Review Article

SOX9 Stem-Cell Factor: Clinical and Functional Relevance in Cancer

Maribel Aguilar-Medina (),¹ Mariana Avendaño-Félix,¹ Erik Lizárraga-Verdugo,¹ Mercedes Bermúdez,¹ José Geovanni Romero-Quintana,¹ Rosalío Ramos-Payan (),¹ Erika Ruíz-García (),² and César López-Camarillo ()³

 ¹ Facultad de Ciencias Químico Biológicas, Universidad Autónoma de Sinaloa, Culiacán, Sinaloa, Mexico
² Laboratorio de Medicina Traslacional y Departamento de Tumores Gastro-Intestinales, Instituto Nacional de Cancerología. CDMX, Mexico
³ Posgrado en Ciencias Genómicas, Universidad Autónoma de la Ciudad de México, CDMX, Mexico

Correspondence should be addressed to Maribel Aguilar-Medina; maribelaguilar@uas.edu.mx and César López-Camarillo; genomicas@yahoo.com.mx

Received 25 January 2019; Accepted 21 February 2019; Published 1 April 2019

Guest Editor: Daniele Vergara

Copyright © 2019 Maribel Aguilar-Medina et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Transcriptional and epigenetic embryonic programs can be reactivated in cancer cells. As result, a specific subset of undifferentiated cells with stem-cells properties emerges and drives tumorigenesis. Recent findings have shown that ectoderm- and endoderm-derived tissues continue expressing stem-cells related transcription factors of the SOX-family of proteins such as SOX2 and SOX9 which have been implicated in the presence of cancer stem-like cells (CSCs) in tumors. Currently, there is enough evidence suggesting an oncogenic role for SOX9 in different types of human cancers. This review provides a summary of the current knowledge about the involvement of SOX9 in development and progression of cancer. Understanding the functional roles of SOX9 and clinical relevance is crucial for developing novel treatments targeting CSCs in cancer.

1. Introduction

Recently, there has been a growing interest in the study of mechanisms leading to the expression of genes involved in developmental and cell differentiation, since they are related to the presence of a special type of tumor cells with a stemness phenotype dubbed as cancer stem-like cells (CSCs). Stem cells regulatory proteins are now being recognized as potential oncogenes because of their ability to regulate CSCs phenotype and maintenance in tumors of diverse types of cancer. Moreover, it has been well accepted that CSCs are the main driving force behind tumor formation and metastasis [1]. CSCs exhibit diverse cell properties including selfrenewal, differentiation capacity, and resistance to apoptosis. Moreover, CSCs are usually resistant to chemotherapy and eventually give rise to recurrence [1, 2]. Sex-determining region Y (SRY)-box 9 protein (SOX9) is a member of the SOX family of transcription factors (TFs) which are developmental regulators that possess high mobility group (HMG) box

DNA binding and transactivation domains [3]. It participates in a variety of functions, such as lineage restriction and terminal differentiation, through precise temporal and spatial expression patterns that differ between particular cell types and tissues [4]. SOX9 gene has been implicated in different types of cancer as an oncogene; however, it also may behave as a tumor suppressor [5, 6]. Recent findings have shown that ectoderm- and endoderm-derived tissues continue to express SOX9 in stem cell pools [7] and evidence also suggests that it may regulate CSCs [8–10]. However, detailed mechanisms need to be elucidated. In this review we aim to condense the knowledge about the involvement of SOX9 in the initiation and progression of different types of cancer and to highlight its potential as a clinical biomarker.

2. SOX Family of Transcription Factors

SOX family of proteins comprise a group of transcriptional regulators containing a highly conserved HMG domain that

was first discovered in SRY protein, a transcription factor involved in mammalian male sex determination [11]. SOX9 is located in chromosome 17 in a 3 Mb region devoid of other protein-coding genes and its expression is complex with individual enhancers directing tissue-specific expression [12]. In general, proteins containing an HMG domain which consists of three α helices with 50% or higher amino acid similarity to the HMG are referred to as SOX proteins (SRYrelated HMG box). Around 20 related SOX proteins have been identified in humans, and they have been grouped based on the structural homology of regions outside of their conserved HMG boxes [7]. SOX proteins bind to ATTGTT consensus or related sequence motifs through their HMG domain [13]. SOX9 belongs to the SoxE subgroup, and its HMG domain induces significant bending at the consensus-binding motif (A/TA/TCAAA/TG) by forming an L-shaped complex in the minor groove of DNA [7]. Members of the SOXE subgroup share regions of significant homology outside the HMG domain and constitute two additional functional domains: a self-dimerization domain and a transactivation domain at the C-terminus [14]. One current model suggests that dimerization is achieved through cooperative binding between the dimerization domain of one SOXE protein and the HMG box of its partner SOXE protein [15]. SOX protein is subject to posttranslational modifications which alter its nuclear import (phosphorylation and acetylation) and its rate of degradation (ubiquitination and sumoylation) [16]. Individual SOX members within a group share biochemical properties and thus have overlapping functions. Conversely, SOX factors from different groups have acquired distinct biological functions despite recognizing the same DNA consensus motif [4]. Target gene selectivity by different SOX factors can be achieved through differential affinity for particular flanking sequences next to consensus SOX sites, homo- or heterodimerization among Sox proteins, posttranslational modifications of SOX factors, or interaction with other cofactors [4]. The expression of SOX genes is frequently subject to autoregulation or control by other SOX members. SOX is also regulated post-transcriptionally by microRNAs [17]. Furthermore, SOX-dependent regulations intersect with signaling networks such as the sonic hedgehog (Shh) [18], Wnt signaling pathways, in which SOX-Gli and SOX- β -catenin interactions are implicated [19]. SOX factors respond to different extracellular signals and interact with a host of intracellular cofactors to control different sets of genes in distinct cell types [20]. Additionally, SOX compete with other transcriptional factors regulating alternative lineages to achieve different cell fates during development. At molecular level, this is often accomplished by directly activating genes that promote their own lineage and repressing genes of alternative lineages [20]. In summary, SOX factors have profound implication in cell fate determination during development, even though recent findings reveal their crucial role in establishing and maintaining stem and progenitor cells [21].

3. Role of SOX9 in Human Cancer

SOX9 has been studied from a developmental point of view, particularly during chondrogenesis and male gonad

genesis. Nevertheless, recent molecular and functional analyses have elucidated an important role in stem cell biology of mesoderm-, ectoderm-, and endoderm-derived tissues and organs [7]. Importantly, SOX9 maintains both adult stem and progenitor cells with high turnover, as in intestine and hair follicles, and it is crucial for postnatal injury repair in endodermic and ectodermic organs [7]. Remarkably, dysregulation of tissue differentiation pathways and stem cell homeostasis contributed to the initiation and progression of cancer. Experimental and clinical data demonstrated an important role for SOX9 in tumorigenesis since it is overexpressed in a wide range of human cancers where its expression correlated with tumor progression and clinical data (Table 1) [73]. In addition, SOX9 interacted with different transcription factors and exhibited several pro-oncogenic characteristics including promotion of proliferation, senescence inhibition, and neoplastic transformation in collaboration with other oncogenes [73]. COSMIC analysis showed that, from a total of 46,601 unique cancer samples, 572 samples have mutations in SOX9 and the most frequent mutation type is missense substitution (38.81%) of which 113 (33.63%) are C>T transitions. Copy number variations (CNV) gain was reported in 108 unique samples and overexpression was present in 509 samples [74, 75]. The versatility of SOX9 may be explained by a combination of posttranscriptional modifications, binding partners, and the tissue type in which it is expressed [7].

3.1. SOX9 Alterations in Hepatocellular Carcinoma. Hepatocellular carcinoma (HCC) is the most common primary liver malignancy. Its genetic complexity relies on interaction between several somatic genomic alterations and diverse etiologies linked to liver diseases by the concerted action of passenger and driver cancer genes. HCC progression is a complex process that implicates accumulative genomic alteration that includes aberrant gene expression, oncogene upregulation, and tumor suppressor downregulation providing favorable conditions for the development of HCC [76-78]. These mechanisms have been associated to several alterations in some important cellular signaling pathways relevant to a therapeutic perspective, such as RAF/MEK/ERK pathway, phosphatidylinositol-3 kinase (PI3K)/AKT/mammalian target of rapamycin (mTOR) pathway, and Wnt/ β -catenin pathway [79]. Nonetheless, the molecular pathogenesis of HCC is still unclear.

SOX9 regulated by Notch determines the timing and structure of bile duct morphogenesis during liver embryogenesis. Besides, not only during development, but also in the adult organs, SOX9 expression levels appear to be crucial for controlling the cell status of the duct cells [80]. *In vitro* analysis has shown that SOX9 expression in HCC cell lines was upregulated in comparison to normal hepatic cell lines; furthermore, it was expressed at higher level in highly metastatic cells lines relative to low metastatic cells. Moreover, downregulation of SOX9 in HCC cell lines decreased invasiveness and migration [81]. Recent studies using SOX9 chromatin immunoprecipitation combined with deep sequencing (ChIP-seq) analysis indicated that SOX9 can activate canonical Wnt/ β -catenin signaling in HCC endowing stemness features through Frizzled-7 [82].

Type of cancer	Status of SOX9	Sox9 participation	References
Hepatocellular carcinoma	overexpression	Related whit poor prognosis Related with poor disease free survival Related with poor overall survival	[22, 23]
Breast cancer	overexpression	Promotes proliferation, tumorigenesis and metastasis Related with poor overall survival	[24]
Bladder cancer	overexpression	Promotes tumorigenesis Related with poor overall survival	[25]
Gastric cancer	overexpression	Promotes chemoresistance Related with poor disease free survival	[26]
Prostate cancer	overexpression	Promotes cell proliferation and apoptosis resistance Related with high clinical stage Related with poor relapse free survival Related with poor overall survival	[27]
Prostate cancer	downregulation	Promotes metastasis Related with advanced clinical stage Related with EGR-positive tumors	[28]
Ovarian cancer	overexpression	Its coexpression with HIF- 2α induces the expression of TUBB3 which is related with poor overall survival	[29]
Pancreatic cancer	overexpression	Promotes chemoresistance	[30]
colorectal cancer	overexpression	Promotes cell proliferation, senescence inhibition and chemoresistance	[31-33]

TABLE 1: SOX9 expression and functions in human cancers.

Besides, the results of a genome-wide transcriptional analysis indicated that TGF β and Wnt/ β -catenin signaling pathways were activated in hepatocholangiocarcinoma (cHCC-CC) [83]. Furthermore, integrative genomics revealed that cHCC-CC shares characteristics of poorly differentiated HCC with stem cell traits and poor prognosis [83]. Interestingly, early biomarkers of biliary commitment such as SOX9, as well as master genes of signaling pathways, which regulate the differentiation of hepatoblasts to cholangiocytes, were induced in cHCC-CC (e.g., TGF β , Wnt, and Notch) [83]. SOX9 overexpression was commonly observed in HCC with high tumor stage and tumor grade tissues. Also, the high expression of SOX9 was linked to a significant trend toward both poorer disease-free survival and poorer overall survival [22]. Besides, poor prognosis of HCC patients has been linked with high SOX9 expression independent of the presence of cirrhosis [23].

3.2. Role of SOX9 in Breast Cancer. Breast cancer is a complex and heterogeneous disease that includes morphological and molecular different entities. Clinical parameters such as tumor size, lymph node involvement, histological grade, age and the expression of estrogen receptor (ER), progesterone receptor (PGR), and epidermal growth factor receptor 2 (HER2) biomarkers are responsible of its high clinical heterogeneity [84]. Mammary glands contain a small subpopulation of cells with a stem cells activity and it is also known that several TFs play pivotal roles in the establishment of cellular states. SLUG and SOX9 play essential roles in induction and maintenance of tumor initiating capacity in breast cancer cells [58]. In breast tumors, SOX9 expression was higher in comparison to normal mammary tissues, which was associated with an increased proliferation and

Ki67 and p53 expression [59–61]. There is also evidence that upregulation of SOX9 affected metastasis and tumorigenesis in breast cancer cells by 5-fold and 40-fold, respectively [9]. Primary tumors with high expression levels of SLUG and SOX9 had a significant lower overall survival rate.

On the other hand, knockout studies have demonstrated that SOX9 was essential for the function of mammary stem/progenitor cell populations. Knockdown of SOX9 resulted in decreased proliferation of mammary stem cells [85]. On the other hand, higher expression of cytoplasmic-SOX9 in breast tumors was significantly associated with ERstatus and decreased overall survival [24]. Altogether, these data indicate that cytoplasmic location of SOX9 was directly related with increased proliferation in breast cancer cell lines. Similarly, cytoplasmic SOX9 expression was directly related to neoplastic progression and its nuclear expression was more common in early stages of differentiation [24].

3.3. The Importance of SOX9 in Bladder Cancer. Bladder cancer (BC) is the ninth most common malignant disease and the thirteenth most frequent cause of cancer death worldwide. Men are more affected than women (3.2:0.9 ratio) and disease incidence increases with age [86]. In previous studies using biopsies of BC, 75% positive immunostaining of SOX9 was observed in the nucleus of cancer cells and the expression was significantly associated with the advanced pathological grade and clinical stage. However, SOX9 immunostaining in the normal bladder tissues occurred mainly in the cytoplasm and nucleus. These findings could indicate that SOX9 may play a promotive role in the progression of BC [25]. On the other hand, epigenetic changes of SOX9 were associated with the aggressiveness of bladder cancer [87]. Methylation of Sox9 promoter gene was identified in a study of 101 BC samples and it was significantly associated with shorter overall survival. Besides, *in vitro* analyses demonstrated that the expression of SOX9 was aberrantly silenced by CpG island promoter hypermethylation in BC [88]. However, Sun et al. (2009) found a hypermethylated state of SOX9 in only 3/82 (3.7%) cases of BC and 2/15 (13.3%) cases of the control in a Chinese cohort [89]. These results indicated the necessity to further compare the methylation profiles between populations, given the discrepancies in this disease as proposed previously [90].

3.4. SOX9 Aberrant Expression in Gastric Cancer. Gastric cancer (GC) is one of the most aggressive malignant tumors worldwide with a high mortality rate, preceded only by lung cancer [91]. Globally, GC is the fourth most common cancer and second leading cause of cancer related mortality with a 5year overall survival rate less than 25% [86]. SOX9 expression has been found in epithelial cells at the proliferative zone of the normal gastric mucosa and bottom area of the intestinal metaplasia of the stomach. Many tumor cells of type I GC are positive for SOX9 [92, 93]. Ectopic expression of β -catenin in AGS and MKN-1 cells induced increased expression of SOX9 [94], whereas its suppression by PPAR γ decreased SOX9 expression in MKN-28, SGC-7901, and BGC-823 cells [95]. Gastrokine 1 (GKN1), a tumor suppressor like protein which expression is lost in gastric tumors (including adenoma and cancer) [96], was responsible for decreased SOX9 expression in AGS and MKN-1 cells. Nevertheless, in GC tissues, nuclear expression of SOX9 was closely associated with GKN1 immuno-negativity suggesting that aberrant SOX9 expression by GKN1 inactivation may be involved in the development of sporadic GC as an early event [94].

Additionally, SOX9 overexpression was correlated with lymph node metastasis and advanced tumoral stages of GC, indicating that it is related to tumor progression by promoting invasion and metastasis [97] and with reduced disease-free survival [26].

Moreover, elevated SOX9 levels were associated with resistance to cisplatin in MKN45 and KATO III cells [26] whereas miR-524-5p inhibited SOX9 expression conferring sensitivity to cisplatin-resistant GC cells by targeting its 3'UTR. This is because resistance to cisplatin in GC cells was associated with the expression of miR-524-5p. Thus, overexpression of miR-524-5p in GC cells enhanced their sensitivity to cisplatin and it depends on the downregulating SOX9 [98]. Also, SOX9 has been correlated with the carcinoembryonic antigen-related cell adhesion molecule 1 (CEACAM1). Their coexpression was detected in normal gastric mucosa, hyperplastic polyp, intestinal metaplasia, gastric intraepithelial neoplasia, and adenocarcinoma, showing highly elevated expression from benign proliferative lesions to malignant lesions, suggesting that SOX9 could change CEACAM1 expression patterns, which might promote the tumor progression [99].

Helicobacter pylori infection has a very important role in GC. In animal model, histopathological changes such as metaplasia of the gastric mucosa after *H. pylori* infection show increased expression of CD44 and SOX9 dependent on IL-1 signaling, suggesting the participation of SOX9 in gastric carcinogenesis [100]. In humans, the risk of developing GC was higher in individuals infected with cytotoxin-associated gene A (cagA)-positive strains or some vacuolating cytotoxin gene A (vacA) allelic combinations causing the loss of crucial features of epithelial differentiation in gastric cells, leading to transformation and tumor formation [101–103]. In this regard, SOX9 was transcriptionally activated following *H. pylori* infection in GC cell and its silencing resulted in an increase of phospho-histone H3- (p-H3-) proliferative cells and spheres formation ability promoted by bacteria. β catenin-silenced cells also presented a marked reduction in p-H3-positive cells when infected with both strains [26]. Conversely, downregulation of SOX9 by promoter methylation was related to GC progression in Epstein Barr Virus-positive biopsies and infected MKN7 cells. SOX9 methylation was detected in 47% of GCs and correlated with low levels of SOX9 protein. Besides, the rate of methylated SOX9 tumors increased and SOX9 expression gradually decreased through the depth of GC invasion. These data strongly suggested that the decrease of SOX9 expression in advanced GC was related with the epigenetic suppression of SOX9 during tumor invasion [104].

3.5. SOX9 Is Involved in Different Types of Pancreatic Cancer. SOX9 regulated by Notch is involved in the maintenance of pancreatic progenitor pools [105]. Furthermore, SOX9 is essential for pancreas development. At the early stages of mouse pancreas development it is expressed in all epithelial cells and its expression is confined to the ductal cells and centroacinar cells as development progresses [80]. Genetic lineage-tracing studies showed that all types of pancreatic epithelial cells including endocrine, acinar, and duct cells express Sox9, suggesting that the all Sox9-expressing cells are a common progenitor of pancreatic epithelial cells [106].

Recently, it was shown that Sox9 is expressed during premalignant and malignant lesions such as mucinous cystic neoplasias (MCNs), intraductal papillary mucinous neoplasias (IPMNs), pancreatic intraepithelial neoplasias (PanINs), and pancreatic ductal adenocarcinoma (PDAC) (Table 2) [34, 107], which is the most common pancreatic cancer and develops from cells lining pancreatic ducts [108]. The evidences suggested that a phenotypic switch converting pancreatic acinar cells to duct-like cells can lead to PanIN [109] and eventually PDAC. Studies about the expression of SOX9 and Hepatocyte Nuclear Factor 6 (HNF6) show that these TFs were expressed in acinar cells. HNF6 induced SOX9 expression, indicating that SOX9 is downstream of HNF6. In acinar-to-ductal metaplasia (ADM), SOX9 was predominantly found in metaplastic cells that displayed ductlike characteristics and it was also found in PanIN [34, 35].

Epidermal growth factor (EGFR) in the PDAC promotes expression of SOX9 as an early event. In this context, pancreatic metaplasia could be also caused by loss of p27 function, a negative regulator of proliferation and a tumor suppressor that inhibits cyclin-CDK activity in the nucleus [110–112]. Besides, K-RAS activation, the earliest known event in pancreatic carcinogenesis [113, 114], may induce p27 mislocalization producing loss of nuclear p27 expression and as a result derepression of SOX9, triggering ADM [36]. The formation of acinar-derived premalignant lesions depends on

	26 11		- <i>1</i> 0 G	c 6
Lesions	Model	Status of SUX9	Effects	References
Acinar-to-ductal metaplasia	Mice	Overexpressed	HNF6 induces Sox9 expression, which is characteristic of ADM in humans	[34, 35]
Acinar-to-ductal metaplasia	Mice	Overexpressed	Aberrant expression of p27 induces the nuclear expression of Sox9	[36]
Mucinous cystic neoplasias, intraductal papillary mucinous neoplasias, pancreatic intraepithelial neoplasias and pancreatic ductal adenocarcinoma	Mice	Overexpressed	SOX9 and Kras co-expression is associated with PDAC initiation	[37]
Pancreatic ductal adenocarcinoma	88 tumors samples of PDAC	Overexpressed	Sox9 and p-Akt double-positive expression is related with an unfavorable prognosis, high TNM and distant metastasis	[38]
Pancreatic ductal adenocarcinoma	Patient-derived tumor organoids with PDAC	Overexpressed in cytoplasm	High expression of Sox9 in cytoplasm is related with a poor DFS, OS, higher tumor grade and worse disease-specific survival compared to patients with nuclear Sox9 expression	[39]
Pancreatic cancer	PANCI and HPAC cell lines of pancreatic cancer	Overexpressed	Sox9 is highly expressed in pancreatic CSCs. Moreover, NF-&B subunit p65 positively regulates SOX9	[40]
Pancreatic ductal adenocarcinoma	HD3 colon cancer cells	Overexpressed	ST6Gal-I induces expression of Sox9, promoting stem-like cell properties	[41]
Pancreatic cancer	PANC-1, Capan-1, BxPC-3, MiaPaCa-2 cell lines of pancreatic cancer	Expression depending on chemoresistance	High level of Sox9 is related to stronger chemoresistance to Gemcitabine	[42]
Pancreatic ductal adenocarcinoma	HPDE cell line of PDAC	Overexpressed	GLI1 induces the transcription of Sox9 promoting stem cell properties	[43]
Pancreatic cancer	PANC1 cell line of pancreatic cancer	Downregulated	Induction of EMT with TGF- β results in low levels of SOX9, FOXA2, and GATA4	[44]
Pancreatic cancer	Mice	Overexpressed	p53-/- mice enhanced sphere formation, increased expression of the stemness regulator Bmil and Klf4, and pancreatic multipotent progenitor markers as Ptf1a, Pdx1, Cpa1, c-myc, Hnf1b and Sox9	[45]
Pancreatic cancer	FG and L3.6pl cell lines of pancreatic cancer under hypoxia	Overexpressed	Under hypoxia conditions, FG cell line expresses high levels of Sox9 and L3.6pl. Besides, WNT, CXCR4, retinoic acid, and FAK signaling pathways are regulated by Sox9 in L3.6pl	[46]
Pancreatic ductal adenocarcinoma and anaplastic pancreatic cancer	6 APC patients and 53 PDAC patients	Overexpressed	PDAC and APC have high expression of Sox9, being the expression of proteins related with CSCs and EMT process higher in APC samples than PDAC	[30]
Intraductal papillary mucinous neoplasm	19 IPMN cases	Overexpressed	SOX9-positive cells were confined to the lower portions of the papillary structures of IPMN	[47]
Solid pseudopapillary tumor	8 samples of SPT	Overexpressed	PDX1 and Sox9 are both expressed in the cytoplasm of SPT cell	[48]

ces

TABLE 2: Sox9 is expressed during premalignant and malignant lesions in pancreatic cancer.

Journal of Oncology

ectopic induction of SOX9, a ductal gene [115]. Moreover, when it is concomitantly expressed with oncogenic K-RAS, SOX9 accelerated the formation of premalignant PDAC lesions [37]. Furthermore, SOX9 and p-AKT double-positive expression was related with an unfavorable prognosis, high TNM, and distant metastasis in PDAC [38, 116].

Studies using patient-derived tumor organoids with PDAC show that in normal pancreas SOX9 is expressed in the nucleus, while, in the organoids with TP53-mutated (R175H) PDAC, it was expressed in cytoplasm. Clinically, high expression of SOX9 in cytoplasm could be related with a poor disease-free survival (DFS), overall survival, higher tumor grade, and worse disease-specific survival compared to patients with nuclear SOX9 [39]. Emerging evidence suggests that CSCs are exclusively tumorigenic and essential drivers for tumor progression and metastasis [117]. Pancreatic CSCs have been identified and characterized using different surface markers: CD44, CD24, EpCAM, CD133, CXCR4, c-Met, and Aldehyde Dehydrogenase-1a1 (ALDH1) [118-121]. SOX9 has been found expressed in the pancreatic CSCs isolated from PANC1 and HPAC cell lines of pancreatic cancer. This population was more capable of initiating tumors in NOD/SCID xenograft model than the population no-invasive (no-CSCs) [40].

Demethylated SOX9 is found in CSCs and plays a crucial role in the invasion process. Also, NF- κ B subunit p65 positively regulates SOX9 expression by directly binding to the SOX9 promoter [40], suggesting that the NF- κ B pathway is one of the most activated pathways in pancreatic CSCs. Another important regulator of SOX9 is the glycosyltransferase ST6Gal-I which adds α 2-6-linked sialic acids to substrate glycoproteins and it is known that its upregulation in cancer cells confers stemness characteristics [41, 122]. Modulating ST6Gal-I expression in pancreatic cancer cells directly altered CSC spheroid growth. In this regard, ST6Gal-I knockdown decreased the levels of SOX9 [41], suggesting that SOX9 expression was regulated by a specific glycosyltransferase, and tumor glycosylation could be a mechanism for functionally shifting cells to a less differentiated, stem-like state.

Interestingly, SOX9 expression in different pancreatic cell lines (PANC-1, Capan-1) was related to stronger chemoresistance to gemcitabine than cells with low SOX9 expression (BxPC-3, MiaPaCa-2). Conversely, SOX9 repression using siRNA recovers the chemosensitivity, affected spheres formation rate, and the proportion of CD44^{high} and CD24^{high} cells. This indicates that the expression of Sox9 plays an important role in chemoresistance by the induction of stemness in pancreatic cancer cells [42].

On the other hand, GLI1, a member of the GLI family of zinc finger transcription factors, is a central regulator of cell fate that is deregulated in diverse tumor types [62–66]. GLI1 signaling impacts multiple cancer-relevant cellular processes, promoting dedifferentiation, the generation of CSCs, tumor progression, and metastasis. GLI1 directly induced the transcription of SOX9 and a positive feedback promoting SOX9-dependent cancer stem cell properties was observed [43].

Epithelial-to-mesenchymal transition (EMT) process is a critical regulator of the CSC phenotype [123, 124]. Tumor growth factor β (TGF β) induces EMT, promoting cancer cell invasion and metastasis [124]. PANC1 cell line stimulated with TGF β 1 showed a significant downregulation of SOX9, FOXA2, and GATA4 master genes [44].

Furthermore, p53-/- mice enhanced sphere formation, increased expression of the stemness regulator BMI1 and KLF4 and pancreatic multipotent progenitor markers such as PTF1A, PDX1, CPA1, c-MYC, HNF1B, and SOX9 [45]. These results can be relevant to understand the relationship between p53 and SOX9 and their importance in the acquisition of EMT characteristics in pancreatic cancer.

Microarray analysis demonstrated that another important issue related with cancer and the CSCs was hypoxia, which induced expression of Sox9 in low metastatic cell line FG, whereas in high metastatic cell line L3.6pl it was found constitutively expressed and was not more inducible under hypoxic conditions [46]. Besides, a subset of transcripts related different networks including WNT, CXCR4, retinoic acid, and FAK signaling pathways were also regulated by SOX9 in the aggressive-metastatic cells, but driven by HIF- 1α in low metastatic cells [46].

Opposite to the oncogenic role of SOX9 in pancreas carcinogenesis studies in tissues corresponding to later stages of tumor development have found downregulation of SOX9 and other master regulators of embryonic development such as GATA4, PDX1, PTF1a, and HNF1b [123, 125–127].

Even though PDAC is the most common type of pancreatic cancer, there are other types of pancreatic tumors with less incidence but more aggressive behavior such as the anaplastic pancreatic cancer (APC), which has been considered a variant of PDAC [67, 68]. There was evidence that PDAC and APC have high expression of Sox9. Also, the expression of proteins related with CSCs and EMT process is higher in APC samples than PDAC, which correlated with aggressiveness of APC [30].

Another type of pancreatic cancer called intraductal papillary mucinous neoplasm (IPMN) is less aggressive than PDAC and APC [47, 128, 129] and exhibits a characteristic expression of SOX9 confined to the lower portions of IPMN which is lost once the neoplasms advance to high-grade dysplasia carcinoma. Cells in the upper portions of IPMN may be, albeit speculative at this point, supplied by the SOX9-positive cells in the lower portions of the neoplasm [47].

Finally, solid pseudopapillary tumor (SPT) is an uncommon type of pancreatic tumor of undetermined origin present especially in children [130]. Studies on samples from pediatric patients with SPT showed that PDX1 and SOX9 were both expressed in the cytoplasm of SPT cells, supporting the hypothesis that tumor cells originate from pancreatic stem cells persisting after the embryonic period [48]. This is relevant since both transcription factors are crucial for pancreatic organogenesis and linked to Wnt/ β catenin signaling pathway [105, 131–133].

3.6. SOX9 Is Required for Prostate Cancer Initiation. The human prostate is composed of prostatic glands with well-defined basal and luminal epithelial cell layers. Cells within the luminal epithelium have a very low rate of proliferation and express high levels of androgen receptor (AR) [69]. In

contrast, basal cells have a higher rate of proliferation, express low or undetectable levels of AR, and are not androgen dependent, playing critical roles in prostate organogenesis, homeostasis, support, and a barrier for the luminal cells. This barrier becomes discontinuous in prostatic intraepithelial neoplasia, which is believed to be a precancerous lesion. The complete loss of the basal cell layer is a defining feature of prostate cancer (PCa) [134].

SOX9 protein was expressed in adult prostate basal epithelium and at the initial stages of bud outgrowth from the urogenital sinus and could play a role in maintaining the committed stem cell phenotype, differentiation, and supporting the overlying luminal epithelium [134–136].

In vivo studies showed that SOX9 was highly expressed during fetal prostate development by epithelial cells expanding into the mesenchyme, suggesting that it may contribute to invasive growth in PCa [137]. Besides, SOX9 expression in prostate cancer cells was regulated by Wnt/ β -catenin signaling, being AR one identified downstream target [134]. In turn, SOX9 positively regulated multiple Wnt pathway genes, including encoded Wnt receptors (frizzled [FZD] and lipoprotein receptor-related protein [LRP] family members) and the downstream β -catenin effector TCF4 [70]. Microarray analysis showed that SOX9 was overexpressed in PCa tissues when compared with noncancerous prostate tissues. Also, SOX9 overexpression was found in PCa tissues with higher clinical stage and was related to lower biochemical recurrence-free survival and overall survival rates [138]. When the SOX9 expression was correlated with overexpressed HIVEP3 (human immunodeficiency virus type I enhancer binding protein 3), the patients also exhibited significantly shorter biochemical recurrence-free survival [139].

Some tumor suppressors have been related with SOX9 participation in PCa. Its overexpression in adult mouse prostate epithelia gives rise to an increase in proliferation and induced early high-grade prostate intraepithelial neoplasia lesions when mice are heterozygous for PTEN (phosphatase and tensin homolog deleted on chromosome 10). This study shows that high levels of SOX9 contributed to regulate proliferation within the prostate epithelia and can cooperate with PTEN loss to accelerate prostate neoplasia [140]. Zbtb7a (also known as pokemon) has been recently reported as an oncosuppressor in PCa since it is lost in a subset of human advanced prostate cancer and facilitates the oncogenic activity of SOX9 during prostate tumorigenesis favoring senescence bypass, increase of proliferation rate, apoptosis resistance, and invasive potential [71].

Interestingly, in human castration resistant PCa samples, nMET was remarkably increased. Androgen deprivation induced endogenous nMET which activates SOX9 and promoted cell proliferation and stem-like cell self-renewal in androgen-nonresponsive PCa cells. This indicates that coupregulation of endogenous nMET and SOX9 upon androgen deprivation may activate cell reprogramming to promote transformation and androgen nonresponsive growth [141].

Fusion genes have a very important role in PCa. SOX9 is a critical downstream effector of ERG in TMPRSS2: ERG fusion-positive PCa, and ERG stimulates SOX9 expression by redirecting AR to a cryptic AR-regulated enhancer in the SOX9 gene [142]. Besides, association between TMPRSS2, ERG positive PCa, and rs1859962 at 17q24 has been demonstrated suggesting a molecular mechanism linking the risk region to the ERG pathway where SOX9 is a downstream target. There is also evidence of a positive correlation between SOX9 gene expression and the rs1859962 risk allele in TMPRSS2: ERG positive tumor tissue [143].

Analysis of tissue microarrays of prostate biopsies samples of patient with metastatic castration-resistant PCa shows 18.3% and 87.3% of patients with positive ERG and SOX9 expression, respectively [27]. Besides, ERG and SOX9 are significant risk factors for lower prostate-specific antigen-Progression Free Survival (PFS), Clinical/Radiological-PFS, and Overall Survival after docetaxel treatment suggesting that ERG and SOX9 are potential biomarkers for prediction to docetaxel treatment in mCRPC patients [27]. Conversely, a gradual decrease of SOX9 has been related to a progression to advanced stage, high Gleason grade, and metastatic growth in ERG-positive cancers and these effects were strictly limited to the subset of prostate cancers harboring PTEN deletions [28].

3.7. Oncogenic Role of SOX9 in Ovarian Cancer. Ovarian cancer is the most lethal gynecological cancer [144]. Carcinomas of the ovarian surface epithelium correspond to 90% of ovarian malignancies and are classified into four main histological subtypes, which have distinct characteristics regarding genetic abnormalities and specific signaling pathways [144]. Sertoli-Leydig cell tumors, ovarian sex-cord stromal tumors, granulosa cell tumors, and primary ovarian tumors constitute carcinomas in ovary [145].

In normal ovarian development, SOX9 has different expression levels, as wells as roles in comparison to other tissues. During the follicular development in early pre-antral follicles there is not expression of SOX9, but the cells surrounding the developing follicles present nuclear expression. These are very important since they have a participation in the production of collagen or laminin fibers which constructs follicular lamina [146].

Little is known about SOX9 role in ovarian cancer. Nonetheless, higher expression of SOX9 has been found in human Sertoli tumor biopsies coexpressed with BCL-2 and Ki-67, being the last the less expressed in well-differentiated cells [147]. This suggests that apoptotic and proliferative properties depend on tumor differentiation stage. Besides, it is known that hypoxia conditions promote Tubulin Beta 3 (TUBB3) expression through HIF-2 α and SOX9. High expression of these genes correlates with shorter overall survival in women with ovarian cancer [29].

3.8. The Role of SOX9 in Colorectal Cancer. Colorectal cancer (CRC) is a major cause of morbidity and mortality throughout the world [148]. It accounts for over 9% of all cancer incidences [149, 150] and it is the third most common cancer worldwide and the fourth most common cause of death [150]. In most patients, death is not caused by the primary tumor, but rather by its metastasis in other organs and associated complications [151].

Paneth cells are a highly specialized population of intestinal epithelial cells located into the crypts [152]; these cells are critical to the control of the intestinal stem cell (ISC) niche and the intestinal barrier [153, 154]. The function of SOX9 in Paneth cells has not been clarified but *in vitro* studies suggest a role in the control of cell differentiation in the intestinal epithelium [155]. *In vitro* and *in vivo* data indicate that Sox9 gene is a transcriptional target of Wnt signaling; this pathway is involved in the regulation of intestinal epithelium homeostasis [156].

Sox9 expression is regulated by TCF4, the main Wnt pathway TF in the intestinal epithelium [155]. This is relevant since mutations in components of the Wnt pathway, including the tumor suppressor APC and β -catenin protein, result in stabilization of β -catenin, which then continuously interacts with TCF4, leading to constitutive activation of target genes [72]. Moreover, targeted mutations of APC or β -catenin are sufficient to initiate tumorigenesis in mouse [157–159], highlighting the importance of the Wnt pathway in the development of cancer.

Recent studies of CRC have found that overexpression of SOX9 *in vitro* and *in vivo* was related to several pro-oncogenic properties, including the ability to promote proliferation, inhibit senescence, and collaborate with other oncogenes in neoplastic transformation [6, 49, 50, 73, 160]. The overexpression of SOX9 is related with recurrent distal truncating mutations as frameshift mutations and nonsense mutation in approximately 11% of CRCs; also, SOX9 mutation is strongly associated with coexistent mutant K-RAS and wild type TP53 [49, 50].

Nevertheless, in the particular case of DLD-1 CRC cell line, which has a heterozygous L142P inactivating mutation of SOX9, the restoration of wild type SOX9 expression results in an oncoprotective activity which inhibits cell growth, clonal capacity, and colonosphere formation while decreasing both the activity of the oncogenic Wnt/ β -catenin signaling pathway and the expression of the c-MYC oncogene [6].

Besides, a truncated version of SOX9 devoid of transactivation domain as a result of retention of the second intron called MiniSOX9 has been discovered in human tumor samples of CRC; this version is expressed at high levels in CRC but it is undetectable in the surrounding healthy tissue. The possible mechanism of MiniSOX9 could be through activation of canonical Wnt target genes and repression of PKC α (Figure 1), two features in favor of oncogenic properties [51].

It has been stablished that overexpression of SOX9 in CRC is associated with β -catenin activation; however, the largest clinical study on SOX9 expression over 188 primary CRC specimens from a Chinese population shows that it does not present significant correlation between SOX9 and β -catenin [31].

SOX9 upregulation is common in colorectal adenoma and cancer and is an independent indicator for an adverse prognosis in CRC [31]. Conversely, low levels of SOX9 at the invasive front of the primary tumor have been shown as an independent predictor of relapse in stage II colon cancer patients (Table 3) [32]. Studies over African Americans CRC cases, diagnosed at earlier ages compared to non-Hispanic withes, have found that SOX9, GATA6, TET1, GLIS1, and FAT1 were differentially hypermethylated in APC mutationnegative CRC; this lack of APC mutation is associated with the early-onset CRC [33].

A recent study about the role and association between SOX9, β -catenin, and PPAR γ in CRC tissues showed that SOX9 and β -catenin were overexpressed whereas PPAR γ was downregulated. Treatment with the synthetic PPAR γ ligand rosiglitazone induced different changes of SOX9 and β -catenin expression and subcellular localization in the colon cancer cell lines Caco2, SW480, HCT116, and HT29. All this data indicated that SOX9, β -catenin, and PPAR γ expression levels were deregulated in the CRC tissue, and in colon cancer cell lines ligand-dependent PPAR γ activation unevenly influences SOX9 and β -catenin expression and subcellular localization, suggesting a variable mechanistic role in colon carcinogenesis [52].

In HT29 and HCT116 cell lines of CRC, SOX9 was recruited by NF-Y to the target genes and interacted with NF-Y on CCAAT promoter sequences. Besides, SOX9 is necessary for the function of NF-Y in activating expression of some cell-cycle regulatory gene expressions such as cyclin B1, cyclin B2, cyclin dependent kinase 1, and topoisomerase II α [53].

Multiples targets of SOX9 have been described. One of these is FOXK2, a transcription factor which promotes the cell proliferation in samples tissues of CRC [54]. Another important target of SOX9 is S100P; both were coexpressed in CRC and the knockdown of SOX9 expression downregulates S100P expression resulting in reduced invasiveness and metastasis of colon cancer cells by inhibiting the activation of receptor for advanced glycation end products (RAGE)/ERK signaling and EMT [55].

Interestingly, hypoxia induced EMT and SOX9 overexpression in CRC cells. SOX9 was able to migrate to nucleus and upregulated the expression of USP47, a deubiquitinating enzyme [56]. Another way to enhance the EMT by SOX9 is the loss of ZFP36 expression, a tumor suppressor [161]. On the other hand, it was demonstrated that SOX9 levels were higher in metastatic SW620 cell line than in primary CRCs SW480 cell line isolated from the same patient. SOX9 is sufficient and necessary for the acquisition and maintenance of CR-CSCs and metastatic traits, properties linked to transcriptional and post-transcriptional regulation. Finally, SOX9-mediated selfrenewal and growth were impaired by the mTOR inhibitor rapamycin [57].

4. Clinical Relevance of SOX9 in Cancer

SOX9 has proven its functional role in various aspects of cancer biology. Besides, research on SOX9 has also investigated its importance in the clinic regarding disease prognosis, relapse, and therapy resistance. For instance, SOX9 overexpression was commonly observed in those HCC high tumor stage and tumor grade tissues. Also, the high expression of SOX9 was linked to a significant trend toward both poorer disease-free survival and poorer overall survival [22]. Moreover, poor prognosis of HCC patients has been linked with high SOX9 expression independent



FIGURE 1: MiniSOX9 has an oncogenic behavior in CRC. (a) In normal conditions canonical Wnt/ β -catenin pathway triggers SOX9 expression resulting in regulation of differentiation and homeostasis in intestinal epithelium. (b) Truncated version of SOX9, MiniSOX9, accumulates in the nucleus to inhibit SOX9 DNA-binding-dependent transcriptional activity and PKC-alpha expression.

of the presence of cirrhosis [23]. In breast cancer, primary tumors that exhibit high expression levels of both SLUG and SOX9 had a significantly lower overall survival rate than the rest of the patients. Thus, SOX9 expression in carcinogenesis and malignity in breast cancer tumors is relevant. Besides, higher expression of cytoplasmic-SOX9 in human breast tumors is significantly associated to ER-status and to decreased overall survival [24]. Previous studies using biopsies of BC have shown that 75% positive immunostaining of SOX9 is observed in the nucleus of cancer cells and this expression is significantly associated with the advanced pathological grade and clinical stage [25]. In the case of GC, SOX9 overexpression has been correlated with lymph node metastasis and advanced tumoral stages, indicating that it is related to tumor progression by promoting invasion and metastasis [97] and with reduced disease-free survival [26]. Analysis of tissue microarrays of prostate biopsies samples of patient with metastatic castration-resistant PCa (mCRPC) showed 18.3% and 87.3% of patients with positive ERG and SOX9 expression, respectively [27]. Besides, ERG and SOX9 are significant risk factors for lower prostate-specific antigenprogression free survival (PFS), Clinical/radiological-PFS, and overall survival after docetaxel treatment suggesting that ERG and SOX9 were potential biomarkers for prediction to docetaxel treatment in mCRPC patients [27]. Conversely, a gradual decrease of SOX9 has been related to a progression to advanced stage, high Gleason grade, and metastatic growth in ERG-positive cancers, and these effects were strictly limited to the subset of prostate cancers harboring PTEN deletions [28].

Clinical relevance of SOX9 in ovarian cancer relies on its coexpression with HIF-2 α under hypoxia conditions, promoting TUBB3 expression. The combined presence of high TUBB3/SOXn levels is associated with a relevant reduction of PFS and overall survival in women with ovarian cancer [29]. In pancreatic cancer, there is evidence that PDAC and APC have high expression of Sox9. Also, the expression of proteins related with CSCs and EMT process is higher in APC samples than PDAC, which correlates with aggressiveness of APC [30]. SOX9 upregulation is common in colorectal adenoma and cancer and is an independent indicator for an adverse prognosis in CRC [31]. Conversely, low levels of SOX9 at the invasive front of the primary tumor have been shown as an independent predictor of relapse in stage II colon cancer patients [32]. Studies over African Americans CRC cases, diagnosed at earlier ages compared to non-Hispanic withes, have found that SOX9, GATA6, TET1, GLIS1, and FAT1 are differentially hypermethylated in APC mutationnegative CRC; this lack of APC mutation was associated with the early-onset CRC [33].

5. Concluding Remarks

Nowadays, we have a solid background about SOX9 function in normal embryonic and adult tissues, and a whole network of regulatory mechanisms depends on and influences SOX9 expression and activity. However, SOX9 expression is also a common characteristic of CSCs. Emerging evidence suggests that CSCs play a crucial role in the development and progression of malignancies. It is already known that

Model	Status of SOX9	Effects	References
353 tumors samples of CRC	SOX9 mutated and WT are overexpressed	Truncating SOX9 mutations are associated with SOX9 overexpression, KRAS mutation, and TP53 wild type	[49, 50]
DLD-1 cell line of CRC	Loss of SOX9 transcriptional activity by L142P mutation	Restoration of wild type SOX9 expression inhibits cell growth, clonal capacity and colonosphere formation; besides, the expression of the c-MYC and the activity of Wnt/ß-catenin signaling pathway are affected	[6]
17 tumors samples of CRC	High levels of SOX9 and MiniSOX9	Overexpression of MiniSOx9 is found in CCR tissues whereas SOX9 is also expressed in normal and adjacent tissues	[51]
188 tumors samples of CRC from Chinese population	Overexpressed	Does not show significant correlation between SOX9 and β -catenin	[31]
144 primary tumors from patients diagnosed in stage II CRC	Downregulated	Low levels of SOX9 have been shown as an independent predictor of relapse in stage II colon cancer patients	[32]
45 tumors samples of CRC from African Americans population	Hypermethylated	SOX9, GATA6, TET1, GLIS1, and FAT1 are differentially hypermethylated in APC-negative CRC	[33]
CaCo2, SW480, HCT116 and HT29 cell lines of CRC	Overexpressed	The synthetic PPAR γ ligand rosiglitazone induces changes of SOX9 and β -catenin expression and subcellular localization	[52]
HT29 and HCT116 cell lines of CRC	Cofactor of NF-Y	SOX9 is necessary for the function of NF-Y in activating expression of cyclin B1, cyclin B2, cyclin dependent kinase 1 and topoisomerase II α	[53]
HCT116, SW480, SW620, DLD-1 cell lines of CRC	Overexpressed	Sox9 promotes proliferation through FOXK2	[54]
HCT116 cell line of CRC	Overexpressed	Sox9 promotes invasiveness and metastasis in CRC through S100P	[55]
CCD 841 CoN, DLD-1, HCT-116, and HT-29 cell lines of CRC under hypoxia	Overexpressed	Sox9 upregulates the expression of USP47 promoting EMT under hypoxia	[56]
SW620 and SW480 cell lines of CRC	Overexpressed	SOX9 mediates the acquisition and maintenance of CR-CSCs	[57]

TABLE 3: SOX9 roles in CRC as oncogene and tumor suppressor.

SOX9 has an adaptable role since it participates in different steps of cancer progression. For instance, SOX9 is very important in the initiation of pancreatic, gastric, and prostate cancer. Conversely, in bladder and colorectal cancer, SOX9 participates in the progression of the disease, whereas it is correlated to metastasis in breast, gastric, pancreatic, and colorectal cancer. Moreover, SOX9 is clinically relevant as it may contribute in diagnosis, prognosis, and therapeutic among diverse types of cancer. This is because its expression levels and location could be cytoplasmic or nuclear depending on the stage, place, and aggressiveness. Thus, it could serve as a potential biomarker. Besides, SOX9 expression levels are related with chemoresistance in gastric, pancreatic, and colorectal cancer and high expression of SOX9 in several solid tumors is related to poor overall survival, biochemical recurrence-free survival, disease-specific survival, and DFS. Even though SOX9 has a pivotal role in different types of cancer, it has been described as an oncogene and as tumor suppressor. In this regard, it is remarkably important to consider that differences in cell lines, animal models, and populations may cause diverse outcomes. Therefore, more work is needed to study SOX9 participation in Wnt/ β -catenin and other pathways, including its relationship with other TFs (Table 4) related with stem-cell maintenance in different types of cancer, in order to elucidate additional mechanisms through which it may function. This is specially required for gaining a better understanding of SOX9 roles in normal and disease states to developing novel cancer therapeutic strategies.

Conflicts of Interest

The authors declare no conflicts of interest.

Type of cancer	TF	Effects	References
Breast cancer	SLUG (SNA12)	Induction and maintenance of tumor initiating capacity in breast cancer cells	[28]
Breast cancer	TP53	Increased proliferation	[59-61]
Pancreatic cancer	HNF6 (ONECUT1)	Produces ectopic expression of Sox9 in acinar cells converting them in ductal cells	[34, 35]
Pancreatic cancer	NF-ĸB	NF-kB subunit p65 positively regulates SOX9 expression by directly binding to the SOX9 promoter	[40]
Pancreatic cancer	GLII	Induces the transcription of SOX9	[62–66]
Pancreatic cancer	PDX1	Co-expressed in the cytoplasm with Sox9 in solid pseudopapillary tumors	[67, 68]
Prostate cancer	AR	Downstream target of SOX9	[69]
Prostate cancer	TCF4	It is positively regulated by SOX9	[20]
Prostate cancer	ZBTB7A (POKEMON)	It is lost in advance prostate cancer facilitating the oncogenic activity of SOX9	[71]
Ovarian cancer	HIF2A (EPAS1)	Hif-2α and Sox9 promote TUBB3 expression. High expression of TUBB3 and SOX9 correlates with shorter overall survival	[29]
Colorectal cancer	TCF4	Positively regulates SOX9	[72]
Colorectal cancer	$PPAR\gamma$	In cell lines ligand-dependent PPAR γ activation unevenly influences SOX9 and β -catenin expression and subcellular localization, suggesting a variable mechanistic role in colon carcinogenesis	[52]
Colorectal cancer	NF-Y	SOX9 is necessary for the function of NF-Y in activating expression of cyclin B1, cyclin B2, cyclin dependent kinase 1 and topoisomerase II α	[53]
Colorectal cancer	FOXK2	Is a SOX9 target and promotes proliferation	[54]

H	
S S	
ž	
I	
ŭ	
Ľ.	
5	
65	
õ.	
5	
÷.	
e	
Š	
2	
5	
- <u>-</u>	
0	
ц	
ŝ	
Z	
Ĕ	
U.	
а,	
4	
q	
0	
÷	
D_	
· 🗖	
5	
Š	
ц	
a	
E	
_	
Ŋ.	
Ľ,	
a	
õ	
š	
S	
a	
q	
ã	
a	
~	
2	
~	
0	
Ś	
Ψ	
Ö	
0	
Ĕ	
9	
щ	
•.•	
4	
ш	
3	
B	
4	
H	

Acknowledgments

The authors acknowledge Consejo Nacional de Ciencia y Tecnología CONACYT (grant 290311) for funding. Mariana Avendaño-Félix (575985), Erik Lizárraga-Verdugo (304939), and Mercedes Bermúdez (220327) received CONACYT fellowships.

References

- B. Beck and C. Blanpain, "Unravelling cancer stem cell potential," *Nature Reviews Cancer*, vol. 13, no. 10, pp. 727–738, 2013.
- [2] A. Kuşoğlu and Ç. Biray Avcı, "Cancer stem cells: a brief review of the current status," *Gene*, vol. 681, pp. 80–85, 2019.
- [3] R. Sekido and R. Lovell-Badge, "Sex determination and SRY: down to a wink and a nudge?" *Trends in Genetics*, vol. 25, no. 1, pp. 19–29, 2009.
- [4] M. Wegner, "All purpose sox: the many roles of sox proteins in gene expression," *The International Journal of Biochemistry & Cell Biology*, vol. 42, no. 3, pp. 381–390, 2010.
- [5] H.-Y. Wang, P. Lian, and P.-S. Zheng, "SOX9, a potential tumor suppressor in cervical cancer, transactivates p21WAF1/CIP1 and suppresses cervical tumor growth," *Oncotarget*, vol. 6, no. 24, pp. 20711–20722, 2015.
- [6] C. Prévostel, C. Rammah-Bouazza, H. Trauchessec et al., "SOX9 is an atypical intestinal tumor suppressor controlling the oncogenic Wnt/ss-catenin signaling," *Oncotarget*, vol. 7, no. 50, pp. 82228–82243, 2016.
- [7] A. Jo, S. Denduluri, B. Zhang et al., "The versatile functions of Sox9 in development, stem cells, and human diseases," *Genes & Diseases*, vol. 1, no. 2, pp. 149–161, 2014.
- [8] S. Luanpitpong, J. Li, A. Manke et al., "SLUG is required for SOX9 stabilization and functions to promote cancer stem cells and metastasis in human lung carcinoma," *Oncogene*, vol. 35, no. 22, pp. 2824–2833, 2016.
- [9] W. Guo, Z. Keckesova, J. L. Donaher et al., "Slug and Sox9 cooperatively determine the mammary stem cell state," *Cell*, vol. 148, no. 5, pp. 1015–1028, 2012.
- [10] M. A. Voronkova, S. Luanpitpong, L. W. Rojanasakul et al., "SOX9 regulates cancer stem-like properties and metastatic potential of single-walled carbon nanotube-exposed cells," *Scientific Reports*, vol. 7, no. 1, p. 11653, 2017.
- [11] A. H. Sinclair, P. Berta, M. S. Palmer et al., "A gene from the human sex-determining region encodes a protein with homology to a conserved DNA-binding motif," *Nature*, vol. 346, no. 6281, pp. 240–244, 1990.
- [12] A. Symon and V. Harley, "SOX9: A genomic view of tissue specific expression and action," *The International Journal of Biochemistry & Cell Biology*, vol. 87, pp. 18–22, 2017.
- [13] G. Badis, M. F. Berger, A. A. Philippakis et al., "Diversity and complexity in DNA recognition by transcription factors," *Science*, vol. 324, no. 5935, pp. 1720–1723, 2009.
- [14] M. Wegner, "From head to toes: the multiple facets of Sox proteins," *Nucleic Acids Research*, vol. 27, no. 6, pp. 1409–1420, 1999.
- [15] Y.-H. Huang, A. Jankowski, K. S. E. Cheah, S. Prabhakar, and R. Jauch, "SOXE transcription factors form selective dimers on non-compact DNA motifs through multifaceted interactions between dimerization and high-mobility group domains," *Scientific Reports*, vol. 5, pp. 10398-10398, 2015.

- [16] H. Sim, A. Argentaro, and V. R. Harley, "Boys, girls and shuttling of SRY and SOX9," *Trends in Endocrinology & Metabolism*, vol. 19, no. 6, pp. 213–222, 2008.
- [17] C. Peng, N. Li, Y.-K. Ng et al., "A unilateral negative feedback loop between miR-200 microRNAs and Sox2/E2F3 controls neural progenitor cell-cycle exit and differentiation," *The Journal of Neuroscience*, vol. 32, no. 38, pp. 13292–13308, 2012.
- [18] T. Oosterveen, S. Kurdija, Z. Alekseenko et al., "Mechanistic differences in the transcriptional interpretation of local and long-range shh morphogen signaling," *Developmental Cell*, vol. 23, no. 5, pp. 1006–1019, 2012.
- [19] P. Bernard and V. R. Harley, "Acquisition of SOX transcription factor specificity through protein-protein interaction, modulation of Wnt signalling and post-translational modification," *The International Journal of Biochemistry & Cell Biology*, vol. 42, no. 3, pp. 400–410, 2010.
- [20] A. Sarkar and K. Hochedlinger, "The SOX family of transcription factors: versatile regulators of stem and progenitor cell fate," *Cell Stem Cell*, vol. 12, no. 1, pp. 15–30, 2013.
- [21] C. E. Scott, S. L. Wynn, A. Sesay et al., "SOX9 induces and maintains neural stem cells," *Nature Neuroscience*, vol. 13, no. 10, pp. 1181–1189, 2010.
- [22] X. Guo, L. Xiong, T. Sun et al., "Expression features of SOX9 associate with tumor progression and poor prognosis of hepatocellular carcinoma," *Diagnostic Pathology*, vol. 7, no. 1, article 44, 2012.
- [23] G. Richtig, A. Aigelsreiter, D. Schwarzenbacher et al., "SOX9 is a proliferation and stem cell factor in hepatocellular carcinoma and possess widespread prognostic significance in different cancer types," *PLoS ONE*, vol. 12, no. 11, p. e0187814, 2017.
- [24] G. Chakravarty, B. Rider, and D. Mondal, "Cytoplasmic compartmentalization of SOX9 abrogates the growth arrest response of breast cancer cells that can be rescued by Trichostatin A treatment," *Cancer Biology & Therapy*, vol. 11, no. 1, pp. 71–83, 2011.
- [25] Y.-P. Wan, M. Xi, H.-C. He et al., "Expression and clinical significance of SOX9 in renal cell carcinoma, bladder cancer and penile cancer," *Oncology Research and Treatment*, vol. 40, no. 1-2, pp. 15–20, 2017.
- [26] J. C. Santos, E. Carrasco-Garcia, M. Garcia-Puga et al., "SOX9 elevation acts with canonical WNT signaling to drive gastric cancer progression," *Cancer Research*, vol. 76, no. 22, pp. 6735– 6746, 2016.
- [27] W. Song, G. Y. Kwon, J. H. Kim et al., "Immunohistochemical staining of ERG and SOX9 as potential biomarkers of docetaxel response in patients with metastatic castration-resistant prostate cancer," *Oncotarget*, vol. 7, no. 50, pp. 83735–83743, 2016.
- [28] C. Burdelski, E. Bujupi, M. C. Tsourlakis et al., "Loss of SOX9 expression is associated with PSA recurrence in ERG-positive and pten deleted prostate cancers," *PLoS ONE*, vol. 10, no. 6, Article ID e0128525, 2015.
- [29] G. Raspaglio, M. Petrillo, E. Martinelli et al., "Sox9 and Hif-2α regulate TUBB3 gene expression and affect ovarian cancer aggressiveness," *Gene*, vol. 542, no. 2, pp. 173–181, 2014.
- [30] K. Miura, K. Kimura, R. Amano et al., "Analysis of the origin of anaplastic pancreatic cancer and the mechanism of its dedifferentiation," *Journal of Hepato-Biliary-Pancreatic Sciences*, vol. 24, no. 3, pp. 176–184, 2017.
- [31] B. Lü, Y. Fang, J. Xu et al., "Analysis of SOX9 expression in colorectal cancer," *American Journal of Clinical Pathology*, vol. 130, no. 6, pp. 897–904, 2008.

- [32] M. L. Marcker Espersen, D. Linnemann, I. J. Christensen, M. Alamili, J. T. Troelsen, and E. Høgdall, "SOX9 expression predicts relapse of stage II colon cancer patients," *Human Pathology*, vol. 52, pp. 38–46, 2016.
- [33] R. M. Xicola, Z. Manojlovic, G. J. Augustus et al., "Lack of APC somatic mutation is associated with early-onset colorectal cancer in African Americans," *Carcinogenesis*, vol. 39, no. 11, pp. 1331–1341, 2018.
- [34] P.-P. Prévot, A. Simion, A. Grimont et al., "Role of the ductal transcription factors HNF6 and Sox9 in pancreatic acinar-toductal metaplasia," *Gut*, vol. 61, no. 12, pp. 1723–1732, 2012.
- [35] E. Hessmann, J.-S. Zhang, N.-M. Chen et al., "NFATc4 Regulates Sox9 gene expression in acinar cell plasticity and pancreatic cancer initiation," *Stem Cells International*, vol. 2016, Article ID 5272498, 11 pages, 2016.
- [36] P. Jeannot, C. Callot, R. Baer et al., "Loss of p27Kip1 promotes metaplasia in the pancreas via the regulation of Sox9 expression," *Oncotarget*, vol. 6, no. 34, pp. 35880–35892, 2015.
- [37] J. L. Kopp, G. von Figura, E. Mayes et al., "Identification of Sox9dependent acinar-to-ductal reprogramming as the principal mechanism for initiation of pancreatic ductal adenocarcinoma," *Cancer Cell*, vol. 22, no. 6, pp. 737–750, 2012.
- [38] S. Xia, Z. Feng, X. Qi et al., "Clinical implication of Sox9 and activated Akt expression in pancreatic ductal adenocarcinoma," *Medical Oncology*, vol. 32, no. 1, p. 358, 2015.
- [39] L. Huang, A. Holtzinger, I. Jagan et al., "Ductal pancreatic cancer modeling and drug screening using human pluripotent stem cell- and patient-derived tumor organoids," *Nature Medicine*, vol. 21, no. 11, p. 1364, 2015.
- [40] L. Sun, L. A. Mathews, S. M. Cabarcas et al., "Epigenetic regulation of SOX9 by the NF-κB signaling pathway in pancreatic cancer stem cells," *Stem Cells*, vol. 31, no. 8, pp. 1454–1466, 2013.
- [41] M. J. Schultz, A. T. Holdbrooks, A. Chakraborty et al., "The tumor-associated glycosyltransferase ST6Gal-I regulates stem cell transcription factors and confers a cancer stem cell phenotype," *Cancer Research*, vol. 76, no. 13, pp. 3978–3988, 2016.
- [42] T. Higashihara, H. Yoshitomi, Y. Nakata et al., "Sex determining region y box 9 induces chemoresistance in pancreatic cancer cells by induction of putative cancer stem cell characteristics and its high expression predicts poor prognosis," *Pancreas*, vol. 46, no. 10, pp. 1296–1304, 2017.
- [43] W. Deng, D. B. Vanderbilt, C. Lin, K. H. Martin, K. M. Brundage, and J. M. Ruppert, "SOX9 inhibits beta-TrCPmediated protein degradation to promote nuclear GL11 expression and cancer stem cell properties," *Journal of Cell Science*, vol. 128, no. 6, pp. 1123–1138, 2015.
- [44] L. G. Kondratyeva, A. A. Sveshnikova, E. V. Grankina et al., "Downregulation of expression of mater genes SOX9, FOXA2, and GATA4 in pancreatic cancer cells stimulated with TGFβ1 epithelial–mesenchymal transition," *Doklady Biochemistry and Biophysics*, vol. 469, no. 1, pp. 257–259, 2016.
- [45] A. V. Pinho, I. Rooman, and F. X. Real, "p53-dependent regulation of growth, epithelial-mesenchymal transition and stemness in normal pancreatic epithelial cells," *Cell Cycle*, vol. 10, no. 8, pp. 1312–1321, 2011.
- [46] P. Camaj, C. Jackel, S. Krebs et al., "Hypoxia-independent gene expression mediated by SOX9 promotes aggressive pancreatic tumor biology," *Molecular Cancer Research*, vol. 12, no. 3, pp. 421–432, 2014.
- [47] F. Meng, K. Takaori, T. Ito et al., "Expression of SOX9 in intraductal papillary mucinous neoplasms of the pancreas," *Pancreas*, vol. 43, no. 1, pp. 7–14, 2014.

- [48] L. Galmiche, S. Sarnacki, V. Verkarre et al., "Transcription factors involved in pancreas development are expressed in paediatric solid pseudopapillary tumours," *Histopathology*, vol. 53, no. 3, pp. 318–324, 2008.
- [49] B. M. Javier, R. Yaeger, L. Wang et al., "Recurrent, truncating Sox9 mutations are associated with sox9 overexpression, KRAS mutation, and TP53 wild type status in colorectal carcinoma," *Oncotarget*, vol. 7, no. 32, pp. 50875–50882, 2016.
- [50] K. S. Al-Kuraya, "Editorial: KRAS and TP53 mutations in colorectal carcinoma," *Saudi Journal of Gastroenterology*, vol. 15, no. 4, pp. 217–219, 2009.
- [51] R. Abdel-Samad, H. Zalzali, C. Rammah et al., "MiniSOX9, a dominant-negative variant in colon cancer cells," *Oncogene*, vol. 30, no. 22, pp. 2493–2503, 2011.
- [52] A. Panza, "Interplay between SOX9, beta-catenin and PPARgamma activation in colorectal cancer," *Biochimica et Biophysica Acta*, vol. 1833, no. 8, pp. 1853-65, Aug 2013 (English).
- [53] Z. Shi, C.-I. Chiang, P. Labhart et al., "Context-specific role of SOX9 in NF-Y mediated gene regulation in colorectal cancer cells," *Nucleic Acids Research*, vol. 43, no. 13, pp. 6257–6269, 2015.
- [54] Y. Qian, S. Xia, and Z. Feng, "Sox9 mediated transcriptional activation of FOXK2 is critical for colorectal cancer cells proliferation," *Biochemical and Biophysical Research Communications*, vol. 483, no. 1, pp. 475–481, 2017.
- [55] Z. Shen, H. Deng, Y. Fang et al., "Identification of the interplay between SOX9 and S100P in the metastasis and invasion of colon carcinoma," *Oncotarget*, vol. 6, no. 24, pp. 20672–20684, 2015.
- [56] B. Choi, S. Park, S. Lee, Y. N. Cha, and Y. Surh, "Hypoxia induces epithelial-mesenchymal transition in colorectal cancer cells through ubiquitin-specific protease 47-mediated stabilization of Snail: A potential role of Sox9," *Scientific Reports*, vol. 7, no. 1, 2017.
- [57] E. Carrasco-Garcia, L. Lopez, P. Aldaz et al., "SOX9-regulated cell plasticity in colorectal metastasis is attenuated by rapamycin," *Scientific Reports*, vol. 6, no. 1, p. 32350, 2016.
- [58] H. Fazilaty, M. Gardaneh, P. Akbari, A. Zekri, and B. Behnam, "SLUG and SOX9 cooperatively regulate tumor initiating niche factors in breast cancer," *Cancer Microenvironment*, vol. 9, no. 1, pp. 71–74, 2016.
- [59] G. Chakravarty, K. Moroz, N. M. Makridakis et al., "Prognostic significance of cytoplasmic SOX9 in invasive ductal carcinoma and metastatic breast cancer," *Experimental Biology* and Medicine, vol. 236, no. 2, pp. 145–155, 2011.
- [60] Q.-Y. Wang et al., "MiR-133b targets Sox9 to control pathogenesis and metastasis of breast cancer," *Cell Death & Disease*, vol. 9, no. 7, pp. 752–2018, 2018.
- [61] B. Lei, Y. Zhang, T. Liu, Y. Li, and D. Pang, "Sox9 upregulation in breast cancer is correlated with poor prognosis and the CD44+/CD24-/low phenotype," *International Journal of Clinical and Experimental Pathology*, vol. 9, no. 7, 2016.
- [62] M. Lauth and R. Toftgård, "Non-canonical activation of GLI transcription factors: Implications for targeted anti-cancer therapy," *Cell Cycle*, vol. 6, no. 20, pp. 2458–2463, 2007.
- [63] A. Ruiz i Altaba, C. Mas, and B. Stecca, "The Gli code: an information nexus regulating cell fate, stemness and cancer," *Trends in Cell Biology*, vol. 17, no. 9, pp. 438–447, 2007.
- [64] B. Stecca and A. Ruiz I Altaba, "Context-dependent regulation of the GLI code in cancer by HEDGEHOG and non-HEDGEHOG signals," *Journal of Molecular Cell Biology*, vol. 2, no. 2, pp. 84–95, 2010.

- [65] J. P. Morris, S. C. Wang, and M. Hebrok, "KRAS, Hedgehog, Wnt and the twisted developmental biology of pancreatic ductal adenocarcinoma," *Nature Reviews Cancer*, vol. 10, no. 10, pp. 683–695, 2010.
- [66] C.-C. Hui and S. Angers, "Gli proteins in development and disease," *Annual Review of Cell and Developmental Biology*, vol. 27, pp. 513–537, 2011.
- [67] O. Strobel, W. Hartwig, F. Bergmann et al., "Anaplastic pancreatic cancer: presentation, surgical management, and outcome," *Surgery*, vol. 149, no. 2, pp. 200–208, 2011.
- [68] C. J. Clark, R. P. Graham, J. S. Arun, W. S. Harmsen, and K. M. Reid-Lombardo, "Clinical outcomes for anaplastic pancreatic cancer: a population-based study," *Journal of the American College of Surgeons*, vol. 215, no. 5, pp. 627–634, 2012.
- [69] G. S. Evans and J. A. Chandler, "Cell proliferation studies in rat prostate I. the proliferative role of basal and secretory epithelial cells during normal growth," *The Prostate*, vol. 10, no. 2, pp. 163– 178, 1987.
- [70] F. Ma, H. Ye, H. H. He et al., "SOX9 drives WNT pathway activation in prostate cancer," *The Journal of Clinical Investigation*, vol. 126, no. 5, pp. 1745–1758, 2016.
- [71] G. Wang, A. Lunardi, J. Zhang et al., "Zbtb7a suppresses prostate cancer through repression of a Sox9-dependent pathway for cellular senescence bypass and tumor invasion," *Nature Genetics*, vol. 45, no. 7, pp. 739–746, 2013.
- [72] V. Korinek, N. Barker, P. J. Morin et al., "Constitutive transcriptional activation by a beta-catenin-Tcf complex in APC^{-/-} colon carcinoma," *Science*, vol. 275, no. 5307, pp. 1784–1787, 1997.
- [73] A. Matheu, M. Collado, C. Wise et al., "Oncogenicity of the developmental transcription factor Sox9," *Cancer Research*, vol. 72, no. 5, pp. 1301–1315, 2012.
- [74] W. S. Institute, "Catalogue of somatic mutations in cancer," https://cancer.sanger.ac.uk/cosmic.
- [75] S. A. Forbes, D. Beare, H. Boutselakis et al., "COSMIC: Somatic cancer genetics at high-resolution," *Nucleic Acids Research*, vol. 45, no. 1, pp. D777–D783, 2017.
- [76] J. Balogh, D. Victor, E. H. Asham et al., "Hepatocellular carcinoma: a review," *Journal of Hepatocellular Carcinoma*, vol. 3, pp. 41–53, 2016.
- [77] W. Lin, Y.-L. Chen, L. Jiang, and J.-K. Chen, "Reduced expression of chemerin is associated with a poor prognosis and a lowed infiltration of both dendritic cells and natural killer cells in human hepatocellular carcinoma," *Clinical Laboratory*, vol. 57, no. 11-12, pp. 879–885, 2011.
- [78] M. Dettmer, P. Itin, P. Miny, M. Gandhi, G. Cathomas, and N. Willi, "Giant ectopic liver, hepatocellular carcinoma and pachydermia-a rare genetic syndrome?" *Diagnostic Pathology*, vol. 6, no. 1, article no. 75, 2011.
- [79] S. Whittaker, R. Marais, and A. X. Zhu, "The role of signaling pathways in the development and treatment of hepatocellular carcinoma," *Oncogene*, vol. 29, no. 36, pp. 4989–5005, 2010.
- [80] Y. Kawaguchi, "Sox9 and programming of liver and pancreatic progenitors," *The Journal of Clinical Investigation*, vol. 123, no. 5, pp. 1881–1886, 2013.
- [81] T.-C. Xue, L. Zhang, Z.-G. Ren et al., "Sex-determination gene SRY potentially associates with poor prognosis but not sex bias in hepatocellular carcinoma," *Digestive Diseases and Sciences*, vol. 60, no. 2, pp. 427–435, 2015.
- [82] C. O.-N. Leung, W.-N. Mak, A. K. L. Kai et al., "Sox9 confers stemness properties in hepatocellular carcinoma through Frizzled-7 mediated Wnt/β-catenin signaling," *Oncotarget*, vol. 7, no. 20, pp. 29371–29386, 2016.

- [83] C. Coulouarn, C. Cavard, L. Rubbia-Brandt et al., "Combined hepatocellular-cholangiocarcinomas exhibit progenitor features and activation of Wnt and TGF β signaling pathways," *Carcinogenesis*, vol. 33, no. 9, pp. 1791–1796, 2012.
- [84] D. W. Cleere, "Triple-negative breast cancer: a clinical update," *Community Oncology*, vol. 7, no. 5, pp. 203–211, 2010.
- [85] G. K. Malhotra, X. Zhao, E. Edwards et al., "The role of Sox9 in mouse mammary gland development and maintenance of mammary stem and luminal progenitor cells," *BMC Developmental Biology*, vol. 14, no. 1, p. 47, 2014.
- [86] J. Ferlay, I. Soerjomataram, R. Dikshit et al., "Cancer incidence and mortality worldwide: sources, methods and major patterns in GLOBOCAN 2012," *International Journal of Cancer*, vol. 136, no. 5, pp. E359–386, 2014.
- [87] C. Vallot, N. Stransky, I. Bernard-Pierrot et al., "A novel epigenetic phenotype associated with the most aggressive pathway of bladder tumor progression," *Journal of the National Cancer Institute*, vol. 103, no. 1, pp. 47–60, 2011.
- [88] A. Aleman, L. Adrien, L. Lopez-Serra et al., "Identification of DNA hypermethylation of SOX9 in association with bladder cancer progression using CpG microarrays," *British Journal of Cancer*, vol. 98, no. 2, pp. 466–473, 2008.
- [89] J. Sun, Z. Chen, T. Zhu et al., "Hypermethylated SFRP1, but none of other nine genes "informative" for western countries, is valuable for bladder cancer detection in Mainland China.," *Journal of Cancer Research and Clinical Oncology*, vol. 135, no. 12, pp. 1717–1727, 2009.
- [90] J. Yu, T. Zhu, Z. Wang et al., "A novel set of DNA methylation markers in urine sediments for sensitive/specific detection of bladder cancer," *Clinical Cancer Research*, vol. 13, no. 24, pp. 7296–7304, 2007.
- [91] H. S. Son, Y. M. Shin, K. K. Park et al., "Correlation between HER2 overexpression and clinicopathological characteristics in gastric cancer patients who have undergone curative resection," *Gastric Cancer*, vol. 14, no. 3, pp. 180–186, 2014.
- [92] W. Yasui, N. Oue, K. Sentani, N. Sakamoto, and J. Motoshita, "Transcriptome dissection of gastric cancer: Identification of novel diagnostic and therapeutic targets from pathology specimens," *Pathology International*, vol. 59, no. 3, pp. 121–136, 2009.
- [93] M. Sashikawa Kimura, H. Mutoh, and K. Sugano, "SOX9 is expressed in normal stomach, intestinal metaplasia, and gastric carcinoma in humans," *Journal of Gastroenterology*, vol. 46, no. 11, pp. 1292–1299, 2011.
- [94] Y. J. Choi, J. H. Song, J. H. Yoon et al., "Aberrant expression of SOX9 is associated with gastrokine 1 inactivation in gastric cancers," *Gastric Cancer*, vol. 17, no. 2, pp. 247–254, 2014.
- [95] X. Ren, D. Zheng, F. Guo et al., "PPARy suppressed Wnt/βcatenin signaling pathway and its downstream effector SOX9 expression in gastric cancer cells," *Medical Oncology*, vol. 32, no. 4, p. 91, 2015.
- [96] J. H. Yoon, J. H. Song, C. Zhang et al., "Inactivation of the Gastrokine 1 gene in gastric adenomas and carcinomas," *The Journal of Pathology*, vol. 223, no. 5, pp. 618–625, 2011.
- [97] C. Zhou, J. Guo, K. Zhu et al., "Elevated expression of SOX9 is related with the progression of gastric carcinoma," *Diagnostic Cytopathology*, vol. 39, no. 2, pp. 105–109, 2011.
- [98] J. Wang, X. Xue, H. Hong et al., "Upregulation of microRNA-524-5p enhances the cisplatin sensitivity of gastric cancer cells by modulating proliferation and metastasis via targeting SOX9," *Oncotarget*, vol. 8, no. 1, pp. 574–582, 2017.

- [99] J.-N. Liu, Y.-M. Shang Guan, Y.-Z. Qi, H.-B. Wang, T.-G. Zhang, and C.-J. Zhou, "The evaluation of SOX9 expression and its relationship with carcinoembryonic antigen-related cell adhesion molecule 1 in gastric neoplastic and nonneoplastic lesions," *Annals of Diagnostic Pathology*, vol. 16, no. 4, pp. 235– 244, 2012.
- [100] T. Serizawa, Y. Hirata, Y. Hayakawa et al., "Gastric metaplasia induced by Helicobacter pylori is associated with enhanced SOX9 expression via interleukin-1 signaling," *Infection and Immunity*, vol. 84, no. 2, pp. 562–572, 2015.
- [101] M. Amieva and R. M. Peek Jr., "Pathobiology of helicobacter pylori-induced gastric cancer," *Gastroenterology*, vol. 150, no. 1, pp. 64–78, 2016.
- [102] R. Wadhwa, S. Song, J. Lee, Y. Yao, Q. Wei, and J. A. Ajani, "Gastric cancer—molecular and clinical dimensions," *Nature Reviews Clinical Oncology*, vol. 10, no. 11, pp. 643–655, 2013.
- [103] M. Sigal, M. E. Rothenberg, C. Y. Logan et al., "Helicobacter pylori activates and expands Lgr5+ stem cells through direct colonization of the gastric glands," *Gastroenterology*, vol. 148, no. 7, pp. 1392–1404.e21, 2015.
- [104] M. Sun, H. Uozaki, R. Hino et al., "SOX9 expression and its methylation status in gastric cancer," *Virchows Archiv*, vol. 460, no. 3, pp. 271–279, 2012.
- [105] P. A. Seymour, K. K. Freude, M. N. Tran et al., "SOX9 is required for maintenance of the pancreatic progenitor cell pool," *Proceedings of the National Acadamy of Sciences of the United States of America*, vol. 104, no. 6, pp. 1865–1870, 2007.
- [106] K. Furuyama, Y. Kawaguchi, H. Akiyama et al., "Continuous cell supply from a Sox9-expressing progenitor zone in adult liver, exocrine pancreas and intestine," *Nature Genetics*, vol. 43, no. 1, pp. 34–41, 2011.
- [107] J. P. T. Morris IV, D. A. Cano, S. Sekine, S. C. Wang, and M. Hebrok, "Beta-catenin blocks Kras-dependent reprogramming of acini into pancreatic cancer precursor lesions in mice," *The Journal of Clinical Investigation*, vol. 120, no. 2, pp. 508–520, 2010.
- [108] A. Gnoni, A. Licchetta, A. Scarpa et al., "Carcinogenesis of pancreatic adenocarcinoma: precursor lesions," *International Journal of Molecular Sciences*, vol. 14, no. 10, pp. 19731–19762, 2013.
- [109] R. H. Hruban, N. V. Adsay, J. Albores-Saavedra et al., "Pancreatic intraepithelial neoplasia: a new nomenclature and classification system for pancreatic duct lesions," *The American Journal* of Surgical Pathology, vol. 25, no. 5, pp. 579–586, 2001.
- [110] M. L. Fero, M. Rivkin, M. Tasch et al., "A syndrome of multiorgan hyperplasia with features of gigantism, tumorigenesis, and female sterility in p27Kip1-deficient Mice," *Cell*, vol. 85, no. 5, pp. 733–744, 1996.
- [111] H. Kiyokawa, R. D. Kineman, K. O. Manova-Todorova et al., "Enhanced growth of mice lacking the cyclin-dependent kinase inhibitor function of P27Kip1," *Cell*, vol. 85, no. 5, pp. 721–732, 1996.
- [112] K. Nakayama, N. Ishida, M. Shirane et al., "Mice lacking p27(Kip1) display increased body size, multiple organ hyperplasia, retinal dysplasia, and pituitary tumors," *Cell*, vol. 85, no. 5, pp. 707–720, 1996.
- [113] C. Navas, I. Hernández-Porras, A. J. Schuhmacher, M. Sibilia, C. Guerra, and M. Barbacid, "EGF receptor signaling is essential for k-ras oncogene-driven pancreatic ductal adenocarcinoma," *Cancer Cell*, vol. 22, no. 3, pp. 318–330, 2012.

- [114] N. Bardeesy and R. A. DePinho, "Pancreatic cancer biology and genetics," *Nature Reviews Cancer*, vol. 2, no. 12, pp. 897–909, 2002.
- [115] M. Delous, C. Yin, D. Shin et al., "sox9b is a key regulator of pancreaticobiliary ductal system development," *PLoS Genetics*, vol. 8, no. 6, Article ID e1002754, 2012.
- [116] I. Vivanco and C. L. Sawyers, "The phosphatidylinositol 3kinase-AKT pathway in human cancer," *Nature Reviews Cancer*, vol. 2, no. 7, pp. 489–501, 2002.
- [117] A. Balic, J. Dorado, M. Alonso-Gómez, and C. Heeschen, "Stem cells as the root of pancreatic ductal adenocarcinoma," *Experimental Cell Research*, vol. 318, no. 6, pp. 691–704, 2012.
- [118] C. Li, D. G. Heidt, P. Dalerba et al., "Identification of pancreatic cancer stem cells," *Cancer Research*, vol. 67, no. 3, pp. 1030–1037, 2007.
- [119] P. C. Hermann, S. L. Huber, T. Herrler et al., "Distinct populations of cancer stem cells determine tumor growth and metastatic activity in human pancreatic cancer," *Cell Stem Cell*, vol. 1, no. 3, pp. 313–323, 2007.
- [120] C. Li, J. Wu, M. Hynes et al., "c-Met is a marker of pancreatic cancer stem cells and therapeutic target," *Gastroenterology*, vol. 141, no. 6, pp. 2218.e5–2227.e5, 2011.
- [121] Z. A. Rasheed, J. Yang, Q. Wang et al., "Prognostic significance of tumorigenic cells with mesenchymal features in pancreatic adenocarcinoma," *Journal of the National Cancer Institute*, vol. 102, no. 5, pp. 340–351, 2010.
- [122] A. F. Swindall, A. I. Londoño-Joshi, M. J. Schultz, N. Fineberg, D. J. Buchsbaum, and S. L. Bellis, "ST6Gal-I protein expression is upregulated in human epithelial tumors and correlates with stem cell markers in normal tissues and colon cancer cell lines," *Cancer Research*, vol. 73, no. 7, pp. 2368–2378, 2013.
- [123] L. G. Kondratyeva, I. P. Chernov, M. V. Zinovyeva, E. P. Kopantzev, and E. D. Sverdlov, "Expression of master regulatory genes of embryonic development in pancreatic tumors," *Doklady Biochemistry and Biophysics*, vol. 475, no. 1, pp. 250–252, 2017.
- [124] C.-H. Heldin, M. Vanlandewijck, and A. Moustakas, "Regulation of EMT by TGFβ in cancer," *FEBS Letters*, vol. 586, no. 14, pp. 1959–1970, 2012.
- [125] M. Reichert and A. K. Rustgi, "Pancreatic ductal cells in development, regeneration, and neoplasia," *The Journal of Clinical Investigation*, vol. 121, no. 12, pp. 4572–4578, 2011.
- [126] H. E. Arda, C. M. Benitez, and S. K. Kim, "Gene regulatory networks governing pancreas development," *Developmental Cell*, vol. 25, no. 1, pp. 5–13, 2013.
- [127] A. M. Handorf and W.-J. Li, "Fibroblast growth Factor-2 primes human mesenchymal stem cells for enhanced chondrogenesis," *PLoS ONE*, vol. 6, no. 7, Article ID e22887, 2011.
- [128] G. Klöppel, O. Basturk, A. M. Schlitter, B. Konukiewitz, and I. Esposito, "Intraductal neoplasms of the pancreas," *Seminars in Diagnostic Pathology*, vol. 31, no. 6, pp. 452–466, 2014.
- [129] W. Kimura, N. Sasahira, T. Yoshikawa, T. Muto, and M. Makuuchi, "Duct-ectatic type of mucin producing tumor of the pancreas new concept of pancreatic neoplasia," *Hepato-Gastroenterology*, vol. 43, no. 9, pp. 692–709, 1996.
- [130] F. T. Bosman, W. H. Organization, and I. A. f. R. o. Cancer, WHO Classification of Tumours of the Digestive System, International Agency for Research on Cancer, 2010.
- [131] D. A. Cano, M. Hebrok, and M. Zenker, "Pancreatic development and disease," *Gastroenterology*, vol. 132, no. 2, pp. 745–762, 2007.

- [132] P. W. Heiser, J. Lau, M. M. Taketo, P. L. Herrera, and M. Hebrok, "Stabilization of beta-catenin impacts pancreas growth," *Development*, vol. 133, no. 10, pp. 2023–2032, 2006.
- [133] M. F. Offield et al., "PDX-1 is required for pancreatic outgrowth and differentiation of the rostral duodenum," *Development*, vol. 122, no. 3, pp. 983–995, 1996.
- [134] H. Wang, N. C. McKnight, T. Zhang, M. L. Lu, S. P. Balk, and X. Yuan, "SOX9 is expressed in normal prostate basal cells and regulates androgen receptor expression in prostate cancer cells," *Cancer Research*, vol. 67, no. 2, pp. 528–536, 2007.
- [135] Z. Huang, P. J. Hurley, B. W. Simons et al., "Sox9 is required for prostate development and prostate cancer initiation," *Oncotarget*, vol. 3, no. 6, pp. 651–663, 2012.
- [136] M. K. Thomsen, J. C. Francis, and A. Swain, "The role of Sox9 in prostate development," *Differentiation*, vol. 76, no. 6, pp. 728– 735, 2008.
- [137] H. Wang, I. Leav, S. Ibaragi et al., "SOX9 is expressed in human fetal prostate epithelium and enhances prostate cancer invasion," *Cancer Research*, vol. 68, no. 6, pp. 1625–1630, 2008.
- [138] W. Zhong, G. Qin, Q. Dai et al., "SOXs in human prostate cancer: implication as progression and prognosis factors," *BMC Cancer*, vol. 12, no. 1, p. 248, 2012.
- [139] G.-Q. Qin, H.-C. He, Z.-D. Han et al., "Combined overexpression of HIVEP3 and SOX9 predicts unfavorable biochemical recurrence-free survival in patients with prostate cancer," *Onco-Targets and Therapy*, vol. 7, pp. 137–146, 2014.
- [140] M. K. Thomsen, L. Ambroisine, S. Wynn et al., "SOX9 elevation in the prostate promotes proliferation and cooperates with PTEN loss to drive tumor formation," *Cancer Research*, vol. 70, no. 3, pp. 979–987, 2010.
- [141] Y. Xie, W. Lu, S. Liu et al., "Crosstalk between nuclear MET and SOX9/β-catenin correlates with castration-resistant prostate cancer," *Molecular Endocrinology*, vol. 28, no. 10, pp. 1629–1639, 2014.
- [142] C. Cai, H. Wang, H. H. He et al., "ERG induces androgen receptor-mediated regulation of SOX9 in prostate cancer," *The Journal of Clinical Investigation*, vol. 123, no. 3, pp. 1109–1122, 2013.
- [143] M. Luedeke et al., "Prostate cancer risk regions at 8q24 and 17q24 are differentially associated with somatic TMPRSS2:ERG fusion status," *Human Molecular Genetics*, vol. 25, no. 24, pp. 5490–5499, 2016.
- [144] C. Ricciardelli and M. K. Oehler, "Diverse molecular pathways in ovarian cancer and their clinical significance," *Maturitas*, vol. 62, no. 3, pp. 270–275, 2009.
- [145] R. J. Kurman, L. H. Ellenson, and B. M. Ronnettm, Blaustein's Pathology of the Female Genital Tract, Springer, 2011.
- [146] H. F. Irving-Rodgers and R. J. Rodgers, "Extracellular matrix of the developing ovarian follicle," *Seminars in Reproductive Medicine*, vol. 24, no. 4, pp. 195–203, 2006.
- [147] P. Papanastasopoulos, M. Repanti, V. Damaskou, V. Bravou, and H. Papadaki, "Investigating differentiation mechanisms of the constituent cells of sex cord-stromal tumours of the ovary," *Virchows Archiv*, vol. 453, no. 5, pp. 465–471, 2008.
- [148] D. M. Parkin, S. L. Whelan, J. Ferlay, L. Teppo, and D. B. Thomas, "Cancer incidence in five continents," *IARC Scientific Publications*, vol. VIII, no. 155, pp. 1–781, 2002 (English).
- [149] P. Boyle and J. S. Langman, "ABC of colorectal cancer epidemiology," *British Medical Journal*, vol. 321, no. 7264, pp. 805–808, 2000.

- [150] U. Testa, E. Pelosi, and G. Castelli, "Colorectal cancer: genetic abnormalities, tumor progression, tumor heterogeneity, clonal evolution and tumor-initiating cells," *Medical Sciences*, vol. 6, no. 2, p. 31, 2018.
- [151] G. Steele Jr. and T. S. Ravikumar, "Resection of hepatic metastases from colorectal cancer: biologic perspectives," *Annals of Surgery*, vol. 210, no. 2, pp. 127–138, 1989.
- [152] L. G. van der Flier and H. Clevers, "Stem cells, self-renewal, and differentiation in the intestinal epithelium," *Annual Review of Physiology*, vol. 71, pp. 241–260, 2009.
- [153] T. E. Adolph, M. F. Tomczak, L. Niederreiter et al., "Paneth cells as a site of origin for intestinal inflammation," *Nature*, vol. 503, no. 7475, pp. 272–276, 2013.
- [154] H. C. Clevers and C. L. Bevins, "Paneth cells: Maestros of the small intestinal crypts," *Annual Review of Physiology*, vol. 75, pp. 289–311, 2013.
- [155] P. Blache, M. Van De Wetering, I. Duluc et al., "SOX9 is an intestine crypt transcription factor, is regulated by the Wnt pathway, and represses the CDX2 and MUC2 genes," *The Journal of Cell Biology*, vol. 166, no. 1, pp. 37–47, 2004.
- [156] P. Bastide, C. Darido, J. Pannequin et al., "Sox9 regulates cell proliferation and is required for Paneth cell differentiation in the intestinal epithelium," *The Journal of Cell Biology*, vol. 178, no. 4, pp. 635–648, 2007.
- [157] R. Fodde, W. Edelmann, K. Yang et al., "A targeted chaintermination mutation in the mouse Apc gene results in multiple intestinal tumors," *Proceedings of the National Acadamy of Sciences of the United States of America*, vol. 91, no. 19, pp. 8969– 8973, 1994.
- [158] H. Shibata, K. Toyama, H. Shioya et al., "Rapid colorectal adenoma formation initiated by conditional targeting of the Apc gene," *Science*, vol. 278, no. 5335, pp. 120–133, 1997.
- [159] N. Harada, Y. Tamai, T.-O. Ishikawa et al., "Intestinal polyposis in mice with a dominant stable mutation of the β-catenin gene," *EMBO Journal*, vol. 18, no. 21, pp. 5931–5942, 1999.
- [160] P. A. Candy, M. R. Phillips, A. D. Redfern et al., "Notchinduced transcription factors are predictive of survival and 5-fluorouracil response in colorectal cancer patients," *British Journal of Cancer*, vol. 109, no. 4, pp. 1023–1030, 2013.
- [161] L. Montorsi, F. Guizzetti, C. Alecci et al., "Loss of zfp36 expression in colorectal cancer correlates to wnt/ β -catenin activity and enhances epithelial-to-mesenchymal transition through upregulation of zeb1, sox9 and macc1," *Oncotarget*, vol. 7, no. 37, pp. 59144–59157, 2016.