The Relation of Inflammaging With Skeletal Muscle Properties in Elderly Men

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Abstract

Aging is associated with a progressive decline of muscle mass and/or the qualitative impairment of the muscle tissue. There is growing evidence of the prominent role of low-grade chronic inflammation in age-related changes in the neuromuscular system. The purpose of the study was to identify the inflammatory mediators responsible for deficit in functional fitness and to explain whether inflammation is related to changes in body composition and the decline of muscle strength in older men. Thirty-three old-aged males (73.5 \pm 6.3 years) and twenty young-aged males $(21.2 \pm 1.3 \text{ years})$ participated in the study. The body composition (bioelectrical impedance analysis), functional capacity (6-min walking test) and knee extension strength (isokinetic test) were estimated. In serum, circulating inflammatory markers H_2O_2 , IL-I β , TNF α , and hsCRP as well as growth factors IGF-I and PDGF^{BB} concentrations were determined (immunoenzymatic methods). The concentrations of H_2O_2 , IL-1 β , TNF α , and hsCRP were significantly higher in older than young men. The growth factors IGF-I and PDGF^{BB} were twofold lower and related to high levels of IL-1 β and TNF α in the elderly. The changes in cytokines and growth factors levels were correlated with age and peak torque (TQ at 60° /s and 180° /s) in the knee extension. The result of the 6-min walking test was inversely correlated with fat mass index (FMI, r = -.983; p < .001). The generation of inflammatory mediators in older men was related to changes in body composition, maximum strength muscle, and age-related changes in skeletal muscle properties responsible for deficit in functional fitness.

Keywords

body composition, cytokines, functional fitness, growth factors, inflammation

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Skeletal muscle contractions empower human body movements and are essential for maintaining stability. Skeletal muscle tissue accounts for almost half of the human body mass and, in addition to its power-generating role, is a crucial factor in maintaining homeostasis. Given its central role in human mobility and metabolic function, any deterioration in the contractile, material, and metabolic properties of skeletal muscle has an extremely important effect on human health (Zembron-Lacny, Dziubek, Rogowski, Skorupka, & Dąbrowska, 2014).

Aging is associated with a progressive decline of muscle mass and/or the qualitative impairment of the muscle tissue. There is growing evidence of the prominent role of low-grade chronic inflammation in age-related changes in the neuromuscular system (Salminen, Kaarniranta, & Kauppinen, 2012). Multiple causes may contribute to inflammaging, such as pro-inflammatory tissue damages, a

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dysfunctional immune system (Deeks, 2011), proinflammatory cytokines secreted by senescent cells, enhanced NF-KB (nuclear factor kappa-light-chainenhancer of activated B cells) activation, and a defective autophagy response system (Salminen et al., 2012). These factors enhance the activation of inflammatory pathways such as the Nalp-3 inflammasome, and then induce the production of cytokines such as interleukin 1 β (IL-1 β), tumor necrosis factor (TNF α), and reactive oxygen species (ROS) system (Cannizzo, Clement, Sahu, Follo, & Santambrogio, 2011; Green, Galluzzi, & Kroemer, 2011; Salminen et al., 2012). Inflammatory mediators, particularly TNF α , are potent stimulants of proteolysis through the ubiquitin-proteasome-dependent system. A significant negative relationship between myosin heavy chain protein synthesis rates and circulating markers of immune response has been observed (Toth, Ades, Tischler, Tracy, & LeWinter, 2006). Visser et al. (2002) demonstrated that for each increase in standard deviation in the TNF α value, a 1.2–1.3 kg reduction is seen in hand grip strength. Reactive oxygen species (ROS) appear to function as second messengers for TNFα in skeletal muscles increased mitochondrial apoptotic susceptibility, and reduced mitochondrial biogenesis (Chabi et al., 2008). The mitochondria constitute the major source of ROS, including superoxide radicals and hydrogen peroxide, which can cause oxidative damage to surrounding structures; particularly vulnerable is the mitochondrial DNA (mtDNA), which is in close proximity to the primary site of ROS production. Oxidation by ROS results in the synthesis of faulty proteins, oxidized lipids, and mtDNA mutations, which may lead to cellular and mitochondrial dysfunction as well as may accelerate apoptosis muscle stem cells, called satellite cells (SCs; Peterson, Johannsen, & Ravussin, 2012).

During the past decade, skeletal muscles have been identified as a secretory organ that produces various molecules, such as insulin growth factor I (IGF-I) and platelet-derived growth factors BB (PDGF^{BB}; Pedersen & Febbraio, 2012). The growth factors play an important role in muscle regeneration due to their ability to stimulate the activation, proliferation, and differentiation of SCs, and also synthesize muscle proteins. The reduction of the number of SCs is associated with the loss muscle mass and strength (Pallafacchina, Blaauw, & Schiaffino, 2013). The level of factors affecting satellite cells activity and muscle function in older adults is unknown. The purpose of the study was threefold: (a) to identify the agerelated changes in skeletal muscle properties responsible for deficits in functional status, (b) to establish the level of inflammation in young and older adults, and (c) to explain whether the changes in body composition and the decline of muscle strength are related to inflammatory mediators.

Table I.	Anthropometric and Body Composition Data in	
Young and	Older Men (Mean \pm SD).	

	Young $n = 20$	Elders n = 33	ANOVA HSD Tukey
Age, years	21.2 ± 1.3	73.5 ± 6.3	p < .001
Height, cm	181.8 ± 7.6	169.3 \pm 6.0	р < .001
Weight, kg	74.0 ± 6.1	76.3 \pm 10.3	p > .05
BMI, kg/m ²	22.5 ± 2.4	26.6 ± 3.1	p < .01
FFM, kg	60.0 ± 4.7	53.8 ± 7.1	p < .001
FFMI, kg/m ²	18.2 ± 1.9	18.7 ± 2.0	p > .05
FM, kg	14.0 ± 2.6	22.5 ± 5.5	p < .001
%FM	18.9 ± 2.7	$\textbf{29.3}\pm\textbf{5.1}$	p < .001
FMI, kg/m ²	4.3 ± 0.8	7.8 ± 1.9	р < .001

Note. ANOVA = one-way analysis of variance; BMI = body massindex; FFM = fat-free mass; FFMI = fat-free mass index; FM = fat mass; FMI = fat mass index.

Methods

Subjects

Thirty-three healthy males between 60 and 88 years old and twenty males between 20 and 24 years old participated in the study (Table 1). The oldest subjects were recruited from the University of the Third Age (U3A) Wroclaw and did not report absolutely any chronic diseases, that is, they represented successful aging (Geard, Reaburn, Rebar, & Dionigi, 2016). Inclusion criteria were: age 60-90 years for older men and age 20-30 years for young men, signed informed consent. Exclusion criteria were: acute infectious disease and chronic systemic diseases (cardiovascular disease, chronic diseases of liver and kidneys, diabetes mellitus, oncologic disease or other serious disease, based on the assessment of the responsible physician, and investigator) and participation in professional sport activities. The current health status and lifestyle of the subjects were estimated by Doctor of Medicine by using of the health history questionnaire (Aåstrand, 1956; Dursine & Moore, 2003).

All subjects were informed of the aim of the study and gave their written consent for participation in the project. The Bioethics Commission at Regional Medical Chamber Zielona Gora, Poland (No. 4/59/2015), in accordance with the Helsinki Declaration, approved the protocol of the study.

Body Composition

Body mass (BM) and body composition (fat-free mass [FFM] and fat mass [FM]) were estimated by a bioelectrical impedance (BIA) method using Tanita Body Composition Analyser MC-980 (Japan) calibrated prior to each test session in accordance to the manufacturer's guidelines. Duplicate measures were taken with the participant in a

standing position; the average value was used for the final analysis. The recurrence of measurement was 98%. The measurements were taken between 7:00 a.m. and 9:00 a.m., before blood sampling. FFM and FM indexes were calculated according to the definition by VanItallie, Yang, Heymsfield, Funk, and Boileau (1990): FFMI = FFM (kg)/height (m²) and FMI = FM (kg)/height (m²). Note that, mathematically, BMI (kg/m²) = FFMI + FMI. Thus, measured FFMI, FMI, and %FM values falling below the values for a BMI of 18.5 kg/m² were defined as low; measured FFMI, FMI, and %FM values falling in the range for BMI between 18.5 kg/m² and 25.0 kg/m² were considered normal, and values above that range were considered high (Bahadori et al., 2006).

Functional Fitness

The 6-min walking test is part of the Senior Fitness Test Protocol (Rikli & Jones, 2013), and is designed to test the functional fitness of older adults. The walking course is laid out in a 50-m rectangular area (dimensions $20 \text{ m} \times 5 \text{ m}$), with cones placed at regular intervals to indicate distance walked. The aim of the test is to walk as quickly as possible for 6 min covering as much ground as possible. Subjects set their own pace (a preliminary trial is useful to practice pacing), and are able to stop for a rest if they desire. Physical activity level, age, and gender were predictive factors for the result of the 6-min walking test (6-min walking distance [6MWD]) of older adults according to the equation by Enright and Sherrill (1998) for males:

$$6MWD = (74.31 \times height_{cm})$$
$$-(5.02 \times age) - (1.76 \times weight_{kg}) - 309$$

Isokinetic Testing

Peak torque (peak TQ) in the knee joint was assessed using the isokinetic digital dynamometer, Multi Joint 3 Biodex System; both lower limbs were tested. Each time the patient was informed how to perform the task. The test relies on flexion and extension of legs at the knee joint with maximal force and measures maximal flexion and extension peak torque for two arc speeds: 60°/s and 180°/s (five movements for each speed and 1-min rest between each set). According to Hoffman (2009), peak TQ has been previously used as an indicator of muscle function.

Blood Sampling

Blood samples were taken from the median cubital vein between 7.00 a.m. and 9.00 a.m. using S-Monovette-EDTA

tubes (Sarstedt, Austria). Within 20 min, they were centrifuged at $1,000 \times g$ and +4 °C for 10 min. Aliquots of plasma were stored at -80 °C.

Inflammatory Markers

The serum hydrogen peroxide (H₂O₂) concentration was determined in duplicate by the colorimetric method using the Oxis Research kit (USA). H₂O₂ detection limit was 6.25 µmol/L. The intra-assay coefficient of variation (CV) for the H₂O₂ kit was <10%. Serum interleukin-1 β (IL-1 β) and tumor necrosis factor α (TNF α) levels were determined in duplicate by enzyme immunoassay methods using Commercial kits R&D Systems (USA). Detection limits for IL-1 β and TNF α were 0.023 pg/ml and 0.038 pg/ml, respectively, and CV for both cytokines was <8.0%. C-reactive protein (hsCRP) concentration was determined in duplicate by DRG ELISA kit (USA). Detection limit was 0.001 mg/L, and CV for the hsCRP kit was <3%.

Growth Factors

Serum insulin-like growth factor (IGF-I) and muscle isoform of platelet-derived growth factor (PDGF^{BB}) were evaluated in duplicate by R&D Systems ELISA kits (USA). Detection limits were 0.026 ng/ml and 20 pg/ml, respectively. The CV for the growth factors kits was <5%.

Statistical Analysis

Statistical calculations were performed using the statistical software Statistica 13.1 (StatSoft Inc., Tulsa, OK, USA). All data were tested for distribution normality using the Shapiro–Wilk test. The values of W for inflammatory markers and growth factors were closed to the one-value; therefore statistical significances were assessed using one-way analysis of variance (ANOVA) and post-hoc test (HSD Tukey). Associations among measured parameters were analyzed using Pearson's linear regression (r coefficient). Statistical significance was set at p < .05. Results are expressed as mean and standard deviation ($x \pm SD$).

Results

The study comprised 53 healthy older and younger men. Over 90% of the elderly were 65 years or older. The generation of inflammatory mediators was related to changes in body composition, maximum strength muscle, and age-related changes in skeletal muscle properties responsible for deficits in functional fitness.

Table 2. Results of Peak Torque (Peak TQ), Peak TQ/BW, Total Work, Average Power, and Agonist/Antagonist Ratio in the Knee Joint at 60°/s and 180°/s (Mean \pm SD).

Young				Elders						
	Peak TQ, Nm	Peak TQ/ BW, %	Total work, J	Average power, W	Agonist/antagonist ratio, %	Peak TQ, Nm	Peak TQ/ BW, %	Total work, J	Average power, W	Agonist/antagonist ratio, %
60°/s								60°/s		
ER	256 ± 52	312 ± 36	1032 ± 233	160 ± 35	64 ± 7	116 ± 27*	$152 \pm 41*$	$513 \pm 81*$	$63 \pm 21*$	56 ± 17
EL	239 ± 44	292 ± 38	$\textbf{933}\pm\textbf{210}$	147 ± 29	64 ± 7	$115 \pm 36^*$	149 ± 44*	530 ± 199*	$65 \pm 25^*$	54 ± 13
FR	162 ± 39	199 ± 24	791 ± 177	117 ± 23	_	$64 \pm 25^*$	83 ± 29*	295 \pm 136*	$35 \pm 18^{*}$	_
FL	152 ± 26	187 ± 32	721 ± 149	108 ± 21	_	62 ± 22*	82 ± 28*	294 ± 112*	$36 \pm 16^*$	_
180°/s								180°/s		
ER	140 ± 28	117 ± 19	1849 ± 415	172 ± 40	83 ± 9	70 ± 20*	92 ± 27*	1152 ± 507*	88 ± 33*	68 ± 19
EL	138 ± 26	169 ± 21	1819 ± 379	172 ± 30	67 ± 21	69 ± 21*	89 ± 21*	1178 ± 524*	88 ± 34*	67 ± 21
FR	118 ± 26	141 ± 16	1762 ± 356	156 ± 27	-	$46 \pm 16^{*}$	60 ± 17*	662 ± 285*	49 ± 25*	_
FL	112 ± 18	137 ± 17	1741 ± 344	156 ± 29	_	44 ± 16*	57 ± 20*	644 ± 287*	47 ± 23*	_

Note. *p < .05 indicates statistically significant differences between younger and older men. ER = extensors of right knee; EL = extensors of left knee; FR = flexors of right knee; FL = flexors of left knee.

Body Composition

Among the older adults, BMI ranged from 19.9 to 32.7 (Table 1). About 12% of investigated seniors were classified as obese and 61% as overweight, 27% had normal weight. In the younger group, 85% of the subjects had a normal BMI. Normal FFMIs were from 16.0 kg/m² to 20.4 kg/m² in the younger males, and from 12.4 kg/m² to 23.0 kg/m² in the older males. The FMI values were 3.1 kg/m² to 5.7 kg/m² in the young males, and from 3.9 kg/m² to 11.3 kg/m² in the older males for the normal BMI ranges. FMI was significantly related to high concentrations of inflammatory markers, that is, IL-1 β (r = .476, p < .001), TNF α (r = .604, p < .001), and hsCRP (r = .710, p < .001). This indicates that a main source of inflammatory molecules may be elevated fat content.

Functional Fitness

The result of the 6MWD equation was 199 ± 49 , which shows good functional status in seniors from U3A. The older men classified as obese and overweight (FMI >8 kg/m²) were distinguished by low functional status (6MWD <150).

Isokinetic Testing

The analysis of mean values for parameters showed that muscle functionality is, on average, 50% lower in the elderly (Table 2). Knee extension and flexion peak TQ, peak TQ/BW (peak torque/body weight), total work and agonist/antagonist ratio measurements obtained at 60°/s and 180°/s in older men were significantly different from young males. The older men with high fat content demonstrated low values of knee extension peaks TQ at 60°/s and 180°/s.

Table 3. Serum Hydrogen Peroxide H₂O₂, Cytokines IL-1 β and TNF α , hsCRP, as well as IGF-I and PDGF^{BB} in Younger and Older Men (Mean \pm SD)

	Young $N = 20$	Elders N = 33	ANOVA HSD Tukey
H ₂ O ₂ , μmol/L IL-1β, pg/ml TNFα, pg/ml hsCRP, mg/L IGF-1, ng/ml PDGF ^{BB} , pg/ml	$\begin{array}{c} 11.53 \pm 3.87 \\ 0.58 \pm 0.17 \\ 0.85 \pm 0.13 \\ 0.12 \pm 0.09 \\ 117 \pm 36 \\ 2162 \pm 551 \end{array}$	$\begin{array}{c} 20.35 \pm 8.16 \\ 1.62 \pm 0.62 \\ 4.74 \pm 1.46 \\ 0.46 \pm 0.07 \\ 64 \pm 22 \\ 954 \pm 294 \end{array}$	p < .01 p < .02 p < .001 p < .001 p < .001 p < .001 p < .001

Note. hsCRP = C-reactive protein; IGF-I = insulin growth factor; PDGF = platelet-derived growth factor.

Inflammatory Markers

There were significant age-related increases in inflammatory molecules (Table 3). H_2O_2 and IL-1 β levels were twofold higher while TNF α level was sixfold higher in the elderly. hsCRP was fourfold higher in older men compared to younger men. All observed inflammatory markers inversely correlated with the knee extension peak TQ at 60°/s and 180°/s (Table 4). This means that inflammation may be a serious reason for impairment of muscles function.

Growth Factors

The growth factors related to muscle regeneration were significantly different in older men compared to younger men (Table 3). IGF-I and PDGF^{BB} levels were reduced twofold in the elderly and inversely correlated with age (for IGF-I r = -0.690, and for PDGF^{BB} r = -0.812; p < .001). The changes in IGF-I and PDGF^{BB} concentrations were also related to the knee extension peak TQ at 60°/s

	H ₂ O ₂ , µmol/L	IL-Iβ, pg/ml	TNFα, pg/ml	hsCRP, mg/L	IGF-I, ng/ml	PDGF ^{BB} , pg/ml
Peak TQ/ER at 60°/s, Nm	539	654	717	852	.713	.785
Peak TQ/ER at 180°/s, Nm	489	637	704	799	.656	.700

Table 4. Relationships (Correlation Coefficients; p < .001) Between Peak TQ, Inflammatory Markers H_2O_2 , IL-1 β , TNF α , and hsCRP as well as IGF-1 and PDGF^{BB}

Note. ER = extensors of right knee; IGF-I = insulin growth factor; PDGF = platelet-derived growth factor.

Table 5. Relationships (Correlation Coefficients; p < .001) Between Pro-Inflammatory Mediators (IL-1 β , TNF α , and hsCRP) and IGF-I and PDGF^{BB}

	IGF-I, ng/ml	PDGF ^{BB} , pg/ml
IL-Iβ, pg/ml	582	550
TNFα, pg/ml	544	683
hsCRP, mg/L	664	778

Note. IGF-I = insulin growth factor; PDGF = platelet-derived growth factor.

and 180°/s (Table 4). Levels of IGF-I and PDGF^{BB} were significantly reduced by high levels of IL-1 β , TNF α and hsCRP (Table 5).

Discussion

Age-related muscle changes are characterized by a gradual loss of spinal motor neurons due to apoptosis, reduced growth factors signaling and protein uptake, elevated amounts of circulating cytokines and ROS generation and so forth. Some denervated muscle fibers are reinnervated through collateral sprouting of nearby surviving motor axons or motor end plates, which results in the formation of enlarged motor units. Consequently, the age-related loss of spinal motor neurons leads to a decline in muscle fiber number and size, resulting in impaired mechanical muscle performance (reduced maximal muscle strength, power, and rate of force development) that translates into a reduced functional capacity during everyday tasks (Aagaard et al., 2001; Zembron-Lacny et al., 2014). These age-related changes in skeletal muscle system are the result of chronic activation of macrophages, which leads to an increase in pro-inflammatory cytokines (Franceschi & Campisi, 2014). The inflammatory mediators such as H_2O_2 , IL-1 β , and TNF α are induced by various stimuli such as bacteria, viruses, and tissue damages (Cannizzo et al., 2011). The increase in pro-inflammatory factors is far less than that seen in acute infection; thus the ageing effects on pro-inflammatory cytokine expression are considered to be a chronic low-grade state (Hansen, Baptiste, Fjeldborg, & Horohov, 2015).

In the study, the levels of H_2O_2 , IL-1 β , TNF α , and hsCRP were several times higher in the elderly, which underlie the low-grade inflammatory status. Meng et al.

(2015) demonstrated that the circulating systemic inflammatory markers are associated with less muscle mass, lower muscle strength, slower walking speed, poorer balance, and lower self-reported functional ability.

The high concentration of hsCRP strongly reduces muscle function and growth factors levels in older men (Tables 4 and 5). While the exact biological actions of hsCRP are not established, its levels predict risk of mobility/disability and accelerated decline in muscle strength and physical performance in older adults (Meng et al., 2015; Verghese, Holtzer, Lipton, & Wang, 2012). The observed fourfold increase in hsCRP concentration in older men proves the presence of low-grade inflammation during ageing. Inflammaging is considered a predictor of fragility and this condition is currently accepted as a pathogenic factor in the development of several agerelated diseases and increased mortality risk. The precise etiology of inflammaging and its potential causal role in contributing to adverse health outcomes remain largely unknown. The identification of pathways that control age-related inflammation across multiple systems is therefore important in order to understand whether treatments that modulate inflammaging may be beneficial in the elderly population (Franceschi & Campisi, 2014; Lohr et al., 2014).

One of the possibility catabolic actions of inflammatory mediators on skeletal muscle is the inhibition of protein synthesis and myogenesis in myoblasts. Earlier studies demonstrated that after IL-1 β stimulation, the total protein level did not increase, but rather synthesis of the acute phase proteins was favored (Weissman, 1990). Studies indicate that TNF α can interfere with muscle growth and regeneration in part by disrupting growth factors signaling pathways (Strle et al., 2004). TNF α also is responsible for triggering the death receptor-mediated apoptosis, which plays a significant role in atrophy of muscle fibers, especially type II fibers (fast), by a decreased number of motor units (Marzetti et al., 2010).

The cytokines IL-1 β and TNF α are involved, not only in muscle mass decrease, but also in increase in fat mass. At the extreme, these two processes lead to a condition known as "sarcopenic obesity." Studies have highlighted that pro-inflammatory cytokines produced by adipose tissue accelerate muscle catabolism, and thus contribute to the vicious circle that initiates and sustains sarcopenia (Schrager et al., 2007). The study confirmed that fat tissue can be source of inflammatory mediators, which enhance age-related muscle loss. Older men with high FMI demonstrated high concentrations of IL-1 β and TNF α .

The increased level of pro-inflammatory cytokines negatively impacted concentrations of investigated growth factors IGF-I and PDGF^{BB}, which play a central role in myofiber hypertrophy and atrophy, and this balance is of critical importance for muscle wasting in ageing. Grounds, Radley, Gebski, and Shavlakadze (2008) suggested that the effects of IL-1 β and TNF α on muscle atrophy may be mediated in part via interference with IGF-I signaling and inhibition of the anabolic signaling cascade downstream of the IGF-I receptor. The long-term increased concentrations of TNF α and IL-1β may induce muscle resistance to IGF-I (O'Connor et al., 2008). Additionally, TNF α may abolish antiapoptotic effects of IGF-I, reducing the survival of differentiated myoblasts (Grounds et al., 2008). Lohr et al. (2014) demonstrated in large size of the sample that increased inflammatory markers, such as hsCRP with low IGF-I may even increase mortality in young and older men and women.

Among many of the stimulatory growth factors, PDGF^{BB} plays a crucial role in myogenic proliferation and differentiation (Pallafacchina et al., 2013). Despite the fact that knowledge about the level of circulating IGF-I in the elderly is well developed, in case of PDGF^{BB} it is quite enigmatic. In general, there are few data describing the influence of ageing on the level of circulating PDGF^{BB}. In addition to confirming that the level of circulating PDGF^{BB} is lower in the elderly, this study has indicated that both IL-1 β and TNF α have a negative influence on this molecule. There is only one dataset concerning the level of circulating PDGF^{BB} in the muscles of the elderly, especially its interaction with IL-1 β or TNF α (Banerjee et al., 2011; Bentzinger, von Maltzahn, & Rudnicki, 2010). Banerjee et al. (2011) have reported that the inflammatory processes can contribute to the PDGF^{BB} profile in the elderly. These results suggest that mechanisms responsible for negative correlations between investigated pro-inflammatory cytokines and PDGF^{BB} are similar to that for IGF-I.

IGF-I and PDGF^{BB} are important downstream mediators of the anabolic effects of growth hormone and their serum levels are inversely correlated with age. Kaplan et al. (2008) demonstrated that the total IGF-I level is associated with strength, mobility, and mortality. Although there is evidence that ageing muscle retains the ability to synthesize IGF-I, ageing may also be associated with attenuation of the ability of exercise to induce an isoform of IGF-I that promotes satellite cell proliferation (Owino, Yang, & Goldspink, 2001). These results indicate an age-related decrease in systemic derived growth factors, which may be responsible, at least in part, for the age-related decline in muscle function. A large number of studies have suggested the implications of cross-talk between pro-inflammatory cytokines and growth factors in skeletal muscle, which is likely the underlying mechanism of sarcopenia (Meng et al., 2015).

The age-related shifts in body composition toward more fat mass, especially the accumulation of more internalized fat storage, and the loss of muscle mass and function increase the risk of injury from sudden falls and developing a wide range of chronic disorders. FFM and FM indexes have been useful for the clinical evaluation of a deficit in fat-free mass with or without excess fat mass for a given age category, complementing the classical concept of BMI in a more qualitative manner. In the present study, an increased fat storage specifically affected FMI but not FFMI in older men as compared to younger men, which corresponds well with studies done by Bahadori et al. (2006) and VanItallie et al. (1990). The majority of seniors demonstrated very good results in the 6-min walking test which corresponds very well with studies done by Enright and Sherrill (1998). Their good functional status could be related to their participation in various physical and health educational forms at the University of the Third Age (U3A). According to Zielinska-Wieczorkowska, Kedziora-Kornatowska, and Ciemnoczolowski (2011), the high life quality of the U3A students significantly denotes the level of their knowledge concerning illnesses, afflictions, depression, and the health benefits of physical activity.

Conclusions

In the current study, the strong dominance of inflammatory mediators H_2O_2 , IL-1 β , TNF α , and hsCRP over the anabolic factors IGF-1 and PDGF^{BB} in the older men were observed, whereas in young men the reverse situation was detected. The enhancement of pro-inflammatory state with age was responsible for deficits in maximum strength muscle and functional capacity. However, it is too early to draw a clear conclusion on a clinically relevant relationship between inflammation and skeletal muscle properies due to the small sample size.

Declaration of Conflicting Interests

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References

- Aagaard, P., Andersen, J. L., Dyhre-Poulsen, P., Leffers, A.-M., Wagner, A., Magnusson, S. P., ... Simonsen, E. B. (2001). A mechanism for increased contractile strength of human pennate muscle in response to strength training: Changes in muscle architecture. *The Journal of Physiology*, *534*(2), 613–623.
- Aåstrand, P.-O. (1956). Human physical fitness with special reference to sex and age. *Physiological Review*, 36(3), 307–335.
- Bahadori, B., Uitz, E., Tonninger-Bahadori, K., Pestemer-Lach, I., Trummer, M., Thonhofer, R., ... Schaflinger, E. (2006).
 Body composition: The Fat-Free Mass Index (FFMI) and the Body Fat Mass Index (BFMI) distribution among the adult Austrian population – Results of a cross-sectional pilot study. *International Journal of Body Composition Research*, 4, 123–128.
- Banerjee, C., Ulloor, J., Dillon, E. L., Dahodwala, Q., Franklin, B., Storer, T., & Montano, M. (2011). Identification of serum biomarkers for aging and anabolic response. *Immunity & Ageing*, 8(1), 5. doi:10.1186/1742-4933-8-5
- Bentzinger, C. F., von Maltzahn, J., & Rudnicki, M. A. (2010). Extrinsic regulation of satellite cell specification. *Stem Cell Research & Therapy*, 1(3), 27.
- Cannizzo, E. S., Clement, C. C., Sahu, R., Follo, C., & Santambrogio, L. (2011). Oxidative stress, inflamm-aging and immunosenescence. *Journal of Proteomics*, 74(11), 2313–2323.
- Chabi, B., Ljubicic, V., Menzies, K. J., Huang, J. H., Saleem, A., & Hood, D. A. (2008). Mitochondrial function and apoptotic susceptibility in aging skeletal muscle. *Aging Cell*, 7(1), 2–12.
- Deeks, S. G. (2011). HIV infection, inflammation, immunosenescence, and aging. *Annual Review of Medicine*, 62, 141– 155.
- Enright, P. L., & Sherrill, D. L. (1998). Reference equations for the six-minute walk in healthy adults. *American Journal of Respiratory and Critical Care Medicine*, 158(5), 1384–1387.
- Franceschi, C., & Campisi, J. (2014). Chronic inflammation (inflammaging) and its potential contribution to age-associated diseases. *The Journals of Gerontology. Series A, Biological Sciences and Medical Sciences*, 69(Suppl 1), S4–S9.
- Geard, D., Reaburn, P., Rebar, A., & Dionigi, R. (2016). Masters athletes: Exemplars of successful aging? *Journal of Aging and Physical Activity*, 25(3), 490–500. doi:10.1123/japa.2016-0050
- Green, D. R., Galluzzi, L., & Kroemer, G. (2011). Mitochondria and the autophagy-inflammation-cell death axis in organismal aging. *Science*, 333(6046), 1109–1112.
- Grounds, M. D., Radley, H. G., Gebski, B. L., Bogoyevitch, M. A., & Shavlakadze, T. (2008). Implications of crosstalk between tumour necrosis factor and insulin-like growth factor-1 signalling in skeletal muscle. *Clinical* and Experimental Pharmacology & Physiology, 35(7), 846–851.
- Hansen, S., Baptiste, K. E., Fjeldborg, J., & Horohov, D. W. (2015). A review of the equine age-related changes in the

immune system: Comparisons between human and equine aging, with focus on lung-specific immune-aging. *Ageing Research Reviews*, 20, 11–23.

- Hoffman, J. (2009). *Norms for fitness, performance, and health.* Champaign, IL: Human Kinetics.
- Kaplan, R. C., McGinn, A. P., Pollak, M. N., Kuller, L., Strickler, H. D., Rohan, T. E., & Psaty, B. M. (2008). Total insulin-like growth factor 1 and insulin-like growth factor binding protein levels, functional status, and mortality in older adults. *Journal of the American Geriatrics Society*, 56(4), 652–660.
- Lohr, J., Grotevendts, A., Nauck, M., Völzke, H., Wallaschofski, H., & Friedrich, N. (2014). Relation of insulin-like growth factor-I and IGF binding protein 3 with markers of inflammation: Results of a population-based study. *Clinical Endocrinology*, 80, 148–154.
- Marzetti, E., Privitera, G., Simili, V., Wohlgemuth, S. E., Aulisa, L., Pahor, M., & Leeuwenburgh, C. (2010). Multiple pathways to the same end: Mechanisms of myonuclear apoptosis in sarcopenia of aging. The *Scientific World Journal*, 10, 340–349.
- Meng, Y., Wu, H., Yang, Y., Du, H., Xia, Y., Guo, X., & Niu, K. (2015). Relationship of anabolic and catabolic biomarkers with muscle strength and physical performance in older adults: A population-based cross-sectional study. *BMC Musculoskeletal Disorders*, 16, doi:10.1186/s12891-015-0654-7
- O'Connor, J. C., McCusker, R. H., Strle, K., Johnson, R. W., Dantzer, R., & Kelley, K. W. (2008). Regulation of IGF-I function by proinflammatory cytokines: At the interface of immunology and endocrinology. *Cell Immunology*, 252(1– 2), 91–110.
- Owino, V., Yang, S. Y., & Goldspink, G. (2001). Agerelated loss of skeletal muscle function and the inability to express the autocrine form of insulin-like growth factor-1 (MGF) in response to mechanical overload. *Federation of European Biochemical Societies Letters*, 505(2), 259–263.
- Pallafacchina, G., Blaauw, B., & Schiaffino, S. (2013). Role of satellite cells in muscle growth and maintenance of muscle mass. *Nutrition, Metabolism and Cardiovascular Diseases*, 23, S12–S18.
- Pedersen, B. K., & Febbraio, M. A. (2012). Muscles, exercise and obesity: Skeletal muscle as a secretory organ. *Nature Reviews Endocrinology*, 8(8), 457–465.
- Peterson, C. M., Johannsen, D. L., & Ravussin, E. (2012). Skeletal muscle mitochondria and aging: A review. *Journal of Aging Research*, 2012, 1–20. doi:10.1155/2012/ 194821
- Rikli, R. E., & Jones, C. J. (2013). Development and validation of criterion-referenced clinically relevant fitness standards for maintaining physical independence in later years. *The Gerontologist*, 53(2), 255–267.
- Salminen, A., Kaarniranta, K., & Kauppinen, A. (2012). Inflammaging: Disturbed interplay between autophagy and inflammasomes. *Aging*, 4(3), 166–175.
- Schrager, M. A., Metter, E. J., Simonsic, E., Ble, A., Bandinelli, S., Lauretani, F., & Ferrucci, L. (2007). Sarcopenic obe-

sity and inflammation in the InCHIANTI study. *Journal of Applied Physiology*, *102*(3), 919–925.

- Strle, K., Broussard, S. R., McCusker, R. H., Shen, W. H., Johnson, R. W., Freund, G. G., ... Kelley, K. W. (2004). Proinflammatory cytokine impairment of insulin-like growth factor-I induced protein synthesis in skeletal muscle myoblasts requires ceramide. *Endocrinology*, 145(10), 4592–4602.
- Toth, M. J., Ades, P. A., Tischler, M. D., Tracy, R. P., & LeWinter, M. M. (2006). Immune activation is associated with reduced skeletal muscle mass and physical function in chronic heart failure. *International Journal of Cardiology*, 109(2), 179–187.
- VanItallie, T. B., Yang, M. U., Heymsfield, S. B., Funk, R. C., & Boileau, R. A. (1990). Height-normalized indices of the body's fat-free mass and fat mass: Potentially useful indicators of nutritional status. *The American Journal of Clinical Nutrition*, 52(6), 953–959.
- Verghese, J., Holtzer, R., Lipton, R. B., Wang, C. (2012). Highsensitivity C-reactive protein and mobility disability in older adults. *Age and Ageing*, 41(4), 541–545.

- Visser, M., Pahor, M., Taaffe, D. R., Goodpaster, B. H., Simonsick, E. M., Newman, A. B., & Harris, T. B. (2002). Relationship of interleukin-6 and tumor necrosis factoralpha with muscle mass and muscle strength in elderly men and women: The health ABC study. *The Journals of Gerontology. Series A, Biological Sciences and Medical Sciences*, 57(5), M326–M332.
- Weissman, C. (1990). The metabolic response to stress: An overview and update. *Anesthesiology*, 73(2), 308–327.
- Zembron-Lacny, A., Dziubek, W., Rogowski, L., Skorupka, E., & Dąbrowska, G. (2014). Sarcopenia: Monitoring, molecular mechanisms, and physical intervention. *Physiological Research*, 63(6), 683–691.
- Zielinska-Wieczorkowska, H., Kedziora-Kornatowska, K., & Ciemnoczolowski, W. (2011). Evaluation of Quality of Life (QoL) of students of the University of Third Age (U3A) on the basis of socio-demographic factors and health status. *Archives of Gerontology and Geriatrics*, 53(2), e198–e202.