# Network of small no-take marine reserves reveals greater abundance and body size of fisheries target species 

Fernanda A. Rolim ${ }^{1,2 \odot}{ }^{1,2}$, Tim Langlois ${ }^{3 థ}$, Pedro F. C. Rodrigues ${ }^{2 \ddagger}$, Todd Bond ${ }^{3 \ddagger}$, Fábio S. Motta ${ }^{\ddagger \ddagger}$, Leonardo M. Neves ${ }^{5 \ddagger}$, Otto B. F. Gadig ${ }^{2 ®}$<br>1 Instituto de Biociências, Universidade Estadual Paulista (UNESP), Campus de Rio Claro, Rio Claro, São Paulo, Brazil, 2 Laboratório de Pesquisa em Elasmobrânquios, Instituto de Biociências, Universidade Estadual Paulista (UNESP), Campus do Litoral Paulista, São Vicente, São Paulo, Brazil, 3 The UWA Oceans Institute and School of Biological Sciences, The University of Western Australia (UWA), Perth, Western Australia, Australia, 4 Laboratório de Ecologia e Conservação Marinha, Instituto do Mar, Universidade Federal de São Paulo (UNIFESP), Santos, São Paulo, Brazil, 5 Departamento de Ciências do Meio Ambiente, Universidade Federal Rural do Rio de Janeiro (UFRRJ), Campus Três Rios, Rio de Janeiro, Brazil<br>- These authors contributed equally to this work.<br>$\ddagger$ These authors also contributed equally to this work.<br>* fernandarolim2 @ gmail.com


#### Abstract

No-take marine reserves (NTRs), i.e. areas with total fishing restrictions, have been established worldwide aiming to promote biodiversity and ecosystem conservation. Brazil has $3.3 \%$ of its exclusive economic zone protected by 73 different NTRs, however, most of them currently lack scientific knowledge and understanding of their ecological role, particularly regarding rocky reefs in subtropical regions. In this context, this study aimed to contrast a network of NTRs with comparable fished sites across a coastal biogeographic gradient to investigate the effect of fishing and habitat variability on the abundance and body size of rocky reef fish. We used Baited Remote Underwater stereo-Video (stereo-BRUVs) and Diver Operated stereo-Video (stereo-DOVs) systems to simultaneously sample reef fish and habitat. Model selection and results identified habitat and biogeographic variables, such as distance from shore, as important predictor variables, explaining several aspects of the fish assemblage. The effect of protection was important in determining the abundance and body size of targeted species, in particular for epinephelids and carangids. Conversely, species richness was correlated with habitat complexity but not with protection status. This is the first study using these survey methods in the Southwestern Atlantic, demonstrating how a network of NTRs can provide benchmarks for biodiversity conservation and fisheries management.


## Introduction

No-take marine reserves (NTRs) have been established worldwide as an important management strategy, mostly aiming to protect marine biodiversity from the effects of fishing and
study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing interests: The authors have declared that no competing interests exist.
other human disturbances [1,2]. It is well documented that these NTRs can provide refuge to marine life, increasing local abundance, species richness, body size and the reproductive capacity of fish [3-6]. Networks of NTRs can be used to investigate effects of fishing across biogeographic gradients, with the aim of estimating benchmarks for conservation and fisheries management. Increased biomass of target species has been recorded inside NTRs, contrasting with open areas where the removal of large carnivores can result in higher abundance of prey species, leading to a trophic reorganization. [7-9].

Extensive research has documented that fish assemblage structure varies with physical, chemical and biological factors across biogeographic and habitat gradients [10-12]. In particular, distance from the coast and topographic complexity have shown increase of species richness, abundance and biomass of reef fish [13-19]. It is therefore important for any investigation of the effects of fishing to control for covariates across NTRs and open areas.

Brazil has 8500 km of coastline and a territorial sea that, together with the Exclusive Economic Zone, encompasses 4 million $\mathrm{km}^{2}$. Of this area, $26.4 \%$ is currently protected by 177 marine protected areas (MPAs), of which 73 are NTRs, representing $3.3 \%$ of the country's marine waters [20]. However, the majority of this protection is in large and remote offshore areas, with only $0.3 \%$ of these NTRs occurring in small to medium-sized protected areas (1$100 \mathrm{~km}^{2}$ ) in coastal waters [20]. The effectiveness of these remote NTRs in terms of achieving conservation objectives has been questioned due to the difficulties of enforcement and monitoring of offshore waters [21,22]. Despite the relatively small sizes of these coastal networks of NTRs, they have potentially high ecological and social value given the greater human impacts occurring in these coastal waters [13,23,24].

Coastal habitats along the northern coast of Brazil (north of $19^{\circ} \mathrm{S}$ ) are dominated by coral reefs, whereas southern regions (between $19-28^{\circ}$ ) are typified by rocky reefs. In general, the Brazilian province shelters a high number of endemic species and biomass of marine organisms [25-28]. In the transition zone between tropical and subtropical-temperate environments $\left(20^{\circ} \mathrm{S}\right.$ to $\left.23^{\circ} \mathrm{S}\right)$, the mosaic of habitat types results in one of the highest species diversity of benthic [29] and reef fish species recorded in Brazil [25,28,30]. These transitional reefs are biologically rich and complex environments, where it is vitally important to establish, enforce and understand the benefits of NTRs. However, the few studies available about the effects of Brazilian NTRs on fish assemblage are concentrated in the northern [31] and southern region [32,33] of the country's coastline, or in offshore islands [34,35], with a lack of studies in the transition zones between tropical and subtropical realms of coastal NTR networks.

Historically, NTRs and reef ecosystems in the Southwestern Atlantic have been assessed using underwater visual census (UVC) (e.g. [31,32,34,35]). Despite the benefits of UVCs, such as being a rapid and effective tool in providing precise data especially about conspicuous and sedentary fish species [36-38], biases involving interobserver variability, underrepresentation of large and mobile species targeted by fisheries, as well as inaccuracy of abundance and size estimates can occur [39-41]. In order to mitigate some of these issues and complement fish assemblage assessments, the use of video-based methods to collect data has been increasingly adopted; aided by rapid advancements in video technology and accessibility to cheaper and higher quality equipment [41,42]. Importantly, methods using such technologies create a permanent record allowing fish identification to be confirmed by experts and revisited when necessary.

Baited Remote Underwater stereo-Video (stereo-BRUV) and Diver Operated stereo-Video (stereo-DOV) are being widely employed to assess diverse aspects of fish assemblages [36,4346]. Stereo-video techniques provide accurate body size and range measurements of individuals from the three-dimensional calibration of imagery [47]. Stereo-BRUV have been found to sample a wide range of species without precluding estimates of herbivorous species [48] and
can be applied across a wide variety of habitats and depths [44]. Also, as a remote sensing technique, it detects large and mobile animals which usually avoid divers and active fishing gears [36,43], but has a range of acknowledged biases and limitations related to the presence of the bait and potential underrepresentation of small-bodied fish species (see Langlois et al. [49] and Goetze et al. [36]). Conversely, the presence of a diver may impact the abundance of fish recorded using stereo-DOVs $[36,50]$, suggesting that the combination of methods is more effective to sample fish assemblages [36,51].

In order to expand knowledge about the ichthyofauna of the Southwestern Atlantic, we applied novel non-destructive methods that complement the traditionally used visual sampling techniques, offering potentially more robust estimates of targeted species among protected and fished areas. The improvement of non-lethal and non-destructive techniques to assess fish assemblage is crucial, especially for sensitive habitats inside protected areas such as reef environments, which shelter a significant amount of endangered and endemic species [32]. Thus, this study is the first assessing fish assemblages using stereo-BRUVs and stereo-DOVs in the Southwestern Atlantic, and aims to contribute to the conservation and fisheries management in the region. Based on this, we aim to investigate the response of the fish assemblage to environmental and habitat variables, as well as the effect of protection among NTRs. We hypothesize that: (1) abundance and body size of targeted fish groups will be greater inside NTRs; whereas (2) non-target fish abundance and species richness will be explained better by habitat and biogeographic variables.

## Material and methods

This study was conducted in accordance with all Brazilian government legislation. This includes Federal Government authorization to observe and assess images within the Tupinambás Ecological Station under the permits \#48259-1, and also authorization from the São Paulo State government (Fundação Florestal), by the Comissão Técnico Científica-COTEC, to develop the research project.

## Study site

The Ecological Station (ESEC) of Tupinambás is a no-take marine reserve (NTR) (corresponding to IUCN Category Ia) located on the northern coast of São Paulo State, Brazil, Southwestern Atlantic. The ESEC was established in 1987 [52] and is divided into two sectors. Sector I is in the archipelago of Alcatrazes ( $24.101^{\circ} \mathrm{S} ; 45.692^{\circ} \mathrm{W}$ ), which is located approximately 43 km from of São Sebastião, São Paulo. This sector has six protected localities, each of them with 1 km of buffer area. Two sets of two of these sites are close enough to overlap, creating four primary areas of protection (Fig 1). Sector II protects Palmas Island ( $23.547^{\circ}$ S; $45.029^{\circ} \mathrm{W}$ ) including two nearby reefs (Palmas Reef and Forno Reef) and Cabras Island ( $23.517^{\circ} \mathrm{S}$; $45.041^{\circ} \mathrm{W}$ ), located 5.7 km and 3.6 km respectively from the coast of Ubatuba, São Paulo.

The open-fishing areas used to test the effects of protection on fish assemblage included Búzios ( $23.804^{\circ} \mathrm{S} ; 45.139^{\circ} \mathrm{W}$ ), Mar Virado ( $23.567^{\circ} \mathrm{S}$; $45.156^{\circ} \mathrm{W}$ ) and Tamanduá (23.597${ }^{\circ} \mathrm{S}$; $45.289^{\circ} \mathrm{W}$ ) islands. These islands are part of a multiple use MPA established in 2008 (Environmental Protection Area-corresponding to IUCN category V). They are located $34 \mathrm{~km}, 2 \mathrm{~km}$, and 0.5 km respectively from the mainland. Small scale fishing, such as angling, spearfishing, longlines, fixed traps and gillnetting, is permitted around Mar Virado and Tamanduá islands, but no industrial fishing that uses pair trawling, driftnet vessels above 20 gross tonnage (GT) or trawling vessels up to 10 GT is allowed. However, only pair trawlers are excluded from fishing in Búzios Island.

ONE


Fig 1. Map of the study area on the southeastern Brazilian coast with the no-take marine reserve Tupinambás Ecological Station in red. The control islands, where fishing activity is permitted (Tamanduá, Mar Virado and Búzios), are also displayed (A, B). No-take areas in detail in the islands of Cabras and Palmas (A) and in Alcatrazes Archipelago (C) with the sample sites represented by the black spots.
https://doi.org/10.1371/journal.pone.0204970.g001

## Sampling

Samples were collected using stereo-DOVs and stereo-BRUVs. Both equipment types are comprised of a metal base bar with two underwater SeaGIS housing (www.seagis.com.au), each with a digital video camera inside. Housings are positioned approximately 700 mm apart, each inwardly converged at 8 degrees. Stereo-BRUVs were deployed from a boat connected by a rope with a surface float, and left on the seafloor for 90 minutes to record fishes and habitat characteristics. The camera base bar was enclosed within a stainless steel frame, and a bait cage with 800 g of mashed sardine (Sardinella brasiliensis) was positioned at the end of a bait arm approximately 1.5 m away from the cameras. Oily bait provide greater sampling efficiency $[53,54]$ due to the odor plume dispersion. Stereo-DOVs used the same camera base bar setup,
with the addition of a handle, allowing divers to swim along a transect. These standard survey methods have been developed and used by many authors worldwide [43,44,55].

Stereo-BRUV and stereo-DOV sampling was undertaken bimonthly at each island for a year (2016-March, May, July, October, November; and 2017-January). Each expedition was approximately 4-6 days long, covering all six islands. Due to the small size of islands and to maintain independence among samples (minimum distance between replicates was at least 250 m [48]), only two stereo-BRUV samples were collected on the leeward side of the islands, totaling 12 stereo-BRUVs at each island after six expeditions. Each stereo-BRUV was deployed at the interface of the rocky reef with the sandy bottom. Water depth ranged from 2-17 m depending on the location of the rock-sand interface at each island and the average water depth sampled was $8.3 \pm 3.6 \mathrm{~m}$.

Stereo-DOV transects were 25 m long and 5 m wide, and swum at the interface of the rocky shore with the sandy bottom, as well as at the shallow zone above the reef. Due to the small size of the islands, sampling was restricted to three transects at the rock-sand interface and three in the shallow zone on each island at each expedition, totaling 36 transects per island at the end of six expeditions. Stereo-DOV transects were surveyed twice. During the first survey, the observer filmed conspicuous species in the water column; during the second survey, the observer focused on the substrate to detect cryptic species (families Blenniidae, Gobiidae, Labrisomidae and Chaenopsidae). The sampling unit therefore included the number and size of both conspicuous and cryptic fish species per transect. This protocol is comparable to that used for underwater visual census in the region to ensure that the species that are more likely to avoid divers are recorded first, whilst small cryptic species are also sampled [14,18,56]. In stereo-DOV samplings, the interface zone presented an average depth of $8.9 \pm 3.8 \mathrm{~m}$ and the shallow zone $4.2 \pm 1.9 \mathrm{~m}$.

## Video analysis

Fish assemblage. Stereo-video systems were calibrated using the CAL software and video analysis was carried out in the EventMeasure software (www.seagis.com.au). The description of the design and calibration of stereo-videos can be found in Harvey and Shortis [47,57]. Fish were identified to the finest taxonomic level possible, counted and measured if they were within 7 m of the stereo-BRUVs and 5 m for stereo-DOVs.

The relative abundance of each species filmed on stereo-BRUVs was recorded as MaxN, defined as the maximum number of individuals of the same species recorded in a single frame from the left camera. This is a conservative approach in order to avoid counting and measuring the same individual more than once. The fork length of individual fish contributing to a species' MaxN was measured when the fish was straight and no more than 45 degrees perpendicular to the cameras. In the stereo-DOV, all fish filmed on the left camera were counted and measured using the same rules. These data are stored on GlobalArchive [58] (globalarchive. org), under the project "Effectiveness of Marine Protected Areas, Brazil", and also available in the supporting information files.

Biomass was calculated for all species using measured fish lengths and length-weight relationship referenced in the FishBase database [59]. If equations for fork length of a species were not available, length-length conversions were used if available. Biomass of species without length-weight information was calculated using equations from a similar species from the same family.

Fish species were classified by broad functional groups based on diet, using information available in the literature [60,61] and FishBase [59]. Groups included: carnivores, piscivores, planktivores, roving herbivores, territorial herbivores, omnivores, sessile invertebrate feeders,

Table 1. Habitat classification based on broad CATAMI Classification scheme [68] and on Wilson et al. [69], used in Baited Remote Underwater stereo-Videos and Diver Operated stereo-Videos images.

| Criteria | Description |  |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: |
| Relief | 0-Flat substrate, sandy, rubble with few features. $\sim 0$ substrate slope |  |  |  |  |
|  | 1 -Some relief features amongst mostly flat substrate/sand/rubble. $<45$ degree substrate slope |  |  |  |  |
|  | 2-Mostly relief features amongst some flat substrate or rubble. $\sim 45$ substrate slope |  |  |  |  |
|  | 3-Good relief structure with some overhangs. $>45$ substrate slope |  |  |  |  |
|  | 4-High structural complexity, fissures and caves. Vertical wall. $\sim 90$ substrate slope |  |  |  |  |
|  | 5-Exceptional structural complexity, numerous large holes and caves. Vertical wall. $\sim 90$ substrate slope |  |  |  |  |
|  | Unknown |  |  |  |  |
| Field of view | Facing up | Limited |  |  |  |
|  | Facing down | Open |  |  |  |
| Broad/Benthos | Ascidians | Consolidated | Open water | Stony corals | Unknown |
|  | Bryozoa | Macroalgae | Sponges | Unconsolidated | Zoanthids |

https://doi.org/10.1371/journal.pone.0204970.t001
mobile invertebrate feeders. Piscivores were pooled with carnivores because there was not enough individuals for statistical analysis. Species were categorized in target and non-target for fisheries in the region according to the literature [18,62-64]. Four families (Epinephelidae, Kyphosidae, Scaridae and Carangidae) identified as abundant or frequent and also targeted by fisheries were selected for analysis.

Habitat characteristics. Habitat classification and complexity (mean relief) were analysed using a single high definition image of each stereo-BRUV deployment and three single frames of each stereo-DOV transect separated by approximately 8 m . This method is shown to be effective to determine reefs structural complexity [65-67]. Images were analyzed in TransectMeasure software (www.seagis.com.au) using a standardised broad habitat classification scheme based on CATAMI [68] to classify benthic composition and based on Wilson et al. [69] to classify relief characteristics (Table 1). Each image was divided into a $5 \times 4$ grid and the dominant habitat type of each square was recorded. The proportion of the total number of grid squares that fell on each category was used to estimate percent cover by sample. For ste-reo-BRUVs, this estimate was based on a single frame per deployment; and for stereo-DOVs it was based on the average of the three replicates per transect. An additional category, 'reef', was formed at the end of the image analysis by pooling macroalgae, stony coral, rock and zoanthids, and is based on the similar broad structure these environments present.

Environmental variables. Environmental variables were recorded at each sampling event. Temperature and salinity were measured using a Castaway CTD (Conductivity, Temperature and Depth) and an average temperature and salinity value was calculated from values recorded at the BRUV or dive depth, and 1 m above and below this. Visibility was estimated using a Secchi disk.

## Data analysis

The influence of habitat characteristics and environmental variables on fish assemblage richness, abundance and biomass was investigated using Generalized Additive Mixed Models (GAMM) [70,71] and a full-subsets multiple regression approach based on the function described by Fisher et al. [72]. GAMMs use smoothing splines to estimate non-parametric additive functions, allowing for overdispersion and correlation in the data [70], which may arise in studies like this.

Models were fitted to untransformed overall abundance, richness and biomass data, as well as to abundance by functional group and by families. Models for biomass by functional group
and by family were also determined, however, as the same trends were found, we decided to report results on abundance only. A prior selection of the predictor variables was made based on their coverage and on the high collinearity between them (Pearson correlation coefficient $r>0.8$ ). As a result, Reef, Rock and Mean relief remained as continuous variables for the analysis. Null variables of the random model included Month, Method, Depth and Visibility, and fixed factors included Distance to shore (two levels: inshore and offshore) and Protection (two levels: no-take and open). Continuous predictor variables were square root transformed to reduce dispersion of data.

Model selection for each response variable was based on the second-order variant of Akaike's Information Criterion suited for small samples ( $\mathrm{AIC}_{C}$ ) [73] and on $\mathrm{AIC}_{c}$ weights $\left(\omega \mathrm{AIC}_{c}\right)$. The best model was the most parsimonious one (with the fewest variables) within two $\mathrm{AIC}_{c}$ units of the lowest $\mathrm{AIC}_{c}$ value $\left(\Delta \mathrm{AIC}_{c}<2\right)$ [74]. Because the effect of protection status, and any interactions, were relevant to the primary hypothesis of this study, models that were within two AICc units of the model with the lowest $\mathrm{AIC}_{c}$ and included protection status, were therefore preferentially investigated ('hypothesis model'). Selected models had their shape and effective degrees of freedom (EDF) examined to ensure they did not overfit the data.

The distributions of fish lengths for key families were compared inside and outside NTRs using Mann-Whitney $U$ test, considering a significant difference as p -values below 0.05 . All analyses were performed using R Language for Statistical Computing [75], with the packages gamm4 [76], mgcv [77], MuMIn [78], doParallel [79] and dplyr [80].

## Results

A total of 23,505 individuals were observed belonging to 126 species of 44 families (detailed list in S1 Table, data in S2 Table). Large schools ( $>100$ ) of sardines (Clupeidae), mullets (Mugil spp.), young scads (Decapterus spp.), young vermilion snapper (Rhomboplites aurorubens) and young grunts (Haemulidae) were excluded from statistical analysis in order to reduce dispersion of data and highlight effects. Not considering these schools, the most abundant and frequent families were grunts (Haemulidae), damselfishes (Pomacentridae), jacks (Carangidae) and snappers (Lutjanidae). The most abundant species were tomtate grunt (Haemulon aurolineatum) (28.1\%), sergeant major (Abudefduf saxatilis) (10.1\%), Brazilian damsel (Stegastes fuscus) (4.7\%), gobies (Coryphopterus spp.) (2.3\%) and squirrelfish (Holocentrus adscensionis) (1.3\%). And the most frequent species were tomtate grunt (64.2\%), Brazilian damsel (54.2\%), sergeant major (49.6\%), porkfish (Anisotremus virginicus) (38.9\%) and dusky grouper (Mycteroperca marginatus) (35.1\%).

Tomtate grunt was listed as a highly common and abundant species in inshore and offshore areas, and in both no-take and fished areas (Table 2). Brazilian damsel and sergeant major were also highly recorded as abundant and frequent, except for the abundance in offshore no-take areas, which was mainly represented by schools of grunts, scads and vermilion snappers (Table 2).

Nineteen species recorded are endemic to the Brazilian Province [28,81-83] and fourteen species are considered threatened (vulnerable/endangered) or near threatened, by the International Union for Conservation of Nature (IUCN) Red List [84] and the Brazilian legislation [85] (detailed list in S1 Table).

The most parsimonious model for total richness included distance to shore and mean relief, whereas for both total abundance and biomass the selected models included protection status and distance to shore (Table 3, Figs 2 and 3). The model for overall abundance was selected based on the primary hypothesis of interest, and was within 2AIC of the top model, but it is interesting to note that mean relief was highly important (Fig 2) and present in the most parsimonious model.

ONE

Table 2. Top five most abundant and frequent species (\% of samples a species was observed) in no-take and open to fisheries areas in inshore and offshore regions.

|  | No-take |  |  |  | Open |  |  |  |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
|  | Abundance (n) |  | Frequency (\%) |  | Abundance (n) |  | Frequency (\%) |  |
| Inshore | Haemulon aurolineatum | 873 | Haemulon aurolineatum | 73 | Haemulon spp. | 651 | Abudefduf saxatilis | 48 |
|  | Abudefduf saxatilis | 604 | Mycteroperca marginatus | 62 | Abudefduf saxatilis | 415 | Stegastes fuscus | 46 |
|  | Decapterus spp. | 500 | Stegastes fuscus | 53 | Stegastes fuscus | 335 | Mycteroperca acutirostris | 35 |
|  | Coryphopterus spp. | 289 | Anisotremus virginicus | 50 | Haemulon aurolineatum | 168 | Haemulon aurolineatum | 26 |
|  | Stegastes fuscus | 265 | Abudefduf saxatilis | 43 | Caranx latus | 96 | Anisotremus virginicus | 22 |
| Offshore | Haemulon aurolineatum | 4336 | Haemulon aurolineatum | 90 | Haemulon aurolineatum | 1231 | Haemulon aurolineatum | 50 |
|  | Decapterus punctatus | 2304 | Pomacanthus paru | 75 | Abudefduf saxatilis | 939 | Abudefduf saxatilis | 56 |
|  | Rhomboplites aurorubens | 1807 | Holocentrus adscensionis | 71 | Haemulon spp. | 500 | Halichoeres poeyi | 33 |
|  | Haemulon spp. | 630 | Kyphosus spp. | 63 | Stegastes fuscus | 285 | Stegastes fuscus | 53 |
|  | Decapterus spp. | 504 | Stegastes fuscus | 60 | Mugil spp. | 264 | Chaetodon striatus | 32 |

https://doi.org/10.1371/journal.pone.0204970.t002

Higher richness of target and non-target species and greater abundance of non-target species were best predicted by increased distance to shore and mean relief, whilst the most parsimonious models for the abundance of target species indicated they were likely to increase with protection and distance to shore (Fig 4). Concerning abundance by functional groups, the most parsimonious models for carnivores/piscivores, planktivores and territorial herbivores all included a positive relationship with protection and mean relief. However, as the variance explained by the model for planktivores was very low ( $\mathrm{R}^{2}<10$ ) (Table 2), it was not represented graphically in Figs 2 and 5. Contrary to the trend found for the other functional groups, the abundance of sessile invertebrate feeders was found to be negatively correlated with protection status and mean relief. For mobile invertebrate feeders, the abundance is likely to increase with protection and distance to shore. The number of herbivores and omnivores was higher in areas further from shore and also on structurally complex reefs (Fig 5). All data used to fit the models is available on S3 Table.

Targeted families Carangidae and Epinephelidae increased with protection and presence of reef, whilst kyphosids were found in greater abundance in areas with more reef and greater distance from shore. Scarid abundance showed a negative correlation with protection and a positive correlation with distance to shore (Fig 6). In terms of body size of these families, the largest individuals were found inside the NTRs, with significant differences (Carangidae: $U=38283$, p-value $<0.001$; Scaridae: $U=4462.0$, p-value $<0.001$; Kyphosidae: $U=6450.5$, pvalue $<0.001$; Epinephelidae: $U=9341.5$, p-value $=0.013)($ Fig 6C, 6F and 6I).

## Discussion

This study is the first to generate fisheries independent data using non-destructive stereovideo methods in the Southwestern Atlantic. Besides, the approach adopted here made it possible to distinguish the effect of fishing from habitat variables on different components of the fish assemblage, demonstrating how NTRs can be used as benchmarks to contribute to resource management and marine conservation.

## The role of no-take marine reserves

Broadly, total abundance and biomass were greater inside no-take areas, a pattern also registered in previous studies [7,86,87]. The assessment of biomass in the marine environment is important and can reveal the health status of an environment especially because it can be used to represent the energy flux, as well as the potential of the ecosystem to provide goods and

Table 3. Top generalised additive mixed models (GAMMs) to predict different aspects of fish assemblage.

| Dependent variables | Best models | $\Delta \mathrm{AIC}_{c}$ | $\triangle \mathrm{BIC}$ | $\omega \mathrm{AIC}_{c}$ | $\omega \mathrm{BIC}$ | $\mathrm{R}^{2}$ | EDF |
| :---: | :---: | :---: | :---: | :---: | :---: | :---: | :---: |
| Overall |  |  |  |  |  |  |  |
| Richness | Distance to shore + Mean relief.by.Distance to shore | 0.00 | 0.00 | 1.00 | 0.89 | 0.57 | 18.75 |
| Abundance | Mean relief + Distance to shore | 0.49 | 0.00 | 0.32 | 0.68 | 0.30 | 8.53 |
|  | Protection + Distance to shore | 0.00 | 1.57 | 0.41 | 0.31 | 0.34 | 8.76 |
|  | Distance to shore + Mean relief.by.Distance to shore | 0.89 | 7.52 | 0.26 | 0.02 | 0.30 | 10.33 |
| Biomass | Protection + Distance to shore | 0.00 | 0.00 | 1.00 | 1.00 | 0.27 | 16.50 |
| Importance to fisheries |  |  |  |  |  |  |  |
| Non-target species richness | Distance to shore + Mean relief.by.Distance to shore | 0.00 | 0.00 | 1.00 | 1.00 | 0.47 | 14.53 |
| Non-target species abundance | Distance to shore + Mean relief.by.Distance to shore | 0.00 | 4.83 | 0.90 | 0.08 | 0.39 | 8.54 |
| Target species richness | Mean relief + Distance to shore | 0.00 | 0.00 | 0.61 | 0.89 | 0.56 | 18.42 |
| Target species abundance | Protection + Distance to shore | 0.00 | 0.00 | 1.00 | 1.00 | 0.25 | 13.55 |
| Abundance by functional group |  |  |  |  |  |  |  |
| Carnivores/Piscivores | Protection + Mean relief.by.Protection | 0.00 | 6.50 | 0.86 | 0.02 | 0.30 | 17.28 |
| Mobile invertebrate feeders | Protection + Distance to shore | 0.00 | 0.00 | 1.00 | 1.00 | 0.23 | 10.21 |
| Sessile invertebrate feeders | Protection | 1.43 | 0.00 | 0.15 | 0.40 | 0.13 | 7.31 |
|  | Protection + Reef.by.Protection | 0.17 | 4.73 | 0.28 | 0.04 | 0.16 | 9.16 |
|  | Protection + Mean relief.by.Protection | 0.00 | 12.54 | 0.31 | 0.00 | 0.11 | 10.92 |
| Omnivores | Mean relief + Distance to shore | 0.00 | 0.00 | 0.90 | 0.96 | 0.31 | 13.38 |
| Planktivores | Protection + Mean relief.by.Protection | 0.00 | 3.54 | 0.92 | 0.14 | 0.03 | 8.81 |
| Roving herbivores | Mean relief + Distance to shore | 0.00 | 0.00 | 0.44 | 0.48 | 0.17 | 6.95 |
|  | Distance to shore + Mean relief.by.Distance to shore | 0.47 | 17.10 | 0.35 | 0.00 | 0.18 | 9.22 |
| Territorial herbivores | Protection + Mean relief.by.Protection | 0.00 | 0.00 | 0.89 | 0.93 | 0.40 | 14.66 |
| Abundance by family |  |  |  |  |  |  |  |
| Epinephelidae | Protection + Reef.by.Protection | 0.00 | 0.00 | 1.00 | 1.00 | 0.27 | 19.49 |
| Kyphosidae | Distance to shore + Reef.by.Distance to shore | 0.00 | 0.00 | 0.53 | 0.42 | 0.15 | 17.92 |
|  | Distance to shore + Rock.by.Distance to shore | 0.30 | 0.58 | 0.46 | 0.31 | 0.16 | 17.94 |
| Scaridae | Protection + Distance to shore | 0.00 | 0.00 | 1.00 | 1.00 | 0.10 | 15.14 |
| Carangidae | Protection + Reef.by.Protection | 0.00 | 0.00 | 1.00 | 1.00 | 0.22 | 20.22 |

$\Delta \mathrm{AICc}=$ Difference between lowest reported corrected Akaike Information Criterion; $\Delta \mathrm{BIC}=$ Bayesian Information Criterion; $\omega \mathrm{AICc}=\mathrm{AICc}$ weights; $\omega \mathrm{BIC}=\mathrm{BIC}$ weights; R2 = variance explained; EDF = effective degrees of freedom. Model selection was based on the most parsimonious model within two units of the lowest AICc which has the fewest variables.
https://doi.org/10.1371/journal.pone.0204970.t003
services [88,89]. Based on this, the results indicate that the NTR in question is protecting natural processes and resources, which are being effectively converted into biomass. Conversely, the opposite was found in areas open to fishing, presenting a decreased ecosystem functioning driven by the selective removal of large individuals $[90,91]$. Higher overall abundance and biomass within NTRs indicates the significant removal of fish by fisheries in the open access areas in the region.

Distance from the coast was an important factor, explaining the higher richness, abundance and biomass recorded in islands further from the coast. This factor has been demonstrated to influence fish assemblages structure in several coral and rocky reefs around the world [92-95] and also in the Brazilian Province [13,15,25,60]. The first hypothesis we raise to explain the higher richness and abundance in offshore islands may be related to the total area of rocky reefs. In the region, offshore reefs are typically deeper and form a larger continuous extensions when compared to inshore reefs that are often interspersed with sandy beaches, probably leading a smaller surface area available for reef fishes. Surface area of reef has been directly


Fig 2. Variable importance scores from full-subset generalised additive mixed models analysis, with $>10 \%$ variance explained shown. $\mathrm{X}=$ Predictor variables within the most parsimonious model for each response variable (see Table 1).
https://doi.org/10.1371/journal.pone.0204970.g002
attributed to fish assemblage structure in some studies. For example, Francini-Filho and Moura [31] found a more pronounced increase of overall biomass over time in areas adjacent to coral reefs that reach deeper water. Furthermore, Roberts and Ormond [96] registered higher species richness with depth, and Gibran and Moura [60] also detected this tendency for rocky reefs in the same region of the present study. These findings might be due to higher availability of resources and a possible lower competition in offshore islands, especially for space [97].

The second hypothesis to explain the higher values of ecological metrics is related to the proximity of anthropogenic activities. The close proximity of human populations to a fish assemblage causes negative effects and is demonstrated worldwide [5,90,98-100]. Areas close to the mainland are easier to access and tend to have more fishing activities. Nearshore waters ( $<50 \mathrm{~m}$ water depth) of the São Paulo state coast, are highly explored by both artisanal and


Fig 3. Plots of the most parsimonious models, with $>\mathbf{1 0} \%$ variance explained shown. (A,B) species richness, (C,D) total abundance and (E,F) total biomass. The dotted line represents $95 \%$ confidence interval.
https://doi.org/10.1371/journal.pone.0204970.g003
industrial fishing fleets, with artisanal, low mobility fleets most dominant in water depth $<20$ m [23]. Coastal regions with high population densities, such as São Paulo, are more exposed to human activities causing disturbances and changes in coastal dynamics, especially concerning the high input of nutrients and pollution through air deposition, river discharges, urban and industrial wastewater effluents, groundwater and surface runoff [24]. These potentially harmful components cause environmental stress and may damage coastal biota directly or indirectly [101]. In addition, areas near the coast also face greater exposure to major developments, such as harbors and marinas, which can also significantly change the coastal landscape, causing degradation of habitats and consequently affecting fish assemblage. Further studies in the region are needed to test these hypotheses in order to determine whether or how much of this pattern is explained by biogeography or anthropogenic activities.

## Target and non-target species

Higher abundance of target species was observed within NTRs, but protection status did not correlate with any differences in the abundance of non-target species. Indeed, studies have shown increased abundance of highly targeted fishes inside no-take NTRs, with lower influence on non-target [5,7,35,102,103], reinforcing evidence of the direct effects of fishing.

## Non-target species



Fig 4. Plots of the most parsimonious models for target and non-target fish assemblage overall characteristics, with $>10 \%$ variance explained shown. On-target species $(A, B)$ richness, $(C, D)$ abundance, (E,F) biomass. And for target species (G,H) richness, (I,J) abundance, (K,L) biomass. The dotted line represents $95 \%$ confidence interval.
https://doi.org/10.1371/journal.pone.0204970.g004
Abundance of target species also increased with greater distance from the shore, which can be related to the increased fisheries activity close to the shore as described above [23].

Conversely, species richness and richness of target and non-target fish, was not related to protection status, being mostly explained by relief. Higher species diversity in more complex environments has been described in the literature [5,14,19], and is likely related to increased availability of food, decreased competition, and lower probability of predator-prey encounters [97,104,105]. Structurally complex environments have higher availability and diversity of niches, accommodating a higher number of species in a small area.

## Fish functional groups

Although functional groups responded differently to fishing pressure, we found evidence that protection status affected the trophic structure of the fish assemblage, since carnivores/piscivores, mobile invertebrate feeders, and territorial herbivores were more abundant within the NTR, whilst sessile invertebrate feeders were less abundant. However, protection was not relevant for omnivores and roving herbivores. The abundance of the carnivores/piscivores functional group, which is comprised of species targeted by fisheries in the region [18,62-64], was higher within NTRs. Even though relief was important, it was relevant only when combined with protection. These results suggests that the NTR is facilitating the recovery of high trophic level organisms, which are usually the first group depleted by fisheries [91,106,107].


Fig 5. Plots of the most parsimonious model for abundance by functional group, with $>\mathbf{1 0 \%}$ variance explained shown. (A,B) Carnivores/piscivores, (C,D) Mobile invertebrate feeders, (E) Sessile invertebrate feeders, (F,G) Omnivores, (H,I) Roving herbivores, (J,K) Territorial herbivores. The dotted line represents $95 \%$ confidence interval.
https://doi.org/10.1371/journal.pone.0204970.g005
Some mobile invertebrate feeder species are targeted by fisheries, but are not considered as important to fisheries as carnivores because of their smaller body size, such as haemulids, labrids and small carangids. Nevertheless, protection was still an important factor to predict abundance of this group, suggesting some fishing pressure, albeit less than highly targeted carnivores/piscivores. This might be related to a depletion of top predators, leading to an exploration of lower trophic levels, as already described worldwide [107], including Brazilian coast [108,109]. The other factor strongly affecting abundance of this group is distance to shore, which may be related to the larger rocky reef surface, offering more resources and, consequently, less competition [97]. This is especially important for small and benthic mobile invertebrate feeders of the families Blenniidae and Serranidae, which live closely associated with the substrate [62]. Similar results were found for omnivores, in which higher abundance is more likely to occur in high complex habitats in offshore islands, probably for the same reasons, since this group encompasses blennies, pomacentrids, pomacanthids and species of the order Tetraodontiformes. Although some species within this category are targeted by fisheries (Mugilidae, Sparidae, Ephippidae), protection was not an important factor to determine abundance. This is probably related to the plasticity of the omnivorous diet, which can enable greater resistance to environmental changes (e.g. [110,111]).

As the abundance of sessile invertebrate feeders was very low in samples, the model was not robust. However, lower abundance found within NTRs and in more complex reefs indicated by the model may be related to the elusive behaviour of these species, which usually hide from divers and may not be recorded. Since these species feed on benthic invertebrates generally

ONE


Fig 6. Plots of the most parsimonious models for abundance, Kernel density plots and boxplots for fork length (mm) for important fishing target families. (A, B,C) Carangidae, (D,E,F) Scaridae, (G,H,I) Kyphosidae and (J,K,L) Epinephelidae. The dotted line represents $95 \%$ confidence interval. * Significant difference. Fish drawings were based on Carvalho-Filho [61].
https://doi.org/10.1371/journal.pone.0204970.g006
associated with hard substrate, we would expect a higher abundance in more topographic complex environments. For planktivores, models did not predict the abundance well, most likely because species in this group show highly variable body sizes, occupying very different niches. For example, fish from Echeneidae and Carangidae families are mobile and large-bodied species, occupying the pelagic environment, whilst the species from Pomacentridae and Pempheridae families are small-bodied species that live associated with burrows and crevices on the rocky reef [62]. Therefore, it was not possible to determine a single robust model to explain abundance of this functional group with the predictor variables used.

Abundance of roving herbivores was related to distance from shore and topographic complexity, which is expected considering its diet, algae and detritus, are mostly found in reef environments [112], which are more likely abundant in larger rocky reefs of offshore islands. This is similar with the results for territorial herbivores, in which protection was only important when combined with topographic complexity. This is also likely, since territorial herbivores, such as damselfishes (Stegastes spp.), are found in complex regions of the reef protecting
colonies of the major components of their diet, primarily fast growing red and green filamentous algae [113,114]. As habitat characteristics were more influential in herbivores abundance than protection status, fisheries effects were not evident for these groups, even though some of them are targeted in the region.

## Targeted fish families

One of the consequences of large removal of individuals by fisheries activities is represented by a rapid decrease in abundance and richness, especially of large bodied target species [91]. Indeed, the effects of fishing on the size of individuals is well described, in which target species reach larger sizes within NTRs [35,99,102,115,116]. The present study corroborates these findings, showing a significantly higher density of larger individuals of target species of the families Epinephelidae, Kyphosidae, Carangidae and Scaridae within protected areas. This also represents an increase in reproduction capacity of these groups in protected areas as larger individuals usually present much higher fecundity [117]. This increases the probability of exporting larvae from NTRs to adjacent areas [31,118] repopulating fished reefs and helping to restock targeted species in fished areas.

Networks of moderate size ( $10-100 \mathrm{~km}^{2}$ ) NTRs have demonstrated to be more effective in resource management and conservation when compared to smaller protected areas [119]. However, small ( $1-5 \mathrm{~km}^{2}$ ) and very small ( $<1 \mathrm{~km}^{2}$ ) areas have been widely implemented and shown to have some advantages, specifically for small bodied and sedentary species with smaller home ranges [120-123]. In particular, individuals of the Epinephelidae family presented a higher abundance with protection and also in complex environments within the very small NTR in question. These species live associated with burrows within rocky reefs [124] and are highly targeted by fisheries, indicating that they may be the group benefiting most from protection, as seen in this study.

The abundance of kyphosids was not related to protection and was more abundant in regions offshore with the greater presence of reef. As this species is considered herbivorous, grazing predominately on macroalgae (Sargassum spp.) associated with rocks [125], we expect to record higher numbers at locations with greater food availability, including offshore areas with more rocky reef. However, larger individuals could be targeted by fishers, resulting in their higher abundance recorded within NTRs. This indicates that the NTRs allow the growth of individuals, and therefore provide greater reproductive capacity for the species.

For the Carangidae family, an effect of protection in abundance was evident, suggesting a high removal, especially of large individuals, in areas open to fisheries. Besides, regardless of being a mobile species, they are frequently found associated with hard structures [45] and even following other species [126], and probably for this reason, individuals of this family have shown to benefit from NTRs in reefs [5,127].

Fish of the Scaridae family showed a higher abundance in fished areas, likely due to the absence of top predators (carnivores/piscivores), since species of this family have been registered to be preyed upon by epinephelids, carangids and muraenids [128]. Even though they were more abundant in fished areas, fish size was smaller, representing a fishing pressure in larger sizes, as also described by Floeter et al. [35]. Also, the abundance of these roving herbivores was higher with distance from the coast, what could be related to the availability of food and lower competition in larger and continuous reefs offshore.

## Conclusions

Brazil shelters the second richest reefs in the Atlantic Ocean [28], and also stands out for the proportion of endemic and endangered species concentrated in small areas [27,28,129].

Therefore, it is crucial to generate information about the role NTRs can play in protecting fish assemblages of this region. In addition, a better understanding of patterns in the effects of fishing on a fish assemblage provides robust metrics for conservation and fisheries management, whilst also providing information on focal species and biological variables most relevant to monitor the effectiveness of NTRs to protect fish assemblages.

The present study presents evidence that very small NTRs $\left(<1 \mathrm{~km}^{2}\right)$ can protect fish assemblages from the direct effects of fishing, increasing abundance and biomass, especially of targeted species, therefore contributing to the management of fisheries resources at a local and regional scale. Some functional groups showed a higher benefit from protection, such as carnivores/piscivores and mobile invertebrate feeders, while others decreased in abundance, such as the Scaridae family. Another outstanding difference is concerning body size, mostly for target species, in which NTRs allow target species to reach larger sizes. The Epinephelidae family showed greater evidence to benefit from these very small NTRs, especially due to its high importance to fisheries and its small home range. However, we recommend that networks of larger NTRs ( $>10 \mathrm{Km}^{2}$ ) should be established in the region, which would provide a more robust framework for investigating and managing the effects of fishing and informing conservation and fisheries management more broadly.

As a concluding remark, our findings show strong influence of protection, distance from the shore and mean relief on fish assemblage characteristics, in which protected areas further from the human influence and with a higher topographic complexity tend to have greater abundance and biomass of fish. Our results highlight the crucial role these areas play in the conservation and recovery of highly valuable commercial stocks to the fishing activity of the region, displaying the importance of keeping and implementing more NTRs in the region. The use of stereo-videos in this study has shown to be effective and feasible in this region, providing valuable and robust information to aid conservation and fisheries management in Brazil.

## Supporting information

S1 Table. List of species found within no-take reserves and fished areas. *Endemic from Brazilian biogeographic province [28, 81-83]; $\mathrm{VU}_{\mathrm{I}}=$ Vulnerable by International Union for Nature Protection (IUCN) Red List [84]; $\mathrm{NT}_{\mathrm{I}}=$ Near threatened by IUCN; $\mathrm{VU}_{\mathrm{Br}}=$ Vulnerable by Brazilian legislation [85]; $\mathrm{CR}_{\mathrm{Br}}=$ Critically endangered by Brazilian Legislation; $\mathrm{Y}=$ Target; $\mathrm{N}=$ Non-target; $\mathrm{N}=$ Abundance; $\mathrm{F} \%=$ Frequency.
(DOCX)
S2 Table. Fish abundance data.
(TXT)
S3 Table. Data used to fit the generalised additive mixed models (GAMMs). (TXT)

## Acknowledgments

The authors would like to acknowledge the Chico Mendes Institute for Biodiversity Conservation (ICMBio) for the permits and field support, and also to the Universo Marinho Dive Company and the staff for sampling assistance and equipment. We are also thankful to Instituto Biopesca for all the funding management and Alfredo Carvalho Filho for the fish drawings.

## Author Contributions

Conceptualization: Fernanda A. Rolim, Tim Langlois, Otto B. F. Gadig.

Data curation: Fernanda A. Rolim, Tim Langlois, Pedro F. C. Rodrigues, Todd Bond.
Formal analysis: Fernanda A. Rolim, Tim Langlois, Todd Bond.
Funding acquisition: Fernanda A. Rolim, Otto B. F. Gadig.
Investigation: Fernanda A. Rolim, Pedro F. C. Rodrigues, Fábio S. Motta, Leonardo M. Neves.
Methodology: Fernanda A. Rolim, Tim Langlois, Pedro F. C. Rodrigues, Fábio S. Motta, Leonardo M. Neves, Otto B. F. Gadig.

Project administration: Fernanda A. Rolim, Pedro F. C. Rodrigues, Otto B. F. Gadig.
Resources: Tim Langlois, Fábio S. Motta, Otto B. F. Gadig.
Supervision: Tim Langlois, Otto B. F. Gadig.
Validation: Fernanda A. Rolim, Tim Langlois, Todd Bond.
Visualization: Fernanda A. Rolim, Tim Langlois, Todd Bond, Fábio S. Motta, Leonardo M. Neves.

Writing - original draft: Fernanda A. Rolim.
Writing - review \& editing: Fernanda A. Rolim, Tim Langlois, Pedro F. C. Rodrigues, Todd Bond, Fábio S. Motta, Leonardo M. Neves, Otto B. F. Gadig.

## References

1. Fox HE, Mascia MB, Basurto X, Costa A, Glew L, Heinemann D, et al. Reexamining the science of marine protected areas: linking knowledge to action: MPA science: linking knowledge to action. Conservation Letters. 2012; 5: 1-10.
2. Flournoy PH. Marine Protected Areas: Tools for Sustaining Ocean Ecosystems. J Int Wildl Law Policy. Routledge; 2003; 6: 137-142.
3. Lester S, Halpern B, Grorud-Colvert K, Lubchenco J, Ruttenberg B, Gaines S, et al. Biological effects within no-take marine reserves: a global synthesis. Mar Ecol Prog Ser. 2009; 384: 33-46.
4. Gell FR, Roberts CM. The fishery effects of marine reserves and fishery closures. Washington, DC.: World Wildlife Fund; 2002.
5. Edgar GJ, Stuart-Smith RD, Willis TJ, Kininmonth S, Baker SC, Banks S, et al. Global conservation outcomes depend on marine protected areas with five key features. Nature. 2014; 506: 216-220. https://doi.org/10.1038/nature13022 PMID: 24499817
6. Worm B, Barbier EB, Beaumont N, Duffy JE, Folke C, Halpern BS, et al. Impacts of biodiversity loss on ocean ecosystem services. Science. 2006; 314: 787-790. https://doi.org/10.1126/science. 1132294 PMID: 17082450
7. Micheli F, Halpern BS, Botsford LW, Warner RR. Trajectories and correlates of community change in no-take marine reserves. Ecol Appl. 2004; 14: 1709-1723.
8. Steneck RS. Human influences on coastal ecosystems: does overfishing create trophic cascades? Trends Ecol Evol. 1998; 13: 429-430. PMID: 21238380
9. Graham NAJ, Evans RD, Russ GR. The effects of marine reserve protection on the trophic relationships of reef fishes on the Great Barrier Reef. Environ Conserv. Cambridge University Press; 2003; 30: 200-208.
10. Krebs CJ. Ecology: The experimental analysis of distribution and abundance. New York: Harper \& Row; 1972.
11. Tokeshi M, Arakaki S. Habitat complexity in aquatic systems: fractals and beyond. Hydrobiologia. 2012; 685: 27-47.
12. Langlois TJ, Radford BT, Van Niel KP, Meeuwig JJ, Pearce AF, Rousseaux CSG, et al. Consistent abundance distributions of marine fishes in an old, climatically buffered, infertile seascape: Abundance distributions of fishes in stable seascapes. Glob Ecol Biogeogr. 09/2012; 21: 886-897.
13. Teixeira-Neves TP, Neves LM, Araújo FG. Hierarchizing biological, physical and anthropogenic factors influencing the structure of fish assemblages along tropical rocky shores in Brazil. Environ Biol Fishes. 2015; 98: 1645-1657.
14. Neves LM, Teixeira-Neves TP, Pereira-Filho GH, Araújo FG. The farther the better: effects of multiple environmental variables on reef fish assemblages along a distance gradient from river influences. PLoS One. 2016; 11: e0166679. https://doi.org/10.1371/journal.pone. 0166679 PMID: 27907017
15. Morais RA, Ferreira CEL, Floeter SR. Spatial patterns of fish standing biomass across Brazilian reefs. J Fish Biol. 2017; https://doi.org/10.1111/jfb. 13482 PMID: 29076535
16. García-Charton JA, Pérez-Ruzafa Á, Sánchez-Jerez P, Bayle-Sempere JT, Reñones O, Moreno D. Multi-scale spatial heterogeneity, habitat structure, and the effect of marine reserves on Western Mediterranean rocky reef fish assemblages. Mar Biol. Springer-Verlag; 2004; 144: 161-182.
17. Parsons DF, Suthers IM, Cruz DO, Smith JA. Effects of habitat on fish abundance and species composition on temperate rocky reefs. Mar Ecol Prog Ser. 2016; 561: 155-171.
18. Floeter SR, Krohling W, Gasparini JL, Ferreira CEL, Zalmon IR. Reef fish community structure on coastal islands of the southeastern Brazil: the influence of exposure and benthic cover. Environ Biol Fishes. 2007; 78: 147-160.
19. Pinheiro HT, Martins AS, Joyeux J-C. The importance of small-scale environment factors to community structure patterns of tropical rocky reef fish. J Mar Biol Assoc U K. Cambridge University Press; 2013; 93: 1175-1185.
20. ICMBio. Cadastro Nacional de Unidades de Conservação. In: Instituto Chico Mendes de Proteção à Natureza [Internet]. 1 Jun 2018 [cited 12 Nov 2018]. Available: http://www.mma.gov.br/areas-protegidas/cadastro-nacional-de-ucs/dados-consolidados.html
21. Giglio VJ, Pinheiro HT, Bender MG, Bonaldo RM, Costa-Lotufo LV, Ferreira CEL, et al. Large and remote marine protected areas in the South Atlantic Ocean are flawed and raise concerns: Comments on Soares and Lucas (2018). Mar Policy. 2018; 96: 13-17.
22. Magris RA, Pressey RL. Marine protected areas: Just for show? Science. 2018; 360: 723-724
23. Imoto RD, Carneiro MH, Ávila-da-Silva AO. Spatial patterns of fishing fleets on the Southeastern Brazilian Bight. Lat Am J Aquat Res. 2016; 44. https://doi.org/10.3856/vol4-issue-fulltext-12
24. von Glasow R, Jickells TD, Baklanov A, Carmichael GR, Church TM, Gallardo L, et al. Megacities and large urban agglomerations in the coastal zone: interactions between atmosphere, land, and marine ecosystems. Ambio. 2013; 42: 13-28. https://doi.org/10.1007/s13280-012-0343-9 PMID: 23076973
25. Floeter SR, Guimaraes RZP, Rocha LA, Ferreira CEL, Rangel CA, Gasparini JL. Geographic variation in reef-fish assemblages along the Brazilian coast. Glob Ecol Biogeogr. Wiley Online Library; 2001; 10: 423-431.
26. Rocha LA. Patterns of distribution and processes of speciation in Brazilian reef fishes. J Biogeogr. Blackwell Science Ltd; 2003; 30: 1161-1171.
27. Moura RL. Brazilian reefs as priority areas for biodiversity conservation in the Atlantic Ocean. Proceeding of the 9th International Coral Reef Symposium, Bali, Indonesia. researchgate.net; 2002. pp. 917-920.
28. Pinheiro HT, Rocha LA, Macieira RM, Carvalho-Filho A, Anderson AB, Bender MG, et al. South-western Atlantic reef fishes: Zoogeographical patterns and ecological drivers reveal a secondary biodiversity centre in the Atlantic Ocean. Treml E, editor. Divers Distrib. 2018; 24: 951-965.
29. Aued AW, Smith F, Quimbayo JP, Cândido DV, Longo GO, Ferreira CEL, et al. Large-scale patterns of benthic marine communities in the Brazilian Province. PLoS One. 2018; 13: e0198452. https://doi. org/10.1371/journal.pone.0198452 PMID: 29883496
30. Pinheiro HT, Madureira JMC, Joyeux J-C, Martins AS. Fish diversity of a southwestern Atlantic coastal island: aspects of distribution and conservation in a marine zoogeographical boundary. Check List. 2015; 11: 1615.
31. Francini-Filho RB, Moura RL. Dynamics of fish assemblages on coral reefs subjected to different management regimes in the Abrolhos Bank, eastern Brazil. Aquat Conserv. 2008; 18: 1166-1179.
32. Anderson A, Bonaldo R, Barneche D, Hackradt C, Félix-Hackradt F, García-Charton J, et al. Recovery of grouper assemblages indicates effectiveness of a marine protected area in Southern Brazil. Mar Ecol Prog Ser. 2014; 514: 207-215.
33. Anderson AB, Batista MB, Gibran FZ, Félix-Hackradt FC, Hackradt CW, García-Charton JA, et al. Habitat use of five key species of reef fish in rocky reef systems of southern Brazil: evidences of MPA effectiveness. Mar Biodivers. 2018; https://doi.org/10.1007/s12526-017-0666-7
34. Ilarri MI, Souza AT, Rosa RS. Community structure of reef fishes in shallow waters of the Fernando de Noronha archipelago: effects of different levels of environmental protection. Mar Freshwater Res. CSIRO Publishing; 2017; 68: 1303-1316.
35. Floeter SR, Halpern BS, Ferreira CEL. Effects of fishing and protection on Brazilian reef fishes. Biol Conserv. 2005; 128: 391-402.
36. Goetze JS, Jupiter SD, Langlois TJ, Wilson SK, Harvey ES, Bond T, et al. Diver operated video most accurately detects the impacts of fishing within periodically harvested closures. J Exp Mar Bio Ecol. 2015; 462: 74-82
37. Minte-Vera CV, de Moura RL, Francini-Filho RB. Nested sampling: an improved visual-census technique for studying reef fish assemblages. Mar Ecol Prog Ser. 2008; 367: 283-293.
38. Murphy HM, Jenkins GP. Observational methods used in marine spatial monitoring of fishes and associated habitats: a review. Mar Freshwater Res. 2010; 61: 236-252.
39. Thompson AA, Mapstone BD. Observer effects and training in underwater visual surveys of reef fishes. Mar Ecol Prog Ser. 1997; 154: 53-63.
40. St. John J, Russ GR, Gladstone W. Accuracy and bias of visual estimates of numbers, size structure and biomass of a coral reef fish. Mar Ecol Prog Ser. Inter-Research Science Center; 1990; 64: 253262.
41. Harvey E, Fletcher D, Shortis M, Kendrick G. A comparison of underwater visual distance estimates made by scuba divers and a stereo-video system: implications for underwater visual census of reef fish abundance. Mar Freshwater Res. 2004; 55: 573-580.
42. Harvey ES, Fletcher D, Shortis M. A comparison of the precision and accuracy of estimates of reef fish lengths determined visually by divers with estimates produced by a stereo-video system. Fish Bull. 2000; 99: 63-71.
43. Cappo M, Harvey E, Shortis M. Counting and measuring fish with baited video techniques-an overview. In: Lyle JM, Furlani DM, Buxton CD, editors. Australian Society for Fish Biology Workshop Proceedings. Tasmania: Australian Society for Fish Biology; 2006. pp. 101-114.
44. Whitmarsh SK, Fairweather PG, Huveneers C. What is big BRUVver up to? Methods and uses of baited underwater video. Rev Fish Biol Fish. Springer International Publishing; 2017; 27: 53-73.
45. Bond T, Partridge JC, Taylor MD, Langlois TJ, Malseed BE, Smith LD, et al. Fish associated with a subsea pipeline and adjacent seafloor of the North West Shelf of Western Australia. Mar Environ Res. 2018; https://doi.org/10.1016/j.marenvres.2018.08.003 PMID: 30107887
46. Mallet D, Pelletier D. Underwater video techniques for observing coastal marine biodiversity: A review of sixty years of publications (1952-2012). Fish Res. 2014; 154: 44-62.
47. Harvey E, Shortis M. A system for stereo-video measurement of sub-tidal organisms. Mar Technol Soc J. 1996; 29: 10-22.
48. Harvey E, Cappo M, Butler J, Hall N, Kendrick G. Bait attraction affects the performance of remote underwater video stations in assessment of demersal fish community structure. Mar Ecol Prog Ser. 2007; 350: 245-254.
49. Langlois TJ, Newman SJ, Cappo M, Harvey ES, Rome BM, Skepper CL, et al. Length selectivity of commercial fish traps assessed from in situ comparisons with stereo-video: Is there evidence of sampling bias? Fish Res. 2015; 161: 145-155.
50. Holmes TH, Wilson SK, Travers MJ, Langlois TJ, Evans RD, Moore GI, et al. A comparison of visual and stereo-video based fish community assessment methods in tropical and temperate marine waters of Western Australia. Limnol Oceanogr Methods. 2013; 11: 337-350.
51. Watson DL, Harvey ES, Anderson MJ, Kendrick GA. A comparison of temperate reef fish assemblages recorded by three underwater stereo-video techniques. Mar Biol. 2005; 148: 415-425.
52. Brazil. Decreto No 94.656, de 20 de Julho de 1987 [Internet]. 1987 [cited 29 Aug 2018]. Available: http://www.planalto.gov.br/
53. Dorman SR, Harvey ES, Newman SJ. Bait effects in sampling coral reef fish assemblages with stereoBRUVs. PLoS One. 2012; 7: e41538. https://doi.org/10.1371/journal.pone.0041538 PMID: 22848522
54. Whitelaw AW, Sainsbury KJ, Dews GJ, Campbell RA. Catching characteristics of four fish-trap types on the North West Shelf of Australia. Mar Freshwater Res. CSIRO Publishing; 1991; 42: 369-382.
55. Langlois T, Williams J, Monk J, Bouchet P, Currey L, Goetze J, et al. Marine sampling field manual for benthic stereo-BRUVS (Baited Remote Underwater Videos). In: Przeslawski R FS, editor. Field Manuals for Marine Sampling to Monitor Australian Waters. National Environmental Science Programme (NESP); 2018. pp. 82-104.
56. Aburto-Oropeza O, Balart EF. Community structure of reef fish in several habitats of a rocky reef in the Gulf of California. Mar Ecol. 2001; 22: 283-305.
57. Harvey ES, Shortis MR. Calibration stability of an underwater stereo-video system: implications for measurement accuracy and precision. Mar Technol Soc J. 1998; 32: 3-17.
58. Langlois TJ, Friedman A, Goetze J, Bond T, Birt MJ, McLean DL, et al. GlobalArchive: An online repository of ecological data and science communication [Internet]. Available: https://globalarchive. org/
59. Froese R, Pauly D. FishBase. Version (06/2018) [Internet]. 2018 [cited 27 Aug 2018]. Available: www. fishbase.org
60. Gibran FZ, Moura RL de. The structure of rocky reef fish assemblages across a nearshore to coastal islands' gradient in Southeastern Brazil. Neotrop Ichthyol. 2012; 10: 369-382.
61. Ferreira CEL, Gonçalves JEA, Coutinho R. Community structure of fishes and habitat complexity on a tropical rocky shore. Environ Biol Fishes. Kluwer Academic Publishers; 2001; 61: 353-369.
62. Carvalho-Filho A. Peixes: Costa Brasileira. São Paulo: Editora Melro; 1999.
63. Rolim FA, Rodrigues PFC, Gadig OBF. Peixes de recife rochoso: Estação Ecológica de Tupinambás. São Paulo: Anolis Books; 2017.
64. Begossi A, Richerson PJ. Biodiversity, family income and ecological niche: A study on the consumption of animal foods on Búzios Island (Brazil). Ecol Food Nutr. Routledge; 1993; 30: 51-61.
65. Bennett K, Wilson SK, Shedrawi G, McLean DL, Langlois TJ. Can diver operated stereo-video surveys for fish be used to collect meaningful data on benthic coral reef communities?: Comparing benthos from video methods. Limnol Oceanogr Methods. 2016; 14: 874-885.
66. Cappo M, Stowar M, Syms C, Johansson C, Cooper T. Fish-habitat associations in the region offshore from James Price Point-a rapid assessment using Baited Remote Underwater Video Stations (BRUVS). J R Soc West Aust. 2011; 94: 303-321.
67. Wilson SK, Babcock RC, Fisher R, Holmes TH, Moore JAY, Thomson DP. Relative and combined effects of habitat and fishing on reef fish communities across a limited fishing gradient at Ningaloo. Mar Environ Res. 2012; 81: 1-11. https://doi.org/10.1016/j.marenvres.2012.08.002 PMID: 22925735
68. Althaus F, Hill N, Ferrari R, Edwards L, Przeslawski R, Schönberg CHL, et al. A standardised vocabulary for identifying benthic biota and substrata from underwater imagery: The CATAMI Classification Scheme. PLoS One. 2015; 10: e0141039. https://doi.org/10.1371/journal.pone. 0141039 PMID: 26509918
69. Wilson SK, Graham NAJ, Polunin NVC. Appraisal of visual assessments of habitat complexity and benthic composition on coral reefs. Mar Biol. 2007; 151: 1069-1076.
70. Lin X, Zhang D. Inference in generalized additive mixed models by using smoothing splines. J R Stat Soc Series B Stat Methodol. 1999; 61: 381-400.
71. Hastie T, Tibshirani R. Generalized Additive Models. Stat Sci. Institute of Mathematical Statistics; 1986; 1: 297-310.
72. Fisher R, Wilson SK, Sin TM, Lee AC, Langlois TJ. A simple function for full-subsets multiple regression in ecology with R. Ecol Evol. 2018; https://doi.org/10.1002/ece3.4134 PMID: 29988441
73. Akaike H. Information theory and an extension of the maximum likelihood principle. In: BN Petrov; F Csaki, editor. 2nd International Symposium on Information Theory. Budapest, Hungary: Akadémiai Kiadó; 1973. pp. 267-281.
74. Burnham KP, Anderson DR. Model selection and multimodel Inference: A practical information-theoretic approach. 2nd Edition. New York: Springer Science \& Business Media; 2003.
75. R Core Team. R Language for Statistical Computing. In: R: A language and environment for statistical computing [Internet]. Vienna, Austria: R Foundation for Statistical Computing; 2018 [cited 8 Aug 2018]. Available: https://www.R-project.org/
76. Wood S, Scheipl F. gamm4: Generalized Additive Mixed Models using "mgcv" and "Ime4" [Internet]. 2017 [cited 27 Aug 2018]. Available: https://CRAN.R-project.org/package=gamm4
77. Wood SN. Generalized additive models: An introduction with R. Boca Raton, FL: CRC Press; 2006.
78. Barton K. MuMIn: Multi-Model Inference [Internet]. 2018 [cited 27 Aug 2018]. Available: https://CRAN. R-project.org/package=MuMIn
79. Microsoft-Corporation, Weston S. doParallel: Foreach Parallel Adaptor for the "parallel" Package [Internet]. 2017. Available: https://CRAN.R-project.org/package=doParallel
80. Wickham H, François R, Henry L, Müller K. dplyr: A Grammar of Data Manipulation [Internet]. 2018 [cited 27 Aug 2018]. Available: https://CRAN.R-project.org/package=dplyr
81. Floeter SR, Gasparini JL. The Southwestern Atlantic reef fish fauna: composition and zoogeographic patterns. J Fish Biol. Wiley Online Library; 2000; 56: 1099-1114.
82. Joyeux JC, Floeter SR, Ferreira CEL, Gasparini JL. Biogeography of tropical reef fishes: the South Atlantic puzzle. J Biogeogr. Wiley; 2001; 28: 831-841.
83. Gasparini JL, Joyeux J-C, Floeter SR. Sparisoma tuiupiranga, a new species of parrotfish (Perciformes: Labroidei: Scaridae) from Brazil, with comments on the evolution of the genus. Zootaxa. 2003; 384: 1-14.
84. IUCN. The IUCN Red List of Threatened Species. Version 2018-2 [Internet]. 2018 [cited 12 Nov 2018]. Available: https://www.iucnredlist.org/
85. MMA. Portaria MMA No 445, de 17 de Dezembro de 2014. In: Ministério do Meio Ambiente, Brasil [Internet]. 2014. Available: http://www.icmbio.gov.br/cepsul/images/stories/legislacao/Portaria/2014/ p_mma_445_2014_lista_peixes_amea\%C3\%A7ados_extin\%C3\%A7\%C3\%A3o.pdf
86. Soler GA, Edgar GJ, Thomson RJ, Kininmonth S, Campbell SJ, Dawson TP, et al. Reef fishes at all trophic levels respond positively to effective marine protected areas. PLoS One. 2015; 10: e0140270. https://doi.org/10.1371/journal.pone.0140270 PMID: 26461104
87. Halpern BS. The impact of marine reserves: do reserves work and does reserve size matter? Ecol Appl. 2003; 13: 117-137.
88. Cardinale BJ, Srivastava DS, Emmett Duffy J, Wright JP, Downing AL, Sankaran M, et al. Effects of biodiversity on the functioning of trophic groups and ecosystems. Nature. Nature Publishing Group; 2006; 443: 989. https://doi.org/10.1038/nature05202 PMID: 17066035
89. Hooper DU, Chapin FS III, Ewel JJ, Hector A, Inchausti P, Lavorel S, et al. Effects of biodiversity on ecosystem functioning: a consensus of current knowledge. Ecol Monogr. 2005; 75: 3-35.
90. Mora C, Aburto-Oropeza O, Bocos AA, Ayotte PM, Banks S, Bauman AG, et al. Global human footprint on the linkage between biodiversity and ecosystem functioning in reef fishes. PLoS Biol. Public Library of Science; 2011; 9: e1000606. https://doi.org/10.1371/journal.pbio. 1000606 PMID: 21483714
91. Crowder LB, Hazen EL, Avissar N, Bjorkland R, Latanich C, Ogburn MB. The Impacts of Fisheries on Marine Ecosystems and the Transition to Ecosystem-Based Management. Annu Rev Ecol Evol Syst. 2008; 39: 259-278.
92. Van Nguyen L, Kim Phan H. Distribution and factors influencing on structure of reef fish communities in Nha Trang Bay Marine Protected Area, South-Central Vietnam. Environ Biol Fishes. 2007; 82: 309.
93. Lecchini D, Adjeroud M, Pratchett MS, Cadoret L, Galzin R. Spatial structure of coral reef fish communities in the Ryukyu Islands, southern Japan. Oceanol Acta. 2003; 26: 537-547.
94. Schultz AL, Malcolm HA, Bucher DJ, Linklater M, Smith SDA. Depth and medium-scale spatial processes influence fish assemblage structure of unconsolidated habitats in a subtropical marine park. PLoS One. journals.plos.org; 2014; 9: e96798. https://doi.org/10.1371/journal.pone.0096798 PMID: 24824998
95. Malcolm HA, Jordan A, Smith SDA. Biogeographical and cross-shelf patterns of reef fish assemblages in a transition zone. Mar Biodivers. 2010; 40: 181-193.
96. Roberts CM, Ormond R. Habitat complexity and coral reef fish diversity and abundance on Red Sea fringing reefs. Mar Ecol Prog Ser. 1987; 41: 1-8.
97. Sale PF. Maintenance of high diversity in coral reef fish communities. Am Nat. [University of Chicago Press, American Society of Naturalists]; 1977; 111: 337-359.
98. Babcock RC, Shears NT, Alcala AC, Barrett NS, Edgar GJ, Lafferty KD, et al. Decadal trends in marine reserves reveal differential rates of change in direct and indirect effects. Proc Natl Acad Sci U S A. 2010; 107: 18256-18261. https://doi.org/10.1073/pnas. 0908012107 PMID: 20176941
99. Edgar GJ, Stuart-Smith RD, Thomson RJ, Freeman DJ. Consistent multi-level trophic effects of marine reserve protection across northern New Zealand. PLoS One. 2017; 12: e0177216. https://doi. org/10.1371/journal.pone. 0177216 PMID: 28542268
100. Langlois TJ, Harvey ES, Meeuwig JJ. Strong direct and inconsistent indirect effects of fishing found using stereo-video: Testing indicators from fisheries closures. Ecol Indic. Elsevier; 2012; 23: 524-534.
101. Adams SM. Assessing cause and effect of multiple stressors on marine systems. Mar Pollut Bull. 2005; 51: 649-657. https://doi.org/10.1016/j.marpolbul.2004.11.040 PMID: 16291182
102. Malcolm HA, Williams J, Schultz AL, Neilson J, Johnstone N, Knott NA, et al. Targeted fishes are larger and more abundant in "no-take" areas in a subtropical marine park. Estuar Coast Shelf Sci. Elsevier; 2018; 212: 118-127.
103. Evans RD, Russ GR. Larger biomass of targeted reef fish in no-take marine reserves on the Great Barrier Reef, Australia. Aquat Conserv. Wiley Online Library; 2004; 14: 505-519.
104. Eklöv P, Diehl S. Piscivore efficiency and refuging prey: the importance of predator search mode. Oecologia. 1994; 98: 344-353. https://doi.org/10.1007/BF00324223 PMID: 28313911
105. Hauzy C, Tully T, Spataro T, Paul G, Arditi R. Spatial heterogeneity and functional response: an experiment in microcosms with varying obstacle densities. Oecologia. 2010; 163: 625-636. https://doi.org/ 10.1007/s00442-010-1585-5 PMID: 20213153
106. Christensen V, Guenette S, Heymans JJ, Walters CJ, Watson R, Zeller D, et al. Hundred-year decline of North Atlantic predatory fishes. Fish Fish. 2003; 4: 1-24.
107. Pauly D, Christensen VV, Dalsgaard J, Froese R, Torres F Jr. Fishing down marine food webs. Science. science.sciencemag.org; 1998; 279: 860-863. PMID: 9452385
108. Freire KMF, Pauly D. Fishing down Brazilian marine food webs, with emphasis on the east Brazil large marine ecosystem. Fish Res. 2010; 105: 57-62.
109. Pinheiro HT, Joyeux JC, Martins AS. Reef fisheries and underwater surveys indicate overfishing of a Brazilian coastal island. Nat Conserv. 2010; 08: 151-159.
110. Bellwood DR, Hoey AS, Ackerman JL, Depczynski M. Coral bleaching, reef fish community phase shifts and the resilience of coral reefs. Glob Chang Biol. 2006; 12: 1587-1594.
111. Pratchett MS, Hoey AS, Wilson SK, Messmer V, Graham NAJ. Changes in biodiversity and functioning of reef fish assemblages following coral bleaching and coral loss. Diversity. Molecular Diversity Preservation International; 2011; 3: 424-452.
112. Francini-Filho RB, Ferreira CM, Coni EOC, De Moura RL, Kaufman L. Foraging activity of roving herbivorous reef fish (Acanthuridae and Scaridae) in eastern Brazil: influence of resource availability and interference competition. J Mar Biol Assoc U K. Cambridge University Press; 2010; 90: 481-492.
113. Ferreira CEL, Gonçalves JEA, Coutinho R, Peret AC. Herbivory by the Dusky Damselfish Stegastes fuscus (Cuvier, 1830) in a tropical rocky shore: effects on the benthic community. J Exp Mar Bio Ecol. 1998; 229: 241-264.
114. Ferreira CM, Coni EOC, Medeiros DV, Sampaio CLS, Reis-Filho JA, Barros F, et al. Community structure of shallow rocky shore fish in a tropical bay of the southwestern Atlantic. Brazil J Oceanogr. Instituto Oceanográfico da Universidade de São Paulo; 2015; 63: 379-396.
115. Watson DL, Anderson MJ, Kendrick GA, Nardi K, Harvey ES. Effects of protection from fishing on the lengths of targeted and non-targeted fish species at the Houtman Abrolhos Islands, Western Australia. Mar Ecol Prog Ser. 2009; 384: 241-249.
116. DeMartini EE, Friedlander AM, Sandin SA, Sala E. Differences in fish-assemblage structure between fished and unfished atolls in the northern Line Islands, central Pacific. Mar Ecol Prog Ser. 2008; 365: 199-215.
117. Jennings S, Kaiser M, Reynolds JD. Marine Fisheries Ecology. John Wiley \& Sons; 2009.
118. Palumbi SR. Marine reserves and ocean neighborhoods: The spatial scale of marine populations and their management. Annu Rev Environ Resour. 2004; 29: 31-68.
119. Halpern BS, Warner RR. Matching marine reserve design to reserve objectives. Proc Biol Sci. 2003; 270: 1871-1878. https://doi.org/10.1098/rspb.2003.2405 PMID: 14561299
120. Bonaldo RM, Pires MM, Guimarães PR Junior, Hoey AS, Hay ME. Small marine protected areas in Fij provide refuge for reef fish assemblages, feeding groups, and corals. PLoS One. 2017; 12: e0170638. https://doi.org/10.1371/journal.pone. 0170638 PMID: 28122006
121. Di Franco A, Plass-Johnson JG, Di Lorenzo M, Meola B, Claudet J, Gaines SD, et al. Linking home ranges to protected area size: The case study of the Mediterranean Sea. Biol Conserv. 2018; 221: 175-181.
122. Afonso P, Fontes J, Santos RS. Small marine reserves can offer long term protection to an endangered fish. Biol Conserv. 2011; 144: 2739-2744.
123. McLaren BW, Langlois TJ, Harvey ES, Shortland-Jones H, Stevens R. A small no-take marine sanctuary provides consistent protection for small-bodied by-catch species, but not for large-bodied, highrisk species. J Exp Mar Bio Ecol. 2015; 471: 153-163.
124. Gibran FZ. Activity, habitat use, feeding behavior, and diet of four sympatric species of Serranidae (Actinopterygii: Perciformes) in Southeastern Brazil. Neotrop Ichthyol. Sociedade Brasileira de Ictiologia; 2007; 5: 387-398.
125. Silvano RAM, Güth AZ. Diet and feeding behavior of Kyphosus spp. (Kyphosidae) in a Brazilian subtropical reef. Braz Arch Biol Technol. Tecpar; 2006; 49: 623-629.
126. Sazima C, Krajewski JP, Bonaldo RM, Sazima I. Nuclear-follower foraging associations of reef fishes and other animals at an oceanic archipelago. Environ Biol Fishes. 2007; 80: 351-361.
127. Santana-Garcon J, Newman SJ, Langlois TJ, Harvey ES. Effects of a spatial closure on highly mobile fish species: an assessment using pelagic stereo-BRUVs. J Exp Mar Bio Ecol. 2014; 460: 153-161.
128. Randall JE. Food habits of reef fishes of the West Indies. Institute of Marine Sciences, University of Miami Coral Gables; 1967; Available: http://www.aoml.noaa.gov/general/lib/CREWS/Cleo/ PuertoRico/prpdfs/randall-habits.pdf
129. Floeter SR, Rocha LA, Robertson DR, Joyeux JC, Smith-Vaniz WF, Wirtz P, et al. Atlantic reef fish biogeography and evolution. J Biogeogr. 2007; 35: 22-47.
