#### **COHORT PROFILE**



# Lifelines NEXT: a prospective birth cohort adding the next generation to the three-generation Lifelines cohort study

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#### **Abstract**

Epidemiological research has shown there to be a strong relationship between preconceptional, prenatal, birth and early-life factors and lifelong health. The Lifelines NEXT is a birth cohort designed to study the effects of intrinsic and extrinsic determinants on health and disease in a four-generation design. It is embedded within the Lifelines cohort study, a prospective three-generation population-based cohort study recording the health and health-related aspects of 167,729 individuals living in Northern Netherlands. In Lifelines NEXT we aim to include 1500 pregnant Lifelines participants and intensively follow them, their partners and their children until at least 1 year after birth. Longer-term follow-up of physical and psychological health will then be embedded following Lifelines procedures. During the Lifelines NEXT study period biomaterials—including maternal and neonatal (cord) blood, placental tissue, feces, breast milk, nasal swabs and urine—will be collected from the mother and child at 10 time points. We will also collect data on medical, social, lifestyle and environmental factors via questionnaires at 14 different time points and continuous data via connected devices. The extensive collection of different (bio)materials from mother and child during pregnancy and afterwards will provide the means to relate environmental factors including maternal and neonatal microbiome composition) to (epi)genetics, health and developmental outcomes. The nesting of the study within Lifelines enables us to include preconceptional transgenerational data and can be used to identify other extended families within the cohort.

 $\textbf{Keywords} \ \ Biobank \cdot Birth \ cohort \cdot Prospective \ study \cdot Microbiome \cdot Transgenerational \ effects \cdot Developmental \ Origins \ of \ Health \ and \ Disease \ (DOHaD)$ 

#### Introduction

Exposures or events in the preconceptional, prenatal, birth and early life period may have lifelong effects on an individual's development and disease susceptibility. An impressive body of evidence in the field of Developmental Origins of Health and Disease (DOHaD) has shown that maternal factors (e.g. physical and psychological health, lifestyle) and environmental factors can modulate the developmental program [1–3]. This modulation could then permanently change the physiology, metabolism, epigenome and microbiome of the child, subsequently affecting healthy development or

increasing susceptibility to (chronic) diseases [4–6]. Developmental programming via epigenetic changes may also have transgenerational effects without changing the genetic code in either the maternal or paternal line [7].

Several birth cohorts have been initiated with comprehensive data collection that include questionnaires as well as biomaterials to identify factors associated with child health and disease susceptibility [8]. However, although the importance of both immune status and the microbiome for the development of diseases has been established in multiple human cross-sectional studies and in animal models, only a few cohort studies include comprehensive immunological and microbiome data [9]. Moreover, very few birth cohorts have included three or more generations in their study design.

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Another unique feature of the Lifelines NEXT data set is the presence of biomaterials from homebirths. In the Netherlands, 13% of children of mothers with low risk of complication are born at home [10]. This provides a unique opportunity to study the influence of homebirth versus birth in the hospital environment (with or without medical interventions) on the development of the newborn child. Previous work has found associations of birth interventions with increased risks of several immune-related diseases [11], but the mechanism(s) underlying this observation remain unclear. Data collected in Lifelines NEXT could assist in uncovering these mechanisms.

Finally, compared to other cohorts, Lifelines NEXT will collect large volumes of breastmilk over a long period of time [8]. This provides the means to study the mechanisms behind the immune-competent proteins in breastmilk, the large amounts of cholesterol it contains and the different molecular structures of its lipid content compared to infant formulas. Cellular trafficking from mother to child ("chimerism") and (favorable) antigen exposition in an immune-modulated neonatal environment have been suggested to be part of mechanisms that have a long-lasting effect on neonatal health and can be studied in Lifelines NEXT [12–15].

Lifelines NEXT is an observational prospective birth cohort with a transgenerational design of up to four generations. Lifelines NEXT is embedded within the previously described Lifelines cohort study [16, 17]. In short, Lifelines is a prospective population-based cohort study comprising approximately 10% of the population of the Northern Netherlands (the provinces of Groningen, Drenthe and Friesland). Lifelines includes 167,729 participants and was designed as a three-generation cohort [16, 17]. Within Lifelines, 112,596 participants (67%) have a known family member within the cohort, with 84,888 (51%) part of two generations and 20,362 participants (12%) part of three generations of Lifelines participants [16]. Moreover, once genetic information becomes available, we anticipate that more extended families will be identified within the cohort given the expected cryptic genetic-relatedness of the more homogenous northern part of the Netherlands [18]. Initially, only children 8 years and older participated in Lifelines, and no data and materials were collected from Lifelines participants who became pregnant. With the initiation of Lifelines NEXT, we are filling this gap and adding a fourth generation to Lifelines.

The primary study objective of Lifelines NEXT is to investigate the effects of early life or pre-conceptional transgenerational events on health in early childhood. Its secondary aim is to correlate genomic, epigenetic, serological, metabolomic, microbiome, medical, social and environmental factors to early life health. Lifelines NEXT will provide unique opportunities to separate non-genetic from genetic familial transmission and to assess (epi)genetic influences

and imprinting. Moreover, Lifelines NEXT can associate exposures in the preconception, prenatal, birth and early life period with healthy development and (chronic) disease susceptibility. The main risk factors of interest include microbiome, (epi)genetic, environmental and lifestyle factors.

This paper describes the infrastructure of the detailed and unique (bio)data collection of Lifelines NEXT, which starts in early pregnancy (as early as 12 weeks gestational age) and follows the offspring extensively up until at least 12 months of age. The infrastructure of Lifelines will be used to extend children's follow up and offers participants the opportunity to enter the regular Lifelines cohort including its standardized data collection of biomaterials and questionnaires [16, 17].

#### **Methods**

### Participants and recruitment strategy

Lifelines NEXT aims to include 1500 pregnant women. Upon inclusion, their partners are also invited to participate. Children are included in the birth cohort on the day they are born.

From 2016 to 2021, Lifelines NEXT will recruit pregnant women in the Northern part of the Netherlands, preferably from 12 weeks gestational age onwards. Eligible women are recruited via midwives or gynecologists, the Lifelines website, Lifelines NEXT social media (announcements on Facebook and Instagram), Lifelines newsletters, pregnancyrelated events and informational meetings. A leaflet containing details about Lifelines NEXT is provided. Pregnant women who consent to participate in the study are contacted via the Lifelines service desk. A research assistant at the service desk then provides more detailed information about the data collection of Lifelines NEXT (Fig. 1) and explains the informed consent procedure. A research nurse is allocated as the primary contact person for each Lifelines NEXT participant and obtains the informed consent at the first home visit. Both parents need to consent that their child will be included in Lifelines NEXT.

# Procedures for gathering and storage of (bio) materials and data

Questionnaire data and extensive biomaterial data will be collected for the mother, father, child and the family home environment (Fig. 1). Data collection starts, preferably, at 12 weeks of gestation. Research nurses will visit participants four times to perform tests and collect (bio)materials. To minimize the burden on the participants, sample collection is performed at the participant's home by the research nurse, by the mother herself or by her maternity care provider.



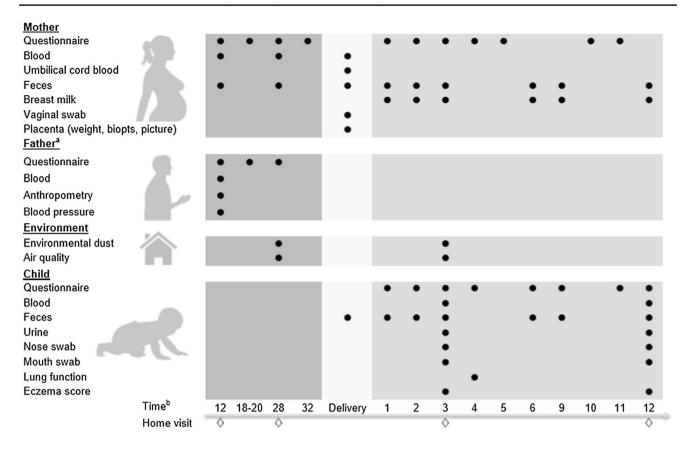


Fig. 1 Timeline of the Lifelines NEXT study indicating the data collection per time point. <sup>a</sup>Only for non-Lifelines fathers. Measurements start at the moment of inclusion. <sup>b</sup>Gestational age (weeks)/child's age (months)

This participant-based sampling program was designed using insights and experience gathered in similar successful program like Lifelines DEEP [19]. To assist participants in this process, Lifelines NEXT provides standardized protocols for the collection and storage of (bio)samples. An overview of all (bio)materials collected during the study is shown in Table 1. Furthermore, questionnaires will be sent digitally by email at standardized time points. Table 2 provides a detailed overview of the measures that longitudinally assesses maternal, neonatal or paternal characteristics in the questionnaires.

Feces and breastmilk are sampled and aliquoted by the participant and stored, along with urine samples, in a freezer in the participant's home ( $-20\,^{\circ}$ C) until they are collected by the research nurse. The nurse ships the frozen samples to the Lifelines laboratory and biobank, where they are processed and then stored at  $-80\,^{\circ}$ C. Venous blood and umbilical cord blood is drawn by allocated professionals, and samples are immediately stored in the refrigerator either at home or at the hospital (EDTA), kept for 30 min at room temperature and then refrigerated (serum), or kept at room temperature (PAXgene collection tubes). Blood samples are processed and divided into aliquots in the laboratory. Other biomaterials surrounding birth are collected and temporarily

stored in a refrigerator (placenta biopsies) or freezer (vaginal swabs) located in either the participant's homes (for homebirths) or at the hospital where the birth took place. These biomaterials are then transferred to the Lifelines laboratory and biobank within 10 h of birth. All materials are stored in barcoded aliquots at  $-80\,^{\circ}\text{C}$  for future research, except for the blood spot collected at 4 months, which is kept at room temperature. All data are stored in a secured data storage environment.

#### Measurements of the mother

During pregnancy and the first year after childbirth, participating women are asked to complete eleven different questionnaires on their physical health, psychological health, reproductive health, lifestyle-related behavior and nutritional intake, and social and working conditions (Table 2). During the first home visit, a venous blood sample is drawn by the research nurse. Additionally, during the prenatal period and the first year post-partum, maternal biomaterials are sampled and stored by the participants themselves and collected at home visits by the research nurse (Fig. 1).

During childbirth, the maternity care provider will collect a vaginal swab, a fecal sample and blood from the mother.



Table 1 Overview of biomaterial samples collected in Lifelines NEXT

Automate   EDTA	Biomaterials	Materials used for collection	Number	Volume (mL)	Sample type <sup>a</sup>	Pregnancy	ancy	Birth	First 3	First year of life (months)	fe (mor	iths)				
1004   EDTA   1   10   Plearm   X   X   X   X   X   X   X   X   X						(week	s) _									
December   December						12	28		_	2	3	9	6	10	11	12
Posture   Posture   1   10   Pasture   X   X   X   X   X   X   X   X   X	Mother															
Serum         1         8.5         Serum         X         X         X           PAXgene         1         6.5         Serum         X	Venous blood	EDTA	_	10	Plasma	×	×	×								
Packgrate   1   6   Stream   2   2   2   2   2   2   2   2   2		Serum	1	8.5	Serum	×	×	×								
Packgene   1   25   Peripheral blood   X   X   X   X   X   X   X   X   X		Serum	1	9	Serum	×	×	×								
Cryotube		PAXgene		2.5	Peripheral blood	×	×									
wath wath wath wath wath wath muturenally Swath Swath (Internall)         Swath Aller         1         Fecess milk         X	Feces	Cryotube	4	2	Feces	×	×	×	×	×	×	×	×			×
wab         Swab         1         Vaginal swab         X		Swab	1		Feces	×	×	×	×	×	×	×	×			×
like         Cryotube         1         9 becental (maternal)         X <t< td=""><td>Vaginal swab</td><td>Swab</td><td>_</td><td></td><td>Vaginal swab</td><td></td><td></td><td>×</td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td></t<>	Vaginal swab	Swab	_		Vaginal swab			×								
Ik   Cryotube   15   2   Breast milk   X   X   X   X   X   X   X   X   X	Placenta (maternal)	RNAlater	2	5	Placenta (maternal)			×								
Lord blood   EDTA   1   10   Plasma   X   X   X   X   X   X   X   X   X	Breast milk	Cryotube	15	2	Breast milk				×	×	×	×	×			×
FOTA blood   EDTA   1   10   Plasma   X   X   X   X   X   X   X   X   X		Swab	1		Breast milk				×	×	×	×	×			×
Serum   Serum   X   Serum   Serum   X	Child															
Serum         8.5         Serum         X           PAXgene         1         2.5         Peripheral blood         X           blood         Blood spot         2         DyBlood Sample         X           lood         EDTA         4         0.5         Plasma         X           lood         EDTA         2         0.5         Plasma         X           Cryouthe         5         2         Plasma         X         X           Swab         1         Nose swab         X         X         X           Swab microbiome         1         Nose swab         X         X         X           Swab microbiome         1         Nose swab         X         X         X           Respot         1         Nose swab         X         X         X           Respot	Umbilical cord blood	EDTA	_	10	Plasma			×								
Serum         1         6         Serum         X           Flood         Blood sport         2         Peripheral blood         X           Bod Sport         2         0.5         Plasma         X           Isod         EDTA         2         0.5         Plasma         X           Isod         EDTA         2         0.5         Plasma         X           Isod         2         0.5         Plasma         X         X           Heparine         2         0.5         Plasma         X         X           Result         2         0.5         Plasma         X         X           Result         2         0.5         Plasma         X         X         X           Cryotube         5         2         Feces         X         X         X         X           Swab         1         1         Nose swab         X         X         X         X         X           Swab microbiome         1         1         1         Nose swab         X         X         X         X           Swab microbiome         1         1         1         1         X         X		Serum	1	8.5	Serum			×								
PAXgene		Serum	_	9	Serum			×								
Plood spot   A		PAXgene	_	2.5	Peripheral blood			×								
EDTA	Capillary blood	Blood spot	2		Dry Blood Sample						×					
Buffy coat   Serum   Swab Diva   1		EDTA	4	0.5	Plasma						×					
Serum					Buffy coat						×					
Serum         2         0.5         Serum         X           Heparine         2         0.6         Plasma         X <td>Venous blood</td> <td>EDTA</td> <td>2</td> <td>0.5</td> <td>Plasma</td> <td></td> <td>×</td>	Venous blood	EDTA	2	0.5	Plasma											×
Serum         2         0.5         Plasma           Heparine         2         0.6         Plasma           KIAAlater         2         5         Placenta (fetal)         X <th< td=""><td></td><td></td><td></td><td></td><td>Buffy coat</td><td></td><td></td><td></td><td></td><td></td><td>×</td><td></td><td></td><td></td><td></td><td>×</td></th<>					Buffy coat						×					×
(fetal)         RNAlater         2         0.6         Plasenta (fetal)         X <t< td=""><td></td><td>Serum</td><td>2</td><td>0.5</td><td>Serum</td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td>×</td></t<>		Serum	2	0.5	Serum											×
(fetal)         RNAlater         2         5         Dlacenta (fetal)         X		Heparine	2	9.0	Plasma											×
Cryotube         5         2         Feces         X <t< td=""><td>Placenta (fetal)</td><td>RNAlater</td><td>2</td><td>5</td><td>Placenta (fetal)</td><td></td><td></td><td>×</td><td></td><td></td><td></td><td></td><td></td><td></td><td></td><td></td></t<>	Placenta (fetal)	RNAlater	2	5	Placenta (fetal)			×								
Swab RNA         1         Feces         X <t< td=""><td>Feces</td><td>Cryotube</td><td>5</td><td>2</td><td>Feces</td><td></td><td></td><td>×</td><td>×</td><td>×</td><td>×</td><td>×</td><td>×</td><td></td><td></td><td>×</td></t<>	Feces	Cryotube	5	2	Feces			×	×	×	×	×	×			×
Swab RNA         1         Nose swab         X           Swab DNA         1         Nose swab         X           Swab microbiome         1         Nose swab         X           Swab microbiome         1         Urine         X           mental         Cotine         X         X           ector         Sheet         2         Dust         X         X           blood         EDTA         1         10         Plasma         X		Swab	1		Feces			×	×	×	×	×	×			×
Swab DNA         1         Nose swab         X           Swab microbiome         1         Nose swab         X           Swab microbiome         1         Mouth swab         X           mental         1         1.2         Urine         X           ector         Sheet         2         Dust         X           olood         EDTA         1         10         Plasma         X	Nose	Swab RNA	_		Nose swab						×					×
Swab microbiome         1         Nose swab         X           Swab microbiome         1         Mouth swab         X           mental         1         1.2         Urine         X           ector         Sheet         2         Dust         X           olood         EDTA         1         10         Plasma         X		Swab DNA	1		Nose swab						×					×
Swab microbiome         1         Mouth swab         X           mental         2         Dust         X           sctor         Sheet         X         X           lood         EDTA         1         10         Plasma         X		Swab microbiome	1		Nose swab						×					×
mental         X           ector         Sheet         2         Dust         X           Alood         EDTA         1         10         Plasma         X	Mouth	Swab microbiome	_		Mouth swab						×					×
mental         Z         Dust         X           ector         Sheet         2         Plasma         X           alood         EDTA         1         10         Plasma         X	Urine	Peespot		1.2	Urine						×					×
ector         Sheet         2         Dust         X           alood         EDTA         1         10         Plasma         X	Environmental															
olood EDTA 1 10 Plasma	Dust collector	Sheet	2		Dust		×				×					
EDTA 1 10 Plasma	Father <sup>b</sup>															
	Venous blood	EDTA	1	10	Plasma	×										



12 10 6 First year of life (months) 9 0 Birth 28 (weeks) 2  $\times \times \times \times$ Peripheral blood Sample typea Plasma Serum Volume (mL) Number Materials used for collection Heparine PAXgene Serum Table 1 (continued) **Biomaterials** 

<sup>a</sup>All materials stored at –80 °C, except for the Dry Blood Sample collected at 4 months (room temperature) bonly for non-Lifelines participants, collected at the moment of inclusion

Immediately after birth, umbilical cord blood and placental tissue are collected according to standardized protocols. The placenta is also weighed and photographed. Maternity care providers also provide a detailed birth report.

#### Measurements of the child

Children are included in the birth cohort on the day they are born. During the first year of follow-up, the parents complete eight different questionnaires on their child's health, development and behavior (Table 2). Seven fecal samples are collected over an interval that starts directly after birth and continues up until the child is 1 year of age. During a home visit at age 4 months, the research nurse will sample capillary blood by a puncture of the child's heel. At age 12 months, venous blood will be sampled at the pediatrics outpatient clinic at the University Medical Center Groningen (UMCG). Additionally, at the 4- and 12-month home visits, a mouth and a nose sample will be collected for microbiome, DNA and RNA profiling (Table 1). The research nurse will also assess the presence of eczema using the cumulative objective scoring index for atopic dermatitis (SCORAD) [20] (Table 2). During a separate home visit at 4 months of age, lung function tests will be performed with the Single Occlusion Technique (WHISTLER) by a dedicated research nurse [21, 22].

# Measurements of the father

If the father consents to participate in Lifelines NEXT and is also a Lifelines participant, no additional (biomaterial) data needs to be collected. However, if the father is not a participant in Lifelines, he will be embedded in the routine data collection of Lifelines [16]. During a home visit, the research nurse will draw blood and perform anthropometric examinations that conform with the Lifelines protocol (Fig. 1, Table 1) [16]. Additionally, throughout the prenatal period, these fathers will complete three different Lifelines baseline questionnaires with an interval of 10 weeks (Table 2) [16].

#### Measurements on the family home environment

Environmental data will be collected by measuring air quality in participant's homes. Airborne dust will be collected with an electrostatic dust fall collector (EDC) [23] placed during a home visit at 28 gestational weeks and at 4-months postpartum (Table 1). The social conditions at home will also be assessed. Additional data will be collected using self-reported questionnaires that include items about the living condition in the house, smoking and the presence of



 Table 2
 Overview of measurement in questionnaires

SWING [24]  MSPSS, SSL [25, 26]  MINI [27]  MAAS, MPAS [28, 29]  LDI, LTE [30, 31]  CEQ [32]	3 [24]	X		ω 4 α × × × ×	6 9		12
swing [24] swind rate versa r towards foods, physical activity and experiences, pre-pregnancy control, reproductive organs, STDs, fertility, pregnancy al factors (e.g. smoking), oral health ry mapped gues, neighbors and level of satisfaction fedness and its effects on functioning str with relationship ners ant satisfaction The counter and prescribed ust ssociated to life events auxing pregnancy and birth CEQ [32]	××			× × × ×			
SWING [24]  uation and vice versa  towards foods, physical activity and experiences,  pre-pregnancy control, reproductive organs, STDs, fertility, pregnancy al factors (e.g. smoking), oral health  ry  MINI [27]  redness and its effects on functioning  er with relationship  rers  ant  LDI, LTE [30, 31]  associated to life events  CEQ [32]	× ×			× × × ×			
pre-pregnancy control, reproductive organs, STDs, fertility, pregnancy al factors (e.g. smoking), oral health  WSPSS, SSL [25, 26]  gues, neighbors and level of satisfaction  MINI [27]  redness and its effects on functioning  err  with relationship  ners  ant  The counter and prescribed  ust  is associated to life events  CEQ [32]	×			× × × ×			
pre-pregnancy control, reproductive organs, STDs, fertility, pregnancy al factors (e.g. smoking), oral health ry sues, neighbors and level of satisfaction tedness and its effects on functioning st with relationship ners ant ust ssociated to life events uring pregnancy and birth		$\times$ $\times$ $\times$ $\times$		× × × ×			
gues, neighbors and level of satisfaction tedness and its effects on functioning or with relationship ners ant ant associated to life events uring pregnancy and birth		$\times$ $\times$ $\times$ $\times$		× × × ×			
satury struction restedness and its effects on functioning artner ction with relationship partners o-infant ings over the counter and prescribed to person the dust the dust the dust the during pregnancy and birth	s, SSL [25, 26] 27] , MPAS [28, 29]	× × × × ×		× × × ×		>	
erestedness and its effects on functioning artner ction with relationship partners o-infant ings over the counter and prescribed it, pets the pets the dust the dust ing presents associated to life events the during pregnancy and birth	27] , MPAS [28, 29]	× × × ×		× × ×		>	
rerestedness and its effects on functioning artner ction with relationship partners o-infant ings over the counter and prescribed i, pets the dust the dust the dust the dust the dust of the counter and birth the during pregnancy and birth	27] , MPAS [28, 29]	× × × ×		× × ×		>	
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ther/partner atisfaction with relationship ween partners where partners ather-to-infant ad feelings ation, over the counter and prescribed adition—dust idation, pets and stress associated to life events e health during pregnancy and birth	, MPAS [28, 29]	× × ×		× ×		>	
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ween partners ther-to-infant d feelings ation, over the counter and prescribed adition—dust ilation, pets und stress associated to life events e health during pregnancy and birth	, MPAS [28, 29]	×		×			
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nattion—dust tilation, pets and stress associated to life events ve health during pregnancy and birth		>		>			
and stress associated to life events ve health during pregnancy and birth		×		×			
;	TE [30, 31]		×			×	
;							
health, pregnancy disorders, environmental factors(e.g. smoking), birth (including place of birth, interventions and mother's experiences)	32]		×				
Food intake FFQ [33]	(3)		×				
Reproductive health—postpartum recovery				×			
Parenting style handling the child and family life situations				×			
Child							
Crying Intensity, duration, week average			×	×	×		×
Gastrointestinal symptoms Rome III [34]	III [34]		×	×	×		×
on, throwing up, defecation pattern and gut complaints							;
Food intake FFQ [33]	[3]		×	×	×		$\times$



Table 2 (continued)

Measurements in questionnaires	Validated instruments used Pregnancy (weeks)	Pregnancy (weeks)	Birth First year of life (months)	ar of lii	e (mon	ths)				
		12 18–20 28 32	1 2	3	4	5 (	6 9	10	11	12
Medication			×	×			×			×
dosage, duration, over the counter and prescribed										
Child health					×					×
health, house, smoking, pets										
Eczema <sup>a</sup>	SCORAD [20]				×					×
Social condition home <sup>a</sup>	HOME [35]				×					
Lung function <sup>a</sup>	WHISTLER [21, 22]				×					
Child care							×			
parent, paid babysitter, grandparents										
Child development	ASQ [36]						×			×
communication, gross and fine motor skills, problem solving, social										
Child development child's behavior	IBQ [37]						×			
Eating behavior									×	
Social and emotional skills	BITSEA [38]									×
feelings and behaviors										
Growth and vaccination										×
Father <sup>b</sup>										
Demographics, work, family		×								
Health	SCL-90 SOM [39]	×								
presence of diseases and disorders										
Physical activity	SQUASH [40]	×								
Medication		×								
dosage, duration, over the counter and prescribed										
Life events	LDI, LTE [30, 31]	×								
difficulties and stress associated to life events										
Diet	FFQ [41]	×								
Gastrointestinal symptoms	Rome III [34]	×								
regurgitation, defecation pattern and gut complaints										
Respiratory health	ECRHS [42]	×								
Allergies, intoxications		×								
Visual function	NEI VFQ-25 [43]	×								
Pain	WPI [44]	×								
Fatigue	CIS [45]	×								
Y 6										

<sup>a</sup>Assessed by research nurse

<sup>&</sup>lt;sup>b</sup>Only for non-Lifelines participants. Items according to the baseline questionnaires from Lifelines, divided in three questionnaires with an interval of 10 weeks, starting at moment of inclusion of the father



pets (Table 2). Finally, regional data on air pollution will be collected by extracting geocoded data on postal code level.

#### The Newborn initiative

Lifelines NEXT participants also have the opportunity to participate in the Newborn initiative, a public–private project that expands the range of phenotypes measured through agreements with companies developing new technologies while looking at local public health issues. In living lab Newborn, prototypes of innovative products that contribute to health can be tested in real-time home situations. Upon additional consent, these data can be used for product development. The Lifelines NEXT data collection will be enriched with data from continuous measurement devices with built-in apps, e.g. the heartbeat of the mother (healthband®), air quality (airvibe®) and behavior of the child (uGrow Baby Monitor®).

#### **Study organization**

Lifelines NEXT is an initiative of the Departments of Genetics, Obstetrics, Laboratory medicine and Pediatrics of the UMCG. The study is embedded in the organizational structure of the Lifelines cohort study and will use its service center, its laboratory for processing of biomaterials and its infrastructure for digital data collection and storage. All collected biomaterials will be stored in labeled tubes in the Lifelines Lifestore, which currently holds more than 5 million biomaterials of Lifelines participants [16].

The project is governed by a steering committee in which the UMCG Departments of Genetics, Pediatrics, Laboratory medicine, Obstetrics, Community and Occupational Health and Midwifery are represented, as well as the Lifelines organization. The chairman of the steering committee leads the project leaders, maintains contact with external sponsors and is accountable to the supervisory board. Moreover, a supervisory board is in charge consisting of the representatives from the three UMCG Departments of Genetics, Laboratory Medicine and Obstetrics and the board of the UMCG Hereditary Metabolic Diseases Fund. Project leaders from the UMCG and Lifelines work closely to manage the study.

Daily management of Lifelines NEXT is carried out by several members of the Lifelines NEXT research team. A Lifelines research assistant is the primary contact person for all the maternity healthcare providers involved who have questions related to (the data collection of) Lifelines NEXT. The research assistant also extensively monitors the Lifelines NEXT study process of each participant and schedules home visits according to the protocol. Outside normal working hours, a member of the birth team, which consists of medical and midwifery students, is available to support data collection at birth. All birth team members have been trained to

collect biomaterials according to protocol, and they arrange the transportation of these biomaterials to the biobank within 3 h. A research nurse is allocated to be the primary contact person for each Lifelines NEXT participant and will perform all the home visits during the study. The research nurse distributes biomaterial collection sets, provides collection instructions and installs devices for environmental data collection. By staying in close contact with the participants, the research nurse acts as a confidant and is instrumental in ensuring that questionnaires are completed and biomaterials collected. Moreover, the research nurse gathers the biomaterials that were collected at home and arranges their transport to the Lifelines laboratory and biobank for processing and storage. All Lifelines NEXT data are stored in a secured data storage environment that utilizes MOLGENIS, a modular suite of web databases for integrating genotype, phenotype and other analyses. Each MOLGENIS database has web user interfaces as well as scriptable interfaces to plug-in R, Java and web services [46]. The handling of data complies with the General Data Protection Regulation (GDPR) [47].

#### Statistical power considerations and study size

Based on the Lifelines add-on study Lifelines DEEP (n=1500), in which integrative analyses have been conducted, a sample size of 1500 subjects will be sufficient to generate novel insights into the preconception, prenatal, birth and early life period [19]. In Lifelines DEEP we had enough power, for instance, to show statistically significant associations between specific microbial compositions and blood lipids [48] and to study various factors influencing microbiome composition and function [49]. These analyses were performed systematically on multiple factors and multiple levels, including microbial diversity, inter-individual distance in composition, individual species and pathway level.

Given a birth rate of 1.6 in the Netherlands [50] and the participation of approximately 40,000 women aged 25–40 years in Lifelines, we expect about 2000 pregnancies per year. Based on previous Lifelines add-on studies, we expect a response rate around 60% [19].

# Harmonization and external database linkages

Harmonization and linkage with baseline and future longitudinally collected Lifelines data, including genomewide genetic data, will be established for all participants in Lifelines NEXT (i.e. the mother, father and child). This will result in data from up to four generations of the women in Lifelines NEXT. Data of the mothers participating in Lifelines NEXT will be linked with the Dutch Perinatal Data Register, Perined. This perinatal registration data includes three separate databases: one for primary



midwife-led care, one for secondary obstetric care and one for neonatal care [11]. These three databases are combined into one national perinatal database via a validated linkage method by the Perinatal Registry Office [51]. The data of the Lifelines NEXT children will be linked with the electronic files of the regional youth and family centers. These centers are entrusted with the national preventive follow-up program of children that monitors the health and development of children from birth until 18 years. This system sees over 95% of all children born in the Netherlands through a comprehensive series of appointments [52]. Moreover, Lifelines NEXT data could be merged and harmonized with data collected by other cohort studies like the Netherlands Kinship Panel Study (NKPS) [53], the birth cohort studying the prevention and incidence of asthma and mite allergy (PIAMA) [54], Trails NEXT [55] and NeoLifeS [56].

# Strengths and limitations

Lifelines NEXT will be a rich resource for research. It is a unique birth cohort that can address critical questions regarding the influence of environmental exposures, social factors, stressors and early-life nutrition on early life development. While the Lifeline NEXT cohort consists of relatively healthy people, it is suitable for research related to (chronic) disease susceptibility (rather than rare diseases). It also enables study of the association of variations in microbiome, metabolomics profiles and (epi) genomics in mothers and children with short- and longerterm health outcomes. The dynamic organization of the study allows us to add additional measurements and to ask new questions during the course of the study. For example, we have already added sampling of feces at all time points in glycerol tubes that allow for later culturing of live bacteria, additional sampling of neonatal feces in week 2 and continuous measurements from connected devices to the study. Further add-on initiatives from additional researchers are also welcomed.

The extended hygiene hypothesis and the EPIgenetic Impact of Childbirth (EPIIC) hypothesis both suggest that factors occurring during the intrapartum and early postnatal period may affect the neonatal immune response or lead to different microbial communities and fetal epigenomic remodeling anomalies [58, 59]. Lifelines NEXT will be able to study different microbial communities of children by metagenomic sequencing, obtaining a comprehensive view of the development of microbial ecosystems in the early life period, as well as its relation to immune, respiratory and metabolic development. It will also be possible to study the change of epigenetic profiles in more detail, for

instance that of genes related to allergy in the first years of life [60].

Another strength of Lifelines NEXT is that it offers the possibility to study interactions of immunological, microbial and metabolic maturation processes with environmental factors such as mode of delivery, maternal and early neonatal feeding patterns, type of nutrition (such as breastmilk or formula), indoor and outdoor environmental factors (e.g. exposure to cigarette smoke, air pollution and allergens), use of medication and infections [12, 13, 15, 61–66].

Finally, little attention has been paid thus far to the viral composition of the microbiome: the virome. Because majority of gut viruses are viruses of bacteria (bacteriophages), they are therefore expected to be a major factor in shaping the human microbiome, and could exert an effect on human physiology [67]. So far, little research has been done looking at the role of bacteriophages in the development of gut ecosystem, and their relation to babies health. Through comprehensive analysis, this relatively new area of the role of the virome can be studied in depth.

However, this study had some limitations. At first there is only a limited amount of data available on fathers that were not in the Lifelines study upon the pregnancy of their partners. To meet the need for background information, several baseline questionnaires of the initial Lifelines cohort study are included and will be completed by those new participants. A challenge is to include a representative sample for the Dutch population. As we know, the Lifelines cohort study had, adjusted for differences in demographic composition, a smaller proportion of low educated participants and smokers. However the population was concluded to be broadly representative for the adult population of the north of the Netherlands [57]. Aiming to maintain generalizability we used similar recruitment techniques within Lifelines NEXT.

#### **Collaboration**

We expect inclusion to be completed in 2021, therefore from 2023 onwards biomaterials and data collected in Lifelines NEXT will be available to other researchers. In the meanwhile the Lifelines NEXT consortium welcomes collaboration with other birth cohorts. For example, Lifelines NEXT data could be harmonized and merged with other European and Canadian birth cohorts as proposed in the EUCAN-Connect project [68]. We also welcome add-on initiatives. The Lifelines NEXT cohort study has an open protocol. Interested researchers can submit an application for an additional study or additional (biomaterial) data collection to the steering committee of Lifelines NEXT for approval. Further information can be requested by e-mail: (lifelinesnext@umcg.nl). The Lifelines website



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(www.Lifelines.nl) provides information about the application process and the data collection, and gives an overview of all available data within Lifelines and publications with Lifelines data.

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#### Compliance with ethical standards

**Conflict of interest** The authors declare no conflict of interest. Although Philips is a partner in the Newborn project, the company has no influence on or participation in the analyses and publication of the results of the data from their devices.

**Informed consent** Informed consent was obtained from all individuals included in the study.

**Research involving human participants** The Lifelines NEXT study was approved by the ethics committee of the University Medical Center Groningen, document number METC UMCG METc2015/600.

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# References

 Barker DJ, Osmond C. Infant mortality, childhood nutrition, and ischaemic heart disease in England and Wales. Lancet. 1986;327:1077-81.

- Barker DJ, Osmond C, Winter PD, Margetts B, Simmonds SJ. Weight in infancy and death from ischaemic heart disease. Lancet. 1989;334:577–80.
- Barker DJ, Godfrey KM, Gluckman PD, Harding JE, Owens JA, Robinson JS. Fetal nutrition and cardiovascular disease in adult life. Lancet. 1993;341:938

  –41.
- Sutton EF, Gilmore LA, Dunger DB, Heijmans BT, Hivert M, Ling C, et al. Developmental programming: state-of-the-science and future directions—Summary from a Pennington Biomedical symposium. Obesity. 2016;24:1018–26.
- Waterland RA, Garza C. Potential mechanisms of metabolic imprinting that lead to chronic disease. Am J Clin Nutr. 1999;69:179–97.
- Godfrey KM, Gluckman PD, Hanson MA. Developmental origins of metabolic disease: life course and intergenerational perspectives. Trends Endocrinol Metab. 2010;21:199–205.
- Aiken CE, Ozanne SE. Transgenerational developmental programming. Hum Reprod Update. 2013;20:63–75.
- Birth cohorts: www.birthcohorts.net (2019). Accessed 20 May 2019.
- Burcelin R. Gut microbiota and immune crosstalk in metabolic disease. Mol Metab. 2016;5:771–81.
- 10. Perined. Perinatale Zorg in Nederland 2016. Utrecht; 2018.
- Peters LL, Thornton C, Jonge A, Khashan A, Tracy M, Downe S, et al. The effect of medical and operative birth interventions on child health outcomes in the first 28 days and up to 5 years of age: a linked data population-based cohort study. Birth. 2018;45:347–57.
- Oosting A, van Vlies N, Kegler D, Schipper L, Abrahamse-Berkeveld M, Ringler S, et al. Effect of dietary lipid structure in early postnatal life on mouse adipose tissue development and function in adulthood. Br J Nutr. 2014;111:215–26.
- Oosting A, Kegler D, Wopereis HJ, Teller IC, van de Heijning Bert JM, Verkade HJ, et al. Size and phospholipid coating of lipid droplets in the diet of young mice modify body fat accumulation in adulthood. Pediatr Res. 2012;72:362.
- Bourlieu C, Michalski M. Structure-function relationship of the milk fat globule. Curr Opin Clin Nutr Metab Care. 2015;18:118-27.
- Plagemann A, Harder T. Breast feeding and the risk of obesity and related metabolic diseases in the child. Metab Syndr Relat Disord. 2005;3:222–32.
- Scholtens S, Smidt N, Swertz MA, Bakker SJ, Dotinga A, Vonk JM, et al. Cohort Profile: lifeLines, a three-generation cohort study and biobank. Int J Epidemiol. 2014;44:1172–80.
- Stolk RP, Rosmalen JG, Postma DS, de Boer RA, Navis G, Slaets JP, et al. Universal risk factors for multifactorial diseases. Eur J Epidemiol. 2008;23:67–74.
- Francioli LC, Menelaou A, Pulit SL, Van Dijk F, Palamara PF, Elbers CC, et al. Whole-genome sequence variation, population structure and demographic history of the Dutch population. Nat Genet. 2014;46:818.
- Tigchelaar EF, Zhernakova A, Dekens JA, Hermes G, Baranska A, Mujagic Z, et al. Cohort profile: lifeLines DEEP, a prospective, general population cohort study in the northern Netherlands: study design and baseline characteristics. BMJ Open. 2015;5:e006772.
- Kunz B, Oranje AP, Labreze L, Stalder J, Ring J, Taieb A. Clinical validation and guidelines for the SCORAD index: consensus report of the European Task Force on Atopic Dermatitis. Dermatology. 1997;195:10–9.
- Katier N, Uiterwaal CS, de Jong BM, Kimpen JL, van der Ent Cornelis K. Feasibility and variability of neonatal and infant lung function measurement using the single occlusion technique. Chest. 2005;128:1822–9.
- van Putte-Katier N, Koopmans M, Uiterwaal CS, de Jong BM, Kimpen JL, Verheij TJ, et al. Relationship between parental lung



- function and their children's lung function early in life. Eur Respir J. 2011;38:664–71.
- Noss I, Wouters IM, Visser M, Heederik DJ, Thorne PS, Brunekreef B, et al. Evaluation of a low-cost electrostatic dust fall collector for indoor air endotoxin exposure assessment. Appl Environ Microbiol. 2008;74:5621–7.
- Geurts SA, Taris TW, Kompier MA, Dikkers JS, Van Hooff ML, Kinnunen UM. Work-home interaction from a work psychological perspective: development and validation of a new questionnaire, the SWING. Work Stress. 2005;19:319–39.
- Zimet GD, Dahlem NW, Zimet SG, Farley GK. The multidimensional scale of perceived social support. J Pers Assess. 1988:52:30–41.
- van Sonderen E. Sociale Steun Lijst–Interacties (SSL-I) en Sociale Steun Lijst-Discrepanties (SSL-D); 1993.
- Sheehan DV, Lecrubier Y, Sheehan KH, Amorim P, Janavs J, Weiller E, et al. The Mini-International Neuropsychiatric Interview (M.I.N.I.): the development and validation of a structured diagnostic psychiatric interview for DSM-IV and ICD-10. J Clin Psychiatry. 1998;59(Suppl 20):2–57.
- Condon JT, Corkindale CJ. The assessment of parent-to-infant attachment: development of a self-report questionnaire instrument. J Reprod Infant Psychol. 1998;16:57–76.
- Condon JT. The assessment of antenatal emotional attachment: development of a questionnaire instrument. Br J Med Psychol. 1993;66:167–83.
- Rosmalen J, Bos EH, De Jonge P. Validation of the Long-term Difficulties Inventory (LDI) and the List of Threatening Experiences (LTE) as measures of stress in epidemiological population-based cohort studies. Psychol Med. 2012;42:2599–608.
- Brugha TS, Cragg D. The list of threatening experiences: the reliability and validity of a brief life events questionnaire. Acta Psychiatr Scand. 1990;82:77–81.
- Dencker A, Taft C, Bergqvist L, Lilja H, Berg M. Childbirth experience questionnaire (CEQ): development and evaluation of a multidimensional instrument. BMC Pregnancy Childbirth. 2010:10:81.
- Streppel MT, de Vries JH, Meijboom S, Beekman M, de Craen AJ, Slagboom PE, et al. Relative validity of the food frequency questionnaire used to assess dietary intake in the Leiden Longevity Study. Nutr J. 2013;12:75.
- Whitehead WE, Palsson OS, Thiwan SM, Talley NJ, Chey W, Irvine EJ, et al. Development and validation of the Rome III diagnostic questionnaire. In: Rome III: the functional gastrointestinal disorders. 3rd ed. McLean, VA: Degnon Associates, Inc; 2006. p. 835–53.
- Rijlaarsdam J, Stevens GW, Van der Ende J, Arends LR, Hofman A, Jaddoe VW, et al. A brief observational instrument for the assessment of infant home environment: development and psychometric testing. Int J Methods Psychiatr Res. 2012;21:195–204.
- Squires J, Potter L, Bricker D. Ages and Stages Questionnaire user's guide. Baltimore: Brookes; 1999.
- Gartstein MA, Rothbart MK. Studying infant temperament via the revised infant behavior questionnaire. Infant Behav Dev. 2003;26:64–86.
- Briggs-Gowan MJ, Carter AS, Irwin JR, Wachtel K, Cicchetti DV. The Brief Infant-Toddler Social and Emotional Assessment: screening for social-emotional problems and delays in competence. J Pediatr Psychol. 2004;29:143–55.
- 39. Derogatis LR, Unger R. Symptom checklist-90-revised. The Corsini encyclopedia of psychology. 2010, p. 1–2.
- Wendel-Vos GW, Schuit AJ, Saris WH, Kromhout D. Reproducibility and relative validity of the short questionnaire to assess health-enhancing physical activity. J Clin Epidemiol. 2003;56:1163–9.

- 41. Brouwer-Brolsma E, Streppel M, van Lee L, Geelen A, Sluik D, van de Wiel A, et al. A national dietary assessment reference database (ndard) for the dutch population: rationale behind the design. Nutrients. 2017;9:1136.
- Burney PG, Luczynska C, Chinn S, Jarvis D. The European community respiratory health survey. Eur Respir J. 1994;7:954

  –60.
- Mangione CM, Lee PP, Gutierrez PR, Spritzer K, Berry S, Hays RD. Development of the 25-list-item national eye institute visual function questionnaire. Arch Ophthalmol. 2001;119:1050–8.
- Wolfe F, Clauw DJ, Fitzcharles M, Goldenberg DL, Häuser W, Katz RS, et al. Fibromyalgia criteria and severity scales for clinical and epidemiological studies: a modification of the ACR Preliminary Diagnostic Criteria for Fibromyalgia. J Rheumatol. 2011;38:1113–22.
- Vercoulen JH, Swanink CM, Fennis JF, Galama JM, van der Meer Jos WM, Bleijenberg G. Dimensional assessment of chronic fatigue syndrome. J Psychosom Res. 1994;38:383–92.
- Pang C, van Enckevort D, de Haan M, Kelpin F, Jetten J, Hendriksen D, et al. MOLGENIS/connect: a system for semi-automatic integration of heterogeneous phenotype data with applications in biobanks. Bioinformatics. 2016;32:2176–83.
- 47. Regulation GDP. Regulation (EU) 2016/679 of the European Parliament and of the Council of 27 April 2016 on the protection of natural persons with regard to the processing of personal data and on the free movement of such data, and repealing Directive 95/46. Off J Eur Union (OJ). 2016;59:294.
- 48. Fu J, Bonder MJ, Cenit MC, Tigchelaar EF, Maatman A, Dekens JA, et al. The gut microbiome contributes to a substantial proportion of the variation in blood lipids novelty and significance. Circ Res. 2015;117:817–24.
- Zhernakova A, Kurilshikov A, Bonder MJ, Tigchelaar EF, Schirmer M, Vatanen T, et al. Population-based metagenomics analysis reveals markers for gut microbiome composition and diversity. Science. 2016;352:565–9.
- CBS Statline: https://opendata.cbs.nl/statline/#/CBS/nl/ (2019).
   Accessed 26 Feb 2019.
- Méray N, Reitsma JB, Ravelli AC, Bonsel GJ. Probabilistic record linkage is a valid and transparent tool to combine databases without a patient identification number. J Clin Epidemiol. 2007;60:883.e11.
- Bos-Veneman NG, Otter M, Reijneveld SA. Using feeding to reduce pain during vaccination of formula-fed infants: a randomised controlled trial. Arch Dis Child. 2018;103:1132–7.
- Dykstra PA, Kalmijn M, Knijn TC, Komter AE, Liefbroer AC, Mulder CH. Questionnaires of the Netherlands Kinship Panel Study, a multi-actor, multi-method panel study on solidarity in family relationships, Wave 1. 2005.
- 54. Wijga AH, Kerkhof M, Gehring U, de Jongste JC, Postma DS, Aalberse RC, et al. Cohort profile: the prevention and incidence of asthma and mite allergy (PIAMA) birth cohort. Int J Epidemiol. 2013;43:527–35.
- Trails NEXT: https://www.trails.nl/en/hoofdmenu/participants/ trails-next (2019). Accessed 20 May 2019.
- NeoLifeS. https://www.umcg.nl/EN/Research/Researchers/Healt hyageing/geras/Paginas/neolifes.aspx (2019). Accessed 20 May 2019.
- Klijs B, Scholtens S, Mandemakers JJ, Snieder H, Stolk RP, Smidt N. Representativeness of the LifeLines Cohort Study. PLoS ONE. 2015;10:e0137203.
- Dahlen HG, Downe S, Wright ML, Kennedy HP, Taylor JY. Childbirth and consequent atopic disease: emerging evidence on epigenetic effects based on the hygiene and EPIIC hypotheses. BMC Pregnancy Childbirth. 2016;16:4.
- Dahlen HG, Kennedy HP, Anderson CM, Bell AF, Clark A, Foureur M, et al. The EPIIC hypothesis: intrapartum effects on



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the neonatal epigenome and consequent health outcomes. Med Hypotheses. 2013;80:656–62.

- Xu C, Bonder MJ, Söderhäll C, Bustamante M, Baïz N, Gehring U, et al. The emerging landscape of dynamic DNA methylation in early childhood. BMC Genomics. 2017;18:25.
- Horta BL, Gigante DP, Lima RC, Barros FC, Victora CG. Birth by caesarean section and prevalence of risk factors for non-communicable diseases in young adults: a birth cohort study. PLoS ONE. 2013:8:e74301.
- Papacleovoulou G, Abu-Hayyeh S, Nikolopoulou E, Briz O, Owen BM, Nikolova V, et al. Maternal cholestasis during pregnancy programs metabolic disease in offspring. J Clin Investig. 2013;123:3172–81.
- 63. Martineau MG, Raker C, Dixon PH, Chambers J, Machirori M, King NM, et al. The metabolic profile of intrahepatic cholestasis of pregnancy is associated with impaired glucose tolerance, dyslipidemia, and increased fetal growth. Diabetes Care. 2015;38:243–8.
- Chu DM, Ma J, Prince AL, Antony KM, Seferovic MD, Aagaard KM. Maturation of the infant microbiome community structure

- and function across multiple body sites and in relation to mode of delivery. Nat Med. 2017;23:314.
- Chu DM, Antony KM, Ma J, Prince AL, Showalter L, Moller M, et al. The early infant gut microbiome varies in association with a maternal high-fat diet. Genome Med. 2016;8:77.
- Sharp GC, Salas LA, Monnereau C, Allard C, Yousefi P, Everson TM, et al. Maternal BMI at the start of pregnancy and offspring epigenome-wide DNA methylation: findings from the pregnancy and childhood epigenetics (PACE) consortium. Hum Mol Genet. 2017;26:4067–85.
- Virgin HW. The virome in mammalian physiology and disease. Cell. 2014;157:142–50.
- EUCAN Connect: https://www.eucanconnect.eu/ (2019).
   Accessed 2 Dec 2019.

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