



The Function of SUMOylation and Its Critical Roles in Cardiovascular Diseases and Potential Clinical Implications

Congcong Du¹, Xu Chen¹, Qi Su¹, Wenbin Lu¹, Qiqi Wang¹, Hong Yuan¹, Zhenzhen Zhang¹, Xiaotong Wang², Hongmei Wu^{1,*} and Yitao Qi^{1,*}

- ¹ Key Laboratory of the Ministry of Education for Medicinal Resources and Natural Pharmaceutical Chemistry, National Engineering Laboratory for Resource Developing of Endangered Chinese Crude Drugs in Northwest of China, College of Life Sciences, Shaanxi Normal University, Xi'an 710119, China; ducongcong@snnu.edu.cn (C.D.); chenxu@snnu.edu.cn (X.C.); suqidream77@163.com (Q.S.); sdluloloo@163.com (W.L.); wang222703789@snnu.edu.cn (Q.W.); yhong@snnu.edu.cn (H.Y.); zhang2877534304@snnu.edu.cn (Z.Z.)
- ² School of Agriculture, Ludong University, Yantai 246011, China; wangxiaotong999@163.com
- * Correspondence: hq8479@snnu.edu.cn (H.W.); qiyitao@snnu.edu.cn (Y.Q.)

Abstract: Cardiovascular disease (CVD) is a common disease caused by many factors, including atherosclerosis, congenital heart disease, heart failure, and ischemic cardiomyopathy. CVD has been regarded as one of the most common diseases and has a severe impact on the life quality of patients. The main features of CVD include high morbidity and mortality, which seriously threaten human health. SUMO proteins covalently conjugate lysine residues with a large number of substrate proteins, and SUMOylation regulates the function of target proteins and participates in cellular activities. Under certain pathological conditions, SUMOylation of proteins related to cardiovascular development and function are greatly changed. Numerous studies have suggested that SUMOylation of substrates plays critical roles in normal cardiovascular development and function. We reviewed the research progress of SUMOylation in cardiovascular development and function, and the regulation of protein SUMOylation may be applied as a potential therapeutic strategy for CVD treatment.

Keywords: SUMOylation; SENP; cardiac development; cardiovascular disease; clinical targets

1. Introduction

Cardiovascular disease (CVD) is a generic term of heart disease caused by a series of factors and is one of the major causes of human death. CVD is a serious threat to human life and has become a hot topic in modern medical research. SUMOylation acts as a dynamic equilibrium protein modification and is ubiquitous in human cells. It has been found that SUMOylation plays important roles in maintaining normal cell functions, and dysregulation of SUMOylation has been observed in the pathogenesis of various heart diseases. There are a large number of target proteins modified by SUMO in the heart, which affect cell proliferation, apoptosis, and metabolism. The imbalance of SUMOylation leads to abnormal cardiac function, resulting in the occurrence of CVD. However, the molecular mechanism of SUMOylation in CVD is still not completely clear. This review describes the latest research of SUMOylation in CVD and aims to provide new ideas and potential therapeutic targets for the treatment of CVD.

1.1. CVD

CVD is a group of diseases caused by many factors, including atherosclerosis, congenital heart disease (CHD), heart failure, and ischemic cardiomyopathy, with a high prevalence, morbidity, and mortality [1]. Atherosclerosis is a progressive inflammatory cardiovascular disease characterized by lipid plaque formation in the artery [2]. The formation and rupture of lipid plaques is one important cause of clinical cardiovascular



Citation: Du, C.; Chen, X.; Su, Q.; Lu, W.; Wang, Q.; Yuan, H.; Zhang, Z.; Wang, X.; Wu, H.; Qi, Y. The Function of SUMOylation and Its Critical Roles in Cardiovascular Diseases and Potential Clinical Implications. *Int. J. Mol. Sci.* 2021, *22*, 10618. https:// doi.org/10.3390/ijms221910618

Academic Editor: Kwang-Hyun Baek

Received: 9 September 2021 Accepted: 28 September 2021 Published: 30 September 2021

Publisher's Note: MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Copyright: © 2021 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (https:// creativecommons.org/licenses/by/ 4.0/). events, including stroke and myocardial infarction, accounting for more than one-quarter of global deaths [3]. CHD is a congenital cardiac structural abnormality, and it is a common congenital defect in infants, accounting for about 1% of all newborns [4]. Atrial septal defects (ASDs) and ventricular septal defects (VSDs) are the most common CHDs, which are characterized by an existing connection between atria and ventricles [5]. Heart failure is a complex disease caused by valve abnormality, myocardial ischemia, coronary artery disease, hypertension, and other pathological changes [6]. Cardiac energy metabolism and mitochondrial biogenesis defects lead to heart failure and cardiac hypertrophy [7]. Acute myocardial infarction (AMI) is an ischemic cardiac disease with acute hypoxia in the myocardial tissue and is one of the leading causes of death. Ischemia/reperfusion (I/R) is an important treatment for rescuing ischemic myocardium from necrosis [8]. Readjustment of blood flow restores oxygen and nutrient supply but may damage the cardiac muscle and induce myocardial I/R injury. Advances in pharmacology and interventional therapy have improved CVD treatment but have not solved the problem from the source. Extensive proteins involved in the pathology of CVD are modified by SUMO proteins, indicating that SUMOylation of substrates plays important roles in the progression of CVD.

1.2. SUMOylation

SUMO (small ubiquitin-like modifier) is ubiquitous in eukaryotes, and the SUMO family contains SUMO1, SUMO2, SUMO3, and SUMO4 [9,10]. SUMO1, SUMO2, and SUMO3 are commonly found [11,12], while SUMO4 is only expressed in the spleen, kidney, and other organs [13]. The identical sequences of SUMO2 and SUMO3 are 97%, so they are difficult to distinguish and commonly referred to as SUMO2/3, while only 47% of the homologous sequences are between SUMO1 and SUMO2/3 [14,15]. SUMO4 was initially shown to be associated with the pathogenesis of type 1 diabetes, but it was later reported that SUMO4 was ineffective in covalent coupling with the substrate due to the presence of proline-90 amino acid residues that prevent maturation of SUMO4 [16].

SUMOylation is accomplished via maturation, activation, conjugation, and ligation with activating enzyme E1 (SAE1/SAE2), conjugating enzyme E2 (UBC9), and ligating enzyme E3 [15]. The C-terminal of SUMO forms a covalent bond with the lysine residues of target proteins through an isopeptide bond. The SUMO precursor is processed to expose the C-terminal di-glycine motif, then the cysteine residue of the E1 subunit forms a high-energy thioester bond with SUMO. The activated SUMO was transferred to the E2-binding enzyme, and linked to the cystine of UBC9 to form the SUMO-E2 intermediate. UBC9 transferred SUMO to the substrate, and the C-terminal glycine of SUMO bound to the lysine residue of the substrate to form the isopeptide bond [17]. Although E1 and E2 are sufficient to complete SUMOylation, the E3 is still required to enhance the specificity of SUMO isoforms and substrates, activate the SUMO binding site, and promote the formation of poly-SUMO chains.

The deSUMOylation is achieved by a family of SENP (Sentrin/SUMO-specific protease) [10,18], which deconjugate the SUMO from target proteins and catalyze the maturation of the SUMO proteins [7]. The SENP family in mammalian cells has six members, which can be divided into three groups. The first group is SENP1 and SENP2, and both have broad substrate specificity [19–24]. The second group is SENP3 and SENP5, and both in the nucleus, but mainly in the nucleolus, tend to deSUMOylation of SUMO2/3. The third group includes SENP6 and SENP7, both with additional ring structures in their catalytic zone. SUMO and SENP2 dynamically regulate the SUMOylation level of target proteins (Figure 1), and SUMOylation is an important dynamic reversible post-translational modification (PTM) to regulate protein stability, subcellular localization, and function [25]. Disorders of SUMOylation affect the normal function of target proteins and lead to the damage of important physiological processes and an abnormal intracellular environment [26].



Figure 1. Dynamic regulation of the SUMOylation pathway. The SUMO precursor protein is matured by SENP and then activated by E1, conjugated by E2, and ligated to substrates by E3. On the other hand, SENP deconjugates SUMO from target proteins. S: SUMO. K: Lysine.

Protein homeostasis is indispensable, especially in cells with low mitotic activity, such as neurons and cardiomyocytes, to maintain cellular function [27,28]. Under certain pathological conditions, SUMOylation-regulated signaling pathways related to cardiovascular development and function are changed [29,30]. SUMOylation affects the function of cardiovascular proteins, emerging as key factors of various CVD progression [29]. In recent years, numerous new SUMOylation targets have emerged, and the important roles of SUMOylation in CVD progression have been continuously discovered [31,32].

2. SUMO Machinery and CVD

2.1. UBC9

UBC9 is the only E2 enzyme responsible for the function of GATA4 in cardiac development and function. UBC9 enhanced the transcriptional activity and altered the nuclear localization of GATA4 [33]. UBC9-mediated SUMOylation of GATA4 on lysine residue 366 (K366) promoted specific gene expression in cardiac pluripotent cells. Knockout of UBC9 in mice stimulated early embryonic death at embryonic day 3.5 [34]. UBC9-mediated SUMOylation induced a higher level of autophagy and increased autophagy flux and the incidence of cardiac development [35]. These studies indicated an important role of UBC9 in cardiac development (Figure 2).

2.2. SUMO1

The SUMOylation was mediated by the SUMO family, and SUMO1 was one of the most important members. SUMO1 plays a critical role in early heart development. Mice with SUMO1 knockout showed the CHD phenotype. Heterozygous and homozygous mice with higher ASD and VSD mortality due to SUMO1 knockout were rescued by cardiac expression of SUMO1 [36]. Thus, SUMO1 played a key role in normal cardiac

development, and decreased SUMOylation activity induces cardiac pathology. The changed SUMOylation levels affect a variety of transcription factors and enzymes that maintain normal cardiac function.

SUMO1 was also found to improve the oxidative stress response during increased cardiac hypertrophy [37]. SUMO1 transfection with adenovirus after TAC, a model of heart failure induced by stress overload, prevented the occurrence of cardiac hypertrophy, blocked the negative effect of hydrogen peroxide on SERCA2a, and decreased the oxidative stress index in mice [38]. These studies indicate that SUMO1 plays important roles in normal cardiac function and suggest that SUMO1 prevents heart failure in a toxic environment (Table 1).



Figure 2. SUMO machinery and CVD. SUMO machinery, including UBC9, SUMO1, SUMO2, SENP2, SENP3, and SENP5, participated in the progression of CVDs, including congenital heart disease, heart failure, and ischemic cardiomyopathy. CHD: congenital heart disease.

Table 1. SUMO machinery and CVD.

SUMO Machinery	Disease	Biophysical and Biological Effects of SUMOylation	
UBC9	CHD	UBC9-mediated SUMOylation induced a higher level of autophagy and increased autophagy flux and the incidence of cardiac development [35,39].	
SUMO1	CHD and HF	SUMO1 plays a key role in normal cardiac development and decreased SUMOylation activity induces cardiac pathology [36].	
SUMO2	HF	Endogenous SUMO2 is important in maintaining normal endothelium-dependent vascular function [40], and SUMO2 upregulation leads to endothelial dysfunction due to cholesterolemia [41].	
SENP2	CHD	Overexpression of SENP2 led to ASD and VSD, reduced proliferation of cardiomyocytes, and induced cardiomyopathy with aging [13].	
SENP3	Ischemic cardiomyopathy	SENP3-mediated deSUMOylation plays an important role in the regulation of myocardial cell survival after ischemic injury [42].	
SENP5	HF	SENP5 level increased in the failing heart, and the increase of mitochondrial division mediated by Drp1 leads to changes in the morphology and structure of cardiac mitochondria, resulting in mitochondrial dysfunction [43].	

CHD: congenital heart disease; HF: heart failure.

2.3. SUMO2

In previous studies, little is known about whether and how SUMO2/3 functions in CVD. It was reported that endogenous SUMO2 was important in maintaining normal

endothelium-dependent vascular function [40], and SUMO2 upregulation disrupts the balance of the vascular environment and leads to endothelial dysfunction due to cholesterolemia [41]. Increased SUMO2 binding protein was found in the heart of SUMO2 transgenic mice, which showed the cardiomyopathy phenotype [41]. These results indicated that SUMO2-mediated SUMOylation maintains a great balance in normal conditions, and the imbalance of SUMOylation leads to certain pathological conditions, including heart failure.

2.4. SENP2

Studies have shown that SENP2 is associated with CHD, cardiac dysfunction, and embryonic development and is a key regulatory factor in early cardiac morphosis [13]. In transgenic mice with cardiac-specific SENP2 expression, overexpressed SENP2 leads to ASDs and VSDs, reduced proliferation of cardiomyocytes, and induced cardiomyopathy with aging [13]. SENP2 deconjugated SUMO from many transcriptional factors that played important roles in cardiac development [36]. SENP2 inhibits the transcriptional activity of NKX2-5 by promoting the uncoupling of SUMO1, affecting cardiac development and function [44]. These results suggested a crucial role of SENP2 in the pathogenesis of CHD.

2.5. SENP3

Interruption of blood supply to the heart is one of the leading causes of disability and death. However, little is known about the molecular events during cardiac ischemia [39] and how these changes function during cell death after reperfusion [45]. SUMOylation of proteins protects cells from several stresses, including ischemia-reperfusion. SENP3 has been reported to be an important determinant factor of cell survival after ischemic infarction [42]. It was found that ischemia-reperfusion significantly reduced the cytoplasmic level of SENP3 and accelerated cell death. It was suggested that SENP3-mediated deSUMOylation plays important roles in regulating myocardial cell survival after ischemic injury.

2.6. SENP5

SENP5 exhibits both endopeptidase and isopeptidase activities [46] and plays important roles in cell proliferation and survival [47,48]. Studies have shown that the level of SENP5, which mainly deconjugates SUMO2/3, is elevated in heart failure [43]. In mitochondria of the normal heart, SUMO1 modified Drp1 to increase its stability [47]. However, in a failing heart, the SENP5 level was increased. The increase of mitochondrial division mediated by Drp1 leads to changes in the morphology and structure of cardiac mitochondria, resulting in mitochondrial dysfunction, thereby inhibiting the proliferation of cardiomyocytes and aggravating apoptosis [43]. These results indicated that SENP5 plays important roles in heart failure and is a potential target for the treatment of heart failure.

3. SUMOylation of Proteins and CVD

3.1. Atherosclerosis

Atherosclerosis is a progressive inflammatory cardiovascular disease characterized by lipid plaque formation in the artery [2,49]. Endothelial cells form the main barrier to vascular permeability, preventing the formation of atherosclerosis [50]. Dysfunction of endothelial cells increases vascular wall permeability, lipid accumulation, and immune cell infiltration in large- and medium-sized artery walls, and subendothelial LDL (low-density lipoprotein) accumulation increases the rate of plaque formation [51,52]. Increasing evidence has shown that SUMOylation plays an important role in regulating inflammation, migration, actin filament remodeling, and apoptosis during the formation of atherosclerosis (Figure 3).



Figure 3. SUMOylation and atherosclerosis. ERK5, NF-κB, *p53*, and PKC were SUMOylated, and SUMOylation of these proteins played important roles in the progression of atherosclerosis. ERK5, extracellular signal-regulated kinase 5; PKC, protein kinase C. S: SUMO.

3.1.1. ERK5 (Extracellular Signal-Regulated Kinase 5)

MAPK (mitogen-activated protein kinases) is a family of serine/threonine kinase composed of four major subfamilies that regulate cellular activities, including cell differentiation, proliferation, and apoptosis. ERK5 (or MAPK7) is one of the most important members of MAPK. ERK5 is not SUMOylated in normal conditions. However, the SUMOylation of ERK5 was induced by d-flow, hyperglycemia, and oxidative stress, and ERK5 SUMOylation induced cell apoptosis and inflammation. SENP2 protects endothelial cells by deSUMOylation of ERK5 to inhibit the pathological change [3]. It has shown that ERK5 activation plays an anti-inflammatory role in the progression of atherosclerosis [53]. It was reported that SUMOylation of ERK5, causing inflammation in the onset and promoting the progression of atherosclerosis [54]. These results suggest that the SUMOylation of ERK5 is one of the most meaningful causes of atherosclerosis, and maintaining the balance of ERK5 SUMOylation is a potential treatment for atherosclerosis.

3.1.2. NF-кВ

NF-κB is a nuclear transcription factor and regulates cytokines and adhesion molecules released by leukocyte accumulation [55]. NF-κB in the cytoplasm binds to the inhibitor of κB (IκB) during the resting state. It was reported that endothelium-specific knockout of NEMO (NF-κB essential modulator) or dominant-negative IκBα inhibits atherosclerotic plaque formation [56]. NF-κB was SUMOylated, and SUMOylation of NF-κB plays a critical role in the pathogenesis of atherosclerosis. SUMO1-mediated SUMOylation of NF-κB inhibits ubiquitination-mediated degradation of IκBα and downregulates NF-κB activation [57]. The modification by SUMO2/3 promoted the detachment of IκBα from NFκB and upregulated NF-κB activation, ultimately inducing atherosclerosis [57,58]. On the other hand, SENP2 negatively regulates inflammation through deSUMOylation of NEMO induced by genotoxic stress and ultimately inhibits atherosclerosis [58]. These results indicated that NF-κB plays important roles in atherosclerosis, and the regulation of NF-κB SUMOylation has become a potential therapeutic target for atherosclerosis.

3.1.3. *p*53

*p*53 is an important tumor suppressor gene that determines cell response to stress conditions, such as apoptosis and cell cycle arrest [59]. When cells are stimulated, nuclear *p*53 is increased, and cell cycle regulators and pro-apoptotic genes are upregulated. The role of *p*53 in atherosclerotic endothelial cells is complicated [60]. In d-flow, PKC mediates the SUMOylation of *p*53 and induces reactive oxygen species (ROS), leading to the transfer of *p*53 from the nucleus to the cytoplasm of endothelial cells. Combined with the anti-apoptotic factor Bax and Bcl-2, SUMOylated *p*53 inhibited the anti-apoptotic and led to apoptosis, ultimately causing the occurrence of atherosclerosis [59]. On the other hand, SENP2 deSUMOylated *p*53 and inhibited atherosclerosis [3]. p90 ribosomal S6 kinase (p90RSK) is a serine/threonine kinase with two kinase functional domains. D-flow activation of *p*53 and ERK5 [61]. These results indicated that deSUMOylation of *p*53 by SENP2 may be an effective strategy to prevent atherosclerosis.

3.1.4. PKC (Protein Kinase C)

PKC isoforms consist of a series of kinases chronically activated by hyperglycemia in diabetic patients and are essential in cell contractility and permeability. PKC is critical in regulating insulin resistance, gluconeogenesis, liver lipid metabolism, and key components of vascular homeostasis and is necessary for atherosclerosis. Changes in the PKC signaling pathway in diabetic patients may lead to various pathological results, thus accelerating DM-mediated atherosclerosis. PKC gene knockout reduced the size of plaques and reduced the production of inflammatory factors [62]. PKC was modified by SUMO1 at K465 and was deSUMOylated by SENP1 [62]. SUMOylation inhibited the function of PKC and played critical roles in the progression of atherosclerosis. The regulation of PKC SUMOylation may become a potential therapeutic target for DM-mediated atherosclerosis.

3.2. CHD (Congenital Heart Disease)

The development of the vertebrate heart begins at the later stage of fetal development and involves a series of complex morphogenetic events, which are strictly regulated by the specific expression of various genes (Table 2) [29]. CHD is a congenital cardiac structural abnormality, and it is a common congenital defect in infants, accounting for about 1% of all newborns [4]. ASDs and VSDs are the most common CHDs, which are characterized by an existing connection between the atria and ventricles [5]. The interaction between different kinds of proteins is the basis of normal cardiac development, and the imbalance of their expression regulated by SUMOylation may lead to changes in protein function and the occurrence of CVD (Figure 4).

Disease	Protein Substrate	Biophysical Function	Biophysical and Biological Effects of SUMOylation
Atherosclerosis	ERK5	ERK5 plays anti-inflammatory role in regulating PPARγ and KLF2 and inhibiting TNFα mediated adhesion gene expression.	SUMOylation of ERK5 significantly inhibits the transcriptional activity of MEF2, causing inflammation in the onset and promoting the progression of atherosclerosis [32].
	NF-ĸB	NF-κB is a nuclear transcription factor and is one of the earliest known cardiac progenitor cell markers.	SUMO1-mediated SUMOylation of NF-κB downregulates its activation, whereas SUMO2/3 leads to the detachment of NF-κB from IκBα and activation [57,58].
	p53	<i>p</i> 53 is an important tumor suppressor gene that regulates cell apoptosis and cell cycle.	SUMOylated <i>p53</i> promotes cell apoptosis and causes the occurrence of atherosclerosis, whereas deSUMOylated <i>p53</i> inhibits atherosclerosis [59].
	РКС	PKC plays an important role in cell growth, apoptosis, inflammation, and contractility of cardiomyocytes.	SUMOylation of PKC inhibits the kinase activity of PKC and accelerates the occurrence of atherosclerosis [4].

Table 2. The roles of SUMOylation of substrates in CVD.

Disease	Protein Substrate	Biophysical Function	Biophysical and Biological Effects of SUMOylation
- - - CHD -	GATA4	GATA4 is one of the most important members of the GATA superfamily and is known to be the primary cause of CHD.	SUMOylation of GATA4 promotes cardiac specific gene expression in pluripotent cells and regulates the development of CHD [63].
	GATA5	GATA5 is an important transcription factor in cardiac development.	SUMOylation of GATA5 restores cardiac insufficiency caused by cardiac dysplasia and plays an important role in cardiac development [64].
	GATA6	GATA6 is one of the most important members of the GATA superfamily and is an important factor in the regulation of cardiac development.	SUMOylation of GATA6 inhibits its transcriptional activity in cardiac development [65].
	MEF2	The MEF2 family contains transcription factors that regulate the activity of muscle-specific genes and involved in cardiac development.	MEF2A, MEF2C, and MEF2D were modified by SUMO, and SUMOylation affects their transcriptional activity [66].
	NKX2-5	NKX2-5 is involved in the differentiation of cardiac progenitor cells and the formation of atrioventricular septum and atrioventricular outflow tract.	SUMOylation promotes the transcriptional activity of NKX2-5 and is associated with the progression of CHD [67].
	SRF	SRF is a member of the DNA binding protein family MADS Box and is involved in cardiac development.	SUMOylation of SRF enhanced activation of c-fos promoter, but inhibited cardiac α -actin promoter, playing critical roles in cardiac development [68].
	TBX2	TBX2 is a T-Box transcription factor and plays vital roles in cardiac cushion development.	TBX2 SUMOylation plays an important role in cardiac development, especially non-atrioventricular myocardium in the atrioventricular and outflow canals [69].
	TBX5	TBX5 is a T-Box transcription factor and plays a critical role in cardiac development.	SUMOylation of TBX5 enhanced the transcriptional activity of TBX5 and enhanced the coordinate regulation with NKX2-5 and GATA4, regulating the progression of cardiac development [70,71].
- HF -	HSF2	HSF2 plays a key role in cardiomyocyte apoptosis and hypertrophy induced by hypertensive angiotensin II.	HSF2 is modified by SUMO1 during HF, and PGCF2 significantly reduced the SUMOylation of HSF2 in spontaneously hypertensive rat hearts [72,73].
	Myocardin	Myocardin regulates the growth of cardiomyocytes by regulating the metabolic activity and plays a key role in the process of cardiomyocyte hypertrophy.	SUMOylation enhanced the activity of myocardin and leads to an increase in cardiac-specific gene expression in cardiomyocytes and promoted myocardin-mediated cardiac hypertrophy [74].
	PARIS	PARIS regulates DNA binding in the promoter region of target genes and plays transcriptional repressor activity.	In cardiac hypertrophy, SUMOylated PARIS inhibited PGC1α transcription to regulate mitochondrial function [75,76].
	PPARy1	PPARγ1 regulates the utilization of cardiac energy substrates.	SUMOylation of PPARγ1 inhibits the transcriptional activity and plays important roles in cardiac hypertrophy [77].
	SERCA2a	SERCA2a regulates the calcium cycle in cardiomyocytes and is responsible for the reuptake of calcium during excitation-contraction coupling.	SUMOylation is essential to maintain the stability and activity of SERCA2a [78,79].
 Ischemic cardiomyopathy 	HDAC4	HDAC4 is a histone deacetylase and is highly expressed in heart, brain, and skeletal muscle.	SUMOylation of HDAC4 mediates ubiquitination and promotes the degradation of HDAC4 in I/R and decreases the production of ROS [80,81].
	HIF1α	$HIF1\alpha$ is an important transcription factor that plays critical roles under hypoxia conditions.	HIF1α SUMOylation regulates the expression of heme oxygenase-1 in rostral ventrolateral medulla (RVLM), whereas its deSUMOylation leads to cell necrosis and apoptosis [82].
	PML	PML is the important organizer of NBs.	PML SUMOylation is critical in PML NBs formation [83] and aggravates ischemia-induced cardiomyocyte apoptosis in vivo [84].

Table 2. Cont.

ERK5, extracellular signal-regulated kinase 5; PKC, protein kinase C; CHD, congenital heart disease; MEF2, myocyte enhancer factor 2; SRF, serum responsive factor; HF, heart failure; HSF2, heat shock transcription factor 2; PARIS, par-kin-interacting substrate; PPAR γ 1, peroxisome proliferation activated receptor γ 1; SERCA2a, sarcoplas-mic/endoplasmic reticulum Ca2+-ATPase 2a; HDAC4, histone deacetylase 4; HIF1 α , hypoxia-inducible factor 1 α ; PML, promyelocytic leukemia.



Figure 4. SUMOylation and CHD. GATA4-6, MEF2, NKX2-5, SRF, TBX2, and TBX5 were SUMOylated, and SUMOylation of these proteins played critical roles in cardiac development, whereas imbalance of SUMOylation caused congenital heart disease. CHD, congenital heart disease; MEF2, myocyte enhancer factor 2; SRF, serum responsive factor. S: SUMO.

3.2.1. GATA4

GATA4 is a transcription factor and one of the most important members of the GATA superfamily of DNA-binding transcription factors, which is reported to be one of the primary causes of CHD [85]. The GATA family plays important roles in regulating cardiac early stage and end-stage development, proliferation and differentiation of cardiac progenitor cells, and cardiac-specific gene expression [86]. GATA4 plays a key role in early heart development as one of the early markers of cardiomyocytes in embryonic development. GATA4 was SUMOylated, and the SUMOylation of GATA4 results in enhanced transcriptional activity and altered nuclear localization [87]. SUMO1 modification of GATA4 on lysine 366 promoted specific gene expression in cardiac pluripotent cells. The SUMOylation of GATA4 may play a role in a complex that regulates a set of genes needed to form cardiac septum. GATA4 mutation was found to be associated with non-syndromic cardiac septal defect [63]. Multiple GATA4 mutations were also found in patients with ASDs and VSDs [88]. Histological analysis suggests that the SUMOylation of GATA4 SUMOylation is a potential therapeutic target for CHD treatment.

3.2.2. GATA5

GATA5 was found to be an important transcriptional factor in zebrafish heart development. GATA5 is reported to be modified by SUMO1 at lysine 324 (K324) and lysine 360 (K360). The SUMOylation sites' mutation affected the transcriptional activity of GATA5 but did not affect the subcellular localization. Studies have shown that K324R mutant GATA5 could rescue the damaged cardiac precursor differentiation, while K360R mutant GATA5 cannot. In addition, the SUMOylation of GATA5 could restore cardiac insufficiency caused by cardiac dysplasia [64]. These results suggest that SUMOylation of GATA5 plays important roles in cardiac development.

3.2.3. GATA6

GATA6 is one of the most important members of the DNA binding transcription factor in the GATA superfamily and is an important factor in the regulation of cardiac development [86]. GATA6 was modified by SUMO1 at lysine 12 (K12), and the specific E3 ligase PIAS1 preferentially promoted the conjugation of SUMO1 to GATA6 via its RING finger domain [20,65]. The SUMOylation of GATA6 inhibited its transcriptional activity but did not affect the subcellular localization [65]. The SUMOylation provides a new regulated mechanism for the modification of GATA6 activity in cardiac development.

3.2.4. MEF2 (Myocyte Enhancer Factor 2)

The MEF2 family is transcription factor that regulates muscle-specific genes involved in cardiac development, including MEF2A, MEF2B, MEF2C, and MEF2D [66]. Reducing the activity of MEF2 inhibits cardiomyocyte differentiation and cardiac development [71]. All these four transcription factors have the SUMO consensus sequence. Previous studies have shown that MEF2A, MEF2C, and MEF2D were modified by SUMO [89]. MEF2A can be modified by SUMO1 in vivo and in vitro. The SUMOylation of MEF2A inhibited its transcriptional activity, but the mutation of lysine 395 to arginine (K395A) in MEF2A enhanced the transcriptional activity of MEF2A [66]. SENP2 enhanced the transcriptional activity of MEF2A in activity dependent-regulation of MEF2A deSUMOylation. Taken together, these results showed that MEF2 played critical roles in embryonic cardiac development.

3.2.5. NKX2-5

NKX2-5 is a cardiac-specific gene that regulates cardiac progenitor cell differentiation, mature and atrioventricular conduction system, and the formation of the atrioventricular outflow [90]. NKX2-5 was SUMOylated by SUMO1 on lysine 51, and SUMO E3 ligase PIASx enhanced the attachment between them [44]. NKX2-5 is modified by SUMO1 and can be modified by SUMO2/3 but to a much lesser degree than SUMO1, and SUMOylation promoted the transcription activity of NKX2-5 [91,92]. SUMOylation regulates NKX2-5 function by changing the interaction between NKX2-5 and related proteins and promotes stable complexes containing NKX2-5. The SUMOylation of NKX2-5 is necessary for normal cardiac development, whereas NKX2-5 mutation was associated with forming a variety of CHD [92]. These results indicate that NKX2-5 SUMOylation plays an important role in CHD and becomes a therapeutic target for CHD.

3.2.6. SRF (Serum Responsive Factor)

SRF is a DNA binding protein family known as MADS Box, which has a highly conserved DNA binding/dimerization [93]. In vertebrates, SRF is mainly expressed in mesoderm and neuroectoderm tissues. SRF plays an important role during cardiac development, particularly in cardiogenesis and mesoderm development [94]. Serum stimulation activates SRF-mediated early gene transcription through various signal transduction pathways, such as newborn heart and smooth muscle-related genes [95]. SRF deficiency decreased cell proliferation and survival rate and increased cell apoptosis [96]. Studies have shown that SRF was SUMOylated by SUMO1 on its lysine residue 147 (K147), and the SUMOylation of SRF enhanced activation of the c-fos promoter but inhibited cardiac α -actin promoter [95]. These results suggest that the regulation of SRF SUMOylation plays critical roles in cardiac development.

3.2.7. TBX2

TBX2 is a member of the T-Box transcription factors family and plays vital roles in cardiac cushion development [97]. Changes in TBX2 activity are associated with a variety of human congenital diseases and cancers. Changes in numerous factors associated with CHD are accompanied by changes in TBX2 [98]. It was reported that common genetic variation in the TBX2 promoter region is strongly associated with the cardiac development and occurrence of CHD [99]. TBX2 is associated with the non-atrioventricular myocardium in

the atrioventricular and outflow canals during cardiac development. TBX2 was identified to be modified by SUMO1, and TBX2 SUMOylation plays an important role in cardiac development, especially non-atrioventricular myocardium in the atrioventricular and outflow canals [100]. Therefore, the SUMOylation of TBX2 plays an important role in cardiac development.

3.2.8. TBX5

TBX5 is a member of the T-Box transcription factor family and plays a critical role in cardiac development [101]. In the early development of the heart, TBX5 acts as an upstream transcriptional activator to promote cardiomyocyte maturation [102]. In the late stage of heart development, TBX5 acts on the heart conduction system and maintains the function of mature cardiomyocytes [70,103]. TBX5 was SUMOylated by SUMO1, and SUMOylation enhanced the transcriptional activity of TBX5 [104]. The E3 ligase PIAS1 specifically enhanced the SUMOylation of TBX5 and enhanced the coordinate regulation with NKX2-5 and GATA4 [104]. These studies suggest the important roles of TBX5 in the progression of cardiac development.

3.3. HF (Heart Failure)

HF is a complex disease caused by valve abnormality, coronary artery disease, hypertension, and other pathological changes [6]. Cardiac energy metabolism and mitochondrial biogenesis defects lead to HF and cardiac hypertrophy [7]. Cardiac hypertrophy is a transitional stage of HF that initiates a process of molecular and structural changes and lead to ventricular remodeling. Initially, persistent cardiac hypertrophy is an adaptive response and increases the risk of HF and sudden death [105]. Treatment with drugs for CVD is effective in reducing left ventricular hypertrophy and related mortality [106], and the SUMOylation pathway is involved in the development of HF and progression of cardiac hypertrophy (Figure 5).



Figure 5. SUMOylation and HF. HSF2, myocardin, PARIS, PPAR γ 1, and SERCA2a were SUMOylated, and SUMOylation of these proteins played crucial roles in the progression of heart failure. HF, heart failure; HSF2, heat shock transcription factor 2; PARIS, parkin-interacting substrate; PPAR γ 1, peroxisome proliferation activated receptor γ 1; SERCA2a, sarcoplasmic/endoplasmic reticulum Ca²⁺-ATPase 2a. S: SUMO.

3.3.1. HSF2 (Heat Shock Transcription Factor 2)

Cardiac hypertrophy is a major feature of early hypertensive heart failure. Insulinlike growth factor II receptor (IGF-IIR) plays key roles in cardiomyocyte hypertrophy induced by hypertensive angiotensin II [72]. IGF-IIR is regulated by HSF2, which is modified by SUMO1 during HF [72]. PGCF2 (polycomb group ring finger 2) significantly reduced the SUMOylation of HSF2 in spontaneously hypertensive rat hearts. Inhibition of HSF2 SUMOylation induced cardiac hypertrophy in hypertensive rats. The treatment by angiotensin-receptor I blocker (ARB) restored the SUMOylation of HSF2 and alleviated cardiac defects in spontaneously hypertensive rats. The studies showed that the PGCF2-SUMO1-HSF2-IGF-IIR pathway had a profound effect on cardiac hypertrophy in patients with hypertensive HF [72]. These results suggested that the regulation of HSF2 SUMOylation is a potential target for the treatment of cardiac hypertrophy.

3.3.2. Myocardin

Myocardin belongs to the SAP superfamily and is only expressed in cardiac smooth muscle during embryonic development and plays a key role in cardiomyocyte hypertrophy [107]. Myocardin affects the growth of cardiomyocytes by regulating the stability and metabolic activity of cardiomyocytes [108]. Myocardin promotes smooth muscle cell differentiation with a coactivator of SRF [109]. Loss of function of myocardin leads to serious defects in vascular smooth muscle development [110]. Abnormal expression of myocardin leads to hypertrophic cardiomyopathy and HF, and its mutation may lead to cardiovascular abnormalities in humans. It was reported that myocardin was SUMOylated by SUMO1 in K445, and SUMOylation of myocardin by SUMO1 and E3 PIAS1 reversed the expression of myocardial genes and enhanced the cardiac hypertrophy of primary neonatal cardiomyocytes [74]. These results indicated the critical roles of myocardin SUMOylation in the development of cardiac hypertrophy.

3.3.3. PARIS (Parkin-Interacting Substrate)

PARIS is regulated by the ubiquitin proteasome system via the E3 ubiquitin ligase parkin [111]. PARIS is a transcriptional repressor containing four zinc finger domains and one KRAB domain to regulate DNA binding in the promoter region of target genes and play transcriptional repressor activity [112]. PARIS was SUMOylated by SUMO1 in K189 and K286 and was deSUMOylated by DJ-1 [75,76]. In cardiac hypertrophy, PARIS was SUMOylated and inhibited PGC1 α transcription. As one of the major regulators of mitochondrial function and biogenesis, the PGC1 α transcription decrease causes mitochondrial dysfunction and ROS accumulation [113]. The accumulation of ROS in cardiac hypertrophy induced the degradation of DJ-1 and augmented the SUMOylation of PARIS, and reduced the promoter activity of PGC1 α [114]. These results indicated the critical roles of PARIS SUMOylation in the regulation of mitochondrial function, and inhibition of the SUMOylation of PARIS may be a potential treatment of cardiac hypertrophy.

3.3.4. PPAR γ 1 (Peroxisome Proliferation-Activated Receptor γ 1)

The utilization of cardiac energy is partially determined by the PPAR transcription factors, including PPAR α , PPAR β , and PPAR γ , and the heterodimer-forming RXR α as well as coactivator PGC-1 α and inhibitor NCoR (nuclear receptor corepressor). The PPAR family regulates the transcription of genes related to energy regulation and lipid metabolism [115]. PPAR γ 1 plays critical roles in the development of myocardial fibrosis and the protection of cardiac oxidative stress. PPAR γ 1 was covalently modified by SUMO1 in K107 and K365 with E3 ligase PIAS1 and PIAS $\alpha\beta$, and SUMOylation inhibited the transcriptional activity of PPAR γ 1 [77,116]. The SUMOylation of PPAR γ 1 regulated NF- κ B in response to hypertrophy and inflammatory stimuli, suggesting a protective role of PPAR γ 1 SUMOylation in cardiac hypertrophy [115]. These results indicated that the SUMOylation of PPAR γ 1 is a potential target for the treatment of cardiac hypertrophy.

3.3.5. SERCA2a (Sarcoplasmic/Endoplasmic Reticulum Ca²⁺-ATPase 2a)

SERCA2a is a key pump that regulates the calcium cycle in cardiomyocytes and is responsible for the reuptake of calcium during excitation-contraction coupling [117]. Reduced activity and expression of SERCA2a are markers of HF in the body [118,119]. It has been demonstrated that SERCA2a can be modified by SUMO1 at lysine 480 and 585, and SUMOylation is essential to maintain the stability and activity of SERCA2a [78]. The content of SUMO1 and SUMOylated SERCA2a was greatly reduced in heart failure. After the knockout of SUMO1, the activity of SERCA2a was also reduced, which accelerated the impairment of heart function and the death of animals [120]. After restoring SUMO1 levels, SERCA2a levels rose, and the heart function of mice and swine went into remission [121,122]. SUMOylation of SERCA2a increased the stability and activity of SERCA2a, and the contractility of the myocardium increased. The SUMOylation of SERCA2a is of great significance for the treatment of HF [123]. There is a small molecule, N106, that can activate E1 ligase and trigger the SUMOylation of SERCA2a and enhance the contractility of cardiomyocytes and improve ventricular function in heart failure mice [124]. These studies indicate that the SUMOylation of SERCA2a is a potential target and a promising treatment for HF.

3.4. Ischemic Cardiomyopathy

AMI (acute myocardial infarction) is ischemic heart disease with acute hypoxia in the myocardial tissue and is one of the leading causes of death [125]. Restoration of reperfusion is the main therapeutic method to limit the size of myocardial infarction, save the viable myocardium, maintain the systolic function of the heart, and delay the occurrence of HF [14,82]. Readjustment of blood flow restores oxygen and nutrient supply but may damage the cardiac muscle and induce myocardial I/R injury. Studies of animal models of AMI have shown that fatal reperfusion injury accounts for half of the myocardial infarction [126]. However, the underlying mechanisms of myocardial I/R injury have not been fully elucidated [127]. SUMOylation has been found to play a significant role in myocardial I/R injury, and several SUMOylated proteins interfere with multiple mechanisms of myocardial I/R injury (Figure 6).



Figure 6. SUMOylation and ischemic cardiomyopathy. HDAC4, HIF1 α , and PML were SUMOylated, and SUMOylation of these proteins played essential roles in developing ischemic cardiomyopathy. HDAC4, histone deacetylase 4; HIF1 α , hypoxia-inducible factor 1 α ; PML, promyelocytic leukemia. S: SUMO.

3.4.1. HDAC4 (Histone Deacetylase 4)

HDAC induces deacetylation, chromatin concentration, and transcription inhibition [128]. According to its function and sequence homology, HDAC can be divided into four types. HDAC4 belongs to class II HDACs and is distributed in the nucleus and cytoplasm. HDAC4 is highly expressed in skeletal muscle, brain, and heart [129–131]. HDAC4 contributes to myocardial I/R injury due to mitochondrial damage and reactive oxygen species (ROS) during reperfusion. Inhibiting HDAC4 expression promoted cardiac repair, improved cardiac function, and inhibited cardiac remodeling [132]. HDAC4 was modified by SUMO1, and SUMOylation of HDAC4 reduced the generation of ROS [80]. The SUMOylation of HDAC4 mediates ubiquitination and promotes the degradation of HDAC4 in I/R, which may be related to the decrease of ROS production after the degradation of HDAC4 [80]. ROS-mediated oxidative stress has been widely recognized as one of the important mechanisms of myocardial I/R injury [133]. These results indicate that the SUMOylation of HDAC4 plays important roles and is a potential therapeutic target for myocardial I/R injury.

3.4.2. HIF1 α (Hypoxia-Inducible Factor 1 α)

HIF1 α is an important transcription factor that plays a critical role in protecting the I/R process by participating in the transcription response under hypoxia conditions. HIF1 α was modified by SUMO1 to regulate the expression of heme oxygenase-1 in the rostral ventrolateral medulla (RVLM), playing a critical role in the regulation of brain death [8]. SENP1-mediated deSUMOylation of HIF1 α leads to cell necrosis and apoptosis and plays an indispensable role in myocardial infarction after I/R in response to cell hypoxia. SUMOylation of HIF1 α reduces its stability and leads to degradation, whereas SENP1 deSUMOylates HIF1 α and improves its stability [134,135]. These studies showed that the decrease of SENP1 levels aggravated the I/R injury of cardiomyocytes by the HIF1 α -dependent pathway, and the overexpression of HIF1 α reversed the worsening effect of SENP1 downregulation on cell death [82]. These results suggest that the SUMOylation of HIF1 α plays an important role in I/R injury and might be a potential target of AMI treatment.

3.4.3. PML (Promyelocytic Leukemia)

PML is the important organizer of nuclear bodies (NBs) and is expressed in several isoforms, including PML I to VII [136]. PML is SUMOylated by SUMO1 and SUMO2/3 and de-SUMOylated by SENP3 [42,137], and SUMOylation is critical in PML NBs formation [83]. Human ring-finger protein 4 (RNF4) is an E3 ubiquitin ligase and targets poly-SUMOylated proteins to degrade by the ubiquitin-proteasome pathway [138]. RNF4 is involved in controlling the degradation of PML and the transcriptional activity of several transcription factors [139,140]. PML is an ROS sensor, and myocardial infarction (MI) generated ROS, and ROS induced the increase of PML SUMOylation and PML-NB formation, leading to p53-dependent cell apoptosis. Overexpression of PML or knockdown of RNF4 enhances p53 recruitment and activation of PML SUMOylation and PML NBs accumulation, which aggravated ischemia-induced cardiomyocyte apoptosis in vivo and cell injury [84]. These results indicated the important roles of PML SUMOylation under oxidative stress is a potential treatment of MI.

3.4.4. Target Protein Network

It has been reported that SUMOylation is enhanced to protect the brain during cardiac ischemia [141,142]. To identify the potential target proteins of SUMOylation in the mouse heart with cardiac I/R, an unbiased proteomic analysis was conducted. There were 102 proteins that significantly interacted with SUMO1, including the composition of the SUMO mechanism and the ubiquitin proteasome system [143,144], multiple subunits of 20S proteasome core particle, and metabolic enzymes involved in glycolytic and oxidative metabolic

pathways [145]. High molecular weight SUMO1 conjugates were significantly reduced 24 h after reperfusion. In total, 15 proteins showed significant changes in SUMOylation at ischemia, and 13 and 12 proteins showed changes in SUMOylation at 1 and 24 h after reperfusion, respectively. Further, 55 proteins were found to be potential SUMO2/3 target proteins, including ubiquitin and proteasome subunits, gene expression regulators and transcription factors, RNA binding proteins and splicing regulators, heat shock proteins, and specific proteins that play important roles in cardiac function. Myocardial ischemia decreased the SUMO level of the whole protein and the number and catalytic activity of de-SUMO enzymes [145]. The heart SUMO2 target protein undergoes a dynamic adaptation process in cardiac I/R. Only one protein was SUMOylated differently at ischemia, and 8 and 11 target proteins were significantly altered 1 and 24 h after reperfusion, respectively. SENP3 is the major deSUMOylation during cardiac I/R and provided potential therapeutic targets for the treatment of cardiac ischemia.

4. Concluding Remarks and Perspective

Cardiovascular development and function are regulated by complex factors and signal pathways, and cardiovascular disorders lead to a series of stress responses [146,147]. SUMOylation is the PTM of proteins by a series of enzyme-linked reactions. Acute cardiac hypoxia induces elevated late sodium current and action potential prolongation in the heart and causes pro-arrhythmic change by SUMOylation of Nav1.5 channels [148]. In the decades since its discovery, studies related to SUMOylation in a variety of diseases have been extensively reported, including cardiovascular development and function [29,30].

Many factors regulated by SUMOylation and the imbalance of protein SUMOylation are involved in different types of CVDs, including atherosclerosis, CHD, HF, and ischemic cardiomyopathy. The previous studies indicated that SUMOylation is closely related to the occurrence and development of CVDs (Figure 7). The disruption of SUMOylation balance leads to severe CVDs. However, the molecular mechanism of SUMOylation in CVD has been preliminarily studied. It is possible to determine endogenous SUMOylated proteins under native conditions and identify endogenous SUMOylation sites on factors important in cardiac functions [149,150]. More SUMOylated proteins will be discovered during cardiovascular development and maintenance of cardiac function with further studies. Although the therapeutic strategies of CVDs have been greatly improved, there is still a lack of accurate, effective, and scientific unified treatment. Target therapy is one favorable way for future medical development and destination of CVD treatment. SUMOylation not only regulates the function of target proteins but also affects other post-translational modification processes. It is possible that SUMOylation co-regulates the same physiological process with other post-translational modifications. Therefore, the synergistic effects of SUMOylation and other post-translational modifications on the progression of CVD should be considered in the clinical treatment. Exploring the dynamic balance between the target of SUMOvlation and also other PTM will help to better understand the biological functions of SUMOylation and the clinical treatment of CVDs.



Figure 7. The SUMOylation of substrates and CVD. SUMOylation of target proteins played critical roles and showed potential clinical implications in CVD, including atherosclerosis, congenital heart disease, heart failure, and ischemic cardiomyopathy. ERK5, extracellular signal-regulated kinase 5; PKC, protein kinase C; MEF2, myocyte enhancer factor 2; SRF, serum responsive factor; HSF2, heat shock transcription factor 2; PARIS, parkin-interacting substrate; PPAR γ 1, peroxisome proliferation activated receptor γ 1; SERCA2a, sarcoplasmic/endoplasmic reticulum Ca²⁺-ATPase 2a; HDAC4, histone deacetylase 4; HIF1 α , hypoxia-inducible factor 1 α ; PML, promyelocytic leukemia.

Author Contributions: Y.Q. and H.W. conceived the idea for the review. C.D. performed the retrieval and collection of relevant literatures. X.C., Q.S., W.L., Q.W., H.Y., Z.Z., and X.W. provided suggestions. C.D., H.W., and Y.Q. wrote the paper. C.D. designed and prepared the figures. All authors have read and agreed to the published version of the manuscript.

Funding: This research was funded by the National Natural Science Foundation of China (81671294 and 81870241 to Y.Q.) and the Fundamental Research Funds for the Central Universities (GK201903066 to H.W.).

Institutional Review Board Statement: Not applicable.

Informed Consent Statement: Not applicable.

Acknowledgments: We are thankful for the financial support of the National Natural Science Foundation of China (81671294 and 81870241 to Y.Q.) and the Fundamental Research Funds for the Central Universities (GK201903066 to H.W.).

Conflicts of Interest: The authors declare no conflict of interest.

References

- Abe, J.I.; Sandhu, U.G.; Hoang, N.M.; Thangam, M.; Quintana-Quezada, R.A.; Fujiwara, K.; Le, N.T. Coordination of Cellular Localization-Dependent Effects of Sumoylation in Regulating Cardiovascular and Neurological Diseases. *Adv. Exp. Med. Biol.* 2017, 963, 337–358. [PubMed]
- Dehnavi, S.; Sadeghi, M.; Penson, P.E.; Banach, M.; Jamialahmadi, T.; Sahebkar, A. The Role of Protein SUMOylation in the Pathogenesis of Atherosclerosis. J. Clin. Med. 2019, 8, 1856. [CrossRef] [PubMed]
- Heo, K.S.; Chang, E.; Le, N.T.; Cushman, H.; Yeh, E.T.; Fujiwara, K.; Abe, J. De-SUMOylation enzyme of sentrin/SUMOspecific protease 2 regulates disturbed flow-induced SUMOylation of ERK5 and p53 that leads to endothelial dysfunction and atherosclerosis. *Circ. Res.* 2013, *112*, 911–923. [CrossRef]

- Ahuja, P.; Sdek, P.; MacLellan, W.R. Cardiac myocyte cell cycle control in development, disease, and regeneration. *Physiol. Rev.* 2007, 87, 521–544. [CrossRef] [PubMed]
- 5. Henning, R.J. Diagnosis and treatment of adults with congenital heart disease. *Future Cardiol.* **2020**, *16*, 317–342. [CrossRef] [PubMed]
- 6. Bui, A.L.; Horwich, T.B.; Fonarow, G.C. Epidemiology and risk profile of heart failure. Nat. Rev. Cardiol. 2011, 8, 30-41. [CrossRef]
- Mendler, L.; Braun, T.; Muller, S. The Ubiquitin-Like SUMO System and Heart Function: From Development to Disease. *Circ. Res.* 2016, 118, 132–144. [CrossRef]
- Chan, J.Y.; Tsai, C.Y.; Wu, C.H.; Li, F.C.; Dai, K.Y.; Sun, E.Y.; Chan, S.H.; Chang, A.Y. Sumoylation of hypoxia-inducible factor-1alpha ameliorates failure of brain stem cardiovascular regulation in experimental brain death. *PLoS ONE* 2011, 6, e17375. [CrossRef]
- 9. Ulrich, H.D. Regulating post-translational modifications of the eukaryotic replication clamp PCNA. *DNA Repair.* **2009**, *8*, 461–469. [CrossRef]
- 10. Yeh, E.T. SUMOylation and De-SUMOylation: Wrestling with life's processes. J. Biol. Chem. 2009, 284, 8223-8227. [CrossRef]
- 11. Matunis, M.J.; Coutavas, E.; Blobel, G. A novel ubiquitin-like modification modulates the partitioning of the Ran-GTPaseactivating protein RanGAP1 between the cytosol and the nuclear pore complex. *J. Cell Biol.* **1996**, *135 Pt 1*, 1457–1470. [CrossRef]
- 12. Mahajan, R.; Delphin, C.; Guan, T.; Gerace, L.; Melchior, F. A small ubiquitin-related polypeptide involved in targeting RanGAP1 to nuclear pore complex protein RanBP2. *Cell* **1997**, *88*, 97–107. [CrossRef]
- 13. Kim, E.Y.; Chen, L.; Ma, Y.; Yu, W.; Chang, J.; Moskowitz, I.P.; Wang, J. Enhanced desumoylation in murine hearts by overexpressed SENP2 leads to congenital heart defects and cardiac dysfunction. *J. Mol. Cell. Cardiol.* **2012**, *52*, 638–649. [CrossRef] [PubMed]
- 14. Zhang, L.; Liu, X.; Sheng, H.; Liu, S.; Li, Y.; Zhao, J.Q.; Warner, D.S.; Paschen, W.; Yang, W. Neuron-specific SUMO knockdown suppresses global gene expression response and worsens functional outcome after transient forebrain ischemia in mice. *Neuroscience* **2017**, *343*, 190–212. [CrossRef] [PubMed]
- 15. Treuter, E.; Gustafsson, J.A. Wrestling rules in transrepression: As easy as SUMO-1, -2, -3? Mol. Cell 2007, 25, 178–180. [CrossRef]
- Bohren, K.M.; Nadkarni, V.; Song, J.H.; Gabbay, K.H.; Owerbach, D. A M55V polymorphism in a novel SUMO gene (SUMO-4) differentially activates heat shock transcription factors and is associated with susceptibility to type I diabetes mellitus. *J. Biol. Chem.* 2004, 279, 27233–27238. [CrossRef]
- 17. Boulanger, M.; Paolillo, R.; Piechaczyk, M.; Bossis, G. The SUMO Pathway in Hematomalignancies and Their Response to Therapies. *Int. J. Mol. Sci.* 2019, 20, 3895. [CrossRef]
- 18. Mukhopadhyay, D.; Dasso, M. Modification in reverse: The SUMO proteases. Trends Biochem. Sci. 2007, 32, 286–295. [CrossRef]
- Cheng, J.; Kang, X.L.; Zhang, S.; Yeh, E.T.H. SUMO-Specific protease 1 is essential for stabilization of HIF1 alpha during hypoxia. *Cell* 2007, 131, 584–595. [CrossRef]
- Kang, X.L.; Qi, Y.T.; Zuo, Y.; Wang, Q.; Zou, Y.Q.; Schwartz, R.J.; Cheng, J.K.; Yeh, E.T.H. SUMO-Specific Protease 2 Is Essential for Suppression of Polycomb Group Protein-Mediated Gene Silencing during Embryonic Development. *Mol. Cell* 2010, *38*, 191–201. [CrossRef]
- 21. Qi, Y.T.; Zuo, Y.; Yeh, E.T.H.; Cheng, J.K. An Essential Role of Small Ubiquitin-like Modifier (SUMO)-specific Protease 2 in Myostatin Expression and Myogenesis. *J. Biol. Chem.* **2014**, *289*, 3288–3293. [CrossRef]
- Qi, Y.T.; Wang, J.X.; Bomben, V.C.; Li, D.P.; Chen, S.R.; Sun, H.; Xi, Y.T.; Reed, J.G.; Cheng, J.K.; Pan, H.L.; et al. Hyper-SUMOylation of the Kv7 Potassium Channel Diminishes the M-Current Leading to Seizures and Sudden Death. *Neuron* 2014, *83*, 1159–1171. [CrossRef]
- Chen, X.; Zhang, S.; Huang, J.; Dong, W.; Xiao, H.; Shao, H.; Cheng, J.; Wu, H.; Qi, Y. Hyper-SUMOylation of K(+) Channels in Sudden Unexplained Death in Epilepsy: Isolation and Primary Culture of Dissociated Hippocampal Neurons from Newborn Mice for Subcellular Localization. *Methods Mol. Biol.* 2018, 1684, 63–71.
- 24. Wu, H.; Chen, X.; Cheng, J.; Qi, Y. SUMOylation and Potassium Channels: Links to Epilepsy and Sudden Death. *Adv. Protein Chem. Struct. Biol.* 2016, 103, 295–321.
- 25. Smits, V.A.; Freire, R. USP7/HAUSP: A SUMO deubiquitinase at the heart of DNA replication. *Bioessays* 2016, 38, 863–868. [CrossRef]
- 26. Bertke, M.M.; Dubiak, K.M.; Cronin, L.; Zeng, E.; Huber, P.W. A deficiency in SUMOylation activity disrupts multiple pathways leading to neural tube and heart defects in Xenopus embryos. *BMC Genom.* **2019**, *20*, 386. [CrossRef]
- 27. Hay, R.T. SUMO: A history of modification. Mol. Cell 2005, 18, 1-12. [CrossRef]
- 28. Johnson, E.S. Protein modification by SUMO. Annu. Rev. Biochem. 2004, 73, 355–382. [CrossRef]
- Gao, J.; Shao, K.; Chen, X.; Li, Z.; Liu, Z.; Yu, Z.; Aung, L.H.H.; Wang, Y.; Li, P. The involvement of post-translational modifications in cardiovascular pathologies: Focus on SUMOylation, neddylation, succinylation, and prenylation. *J. Mol. Cell. Cardiol.* 2020, 138, 49–58. [CrossRef]
- Zhang, L.; Yang, T.H.; Li, D.W. Roles of SUMOylation in Heart Development and Cardiovascular Diseases. Curr. Mol. Med. 2017, 16, 877–884. [CrossRef]
- 31. McMurray, J.J.; Smith, G.L. Calcium handling in the failing heart and SUMO—Weighing the evidence. *N. Engl. J. Med.* **2011**, *365*, 1738–1739. [CrossRef]
- 32. Abe, J.; Manabe, I.; Aikawa, M.; Aikawa, E. Cardiovascular Inflammation 2012: Reactive Oxygen Species, SUMOylation, and Biomarkers in Cardiovascular Inflammation. *Int. J. Inflam.* **2013**, 2013, 953463. [CrossRef] [PubMed]

- 33. Gupta, M.K.; McLendon, P.M.; Gulick, J.; James, J.; Khalili, K.; Robbins, J. UBC9-Mediated Sumoylation Favorably Impacts Cardiac Function in Compromised Hearts. *Circ. Res.* **2016**, *118*, 1894–1905. [CrossRef] [PubMed]
- 34. Nacerddine, K.; Lehembre, F.; Bhaumik, M.; Artus, J.; Cohen-Tannoudji, M.; Babinet, C.; Pandolfi, P.P.; Dejean, A. The SUMO pathway is essential for nuclear integrity and chromosome segregation in mice. *Dev. Cell* **2005**, *9*, 769–779. [CrossRef]
- 35. Lee, Y.J.; Mou, Y.; Maric, D.; Klimanis, D.; Auh, S.; Hallenbeck, J.M. Elevated global SUMOylation in Ubc9 transgenic mice protects their brains against focal cerebral ischemic damage. *PLoS ONE* **2011**, *6*, e25852. [CrossRef]
- 36. Wang, J.; Chen, L.; Wen, S.; Zhu, H.; Yu, W.; Moskowitz, I.P.; Shaw, G.M.; Finnell, R.H.; Schwartz, R.J. Defective sumoylation pathway directs congenital heart disease. *Birth Defects Res. A Clin. Mol. Teratol.* **2011**, *91*, 468–476. [CrossRef]
- Pai, P.; Shibu, M.A.; Chang, R.L.; Yang, J.J.; Su, C.C.; Lai, C.H.; Liao, H.E.; Viswanadha, V.P.; Kuo, W.W.; Huang, C.Y. ERbeta targets ZAK and attenuates cellular hypertrophy via SUMO-1 modification in H9c2 cells. *J. Cell. Biochem.* 2018, 119, 7855–7864. [CrossRef]
- Lee, A.; Jeong, D.; Mitsuyama, S.; Oh, J.G.; Liang, L.; Ikeda, Y.; Sadoshima, J.; Hajjar, R.J.; Kho, C. The role of SUMO-1 in cardiac oxidative stress and hypertrophy. *Antioxid. Redox Signal.* 2014, 21, 1986–2001. [CrossRef] [PubMed]
- Haindl, M.; Harasim, T.; Eick, D.; Muller, S. The nucleolar SUMO-specific protease SENP3 reverses SUMO modification of nucleophosmin and is required for rRNA processing. *EMBO Rep.* 2008, *9*, 273–279. [CrossRef]
- 40. Kim, Y.R.; Jacobs, J.S.; Li, Q.; Gaddam, R.R.; Vikram, A.; Liu, J.; Kassan, M.; Irani, K.; Kumar, S. SUMO2 regulates vascular endothelial function and oxidative stress in mice. *Am. J. Physiol. Heart Circ. Physiol.* **2019**, 317, H1292–H1300. [CrossRef]
- 41. Kim, E.Y.; Zhang, Y.; Ye, B.; Segura, A.M.; Beketaev, I.; Xi, Y.; Yu, W.; Chang, J.; Li, F.; Wang, J. Involvement of activated SUMO-2 conjugation in cardiomyopathy. *Biochim. Biophys. Acta* 2015, *1852*, 1388–1399. [CrossRef] [PubMed]
- Han, Y.; Huang, C.; Sun, X.; Xiang, B.; Wang, M.; Yeh, E.T.; Chen, Y.; Li, H.; Shi, G.; Cang, H.; et al. SENP3-mediated de-conjugation of SUMO2/3 from promyelocytic leukemia is correlated with accelerated cell proliferation under mild oxidative stress. *J. Biol. Chem.* 2010, 285, 12906–12915. [CrossRef] [PubMed]
- 43. Kim, E.Y.; Zhang, Y.; Beketaev, I.; Segura, A.M.; Yu, W.; Xi, Y.; Chang, J.; Wang, J. SENP5, a SUMO isopeptidase, induces apoptosis and cardiomyopathy. J. Mol. Cell. Cardiol. 2015, 78, 154–164. [CrossRef]
- 44. Wang, J.; Zhang, H.; Iyer, D.; Feng, X.H.; Schwartz, R.J. Regulation of cardiac specific nkx2.5 gene activity by small ubiquitin-like modifier. *J. Biol. Chem.* 2008, 283, 23235–23243. [CrossRef]
- 45. Rawlings, N.; Lee, L.; Nakamura, Y.; Wilkinson, K.A.; Henley, J.M. Protective role of the deSUMOylating enzyme SENP3 in myocardial ischemia-reperfusion injury. *PLoS ONE* **2019**, *14*, e0213331. [CrossRef] [PubMed]
- 46. Gong, L.; Yeh, E.T. Characterization of a family of nucleolar SUMO-specific proteases with preference for SUMO-2 or SUMO-3. *J. Biol. Chem.* **2006**, *281*, 15869–15877. [CrossRef]
- Zunino, R.; Schauss, A.; Rippstein, P.; Andrade-Navarro, M.; McBride, H.M. The SUMO protease SENP5 is required to maintain mitochondrial morphology and function. J. Cell Sci. 2007, 120 Pt 7, 1178–1188. [CrossRef]
- 48. Zunino, R.; Braschi, E.; Xu, L.; McBride, H.M. Translocation of SenP5 from the nucleoli to the mitochondria modulates DRP1dependent fission during mitosis. *J. Biol. Chem.* **2009**, *284*, 17783–17795. [CrossRef]
- 49. Bjorkbacka, H.; Nilsson, J. Innate immunity in atherosclerosis. J. Innate Immun. 2010, 2, 305–306. [CrossRef] [PubMed]
- 50. Heo, K.S.; Lee, H.; Nigro, P.; Thomas, T.; Le, N.T.; Chang, E.; McClain, C.; Reinhart-King, C.A.; King, M.R.; Berk, B.C.; et al. PKCzeta mediates disturbed flow-induced endothelial apoptosis via p53 SUMOylation. *J. Cell Biol.* **2011**, *193*, 867–884. [CrossRef]
- 51. Nicorescu, I.; Dallinga, G.M.; de Winther, M.P.J.; Stroes, E.S.G.; Bahjat, M. Potential epigenetic therapeutics for atherosclerosis treatment. *Atherosclerosis* 2019, 281, 189–197. [CrossRef] [PubMed]
- 52. Lozano, R.; Naghavi, M.; Foreman, K.; Lim, S.; Shibuya, K.; Aboyans, V.; Abraham, J.; Adair, T.; Aggarwal, R.; Ahn, S.Y.; et al. Global and regional mortality from 235 causes of death for 20 age groups in 1990 and 2010: A systematic analysis for the Global Burden of Disease Study 2010. *Lancet* 2012, *380*, 2095–2128. [CrossRef]
- Woo, C.H.; Shishido, T.; McClain, C.; Lim, J.H.; Li, J.D.; Yang, J.; Yan, C.; Abe, J. Extracellular signal-regulated kinase 5 SUMOylation antagonizes shear stress-induced antiinflammatory response and endothelial nitric oxide synthase expression in endothelial cells. *Circ. Res.* 2008, 102, 538–545. [CrossRef]
- 54. Sato, M.; Kawai-Kowase, K.; Sato, H.; Oyama, Y.; Kanai, H.; Ohyama, Y.; Suga, T.; Maeno, T.; Aoki, Y.; Tamura, J.; et al. c-Src and hydrogen peroxide mediate transforming growth factor-beta1-induced smooth muscle cell-gene expression in 10T1/2 cells. *Arter. Thromb. Vasc. Biol.* **2005**, *25*, 341–347. [CrossRef]
- 55. Bornfeldt, K.E.; Tabas, I. Insulin resistance, hyperglycemia, and atherosclerosis. *Cell Metab.* **2011**, *14*, 575–585. [CrossRef] [PubMed]
- Gareus, R.; Kotsaki, E.; Xanthoulea, S.; van der Made, I.; Gijbels, M.J.; Kardakaris, R.; Polykratis, A.; Kollias, G.; de Winther, M.P.; Pasparakis, M. Endothelial cell-specific NF-kappaB inhibition protects mice from atherosclerosis. *Cell Metab.* 2008, *8*, 372–383. [CrossRef]
- Chang, E.; Abe, J.I. Kinase-SUMO networks in diabetes-mediated cardiovascular disease. *Metabolism* 2016, 65, 623–633. [CrossRef] [PubMed]
- 58. Desterro, J.M.; Rodriguez, M.S.; Hay, R.T. SUMO-1 modification of IkappaBalpha inhibits NF-kappaB activation. *Mol. Cell* **1998**, 2, 233–239. [CrossRef]
- 59. Mihara, M.; Erster, S.; Zaika, A.; Petrenko, O.; Chittenden, T.; Pancoska, P.; Moll, U.M. p53 has a direct apoptogenic role at the mitochondria. *Mol. Cell* **2003**, *11*, 577–590. [CrossRef]

- 60. Teodoro, J.G.; Parker, A.E.; Zhu, X.; Green, M.R. p53-mediated inhibition of angiogenesis through up-regulation of a collagen prolyl hydroxylase. *Science* **2006**, *313*, 968–971. [CrossRef]
- Heo, K.S.; Le, N.T.; Cushman, H.J.; Giancursio, C.J.; Chang, E.; Woo, C.H.; Sullivan, M.A.; Taunton, J.; Yeh, E.T.; Fujiwara, K.; et al. Disturbed flow-activated p90RSK kinase accelerates atherosclerosis by inhibiting SENP2 function. *J. Clin. Investig.* 2015, 125, 1299–1310. [CrossRef]
- 62. Tabit, C.E.; Shenouda, S.M.; Holbrook, M.; Fetterman, J.L.; Kiani, S.; Frame, A.A.; Kluge, M.A.; Held, A.; Dohadwala, M.M.; Gokce, N.; et al. Protein kinase C-beta contributes to impaired endothelial insulin signaling in humans with diabetes mellitus. *Circulation* **2013**, *127*, 86–95. [CrossRef]
- Dentice, M.; Cordeddu, V.; Rosica, A.; Ferrara, A.M.; Santarpia, L.; Salvatore, D.; Chiovato, L.; Perri, A.; Moschini, L.; Fazzini, C.; et al. Missense mutation in the transcription factor NKX2-5: A novel molecular event in the pathogenesis of thyroid dysgenesis. *J. Clin. Endocrinol. Metab.* 2006, *91*, 1428–1433. [CrossRef] [PubMed]
- 64. Wen, B.; Yuan, H.; Liu, X.; Wang, H.; Chen, S.; Chen, Z.; de The, H.; Zhou, J.; Zhu, J. GATA5 SUMOylation is indispensable for zebrafish cardiac development. *Biochim. Biophys. Acta Gen. Subj.* **2017**, *1861*, 1691–1701. [CrossRef] [PubMed]
- 65. Chen, H.; Sun, W.; Zhu, J.; Yuan, H.; Chu, M.; Wen, B. Modification of cardiac transcription factor Gata6 by SUMO. *Biochimie* 2020, 170, 212–218. [CrossRef] [PubMed]
- Lu, H.; Liu, B.; You, S.; Chen, L.; Dongmei, Q.; Gu, M.; Lu, Y.; Chen, Y.; Zhang, F.; Yu, B. SENP2 regulates MEF2A de-SUMOylation in an activity dependent manner. *Mol. Biol. Rep.* 2013, 40, 2485–2490. [CrossRef] [PubMed]
- 67. Tanaka, M.; Chen, Z.; Bartunkova, S.; Yamasaki, N.; Izumo, S. The cardiac homeobox gene Csx/Nkx2.5 lies genetically upstream of multiple genes essential for heart development. *Development* **1999**, *126*, 1269–1280. [CrossRef] [PubMed]
- Ikeda, T.; Hikichi, T.; Miura, H.; Shibata, H.; Mitsunaga, K.; Yamada, Y.; Woltjen, K.; Miyamoto, K.; Hiratani, I.; Yamada, Y.; et al. Srf destabilizes cellular identity by suppressing cell-type-specific gene expression programs. *Nat. Commun.* 2018, *9*, 1387. [CrossRef]
- Iwanicka-Pronicka, K.; Socha, M.; Jedrzejowska, M.; Krajewska-Walasek, M.; Jamsheer, A. Life-threatening cardiac episode in a Polish patient carrying contiguous gene microdeletion of the TBX5 and the TBX3 genes. *Springerplus* 2016, *5*, 1638. [CrossRef] [PubMed]
- Zhang, Z.; Zhang, W.; Nam, Y.J. Stoichiometric optimization of Gata4, Hand2, Mef2c, and Tbx5 expression for contractile cardiomyocyte reprogramming. *Sci. Rep.* 2019, *9*, 14970. [CrossRef]
- 71. Shalizi, A.; Bilimoria, P.M.; Stegmuller, J.; Gaudilliere, B.; Yang, Y.; Shuai, K.; Bonni, A. PIASx is a MEF2 SUMO E3 ligase that promotes postsynaptic dendritic morphogenesis. *J. Neurosci.* 2007, *27*, 10037–10046. [CrossRef] [PubMed]
- Huang, C.Y.; Kuo, C.H.; Pai, P.Y.; Ho, T.J.; Lin, Y.M.; Chen, R.J.; Tsai, F.J.; Vijaya Padma, V.; Kuo, W.W.; Huang, C.Y. Inhibition of HSF2 SUMOylation via MEL18 upregulates IGF-IIR and leads to hypertension-induced cardiac hypertrophy. *Int. J. Cardiol.* 2018, 257, 283–290. [CrossRef] [PubMed]
- Xu, Q.; Jiang, Y.; Yin, Y.; Li, Q.; He, J.; Jing, Y.; Qi, Y.T.; Xu, Q.; Li, W.; Lu, B.; et al. A regulatory circuit of miR-148a/152 and DNMT1 in modulating cell transformation and tumor angiogenesis through IGF-IR and IRS1. *J. Mol. Cell Biol.* 2013, *5*, 3–13. [CrossRef] [PubMed]
- 74. Xing, W.; Zhang, T.C.; Cao, D.; Wang, Z.; Antos, C.L.; Li, S.; Wang, Y.; Olson, E.N.; Wang, D.Z. Myocardin induces cardiomyocyte hypertrophy. *Circ. Res.* 2006, *98*, 1089–1097. [CrossRef] [PubMed]
- 75. Bonifati, V.; Rizzu, P.; Squitieri, F.; Krieger, E.; Vanacore, N.; van Swieten, J.C.; Brice, A.; van Duijn, C.M.; Oostra, B.; Meco, G.; et al. DJ-1 (PARK7), a novel gene for autosomal recessive, early onset parkinsonism. *Neurol. Sci.* 2003, 24, 159–160. [CrossRef] [PubMed]
- Bonifati, V.; Rizzu, P.; van Baren, M.J.; Schaap, O.; Breedveld, G.J.; Krieger, E.; Dekker, M.C.; Squitieri, F.; Ibanez, P.; Joosse, M.; et al. Mutations in the DJ-1 gene associated with autosomal recessive early-onset parkinsonism. *Science* 2003, 299, 256–259.
 [CrossRef]
- 77. Ohshima, T.; Koga, H.; Shimotohno, K. Transcriptional activity of peroxisome proliferator-activated receptor gamma is modulated by SUMO-1 modification. *J. Biol. Chem.* **2004**, *279*, 29551–29557. [CrossRef]
- Kho, C.; Lee, A.; Jeong, D.; Oh, J.G.; Chaanine, A.H.; Kizana, E.; Park, W.J.; Hajjar, R.J. SUMO1-dependent modulation of SERCA2a in heart failure. *Nature* 2011, 477, 601–605. [CrossRef]
- Oh, J.G.; Watanabe, S.; Lee, A.; Gorski, P.A.; Lee, P.; Jeong, D.; Liang, L.; Liang, Y.; Baccarini, A.; Sahoo, S.; et al. miR-146a Suppresses SUMO1 Expression and Induces Cardiac Dysfunction in Maladaptive Hypertrophy. *Circ. Res.* 2018, 123, 673–685. [CrossRef]
- Du, J.; Zhang, L.; Zhuang, S.; Qin, G.J.; Zhao, T.C. HDAC4 degradation mediates HDAC inhibition-induced protective effects against hypoxia/reoxygenation injury. J. Cell. Physiol. 2015, 230, 1321–1331. [CrossRef]
- Ghosh, T.K.; Aparicio-Sanchez, J.J.; Buxton, S.; Brook, J.D. HDAC4 and 5 repression of TBX5 is relieved by protein kinase D1. *Sci. Rep.* 2019, *9*, 17992. [CrossRef] [PubMed]
- 82. Gu, J.; Fan, Y.; Liu, X.; Zhou, L.; Cheng, J.; Cai, R.; Xue, S. SENP1 protects against myocardial ischaemia/reperfusion injury via a HIF1alpha-dependent pathway. *Cardiovasc. Res.* 2014, *104*, 83–92. [CrossRef] [PubMed]
- Ishov, A.M.; Sotnikov, A.G.; Negorev, D.; Vladimirova, O.V.; Neff, N.; Kamitani, T.; Yeh, E.T.; Strauss, J.F., 3rd; Maul, G.G. PML is critical for ND10 formation and recruits the PML-interacting protein daxx to this nuclear structure when modified by SUMO-1. J. Cell Biol. 1999, 147, 221–234. [CrossRef] [PubMed]

- 84. Qiu, F.; Han, Y.; Shao, X.; Paulo, P.; Li, W.; Zhu, M.; Tang, N.; Guo, S.; Chen, Y.; Wu, H.; et al. Knockdown of endogenous RNF4 exacerbates ischaemia-induced cardiomyocyte apoptosis in mice. *J. Cell. Mol. Med.* **2020**, *24*, 9545–9559. [CrossRef] [PubMed]
- 85. Evans, T. Regulation of Cardiac Gene Expression by GATA-4/5/6. *Trends Cardiovasc. Med.* **1997**, *7*, 75–83. [CrossRef]
- Winick, J.; Abel, T.; Leonard, M.W.; Michelson, A.M.; Chardon-Loriaux, I.; Holmgren, R.A.; Maniatis, T.; Engel, J.D. A GATA family transcription factor is expressed along the embryonic dorsoventral axis in Drosophila melanogaster. *Development* 1993, 119, 1055–1065. [CrossRef]
- 87. Qiu, C.; Wang, Y.; Zhao, H.; Qin, L.; Shi, Y.; Zhu, X.; Song, L.; Zhou, X.; Chen, J.; Zhou, H.; et al. The critical role of SENP1-mediated GATA2 deSUMOylation in promoting endothelial activation in graft arteriosclerosis. *Nat. Commun.* **2017**, *8*, 15426. [CrossRef]
- 88. Liu, X.Y.; Yang, Y.Q.; Ma, J.; Lin, X.P.; Zheng, J.H.; Bai, K.; Chen, Y.H. Novel GATA4 mutations identified in patients with congenital atrial septal defects. *Zhonghua Xin Xue Guan Bing Za Zhi* **2010**, *38*, 724–727.
- 89. Riquelme, C.; Barthel, K.K.; Liu, X. SUMO-1 modification of MEF2A regulates its transcriptional activity. *J. Cell. Mol. Med.* 2006, 10, 132–144. [CrossRef]
- 90. Clark, C.D.; Lee, K.H. Second heart field-specific expression of Nkx2-5 requires promoter proximal interaction with Srf. *Mech. Dev.* **2020**, *162*, 103615. [CrossRef]
- 91. Lyons, I.; Parsons, L.M.; Hartley, L.; Li, R.; Andrews, J.E.; Robb, L.; Harvey, R.P. Myogenic and morphogenetic defects in the heart tubes of murine embryos lacking the homeo box gene Nkx2-5. *Genes Dev.* **1995**, *9*, 1654–1666. [CrossRef] [PubMed]
- 92. Kim, E.Y.; Chen, L.; Ma, Y.; Yu, W.; Chang, J.; Moskowitz, I.P.; Wang, J. Expression of sumoylation deficient Nkx2.5 mutant in Nkx2.5 haploinsufficient mice leads to congenital heart defects. *PLoS ONE* **2011**, *6*, e20803. [CrossRef]
- 93. Norman, C.; Runswick, M.; Pollock, R.; Treisman, R. Isolation and properties of cDNA clones encoding SRF, a transcription factor that binds to the c-fos serum response element. *Cell* **1988**, *55*, 989–1003. [CrossRef]
- 94. Johansen, F.E.; Prywes, R. Serum response factor: Transcriptional regulation of genes induced by growth factors and differentiation. *Biochim. Biophys. Acta* 1995, 1242, 1–10. [CrossRef]
- 95. Matsuzaki, K.; Minami, T.; Tojo, M.; Honda, Y.; Uchimura, Y.; Saitoh, H.; Yasuda, H.; Nagahiro, S.; Saya, H.; Nakao, M. Serum response factor is modulated by the SUMO-1 conjugation system. *Biochem. Biophys. Res. Commun.* 2003, 306, 32–38. [CrossRef]
- 96. Niu, Z.; Yu, W.; Zhang, S.X.; Barron, M.; Belaguli, N.S.; Schneider, M.D.; Parmacek, M.; Nordheim, A.; Schwartz, R.J. Conditional mutagenesis of the murine serum response factor gene blocks cardiogenesis and the transcription of downstream gene targets. *J. Biol. Chem.* **2005**, *280*, 32531–32538. [CrossRef]
- 97. Di Gregorio, A. T-Box Genes and Developmental Gene Regulatory Networks in Ascidians. *Curr. Top. Dev. Biol.* 2017, 122, 55–91. [PubMed]
- Pang, S.; Liu, Y.; Zhao, Z.; Huang, W.; Chen, D.; Yan, B. Novel and functional sequence variants within the TBX2 gene promoter in ventricular septal defects. *Biochimie* 2013, 95, 1807–1809. [CrossRef]
- Zhang, R.R.; Cai, K.; Liu, L.; Yang, Q.; Zhang, P.; Gui, Y.H.; Wang, F. A regulatory variant in TBX2 promoter is related to the decreased susceptibility of congenital heart disease in the Han Chinese population. *Mol. Genet. Genom. Med.* 2019, 7, e00530. [CrossRef] [PubMed]
- 100. Huber, P.; Crum, T.; Clary, L.M.; Ronan, T.; Packard, A.V.; Okkema, P.G. Function of the C. elegans T-box factor TBX-2 depends on SUMOylation. *Cell. Mol. Life Sci.* 2013, 70, 4157–4168. [CrossRef]
- 101. Steimle, J.D.; Moskowitz, I.P. TBX5: A Key Regulator of Heart Development. Curr. Top. Dev. Biol. 2017, 122, 195–221.
- Jia, Y.; Chang, Y.; Guo, Z.; Li, H. Transcription factor Tbx5 promotes cardiomyogenic differentiation of cardiac fibroblasts treated with 5-azacytidine. J. Cell. Biochem. 2019, 120, 16503–16515. [CrossRef] [PubMed]
- 103. Kathiriya, I.S.; Rao, K.S.; Iacono, G.; Devine, W.P.; Blair, A.P.; Hota, S.K.; Lai, M.H.; Garay, B.I.; Thomas, R.; Gong, H.Z.; et al. Modeling Human TBX5 Haploinsufficiency Predicts Regulatory Networks for Congenital Heart Disease. *Dev. Cell* 2021, 56, 292–309.e9. [CrossRef]
- 104. Beketaev, I.; Kim, E.Y.; Zhang, Y.; Yu, W.; Qian, L.; Wang, J. Potentiation of Tbx5-mediated transactivation by SUMO conjugation and protein inhibitor of activated STAT 1 (PIAS1). *Int. J. Biochem. Cell Biol.* **2014**, *50*, 82–92. [CrossRef]
- 105. Frey, N.; Olson, E.N. Cardiac hypertrophy: The good, the bad, and the ugly. Annu. Rev. Physiol. 2003, 65, 45–79. [CrossRef]
- 106. Booz, G.W. Putting the brakes on cardiac hypertrophy: Exploiting the NO-cGMP counter-regulatory system. *Hypertension* **2005**, 45, 341–346. [CrossRef] [PubMed]
- 107. Miano, J.M. Myocardin in biology and disease. J. Biomed. Res. 2015, 29, 3-19.
- 108. Trembley, M.A.; Quijada, P.; Agullo-Pascual, E.; Tylock, K.M.; Colpan, M.; Dirkx, R.A., Jr.; Myers, J.R.; Mickelsen, D.M.; de Mesy Bentley, K.; Rothenberg, E.; et al. Mechanosensitive Gene Regulation by Myocardin-Related Transcription Factors Is Required for Cardiomyocyte Integrity in Load-Induced Ventricular Hypertrophy. *Circulation* 2018, 138, 1864–1878. [CrossRef] [PubMed]
- 109. Cen, B.; Selvaraj, A.; Prywes, R. Myocardin/MKL family of SRF coactivators: Key regulators of immediate early and muscle specific gene expression. *J. Cell. Biochem.* **2004**, *93*, 74–82. [CrossRef]
- 110. Wang, D.Z.; Olson, E.N. Control of smooth muscle development by the myocardin family of transcriptional coactivators. *Curr. Opin. Genet. Dev.* **2004**, *14*, 558–566. [CrossRef]
- 111. Shin, J.H.; Ko, H.S.; Kang, H.; Lee, Y.; Lee, Y.I.; Pletinkova, O.; Troconso, J.C.; Dawson, V.L.; Dawson, T.M. PARIS (ZNF746) repression of PGC-1alpha contributes to neurodegeneration in Parkinson's disease. *Cell* **2011**, *144*, 689–702. [CrossRef]
- Nishida, T.; Yamada, Y. SUMOylation of the KRAB zinc-finger transcription factor PARIS/ZNF746 regulates its transcriptional activity. *Biochem. Biophys. Res. Commun.* 2016, 473, 1261–1267. [CrossRef]

- 113. Dikalov, S.I.; Nazarewicz, R.R. Angiotensin II-induced production of mitochondrial reactive oxygen species: Potential mechanisms and relevance for cardiovascular disease. *Antioxid. Redox Signal.* **2013**, *19*, 1085–1094. [CrossRef]
- 114. Mukherjee, D.; Chander, V.; Bandyopadhyay, A. PARIS-DJ-1 Interaction Regulates Mitochondrial Functions in Cardiomyocytes, Which Is Critically Important in Cardiac Hypertrophy. *Mol. Cell. Biol.* **2020**, *41*, e00106-20. [CrossRef] [PubMed]
- 115. Wadosky, K.M.; Willis, M.S. The story so far: Post-translational regulation of peroxisome proliferator-activated receptors by ubiquitination and SUMOylation. *Am. J. Physiol. Heart Circ. Physiol.* **2012**, 302, H515–H526. [CrossRef] [PubMed]
- 116. Pascual, G.; Fong, A.L.; Ogawa, S.; Gamliel, A.; Li, A.C.; Perissi, V.; Rose, D.W.; Willson, T.M.; Rosenfeld, M.G.; Glass, C.K. A SUMOylation-dependent pathway mediates transrepression of inflammatory response genes by PPAR-gamma. *Nature* 2005, 437, 759–763. [CrossRef] [PubMed]
- 117. Gwathmey, J.K.; Copelas, L.; MacKinnon, R.; Schoen, F.J.; Feldman, M.D.; Grossman, W.; Morgan, J.P. Abnormal intracellular calcium handling in myocardium from patients with end-stage heart failure. *Circ. Res.* **1987**, *61*, 70–76. [CrossRef] [PubMed]
- 118. del Monte, F.; O'Gara, P.; Poole-Wilson, P.A.; Yacoub, M.; Harding, S.E. Cell geometry and contractile abnormalities of myocytes from failing human left ventricle. *Cardiovasc. Res.* **1995**, *30*, 281–290. [CrossRef]
- 119. Hasenfuss, G. Alterations of calcium-regulatory proteins in heart failure. Cardiovasc. Res. 1998, 37, 279–289. [CrossRef]
- 120. Kho, C.; Lee, A.; Jeong, D.; Oh, J.G.; Gorski, P.A.; Fish, K.; Sanchez, R.; DeVita, R.J.; Christensen, G.; Dahl, R.; et al. Small-molecule activation of SERCA2a SUMOylation for the treatment of heart failure. *Nat. Commun.* **2015**, *6*, 1–11. [CrossRef] [PubMed]
- 121. Bian, X.; Xu, J.; Zhao, H.; Zheng, Q.; Xiao, X.; Ma, X.; Li, Y.; Du, X.; Liu, X. Zinc-Induced SUMOylation of Dynamin-Related Protein 1 Protects the Heart against Ischemia-Reperfusion Injury. *Oxid. Med. Cell. Longev.* **2019**, 2019, 1232146. [CrossRef]
- 122. Tilemann, L.; Lee, A.; Ishikawa, K.; Aguero, J.; Rapti, K.; Santos-Gallego, C.; Kohlbrenner, E.; Fish, K.M.; Kho, C.; Hajjar, R.J. SUMO-1 gene transfer improves cardiac function in a large-animal model of heart failure. *Sci. Transl. Med.* 2013, *5*, 211ra159. [CrossRef]
- 123. Shah, S.J.; Wasserstrom, J.A. SERCA2a gene therapy for the prevention of sudden cardiac death: A future theranostic for heart failure? *Circulation* **2012**, 126, 2047–2050. [CrossRef]
- 124. Samuel, T.J.; Rosenberry, R.P.; Lee, S.; Pan, Z. Correcting Calcium Dysregulation in Chronic Heart Failure Using SERCA2a Gene Therapy. *Int. J. Mol. Sci.* 2018, 19, 1086. [CrossRef]
- 125. Lowenstein, C.J. Myocardial reperfusion injury. N. Engl. J. Med. 2007, 357, 1121–1135.
- Frank, A.; Bonney, M.; Bonney, S.; Weitzel, L.; Koeppen, M.; Eckle, T. Myocardial ischemia reperfusion injury: From basic science to clinical bedside. *Semin. Cardiothorac. Vasc. Anesth.* 2012, *16*, 123–132. [CrossRef] [PubMed]
- 127. Chen, J.; Luo, Y.; Wang, S.; Zhu, H.; Li, D. Roles and mechanisms of SUMOylation on key proteins in myocardial ischemia/reperfusion injury. J. Mol. Cell. Cardiol. 2019, 134, 154–164. [CrossRef]
- 128. Kuo, M.H.; Allis, C.D. Roles of histone acetyltransferases and deacetylases in gene regulation. *Bioessays* **1998**, 20, 615–626. [CrossRef]
- 129. Fischle, W.; Emiliani, S.; Hendzel, M.J.; Nagase, T.; Nomura, N.; Voelter, W.; Verdin, E. A new family of human histone deacetylases related to Saccharomyces cerevisiae HDA1p. *J. Biol. Chem.* **1999**, 274, 11713–11720. [CrossRef] [PubMed]
- Grozinger, C.M.; Hassig, C.A.; Schreiber, S.L. Three proteins define a class of human histone deacetylases related to yeast Hda1p. Proc. Natl. Acad. Sci. USA 1999, 96, 4868–4873. [CrossRef] [PubMed]
- 131. Wang, A.H.; Bertos, N.R.; Vezmar, M.; Pelletier, N.; Crosato, M.; Heng, H.H.; Th'ng, J.; Han, J.; Yang, X.J. HDAC4, a human histone deacetylase related to yeast HDA1, is a transcriptional corepressor. *Mol. Cell. Biol.* **1999**, *19*, 7816–7827. [CrossRef]
- 132. Zhang, L.X.; DeNicola, M.; Qin, X.; Du, J.; Ma, J.; Tina Zhao, Y.; Zhuang, S.; Liu, P.Y.; Wei, L.; Qin, G.; et al. Specific inhibition of HDAC4 in cardiac progenitor cells enhances myocardial repairs. *Am. J. Physiol. Cell Physiol.* **2014**, 307, C358–C372. [CrossRef]
- Rodrigo, R.; Fernandez-Gajardo, R.; Gutierrez, R.; Matamala, J.M.; Carrasco, R.; Miranda-Merchak, A.; Feuerhake, W. Oxidative stress and pathophysiology of ischemic stroke: Novel therapeutic opportunities. CNS Neurol. Disord. Drug Targets 2013, 12, 698–714. [CrossRef] [PubMed]
- 134. Huang, C.; Han, Y.; Wang, Y.; Sun, X.; Yan, S.; Yeh, E.T.; Chen, Y.; Cang, H.; Li, H.; Shi, G.; et al. SENP3 is responsible for HIF-1 transactivation under mild oxidative stress via p300 de-SUMOylation. *EMBO J.* 2009, 28, 2748–2762. [CrossRef] [PubMed]
- 135. Wang, Y.; Yang, J.; Yang, K.; Cang, H.; Huang, X.Z.; Li, H.; Yi, J. The biphasic redox sensing of SENP3 accounts for the HIF-1 transcriptional activity shift by oxidative stress. *Acta Pharmacol. Sin.* **2012**, *33*, 953–963. [CrossRef] [PubMed]
- 136. Maroui, M.A.; Kheddache-Atmane, S.; El Asmi, F.; Dianoux, L.; Aubry, M.; Chelbi-Alix, M.K. Requirement of PML SUMO interacting motif for RNF4- or arsenic trioxide-induced degradation of nuclear PML isoforms. *PLoS ONE* 2012, 7, e44949. [CrossRef]
- 137. Niwa-Kawakita, M.; Ferhi, O.; Soilihi, H.; Le Bras, M.; Lallemand-Breitenbach, V.; de The, H. PML is a ROS sensor activating p53 upon oxidative stress. *J. Exp. Med.* **2017**, *214*, 3197–3206. [CrossRef]
- Plechanovova, A.; Jaffray, E.G.; McMahon, S.A.; Johnson, K.A.; Navratilova, I.; Naismith, J.H.; Hay, R.T. Mechanism of ubiquitylation by dimeric RING ligase RNF4. *Nat. Struct. Mol. Biol.* 2011, 18, 1052–1059. [CrossRef]
- Guo, B.; Sharrocks, A.D. Extracellular signal-regulated kinase mitogen-activated protein kinase signaling initiates a dynamic interplay between sumoylation and ubiquitination to regulate the activity of the transcriptional activator PEA3. *Mol. Cell. Biol.* 2009, 29, 3204–3218. [CrossRef]
- 140. Keith, B.; Johnson, R.S.; Simon, M.C. HIF1alpha and HIF2alpha: Sibling rivalry in hypoxic tumour growth and progression. *Nat. Rev. Cancer* **2011**, *12*, 9–22. [CrossRef]

- 141. Yang, W.; Sheng, H.; Thompson, J.W.; Zhao, S.; Wang, L.; Miao, P.; Liu, X.; Moseley, M.A.; Paschen, W. Small ubiquitin-like modifier 3-modified proteome regulated by brain ischemia in novel small ubiquitin-like modifier transgenic mice: Putative protective proteins/pathways. *Stroke* 2014, 45, 1115–1122. [CrossRef]
- 142. Gao, L.; Zhao, Y.; He, J.; Yan, Y.; Xu, L.; Lin, N.; Ji, Q.; Tong, R.; Fu, Y.; Gao, Y.; et al. The desumoylating enzyme sentrin-specific protease 3 contributes to myocardial ischemia reperfusion injury. *J. Genet. Genom.* **2018**, *45*, 125–135. [CrossRef]
- 143. Gartner, A.; Muller, S. PML, SUMO, and RNF4: Guardians of nuclear protein quality. Mol. Cell 2014, 55, 1–3. [CrossRef] [PubMed]
- 144. Keiten-Schmitz, J.; Wagner, K.; Piller, T.; Kaulich, M.; Alberti, S.; Muller, S. The Nuclear SUMO-Targeted Ubiquitin Quality Control Network Regulates the Dynamics of Cytoplasmic Stress Granules. *Mol. Cell* **2020**, *79*, 54–67.e7. [CrossRef] [PubMed]
- 145. Hotz, P.W.; Wiesnet, M.; Tascher, G.; Braun, T.; Muller, S.; Mendler, L. Profiling the Murine SUMO Proteome in Response to Cardiac Ischemia and Reperfusion Injury. *Molecules* 2020, 25, 5571. [CrossRef] [PubMed]
- 146. Gruber, P.J. Cardiac development: New concepts. Clin. Perinatol. 2005, 32, 845-855, vii. [CrossRef] [PubMed]
- 147. Glynne-Jones, P.; Boltryk, R.J.; Hill, M.; Zhang, F.; Dong, L.; Wilkinson, J.S.; Melvin, T.; Harris, N.R.; Brown, T. Flexible acoustic particle manipulation device with integrated optical waveguide for enhanced microbead assays. *Anal. Sci.* 2009, 25, 285–291. [CrossRef]
- 148. Plant, L.D.; Xiong, D.; Romero, J.; Dai, H.; Goldstein, S.A.N. Hypoxia Produces Pro-arrhythmic Late Sodium Current in Cardiac Myocytes by SUMOylation of NaV1.5 Channels. *Cell Rep.* **2020**, *30*, 2225–2236.e4. [CrossRef]
- Lumpkin, R.J.; Gu, H.; Zhu, Y.; Leonard, M.; Ahmad, A.S.; Clauser, K.R.; Meyer, J.G.; Bennett, E.J.; Komives, E.A. Site-specific identification and quantitation of endogenous SUMO modifications under native conditions. *Nat. Commun.* 2017, *8*, 1171. [CrossRef]
- 150. Hendriks, I.A.; Lyon, D.; Su, D.; Skotte, N.H.; Daniel, J.A.; Jensen, L.J.; Nielsen, M.L. Site-specific characterization of endogenous SUMOylation across species and organs. *Nat. Commun.* **2018**, *9*, 2456. [CrossRef]