

Complete Genome Sequence of Klebsiella pneumoniae Myophage May

Katherine T. Nguyen,a Rachele Bonasera,a Garret Benson,a Adriana C. Hernandez-Morales,a Jason J. Gill,a Mei Liua

aCenter for Phage Technology, Texas A&M University, College Station, Texas, USA

ABSTRACT May is a newly isolated myophage that infects multidrug-resistant strains of Klebsiella pneumoniae, a pathogen that is associated with antibiotic-resistant infections in humans. The genome of May has been shown to be similar to that of phage Vi01.

K *kebsiella pneumoniae* is linked to respiratory system infections, abscesses, and
bacteremia and is a growing global health concern as antibiotic-resistant strains continue to emerge, including the multidrug-resistant strains carrying the bla_{KPC} car-bapenemase [\(1\)](#page-1-0). Phages infecting K. pneumoniae may provide treatment options for these infections.

Phage May was isolated from wastewater influent collected in Bryan, Texas, in 2016 using a bla_{Kpc} -containing K. pneumoniae sequence type 258 (ST258) clinical isolate as the host. Host bacteria were cultured on tryptic soy broth or agar (Difco) at 37°C with aeration. Phages were isolated and propagated by the soft agar overlay method [\(2\)](#page-1-1). Phage genomic DNA was prepared using a modified Promega Wizard DNA cleanup kit protocol as described previously [\(3\)](#page-1-2). Pooled indexed DNA libraries were prepared using the Illumina TruSeq Nano LT kit, and the sequence was obtained from the Illumina MiSeq platform using the MiSeq V2 500-cycle reagent kit following the manufacturer's instructions, producing 483,323 paired-end reads for the index containing the phage genome. FastQC 0.11.5 [\(https://www.bioinformatics.babraham.ac.uk/projects/fastqc/\)](https://www.bioinformatics.babraham.ac.uk/projects/fastqc/) and FastX Toolkit 0.0.14 [\(http://hannonlab.cshl.edu/fastx_toolkit/\)](http://hannonlab.cshl.edu/fastx_toolkit/) were used for read quality control, and reads were assembled using SPAdes 3.5.0 [\(4\)](#page-1-3). The assembled genome was closed by PCR using primers (5'-GGAAATGTCCGGCTGAAGATA-3' and 5'-GC TGACGCAGTGTGAAATTG-3') facing off the ends of the assembled contig and Sanger sequencing of the resulting product, with the contig sequence manually corrected to match the resulting Sanger sequencing read. The protein-coding genes were detected with GLIMMER 3.0 and MetaGeneAnnotator 1.0 with manual correction [\(5,](#page-1-4) [6\)](#page-1-5), and the tRNAs were identified with ARAGORN 2.36 [\(7\)](#page-1-6). The gene functions were predicted by similarity detection using BLASTp 2.2.28 [\(8\)](#page-1-7), and the presence of conserved domains was determined by InterProScan 5.15-5.40 [\(9\)](#page-1-8). The Center for Phage Technology (CPT) Galaxy [\(10\)](#page-1-9) and Web Apollo interfaces [\(11\)](#page-1-10) were used for all analyses using default settings [\(cpt.tamu.edu\)](http://cpt.tamu.edu).

The genome of phage May was assembled into a closed contig of 159,631 bp with 380.5-fold coverage and a G-C content of 46.8%. The genome contains 214 proteincoding genes with a coding density of 92.7%. The May genome encodes proteins common to many other myophages of the T4 superfamily. A tape measure protein was identified in phage May, but a tape measure chaperone was not detected upstream. An N-acetylmuramidase-type endolysin (IPR024408) was identified with a predicted N-terminal peptidoglycan-binding domain (IPR002477); however, no holin or spanin complex could be identified. Using the progressiveMauve algorithm 2.4.0 [\(12\)](#page-1-11), May was determined to be 79.8% similar to phage 0507-KN2-1 [\(NC_022343\)](https://www.ncbi.nlm.nih.gov/nuccore/NC_022343), another phage that infects K. pneumoniae. Phage May also shares 178 proteins with phage Vi01, as **Citation** Nguyen KT, Bonasera R, Benson G, Hernandez-Morales AC, Gill JJ, Liu M. 2019. Complete genome sequence of Klebsiella pneumoniae myophage May. Microbiol Resour Announc 8:e00252-19. [https://doi.org/10.1128/](https://doi.org/10.1128/MRA.00252-19) [MRA.00252-19.](https://doi.org/10.1128/MRA.00252-19)

Editor Catherine Putonti, Loyola University Chicago

Copyright © 2019 Nguyen et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0](https://creativecommons.org/licenses/by/4.0/) [International license.](https://creativecommons.org/licenses/by/4.0/)

Address correspondence to Mei Liu, [meiliu@tamu.edu.](mailto:meiliu@tamu.edu)

Received 4 March 2019 **Accepted** 6 April 2019 **Published** 9 May 2019

determined by a BLASTp search ($E < 10^{-5}$) using the NCBI nonredundant (nr) database, and the arrangement of gene modules within the May genome is similar to that of Vi01 [\(13\)](#page-1-12). This suggests that phage May is Vi01-like, a member of a group of large myophages that are distantly related to phage T4. A defining feature among Vi01-like phages and consistent in the May genome is the presence of a cluster of predicted tail spike genes [\(14\)](#page-1-13). The arrangement of four tail spike genes has been predicted to produce umbrella-shaped tail structures [\(15\)](#page-1-14).

Data availability. The genome sequence of phage May was deposited under the GenBank accession no. [MG428991.](https://www.ncbi.nlm.nih.gov/nuccore/MG428991) The associated BioProject, SRA, and BioSample accession numbers are [PRJNA222858,](https://www.ncbi.nlm.nih.gov/bioproject/PRJNA222858) [SRR8788203,](https://www.ncbi.nlm.nih.gov/sra/SRR8788203) and [SAMN11259659,](https://www.ncbi.nlm.nih.gov/biosample/SAMN11259659) respectively.

ACKNOWLEDGMENTS

This work was supported by funding from the National Science Foundation (award no. EF-0949351 and DBI-1565146) and by the National Institutes of Health (NIAID award no. AI121689). Additional support came from the Center for Phage Technology (CPT), an Initial University Multidisciplinary Research Initiative supported by Texas A&M University and Texas AgriLife, and from the Texas A&M University Department of Biochemistry and Biophysics.

We thank Thomas Walsh, Weill Cornell Medical School, for the provision of bacterial isolates. We are grateful for the advice and support of the CPT staff.

This announcement was prepared in partial fulfillment of the requirements for BICH464 Phage Genomics, an undergraduate course at Texas A&M University.

REFERENCES

- 1. Gomez-Simmonds A, Uhlemann AC. 2017. Clinical implications of genomic adaptation and evolution of carbapenem-resistant Klebsiella pneumoniae. J Infect Dis 215:S18-S27. [https://doi.org/10.1093/infdis/](https://doi.org/10.1093/infdis/jiw378) iiw378.
- 2. Adams MK. 1959. Bacteriophages. Interscience Publishers, Inc., New York, NY.
- 3. Summer EJ. 2009. Preparation of a phage DNA fragment library for whole genome shotgun sequencing. Methods Mol Biol 502:27– 46. [https://doi.org/10.1007/978-1-60327-565-1_4.](https://doi.org/10.1007/978-1-60327-565-1_4)
- 4. Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol 19:455– 477. [https://doi.org/10.1089/cmb.2012.0021.](https://doi.org/10.1089/cmb.2012.0021)
- 5. Delcher AL, Harmon D, Kasif S, White O, Salzberg SL. 1999. Improved microbial gene identification with GLIMMER. Nucleic Acids Res 27: 4636 – 4641. [https://doi.org/10.1093/nar/27.23.4636.](https://doi.org/10.1093/nar/27.23.4636)
- 6. Noguchi H, Taniguchi T, Itoh T. 2008. MetaGeneAnnotator: detecting species-specific patterns of ribosomal binding site for precise gene prediction in anonymous prokaryotic and phage genomes. DNA Res 15:387–396. [https://doi.org/10.1093/dnares/dsn027.](https://doi.org/10.1093/dnares/dsn027)
- 7. Laslett D, Canback B. 2004. ARAGORN, a program to detect tRNA genes and tmRNA genes in nucleotide sequences. Nucleic Acids Res 32:11–16. [https://doi.org/10.1093/nar/gkh152.](https://doi.org/10.1093/nar/gkh152)
- 8. Camacho C, Coulouris G, Avagyan V, Ma N, Papadopoulos J, Bealer K, Madden TL. 2009. BLAST+: architecture and applications. BMC Bioinformatics 10:421. [https://doi.org/10.1186/1471-2105-10-421.](https://doi.org/10.1186/1471-2105-10-421)
- 9. Jones P, Binns D, Chang HY, Fraser M, Li W, McAnulla C, McWilliam H, Maslen J, Mitchell A, Nuka G, Pesseat S, Quinn AF, Sangrador-Vegas A,

Scheremetjew M, Yong SY, Lopez R, Hunter S. 2014. InterProScan 5: genome-scale protein function classification. Bioinformatics 30: 1236 –1240. [https://doi.org/10.1093/bioinformatics/btu031.](https://doi.org/10.1093/bioinformatics/btu031)

- 10. Cock PJ, Gruning BA, Paszkiewicz K, Pritchard L. 2013. Galaxy tools and workflows for sequence analysis with applications in molecular plant pathology. PeerJ 1:e167. [https://doi.org/10.7717/peerj.167.](https://doi.org/10.7717/peerj.167)
- 11. Lee E, Helt GA, Reese JT, Munoz-Torres MC, Childers CP, Buels RM, Stein L, Holmes IH, Elsik CG, Lewis SE. 2013. Web Apollo: a Web-based genomic annotation editing platform. Genome Biol 14:R93. [https://doi](https://doi.org/10.1186/gb-2013-14-8-r93) [.org/10.1186/gb-2013-14-8-r93.](https://doi.org/10.1186/gb-2013-14-8-r93)
- 12. Darling AE, Mau B, Perna NT. 2010. progressiveMauve: multiple genome alignment with gene gain, loss and rearrangement. PLoS One 5:e11147. [https://doi.org/10.1371/journal.pone.0011147.](https://doi.org/10.1371/journal.pone.0011147)
- 13. Pickard D, Toribio AL, Petty NK, van Tonder A, Yu L, Goulding D, Barrell B, Rance R, Harris D, Wetter M, Wain J, Choudhary J, Thomson N, Dougan G. 2010. A conserved acetyl esterase domain targets diverse bacteriophages to the Vi capsular receptor of Salmonella enterica serovar Typhi. J Bacteriol 192:5746 –5754. [https://doi.org/10.1128/JB.00659-10.](https://doi.org/10.1128/JB.00659-10)
- 14. Hooton SP, Timms AR, Rowsell J, Wilson R, Connerton IF. 2011. Salmonella Typhimurium-specific bacteriophage PhiSH19 and the origins of species specificity in the Vi01-like phage family. Virol J 8:498. [https://doi](https://doi.org/10.1186/1743-422X-8-498) [.org/10.1186/1743-422X-8-498.](https://doi.org/10.1186/1743-422X-8-498)
- 15. Adriaenssens EM, Ackermann HW, Anany H, Blasdel B, Connerton IF, Goulding D, Griffiths MW, Hooton SP, Kutter EM, Kropinski AM, Lee JH, Maes M, Pickard D, Ryu S, Sepehrizadeh Z, Shahrbabak SS, Toribio AL, Lavigne R. 2012. A suggested new bacteriophage genus: "Viunalikevirus." Arch Virol 157:2035–2046. [https://doi.org/10.1007/s00705-012](https://doi.org/10.1007/s00705-012-1360-5) [-1360-5.](https://doi.org/10.1007/s00705-012-1360-5)