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Data Article

Data on Western blot and ELISA analysis of medaka (*Oryzias latipes*) follicle-stimulating hormone (Fsh) and luteinizing hormone (Lh) using recombinant proteins expressed with *Pichia pastoris*



Susann Burow^a, Romain Fontaine^{a,1}, Kristine von Krogh^{a,1}, Ian Mayer^b, Rasoul Nourizadeh-Lillabadi^a, Lian Hollander-Cohen^c, Yaron Cohen^c, Michal Shpilman^c, Berta Levavi-Sivan^c, Finn-Arne Weltzien^{a,*}

^a Department of Basic Sciences and Aquatic Medicine, Faculty of Veterinary Medicine, Norwegian University of Life Sciences, 0454 Oslo, Norway

^b Department of Production Animal Clinical Sciences, Faculty of Veterinary Medicine, Norwegian University of Life Sciences, 0454 Oslo, Norway

^c Department of Animal Sciences, Faculty of Agriculture, Food and Environment, The Hebrew University, Rehovot 76100, Israel

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ABSTRACT

The gonadotropins follicle-stimulating hormone (Fsh) and luteinizing hormone (Lh) play essential roles in vertebrate reproduction. This article presents data on molecular weight validation of recombinant medaka (*Oryzias latipes*) (md) gonadotropins Fsh β (mdFsh β), Lh β (mdLh β), Fsh $\beta\alpha$ (mdFsh $\beta\alpha$), and Lh $\beta\alpha$ (mdLh $\beta\alpha$) generated by *Pichia pastoris*, as well as data on a validation of produced antibodies against Fsh β and Lh β by Western blot analysis. Furthermore, the article includes data on Fsh and Lh protein levels in male medaka pituitaries using recombinant mdFsh $\beta\alpha$ and mdLh $\beta\alpha$ within enzyme-linked immunosorbent assays (ELISAs), in which protein amounts were analyzed related to body weight and age of the fish. This dataset is associated with the research article entitled “Medaka Follicle-stimulating hormone (Fsh) and

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* Corresponding author.

E-mail address: finn-arne.weltzien@nmbu.no (F.-A. Weltzien).

¹ These authors contributed equally

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Luteinizing hormone (Lh): Developmental profiles of pituitary protein and gene expression” (Burow et al., in press).

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Specifications table

Subject area	Biology
More specific subject area	Physiology, Neuroendocrinology
Type of data	Image (Western blot analysis), Graph (ELISA)
How data was acquired	Data for validation of recombinant proteins and antibodies were acquired through Western blot, data for protein levels were obtained through ELISA using microplate spectrophotometer.
Data format	Analyzed
Experimental factors	Prior to Western blot analysis, <i>N</i> -glycosidase F was used to produce deglycosylated proteins by hydrolyzing all types of <i>N</i> -glycan chains.
Experimental features	Validation of medaka recombinant proteins and antibodies, and generation of pituitary Fsh and Lh levels in male medaka was performed.
Data source location	Department of Basic Sciences and Aquatic Medicine, Faculty of Veterinary Medicine, Norwegian University of Life Sciences, 0454 Oslo, Norway.
Data accessibility	Data are presented in this article.
Related research article	Burow, S., Fontaine, R., von Krogh, K., Mayer, I., Nourizadeh-Lillabadi, R., Hollander-Cohen, L., Cohen, Y., Shpilman, M., Levavi-Sivan, B., Weltzien, F.A., Medaka Follicle-stimulating hormone (Fsh) and Luteinizing hormone (Lh): Developmental profiles of pituitary protein and gene expression levels, <i>Gen. Comp. Endocrinol.</i> (in press) [1].

Value of the data

- The establishment of competitive ELISAs using recombinant medaka gonadotropins to quantify the content of Fsh and Lh, for the first time, extends the accessibility of quantitative methods for medaka and enables advanced functional studies on gonadotropin physiology in fish.
- The generated ELISA data determining pituitary Fsh and Lh protein levels in male fish during development in this article represent valuable data and a tool for future studies, since investigations in male fish during puberty are quite limited until today.
- The data on Fsh and Lh protein levels in male medaka pituitaries using recombinant mdFsh $\beta\alpha$ and mdLh $\beta\alpha$ reveal that body weight explains the variance in the dependent variable (gonadotropin) better compared to age of the fish for Fsh β and Lh β . In addition, body weight is indicated to explain the variance in the dependent variable for Lh β better compared to Fsh β .
- The generation of specific antibodies against medaka Fsh β and Lh β presented here will be a valuable tool for future experiments on gonadotropins in medaka, an important model organism in biology.

1. Data

The data on characterization of recombinant medaka (md) gonadotropins Fsh β (mdFsh β) (Fig. 1A), Lh β (mdLh β) (Fig. 1B), Fsh $\beta\alpha$ (mdFsh $\beta\alpha$) (Fig. 1C), and Lh $\beta\alpha$ (mdLh $\beta\alpha$) (Fig. 1D) by immunoreacting

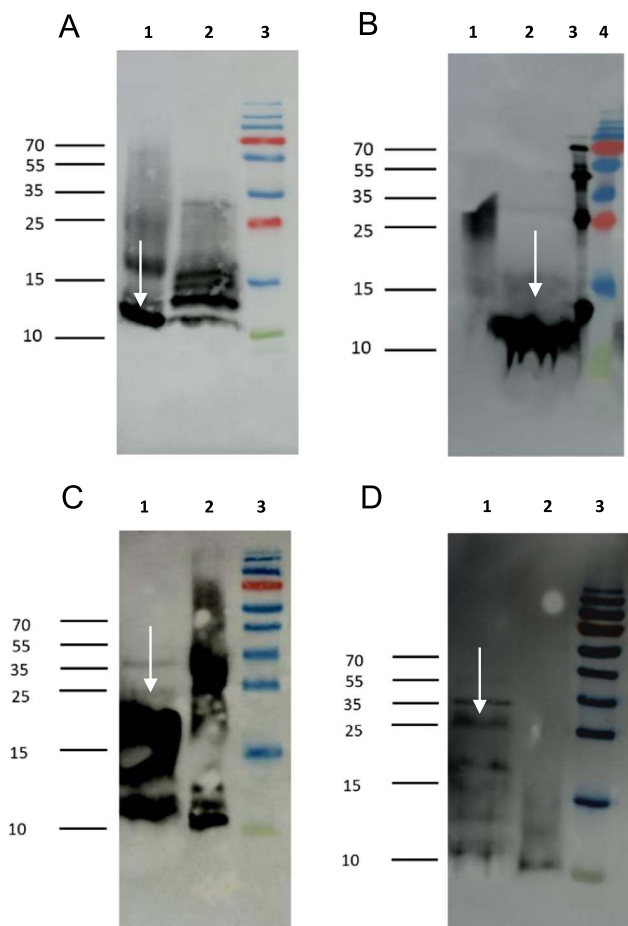


Fig. 1. Molecular weight validation of mdFsh β , mdLh β , mdFsh $\beta\alpha$, and mdLh $\beta\alpha$ expressed with *P. pastoris* by Western blot analysis. Supernatants of transformed *P. pastoris* cultures were separated by SDS-PAGE and immunoreacted with antibodies against His. PageRuler Plus Prestained Protein Ladder lane 3 (A), lane 4 (B), lane 3 (C), lane 3 (D). His-tagged Protein Standard lane 3 (B). The Western blot confirmed the expected molecular weight of A) mdFsh β , Lane 2 represents mdFsh β ; Lane 1 represents deglycosylated mdFsh β . B) mdLh β , Lane 1 represents mdLh β ; Lane 2 represents deglycosylated mdLh β C) mdFsh $\beta\alpha$, Lane 2 represents mdFsh $\beta\alpha$; Lane 1 represents deglycosylated mdFsh $\beta\alpha$ D) mdLh $\beta\alpha$, Lane 2 represents mdLh $\beta\alpha$; Lane 1 represents deglycosylated mdLh $\beta\alpha$. White arrows indicate protein bands after deglycosylation with PNGase F.

them against the His-tag demonstrated a clear validation since all recombinant proteins were successfully detected with His-tail antibodies, and their molecular sizes derived from Western blots were in accordance with the calculated estimates (according to sequence). Under reducing conditions, mdFsh β and mdFsh $\beta\alpha$ were detected as bands of 14–16 kDa (Fig. 1A) and 25–30 kDa (Fig. 1C), respectively, and after deglycosylation with PNGase F as bands of 12–14 kDa (Fig. 1A) and 24–25 kDa (Fig. 1C), respectively. This is in accordance with the calculated molecular weight without glycosylation residues for mdFsh β (13 kDa) and for mdFsh $\beta\alpha$ (25 kDa). Under reducing conditions, mdLh β and mdLh $\beta\alpha$ had a molecular weight of 15 kDa (Fig. 1B) and 35 kDa (Fig. 1D), respectively, and after deglycosylation 12–14 kDa (Fig. 1B) and 27–28 kDa (Fig. 1D), respectively. Again, this was in accordance with the expectation for deglycosylated mdLh β (15 kDa) and mdLh $\beta\alpha$ (28 kDa).

Western blot analysis of antibodies produced against medaka Fsh β and Lh β revealed specificity and absence of cross-reactions as all recombinant proteins mdFsh β , mdLh β , mdFsh $\beta\alpha$, and mdLh $\beta\alpha$ were detected exclusively with antibodies against either medaka Fsh β (Fig. 2A, B) or Lh β (Fig. 2C, D). Under

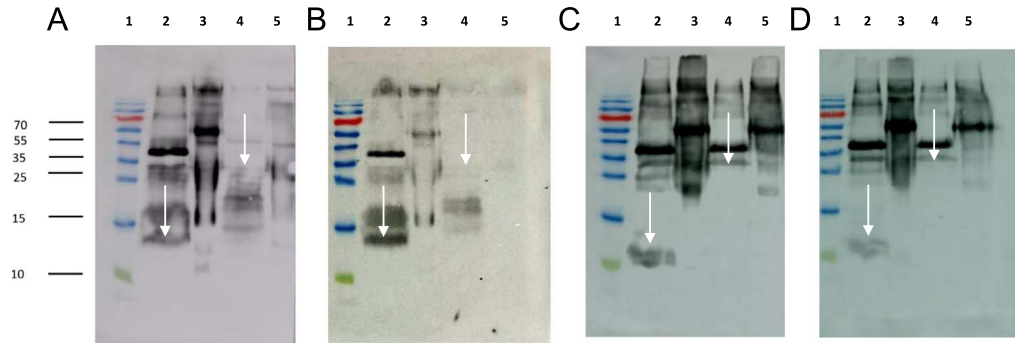


Fig. 2. Validation of recombinant proteins mdFsh β , mdLh β , mdFsh $\beta\alpha$, and mdLh $\beta\alpha$ expressed with *P. pastoris* by Western blot analysis. Supernatants of transformed *P. pastoris* cultures were separated by SDS-PAGE and immunoreacted with antibodies against mdFsh β (2A, 2B) and mdLh β (2C, 2D). First lane represents PageRuler Plus Prestained Protein Ladder. The Western blot confirmed that the antibodies detected the correct proteins, and verified the absence of cross-reactions. A and B) mdFsh β and mdFsh $\beta\alpha$, Antibody against mdFsh β , Dilution 1:100.000 (2A) and 1:600.000 (2B); Lane 3 represents mdFsh β , lane 5 represents mdFsh $\beta\alpha$; Lanes 2 and 4 represent deglycosylated samples of those shown in lanes 3 and 5 respectively. C and D) mdLh β and mdLh $\beta\alpha$, Antibody against mdLh β , Dilution 1:100.000 (2C) and 1:600.000 (2D); Lane 3 represents mdLh β , lane 5 represents mdLh $\beta\alpha$; Lanes 2 and 4 represent deglycosylated samples of those shown in lanes 3 and 5 respectively. White arrows indicate protein bands after deglycosylation with PNGase F.

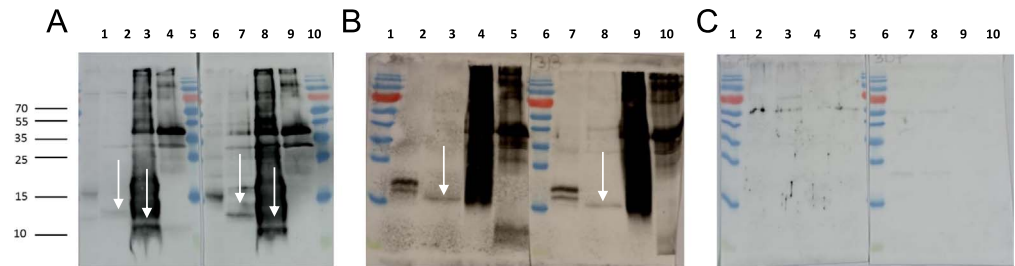


Fig. 3. Validation of the produced antibodies against mdFsh β and mdLh β and characterization of medaka pituitary extract, mdFsh β , and mdLh β expressed with *P. pastoris* by Western blot analysis. Supernatants of transformed *P. pastoris* cultures were separated by SDS-PAGE and immunoreacted with antibodies against mdFsh β (3A) and mdLh β (3B) and with medaka pre-immune serum (3C). The Western blot confirmed that the antibodies detected proteins of the right size in medaka pituitaries. A) Antibody against mdFsh β , Dilution 1:2000; Lane 1 (Rabbit 1 (R1)) and 6 (Rabbit 2 (R2)) represent medaka pituitary extract, lane 2 (R1) and 7 (R2) represent medaka pituitary extract after deglycosylation; Lanes 3 (R1) and 8 (R2) represent mdFsh β after deglycosylation; Lanes 4 (R1) and 9 (R2) represent deglycosylated samples of mdLh β . Lane 5 and 10 represent PageRuler Plus Prestained Protein Ladder. B) Antibody against mdLh β , Dilution 1:2000; Lane 2 (R1) and 7 (R2) represent medaka pituitary extract, lane 3 (R1) and 8 (R2) represent medaka pituitary extract after deglycosylation; Lanes 4 (R1) and 9 (R2) represent mdLh β after deglycosylation; Lanes 5 (R1) and 10 (R2) represent deglycosylated samples of mdFsh β . Lane 1 and 6 represent PageRuler Plus Prestained Protein Ladder. C) The Western blot confirmed the validation of the produced antibodies and verified that the plasma taken before the final injections did not react with mdFsh β and mdLh β . Medaka pituitary extract, mdFsh β , and mdLh β were immunoreacted against medaka pre-immune serum as a negative control (test bleeding). Negative control: Pre-immune serum of Rabbit 1 (3C, lane 1 to 5) and Rabbit 2 (3C, lane 6 to 10); Lane 2 (R1) and 7 (R2) represent medaka pituitary extract, lane 3 (R1) and 8 (R2) represent medaka pituitary extract after deglycosylation; Lanes 4 (R1) and 9 (R2) represent mdFsh β after deglycosylation; Lanes 5 (R1) and 10 (R2) represent deglycosylated samples of mdLh β . Lane 1 and 6 represent PageRuler Plus Prestained Protein Ladder. White arrows indicate protein bands after deglycosylation with PNGase F.

reducing conditions and after deglycosylation, mdFsh β and mdFsh $\beta\alpha$ were determined as bands of 12–13 kDa and 23–25 kDa (Fig. 2A, B), respectively. mdLh β was revealed after deglycosylation very weakly as a band of 12–13 kDa, and mdLh $\beta\alpha$ was observed as a band of 27–29 kDa (Fig. 2C, D).

When using the antibodies on medaka pituitary extracts, native mdFsh β (Fig. 3A) and mdLh β (Fig. 3B) could be detected. Using the mdFsh β antibody, bands of approximately 13 kDa were revealed for mdFsh β (Fig. 3A). When using the mdLh β antibody, there was no clean band for mdLh β due to very strong signals (Fig. 3B). No bands were revealed for mdLh β with the mdFsh β antibody (Fig. 3A) and no

bands for mdFsh β using the mdLh β antibody (Fig. 3B). When medaka pituitary extract, recombinant mdFsh β , or recombinant mdLh β were immunoreacted against rabbit pre-immune serum as a negative control (test bleeding), there was no specific band observed (Fig. 3C).

Furthermore, this article provides data on Fsh and Lh protein levels in pituitaries from juvenile and adult male medaka that were obtained by enzyme-linked immunosorbent assay (ELISA). The data have been analyzed as a function of body weight (Fsh Fig. 4A, Lh Fig. 4C) and age of the fish (Fsh Fig. 4B, Lh Fig. 4D). Body weight ($R^2 = 0,3276$; Fig. 4A) explains the variance in the dependent variable (gonadotropin) better compared to age of the fish ($R^2 = 0,2499$; Fig. 4B) or body length (protein levels in relation to body length has been shown in Burow et al. [1]) for Fsh β using a linear trendline. As for Fsh β , body weight ($R^2 = 0,6221$; Fig. 4C) explains the variance in the dependent variable better compared to age ($R^2 = 0,524$; Fig. 4D) for Lh β using a power trendline. Notably, the R^2 s are higher for Lh β than for Fsh β , indicating that body weight explains the variance in the dependent variable for Lh β better compared to Fsh β . Since none of the R^2 is close to 1, a correlation of Fsh/Lh levels to either body weight or age of the fish is not indicated.

2. Experimental design, materials and methods

2.1. Animals

Japanese medaka (*Oryzias latipes*) of the dr-R strain were kept in re-circulating systems with light-dark cycle of L14:D10 and water temperature of 28 ± 1 °C. Embryos were incubated in embryo culture medium (E3; 5 mM NaCl, 0.17 mM KCl, 0.33 mM CaCl₂, 0.33 mM MgSO₄ (all Sigma-Aldrich, St. Louis, U.S.A.)), and kept at 26 °C until hatching and transfer to system tanks. The fish were fed three times per day with a combination of dry feed and live brine shrimp nauplii larvae (*Artemia salina*). Fish were raised under the same conditions with regard to temperature, photoperiod, food, tank size, and density. Handling, husbandry and use of fish were according to the guidelines and requirements for the care and welfare of research animals of the Norwegian Animal Health Authority and of the Norwegian University of Life Sciences. The work of the present article has been carried out in accordance with the EU Directive 2010/63/EU for animal experiments and Uniform Requirements for manuscripts submitted to Biomedical journals, and informed consent was obtained for experimentation with animal subjects.

2.2. Production and purification of recombinant gonadotropins mdFsh β , mdLh β , mdFsh $\beta\alpha$, and mdLh $\beta\alpha$, generation of specific antibodies for mdFsh β and mdLh β , and Western blot analysis

Generation of recombinant proteins was conducted using the methylotrophic yeast *Pichia pastoris* (*P. pastoris*) expression system, generally according to Kasuto and Levavi-Sivan [2] and Yom-Din et al. [3], and described in detail in Burow et al. [1]. Synthesis of genes for medaka *fshb* (Accession Number NM_001309017.1), *lhb* (Accession Number AB541982.1), *fshba*, and *lhba* (*gpa*; Accession Number NM_001122906) was outsourced to GenScript, New Jersey, U.S.A. For each construct gene expression cassettes were generated with *P. pastoris* codon optimized DNA sequence. Polyclonal antisera against recombinant mdFsh β and mdLh β were produced following a procedure according to Aizen et al. [4], which is reported in detail in Burow et al. [1].

For molecular weight validation, the purified recombinant proteins were analyzed by Western blot analysis using anti-His (diluted 1:2000), generally according to Yom-Din et al. [3]. To validate the produced antibodies, the recombinant proteins and medaka pituitary extract were visualized using anti-mdFsh β , or anti-mdLh β (both diluted 1:2000, 1:100000, 1:600000) antisera. To confirm that the plasma of the rabbit before the final injections did not react with mdFsh β and mdLh β , a Western blot using medaka pre-immune serum as a negative control against medaka pituitary extract, mdFsh β , and mdLh β was performed.

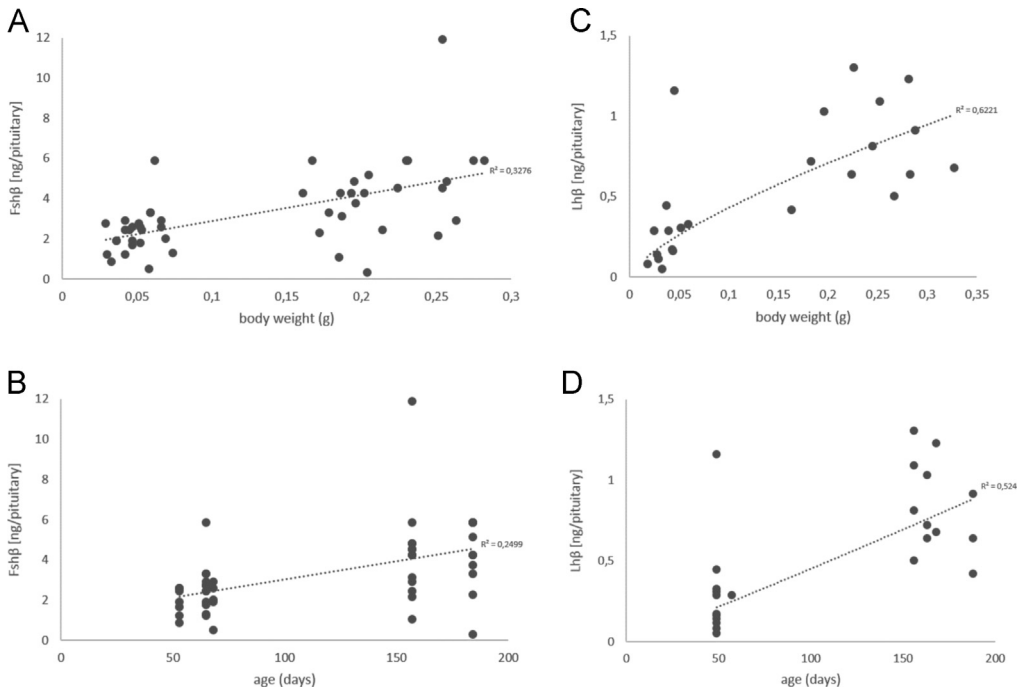


Fig. 4. Profile of Fsh and Lh protein levels in pituitaries from juvenile and adult male medaka as a functional study obtained by ELISA. Quantified amounts of Fsh β (Fig. 4A, B) and Lh β (Fig. 4C, D) (in ng/pituitary). Data have been analyzed as a function of body weight (Fsh Fig. 4A, linear trendline; Lh Fig. 4C, power trendline), and age of the fish (Fsh Fig. 4B, linear trendline; Lh Fig. 4D, power trendline). Body weight ($R^2 = 0,3276$; Fig. 4A) explains the variance in the dependent variable (gonadotropin) better compared to age of the fish ($R^2 = 0,2499$; Fig. 4B) for Fsh β using a linear trendline. As for Fsh β , body weight ($R^2 = 0,6221$; Fig. 4C) explains the variance in the dependent variable better compared to age ($R^2 = 0,524$; Fig. 4D) for Lh β using a power trendline. Comparing the R^2 of body weight and age between the Fsh β and Lh β profiles, it is important to note that the R^2 's are higher for Lh β than for Fsh β , indicating that body weight explains the variance in the dependent variable for Lh β better compared to Fsh β .

2.3. Quantification of Fsh and Lh in male medaka pituitaries using ELISA

To quantify the content of Fsh and Lh protein levels in male medaka pituitaries, the ELISA methodology described in Burow et al. [1] was performed. For the profile of Fsh, pituitaries from 24 juvenile males with standard length (SL) between 12 mm and 16.5 mm, and of 24 adult males between 21 mm and 25.5 mm were used. Pituitaries from 12 juvenile males with SL between 12 mm and 16 mm, and of 12 adult males between 22.5 mm and 26.5 mm were dissected for the profile of Lh. For both Fsh and Lh 1 pituitary in 40 μ l 0.1% BSA in PBST per biological replicate was used. Within the two groups juveniles and adults, body weight and age were measured, and protein amounts were analyzed related to body weight and age of the fish.

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Transparency document. Supporting information

Transparency data associated with this article can be found in the online version at <https://doi.org/10.1016/j.dib.2019.01.034>.

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