studies.

In this study, the cytotoxic activity of the extract of *C. cylindracea*, and two isolates caulerpin and monomethyl caulerpinate were also investigated against the aggressive breast cancer cell line MDA-MB-231 for the first time besides breast cell line MCF-7. In fact, in a previous studies, caulerpin was evaluated by Li *et al.* against a panel of human cancer cell lines, and IC_{50} values were found to be 4.67 (human leukemia: K562), 4.20 (human lung: A549), 1.95 (human cervical: HeLa), 4.04 (human colon: HT29), 3.71 (human breast: SK-BR-3), and 0.72 (human liver: Huh7) μM ,³⁹ but not on MDA-MB-231 and MCF-7 breast cell lines.

2 Results and discussion

2.1 Structure elucidation of the isolated compounds

Chemical investigation of the CH_2Cl_2 -MeOH extract of *C. cylindracea* afforded four known (1–4) compounds.

Compound 1 was isolated as an orange-red amorphous solid needles; mp. 315.5–317 °C. Its molecular formula was determined to be $\text{C}_{24}\text{H}_{17}\text{N}_2\text{O}_4$ corresponding to an $[\text{M}-\text{H}]^-$ ion peak at m/z 397.11945 (calcd for $\text{C}_{24}\text{H}_{17}\text{N}_2\text{O}_4$, 397.11880) in the HR-MS spectrum with the combination of ^1H and ^{13}C -NMR experimental data (ESI⁺). The ^{13}C NMR (BB) experiment implied the presence of twelve carbon atoms suggesting that this pigment is a dimer, exhibited only 11 signals owing to C-2 symmetry which verified by the Orbitrap high resolution mass spectrometer.

The ^{13}C NMR spectrum (125 MHz, CDCl_3 , Table 1) of compound 1 showed the presence of a methyl ester carbonyl in the molecule with two signals resonated at δ 166.69 ($\text{OaC} - \text{O}$) and at δ 52.55 ($-\text{OCH}_3$). Aromatic carbons were observed at δ 142.82, 137.72, 132.87, 128.16, 125.48, 123.40, 120.75, 118.07, 112.49 and 111.55. The ^1H -NMR data indicated the presence of two methyl signals at δ 3.90 (6H, s). The chemical shift value at δ 9.20 (2H, s) is attributed the presence of N-H groups. A chemical shift observed at δ 8.06 is assigned to the two proton signals as singlets of an eight-member ring. A pair of doublet of triplet at δ 7.31 (2H, dt, 8.1, 0.9 Hz, 1H), and δ 7.43 (2H, dq, 8.0, 0.8 Hz, 1H), and a signal corresponding to two protons at δ 7.10 (2H, ddd, 8.0, 7.0, 1.0 Hz), and δ 7.18 (2H, ddd, 8.1, 7.0, 1.1 Hz) proposed the bis-indole skeleton of the structure, based on all NMR (Table 1), and mass results and literature data,^{31,40,41} structure of the isolated compound 1 was found to be identical to caulerpin (Fig. 1).

Compound 2 was isolated as an orange-red amorphous solid; mp. 160–162 °C (decomposition). Its molecular ion peak observed at m/z (rel. int.) 384.11128 (26) in the HRESI-MS which corresponded to a molecular formula $\text{C}_{23}\text{H}_{16}\text{N}_2\text{O}_4$. But, $[\text{M}-\text{H}]^-$

Table 1 ^1H - and ^{13}C -NMR data (500 MHz and 125 MHz) for compounds 1 and 2

| Pos. | Caulerpin (1) (in CDCl_3) | | Monomethyl caulerpinate (2) (in CD_3OD) | | HMBC (H \rightarrow C) |
|--------|-------------------------------------|-------------------------------------|--|-------------------------------------|--------------------------|
| | δ_{C} | $\delta_{\text{H,mult.}}$ (J Hz) | δ_{C} | $\delta_{\text{H,mult.}}$ (J Hz) | |
| 1 (N) | — | 9.20 s (2H) | — | — | — |
| 2 | 132.87 | 8.06 s (2H) | 131.60 | — | — |
| 3 | 112.49 | — | 112.64 | — | — |
| 3a | 128.16 | — | 127.99 | — | — |
| 4 | 118.07 | 7.43 dt (2H, 8.0, 0.8) | 116.99 | 7.32 dt (1H, 7.9, 1.1) | C5, C7, C7a |
| 5 | 120.75 | 7.10 ddd (2H, 8.0, 7.0, 1.0) | 121.82 | 6.94 dtd (1H, 7.9, 7.0, 1.0) | C3a, C7 |
| 6 | 123.40 | 7.18 ddd (2H, 8.1, 7.0, 1.1) | 119.03 | 7.02 dddd (1H, 8.2, 7.2, 2.4, 1.2) | C4, C7a |
| 7 | 111.55 | 7.31 dt (2H, 8.1, 0.9 Hz) | 110.69 | 7.22 dt (1H, 8.1, 0.9) | C6, C3a |
| 7a | 137.72 | — | 137.66 | — | — |
| 8 | 142.82 | 8.06 s (2H) | 135.11 | 7.76 s (1H) | C10, C2', C2, C3 |
| 9 | 125.48 | — | 133.78 | — | — |
| 10 | 166.68 | — | 173.22 | — | — |
| 11 | 52.55 | 3.90 s (6H) | — | — | — |
| 1' (N) | — | — | — | — | — |
| 2' | — | — | 136.77 | — | — |
| 3' | — | — | 110.59 | — | — |
| 3a' | — | — | 128.39 | — | — |
| 4' | — | — | 117.52 | 7.41 dt (1H, 7.9, 1.0) | C5', C7a' |
| 5' | — | — | 121.82 | 6.94 dtd (1H, 7.9, 7.0, 1.0) | C3a', C7' |
| 6' | — | — | 119.49 | 7.02 ddd (1H, 8.1, 7.0, 1.1) | C4', C7a' |
| 7' | — | — | 111.06 | 7.25 dt (1H, 8.1, 0.9) | C6', C3a' |
| 7a' | — | — | 138.02 | — | — |
| 8' | — | — | 142.88 | 8.12 s (1H) | C10', C2', C2, C9' |
| 9' | — | — | 125.13 | — | — |
| 10' | — | — | 167.18 | — | — |
| 11' | — | — | 51.21 | 3.80 s (3H) | C10' |

peak of compound 2 were observed at m/z 383.10364 (100) as the main ion (calcd for $\text{C}_{23}\text{H}_{15}\text{N}_2\text{O}_4$, 383.10320) in the HRESI-MS spectrum. The fragment ions were observed at m/z (rel. int.)

369.08786 $[\text{M}-\text{CH}_3]^+$ (16), 353.34222 $[\text{M}-\text{OCH}_3]^+$ (2), 325.18411 $[\text{M}-\text{OCOCH}_3]^+$ (6). Its 1D- and 2D-NMR spectra were very similar to those of compound 1. However, the main difference of compound 2 from caulerpin (1) was observed in the ^1H - and ^{13}C -NMR spectra (Table 1) due to the lack of one of the methyl signals (ESI $^+$). While in caulerpin, the molecule has a symmetry axis, in compound 2 this symmetry axis was not present anymore due to one of the ester groups converted into acid group. Therefore, acid carbonyl-C was observed at δ 173.22 while ester carbonyl resonated at δ 167.18. In addition, the proton signals H-8 and H-8' were differed appearing at δ 7.76 and 8.12, each showed three-bond away correlations with C-2 and C-2' in the HMBC (Table 1, Fig. 2) experiment. In addition, the rest of C-signals of compound 2 were observed

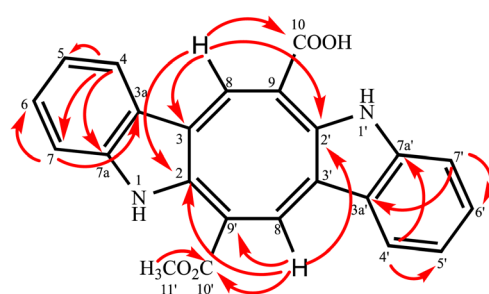
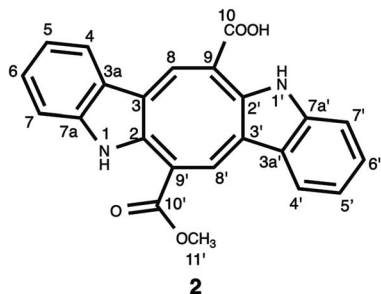
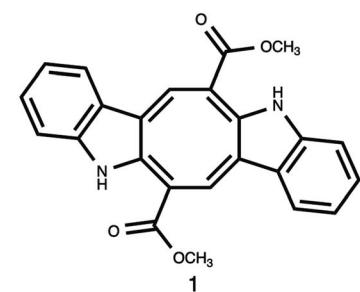


Fig. 1 Chemical structure of the isolated bis-indole alkaloids (1–2).

Fig. 2 HMBC correlation of the monomethyl caulerpinate.

separately, except a C signal resonated at δ 121.82 for two carbon atoms (C-5, 5'). As a result, totally 22C signals resonated for 23 carbon atoms consisting of 10 aromatic methine C's, 10 quaternary C's, 2 carbonyl C's (an acid and an ester), and a methoxy carbon.

Thus, structure of compound 2 was elucidated as mono-methyl caulerpinate considering 1D- and 2D-NMR and HRMS spectroscopic analyses, isolated from a marine green-alga *Caulerpa cylindracea*. In fact, it was first isolated from *C. racemosa*,²² but, the previous ¹H-NMR data were not given sufficiently, even no ¹³C-NMR data were present. The ¹H-NMR and ¹³C-NMR spectra of monomethyl caulerpinate (2) were taken in MeOD-d₄, for the first time, herein.

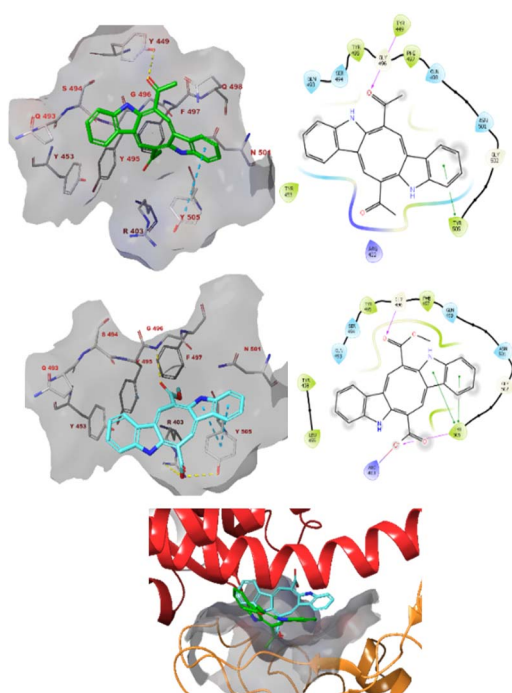


Fig. 3 The docked pose caulerpin (top) and monomethyl caulerpinate (middle) with the SARS-CoV-2 spike protein (pdb 6m0j). The ligand interacts with the pocket on spike to which ACE2 also binds (bottom). Hydrogen bonds are indicated in yellow dashed (3D) or purple solid (2D) plots) lines. π - π stackings are shown in blue dashed (3D) and green solid (2D) plots) lines.

2.2 Caulerpin and monomethyl caulerpinate inhibited spike binding but not main protease activity

The purpose of main protease inhibition assay was to assess whether caulerpin and mono methyl caulerpinate inhibit the main protease enzyme which is important in replication of virus.

Caulerpin did not show any significant enzyme inhibition at any of concentrations. The all concentrations were tested two separate studies at triplicates, 10 nM of caulerpin also prevented the fluorescence measurement. The reason might be due to autofluorescence characteristics of caulerpin that can mask the fluorescence at 460 nm (Fig. 4a). Monomethyl caulerpinate caused slight decrease in main protease activity at higher concentrations. Maximum tested concentration 100 μ M of mono methyl caulerpinate reduced 22.4% enzyme activity, SARS-CoV-2 virus (Fig. 4b). Viral spike and human ACE2 protein which was not significant. Additionally, no other concentrations significantly reduced the main protease enzyme activity of binding were tested to understand whether caulerpin and monomethyl caulerpinate can inhibit the interaction between these proteins. Both of the molecules significantly inhibited the spike-ACE2 binding ($p < 0.001$). Half maximal inhibitory concentration (IC_{50}) of caulerpin was 87.5 nM and IC_{50} of monomethyl caulerpinate was 190.2 nM (S17, S18) maximum inhibition at 100 μ M concentration for caulerpin was 59% (Fig. 4a) and 66% (Fig. 4b) for monomethyl caulerpinate compound. Overall, it can be stated that caulerpin and mono methyl caulerpinate can significantly inhibit the spike-ACE2 binding, but not main protease activity. Therefore, both compounds can be used to prevent SARS-CoV-2 infection rather than directly targeting virus replication.

2.3 Molecular modelling studies

Caulerpin and monomethyl caulerpinate were docked into two different crystal structures of the SARS-CoV-2 spike protein receptor binding domain.⁴²

The study revealed that both compounds could interact with the pocket of the spike protein with which ACE2 also interacts and as such interfere with the binding of the latter (Fig. 3). Caulerpin forms a hydrogen bond with Tyr449 and π - π stacking with Tyr505. On the other hand, monomethyl caulerpinate forms more extensive interactions with the binding pocket, including hydrogen bonds with the sidechains of Arg403 and

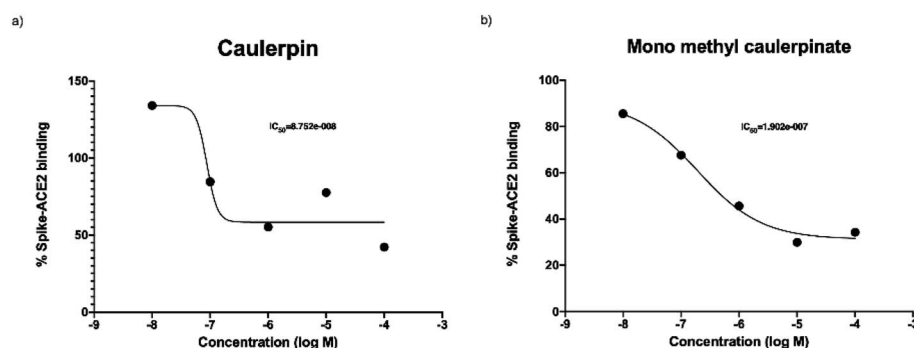


Fig. 4 Spike and ACE2 binding activity of (a) caulerpin and (b) monomethyl caulerpinate at different concentrations.

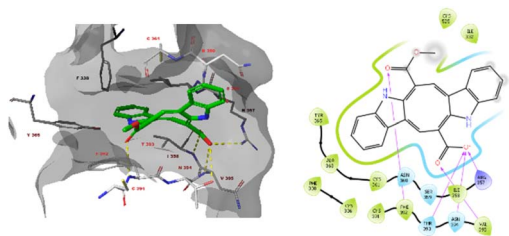


Fig. 5 The docked pose monomethyl caulerpinate with the SARS-CoV-2 spike protein (pdb 7l4z). Hydrogen bonds are indicated in yellow dashed (3D) or purple solid (2D plots) lines. π - π stackings are shown in blue dashed (3D) and green solid (2D plots) lines.

Tyr505 and the backbone of Gly496. In addition, the anionic carboxylic acid tail of the ligand forms electrostatic interactions with Arg403 and π - π stacking with Tyr505 are formed.

Furthermore, monomethyl caulerpinate was able to interact with the pocket of the spike protein where the cyclic peptide inhibitor was bound in the 7l4z cocrystal structure (Fig. 5). The ligand forms hydrogen bonds with the sidechains of Arg357 and Thr393 and with the backbones of Phe392, Asn394 and Val395.

The modelling studies suggest that especially monomethyl caulerpinate and possibly also caulerpin may bind to different sites on the SARS-CoV-2 spike protein and interfere with the formation of the spike-ACE2 complex which is necessary to viral cell entry.

2.4 Strong cytotoxic activity profile

It is worthy that the IC_{50} values of the alga extract of *C. cylindracea* showed no toxic effect in the healthy fibroblast cell line (CCD-1079Sk) as well as in cancer cell lines, which indicates that the possible use as food of this *Caulerpa* species (Table 2). On the other hand, the isolated two bis-indole alkaloids from *C. cylindracea* showed strong activity both in the MDA-MB-231 and the MCF-7 breast cell lines. Moreover, monomethyl caulerpinate showed high IC_{50} value ($424.6 \pm 0.47 \mu\text{g mL}^{-1}$) on the healthy cell lines, particularly after 24 hours incubation that indicated healthy cells remain viable, almost two-fold higher than the standard drug doxorubicin. Thus, monomethyl caulerpinate exhibited almost six times higher viability compared to that of caulerpin. In this work, two bis-indole alkaloids

isolated from *C. cylindracea* tested for the first time against both the MCF-7 and the aggressive MDA-MB-231 cell lines. These results indicate that both compounds have a high potential to be lead drugs in the treatment of aggressive breast cancer.

3 Experimental

3.1 General experimental procedures

NMR data were acquired using Bruker Magnet System 500/54 Ascend 500 NMR spectrometer ($^1\text{H-NMR}$ 500 MHz, $^{13}\text{C-NMR}$ 125 MHz). HRESI-MS analyses were carried out by Thermo Orbitrap Q-Exactive mass spectrometer at Drug Application and Research Center, Bezmialem Vakif University, Istanbul. Melting points were recorded on a STUART SMP40. All the NMR solvents were obtained from Merck. Antiviral activity was analyzed by using multimode microplate reader (HIDEX Sense) for SARS-CoV-2 assays (BPS Biosciences, MBP-tagged Assay Kit #79955-2, ACE2 Inhibitor Screening Assay Kit, #79931).

3.2 Alga material and extraction procedures

3.2.1 Seaweed collection. The green alga *Caulerpa cylindracea* (Fig. 6) was collected from the Dardanelles (Çanakkale Boğazi) at coordinates of the $40^{\circ}4'26.86''\text{N}$ $26^{\circ}21'28.08''\text{E}$ in August 2017 at 5–10 m depth by SCUBA diving in the north Aegean Sea, which is a 61 km long and 1.2–6.4 km wide (narrow) strait. The Dardanelles, also known as the Strait of Gallipoli, is the boundary between Asian and European Turkey. A voucher specimen number E107 is deposited at the Faculty of Aquatic Sciences and Fisheries, Akdeniz University, Antalya, Turkey, where its identity was confirmed by Dr Emine Sukran Okudan.

3.2.2 Extraction and isolation. The shade dried and pulverized alga (225.0 g) was extracted with CH_2Cl_2 -MeOH (1 : 1, 2.4 L \times 10 days) at room temperature. The solution was concentrated to give an extract EEL-CC (8.62 g), by using a rotary evaporator under reduced pressure at 38–40 $^{\circ}\text{C}$ (yield 3.83%).

The extract (EEL-CC, 8.0 g) was applied to the column chromatography (CC) over a silica gel column (70 cm \times 5 cm), [silica gel pore size; 60 Å (70–230 mesh)]. The elution was started using hexane, followed by increasing 25% polarity of CH_2Cl_2 , then after 100% CH_2Cl_2 elution, polarity increased using acetone as a gradient, and then MeOH was used as final gradient to afford totally 23 fractions (EEL-CC-1—EEL-CC-23).

Table 2 Cytotoxic activity results of the isolated compounds and the extract from *C. cylindracea* Sonder (IC_{50})

| | Cell lines | | | | | |
|--|-------------------|-------------------|-------------------|-------------------|-------------------|-------------------|
| | CCD-1079Sk | | MDA-MB-231 | | MCF7 | |
| | 24 h | 48 h | 24 h | 48 h | 24 h | 48 h |
| <i>C. cylindracea</i> extract ^a | 465.87 \pm 1.87 | 295.91 \pm 0.25 | 369.12 \pm 1.18 | 245.45 \pm 0.24 | 346.58 \pm 2.56 | 217.64 \pm 0.65 |
| Caulerpin | 77.44 \pm 0.12 | 36.2 \pm 0.69 | 11.85 \pm 1.03 | 14.68 \pm 0.51 | 15.68 \pm 0.38 | 13.82 \pm 0.72 |
| Monomethyl caulerpinate | 424.6 \pm 0.47 | 53.97 \pm 0.86 | 13.96 \pm 0.52 | 20.59 \pm 0.92 | 16.82 \pm 1.12 | 17.28 \pm 1.32 |
| Doxorubicin | 284.5 \pm 0.04 | — | 62.8 \pm 0.06 | — | 42.39 \pm 0.08 | — |

^a IC_{50} value is calculated as $\mu\text{g mL}^{-1}$ for only the extract, for the others given as μM .



Fig. 6 *Caulerpa cylindracea* Sonder (in Dardanelles-Turkey).

Fr. EEL-CC-8 (377.5 mg) was subjected to a silica gel column (70 cm × 2.2 cm) eluting with a solvent mixture [hexane : CH₂Cl₂ (1 : 4)] to isolate compound 1 (caulerpin, 70 mg). Frs. EEL-CC-18 and EEL-CC-19 were combined (440.0 mg), then they were uploaded to a silica gel column (80 cm × 2.2 cm) and eluted with CHCl₃ : MeOH (9 : 1) to give 33 fractions. Fr. EEL-CC-18-29 was re-chromatographed on a Sephadex LH-20 column (60 cm × 2 cm) using 100% MeOH to afford compound 2 (monomethyl caulerpinate, 9.5 mg). Furthermore, Fr. EEL-CC-7 (918.6 mg) was subjected to a silica gel column (80 cm × 2.2 cm) and eluted with a mixture [hexane : diethyl ether (3 : 1)] to isolate compound 3 (β-sitosterol, 75 mg). Fr. EEL-CC-14-4 (108.2 mg) was uploaded to a silica gel column (50 cm × 2.3 cm) and eluted with hexane : acetone (4 : 1) to give compound 4 (palmitic acid, 30 mg).

3.3 Anti-viral activity

3.3.1 Main protease (3CL) enzyme inhibition assay. The main protease (3CL) screening assay was used to test the inhibitory potential of caulerpin (compound 1) and monomethyl caulerpinate (compound 2). Compounds 1 and 2 were solubilized to 100 mM stock solutions by using dimethyl sulfoxide (DMSO). Compounds were diluted to final concentrations of 10 nM, 100 nM, 1 μM, 10 μM and 100 μM with assay buffer. Maximum DMSO concentration was 1% and 150 ng of 3CL protease enzyme added to each well. GC376 provided by the assay kit which is a 3CL-main protease inhibitor, was used as inhibitory control of the assay. Results were relatively compared with the GC376 activity. Final concentration of 50 μM 3CL-main protease substrate was added to the mixture and incubated for 4 hours at room temperature. Microplate reader was used to measure fluorescence at 360 nm excitation and 460 nm emission wavelength. Data were calculated based on the formula: fluorescence values were subtracted from GC376 inhibitor control fluorescence value and was set as zero percent activity and the fluorescence value from no inhibitor control was set as 100% activity. The data were shown with GraphPad Prism 8.0 software.

3.3.2 Spike-ACE2 binding inhibition assay. Inhibition of the viral spike and human ACE2 protein binding was assessed by using SARS-CoV-2 spike: ACE2 inhibitor screening assay. The tested concentrations of caulerpin and monomethyl caulerpinate were prepared as described. Protocol was followed as manufacturer suggested: first, wells were coated with SARS-CoV-

2 spike protein and then incubated overnight at 4 °C. Next day, plate was washed with assay buffer several times, blocking was done with blocking buffer for 1 hour. Compounds 1 and 2 were added to the spike protein coated wells at different concentrations and, then incubated at room temperature with slow shaking for 3 hours in presence of 2.5 ng μL⁻¹ ACE2-His protein. As the final step, chemiluminescence was produced by adding anti His-HRP and then HRP substrate and measured by microplate reader.

3.4 Docking studies against the receptor binding domain of SARS-CoV-2 spike protein

The crystal structures of SARS-CoV-2 spike protein in complex with ACE2 (pdb 6m0j) and a cyclic peptide inhibitor (pdb 7l4z) were obtained from the RCSB Protein Data Bank. Subsequently, the structure was prepared using the protein preparation tool of Schrödinger (v2022-3, Schrödinger, Inc., New York, USA). All water and buffer molecules were omitted. Subunit A was retained and all other subunits, if present, were omitted. Subsequently, hydrogen atoms were added (pH 7.4), and the system was minimized using the OPLS4 forcefield.

The three-dimensional structures of caulerpin and monomethyl caulerpinate were prepared using the LigPrep tool of Schrödinger, protonated according to pH 7.4 and minimized with the OPLS4 forcefield. Subsequently, all ligands were docked into the binding sites of the target enzymes. The binding sites have been assigned as all residues at the spike-ACE2 interaction interface (pdb 6m0j) or within 5 Å of the cocrystallized ligand (pdb 7l4z). Docking was performed using the Glide tool of Schrödinger with the SP settings. The three highest scoring poses were obtained for each ligand and the poses were subsequently minimized using the Prime tool and MM-GBSA forcefield. To this end, the ligand and all residues within 5.5 Å were unrestrained.

3.5 Viability/cytotoxic activity

In vitro cytotoxic activities were determined by measuring cell proliferation at 6 concentrations and two incubation times (24, 48 h) using the MTT (3-(4,5-dimethylthiazol-2-yl)-2,3-diphenyltetrazolium).

Cytotoxic activity was performed against three different mammalian cell lines; the healthy cell line (CCD fibroblast) and two the cytotoxic breast cell lines; MCF7 and MDA-MB-231. A 96-well plate was seeded with 5000 cells per well (50% cell occupancy) in Dulbecco's modified Eagle's (DMEM) medium containing 10% FBS, L-glutamine, 100 units per mL penicillin or 100 μg mL⁻¹ streptomycin. They were allowed to adhere to the surface by keeping them at 37 °C overnight. The next day, the medium was aspirated and 100 μL of the new medium was added and the compounds prepared at six different concentrations were added to the cells. After incubation for different times, 10 μL of MTT was added and incubated 5% CO₂ for 3 hours in the dark and at 37 °C. After incubation, the absorption at 570 nm was measured with a spectrophotometer. These tests were repeated 3 times for each concentration, the data were

compared and plotted according to the concentration, and the IC₅₀ values were determined.⁴³

4 Conclusions

Caulerpin and mono methyl caulerpinate were isolated from the green alga *Caulerpa cylindracea* species for the first time. Both bis-indole alkaloids, but especially monomethyl caulerpinate, show strong cytotoxic effects against breast cancer cell lines MDA-MB-231 and MCF-7. In addition, both alkaloids inhibit the formation of the spike-ACE2 complex in *in vitro* assays, and the possible binding interactions of both compounds have been investigated by docking studies revealing possible allosteric and competitive interference with ACE2 binding to spike. Thus, these compounds may be further investigated as possible leads to prevent SARS-CoV-2 infection.

Author contributions

Ebru Erol: extraction, isolation and structural elucidation of the compounds, methodology, investigation, writing – original draft. Muge Didem Orhan and Timucin Avsar: testing of antiviral activity, writing – original draft. Atilla Akdemir: molecular modelling studies. Emine Sukran Okudan: identification of the alga. Gulbahar O. Alim Toraman: testing of cytotoxicity activity. Gulacti Topcu: resources, structural elucidation of the compounds, writing – original draft, conceptualization, data curation.

Conflicts of interest

There are no conflicts to declare.

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References

- H. R. A. El-Mageed, D. A. Abdelrheem, S. A. Ahmed, A. A. Rahman, K. N. M. Elsayed, S. A. Ahmed, A. A. EL-Bassuony and H. S. Mohamed, *Struct. Chem.*, 2021, **32**, 1415–1430.
- N. M. Tam, M. Q. Pham, H. T. Nguyen, N. D. Hong, N. K. Hien, D. T. Quang, H. T. Thu Phung and S. T. Ngo, *RSC Adv.*, 2021, **11**, 22206–22213.
- A. I. Elshamy, T. A. Mohamed, M. A. A. Ibrahim, M. A. M. Atia, T. Yoneyama, A. Umeyama and M. E. F. Hegazy, *RSC Adv.*, 2021, **11**, 20151–20163.
- G. Topcu, H. SenoL, G. O. Alim Toraman and V. M. Altan, *Bezmialem Sci.*, 2020, **8**, 131–139.
- L. J. Villarreal-Gómez, I. E. Soria-Mercado, G. Guerra-Rivas and N. E. Ayala-Sánchez, *Rev. Biol. Mar. Oceanogr.*, 2010, **45**, 267–275.
- M. I. Rushdi, I. A. M. Abdel-Rahman, E. Z. Attia, W. M. Abdelraheem, H. Saber, H. A. Madkoure, E. Amin, H. M. Hassan, U. R. Abdelmohsen and S. Afr, *J. Bot.*, 2020, 226–241.
- G. S. Belton, S. G. A. Draisma, W. F. Prud'homme van Reine, J. M. Huisman and C. F. D. Gurgel, *Phycologia*, 2019, **58**, 234–253.
- K. Senthilkumar and S. K. Kim, *Adv. Food Nutr. Res.*, 2014, **72**, 195–213.
- K. Chakraborty, D. Joseph, M. Joy and V. K. Raola, *Food Chem.*, 2016, **212**, 778–788.
- K. Hardouin, G. Bedoux, A. S. Burlot, C. Donnay-Moreno, J. P. Bergé, P. Nyvall-Collén and N. Bourgoignon, *Algal Res.*, 2016, **16**, 233–239.
- S. Ghosh, T. Sarkar, S. Pati, Z. A. Kari, H. A. Edinur and R. Chakraborty, *Front. Mar. Sci.*, 2022, **9**, 585–598.
- Y. J. Yang, S. J. Nam, G. Kong and M. K. Kim, *Br. J. Nutr.*, 2010, **103**, 1345–1353.
- Y. Matsumura, *Asia Pac. J. Clin. Nutr.*, 2001, **10**, 40–47.
- J. Terrados and J. A. Lopez-Jimenez, *Biochem. Mol. Biol. Int.*, 1996, **39**, 863–869.
- M. Blažina, L. Iveša and M. Najdek, *Eur. J. Phycol.*, 2009, **44**, 183–189.
- M. Kumar, V. Gupta, P. Kumari, C. R. K. Reddy and B. Jha, *J. Food Compos. Anal.*, 2011, **24**, 270–278.
- L. Stabili, S. Frascchetti, M. I. Acquaviva, R. A. Cavallo, Sa. A. De Pascali, F. P. Fanizzi, C. Gerardi, M. Narracci and L. Rizzo, *Mar. Drugs*, 2016, **14**, 210.
- G. Aguilar-Santos, *J. Chem. Soc. C*, 1970, 842–843.
- B. C. Maiti, R. H. Thomson and M. Mahendran, *J. Chem. Res., Synop.*, 1978, **4**, 126–127.
- R. J. Capon, E. L. Ghisalberti and P. R. Jefferies, *Phytochemistry*, 1983, **22**, 1465–1467.
- V. J. Paul, M. M. Littler, D. S. Littler and W. Fenical, *J. Chem. Ecol.*, 1987, **13**, 1171–1185.
- A. S. R. Anjaneyulu, C. V. S. Prakash and U. V. Mallavadhani, *Phytochemistry*, 1991, **30**, 3041–3042.
- J. Y. Su, Y. Zhu, L. M. Zeng and X. H. Xu, *J. Nat. Prod.*, 1997, **60**, 1043–1044.
- L. D. Napoli, S. Magno, L. Mayol and E. Novellino, *Experientia*, Birkhauser Verlag, CH-4010 Basel, Switzerland, 1983, vol. 39, pp. 141–143.
- C. S. Vairappan, *Asian J. Microbiol., Biotechnol. Environ. Sci.*, 2004, **6**, 197–201.
- S. C. Mao, Y. W. Guo and X. Shen, *Bioorg. Med. Chem. Lett.*, 2006, **16**, 2947–2950.
- V. Kamath and A. Pai, *Int. J. Green Pharm.*, 2018, **12**, S46–S50.
- K. C. Güven, A. Percot and E. Sezik, *Mar. Drugs*, 2010, **8**, 269–284.
- M. B. Govenkar and S. Wahidulla, *Phytochemistry*, 2000, **54**, 979–981.
- J. Ara, V. Sultana, A. Tariq, D. Taj, M. Azam and V. U. Ahmed, *J. Chem. Soc. Pak.*, 2019, **41**, 379–382.
- D. A. Abdelrheem, H. R. Abd El-Mageed, H. S. Mohamed, A. A. Rahman, K. N. M. Elsayed and S. A. Ahmed, *J. Biomol. Struct. Dyn.*, 2021, **39**, 5137–5147.

- 32 G. Topcu, Z. Aydogmus, S. Imre, A. C. Gören, J. M. Pezzuto, J. A. Clement and D. G. I. Kingston, *J. Nat. Prod.*, 2003, **66**, 1505–1508.
- 33 J. Lunagariya, P. Bhadja, S. Zhong, R. Vekariya and S. Xu, *Mini-Rev. Med. Chem.*, 2017, **19**, 751–761.
- 34 A. M. M. Lucena, C. R. M. Souza, J. T. Jales, P. M. M. Guedes, G. E. C. de Miranda, A. M. A. de Moura, J. X. Araujo-Junior, G. J. Nascimento, K. C. Scortecchi, B. V. O. Santos and J. T. Souto, *Mar. Drugs*, 2018, **16**, 1–18.
- 35 É. T. De Souza, D. P. De Lira, A. C. De Queiroz, D. J. C. Da Silva, A. B. De Aquino, E. A. Campessato Mella, V. P. Lorenzo, G. E. C. De Miranda, J. X. De Araújo-Júnior, M. C. De Oliveira Chaves, J. M. Barbosa-Filho, P. F. De Athayde-Filho, B. V. De Oliveira Santos and M. S. Alexandre-Moreira, *Mar. Drugs*, 2009, **7**, 689–704.
- 36 N. Regina, P. Vieira, M. S. Ribeiro, R. C. Villaça, W. Ferreira, A. M. Pinto, V. L. Teixeira, C. Cirne-santos and C. N. P. Izabel, *Rev. Bras. Farmacogn.*, 2011, **22**, 861–867.
- 37 I. C. P. P. Priscilla, O. Esteves, M. C. de Oliveira, C. de Souza Barros, C. C. Cirne-Santos and V. T. Laneuvlille, *Nat. Prod. Commun.*, 2019, **14**, 1–6.
- 38 D. A. Abdelrheem, S. A. Ahmed, H. R. Abd El-Mageed, H. S. Mohamed, A. A. Rahman, K. N. M. Elsayed and S. A. Ahmed, *J. Environ. Sci. Health, Part A: Toxic/Hazard. Subst. Environ. Eng.*, 2020, **55**, 1373–1386.
- 39 H. Li, X. Liao, Y. Sun, R. Zhou, W. Long, L. Li, L. Gu and S. Xu, *ChemistrySelect*, 2018, **3**, 12406–12409.
- 40 C. I. Canché Chay, R. G. Cansino, C. I. Espitia Pinzón, R. O. Torres-Ochoa and R. Martínez, *Mar. Drugs*, 2014, **12**, 1757–1772.
- 41 A. M. M. Lucena, C. R. M. Souza, J. T. Jales, P. M. M. Guedes, G. E. C. de Miranda, A. M. A. de Moura, J. X. Araújo-Júnior, G. J. Nascimento, K. C. Scortecchi, B. V. O. Santos and J. T. Souto, *Mar. Drugs*, 2018, **16**, 318.
- 42 S. Durdagi, T. Avsar, M. D. Orhan, M. Serhatli, B. K. Balcioglu, H. U. Ozturk, A. Kayabolen, Y. Cetin, S. Aydinlik, T. Bagci-Onder, S. Tekin, H. Demirci, M. Guzel, A. Akdemir, S. Calis, L. Oktay, I. Tolu, Y. E. Butun, E. Erdemoglu, A. Olkan, N. Tokay, Ş. Işık, A. Ozcan, E. Acar, S. Buyukkilic and Y. Yumak, *Mol. Ther.*, 2022, **30**, 963–974.
- 43 T. Mosmann, *J. Immunol. Methods*, 1983, **65**, 55–63.