

Implicating androgen excess in propagating metabolic disease in polycystic ovary syndrome

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Abstract: Polycystic ovary syndrome (PCOS) has been traditionally perceived as a reproductive disorder due to its most common presentation with menstrual dysfunction and infertility. However, it is now clear that women with PCOS are at increased risk of metabolic dysfunction, from impaired glucose tolerance and type 2 diabetes mellitus to nonalcoholic fatty liver disease and cardiovascular disease. PCOS is characterised by androgen excess, with cross-sectional data showing that hyperandrogenism is directly complicit in the development of metabolic complications. Recent studies have also shown that C11-oxy_{C19} androgens are emerging to be clinically and biochemically significant in PCOS, thus emphasising the importance of understanding the impact of both classic and C11-oxy_{C19} androgens on women's health. Here we discuss androgen metabolism in the context of PCOS, and dissect the role played by androgens in the development of metabolic disease through their effects on metabolic target tissues in women.

Keywords: adipose tissue, androgens, C11-oxy C19 androgens, diabetes, metabolic disease, obesity, PCOS

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Introduction

Polycystic ovary syndrome (PCOS) is one of the most common endocrine conditions, affecting approximately 10% of all women across the globe.¹ Women with PCOS commonly present with menstrual irregularities, infertility and signs and symptoms of androgen excess. PCOS has been traditionally perceived as a predominantly reproductive disorder, with minimal focus on long-term metabolic complications. However, recent studies have shown that PCOS is associated with significant health consequences in affected women,² representing a lifelong metabolic disorder (Figure 1).

Although controversies persist with regard to diagnostic criteria for PCOS, androgen excess remains a fundamental biological and diagnostic feature of the disorder.^{3,4} This is recognised in the 2003 Rotterdam Consensus criteria, with androgen excess featuring as a defining criterion alongside irregular periods and polycystic appearances of the ovaries at ultrasound. Following

this, the Androgen Excess Society recommended hyperandrogenism as a mandatory criterion for the diagnosis of PCOS, further highlighting PCOS as a disorder of androgen excess.^{4–6} In cross-sectional clinical phenotyping as well as population studies, androgen excess is a major driver of metabolic risk in PCOS, with the presence and severity of androgen excess closely correlating with surrogate parameters of metabolic risk, including insulin resistance.⁷ Women with PCOS have been shown to be at increased risk of metabolic disease including type 2 diabetes mellitus (T2DM), nonalcoholic fatty liver disease (NAFLD), and cardiovascular disease (CVD).⁸

Most studies on PCOS have focused on classic C₁₉ androgens, their precursors and/or their downstream urinary metabolites for biochemical assessment of androgen excess. However, the newly described, adrenal-derived C11-oxy C₁₉ androgen subclass is emerging to be clinically and biochemically significant in the context of PCOS-related

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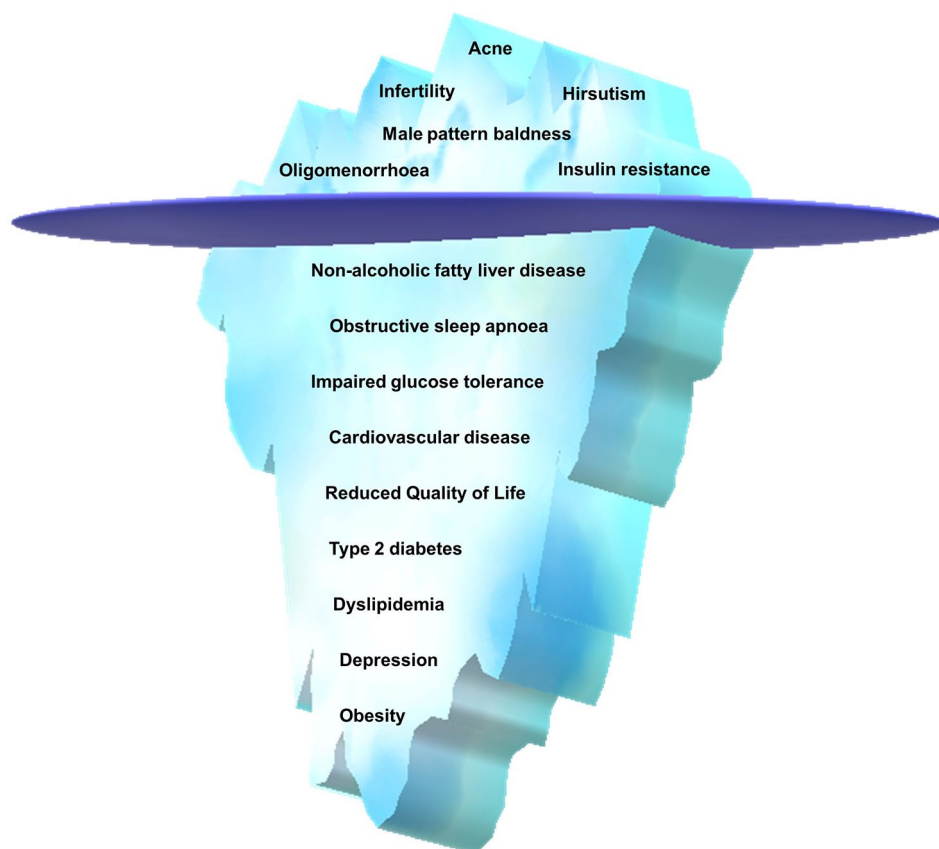


Figure 1. Iceberg phenomenon: PCOS has been traditionally perceived as a primarily reproductive disorder. On a superficial level, clinical consequences of this condition include subfertility, irregular menses and signs and symptoms of androgen excess. However, clinicians should be aware of the significantly increased risk of metabolic complications across the female lifespan in women with PCOS, underpinned by androgen excess and insulin resistance.

androgen excess.⁹ In our recent study, we found that C11-oxy C₁₉ androgens are the majority of circulating serum androgens in women with PCOS¹⁰, thus emphasising the importance of improving our understanding of the impact of both classic and C11-oxy C₁₉ androgens on women's health. Furthermore, circulating levels of the active C11-oxy C₁₉ androgen 11-ketotestosterone (11KT) have recently been shown to remain consistent throughout the female lifespan,¹¹ contrary to classic androgen concentrations, which decrease post-menopause. However, recent study by Stratakis *et al.*¹² analysing steroid hormones of adolescents and young women with PCOS and adrenocortical dysfunction using UPC2-MS/MS found C19 steroids [A4, testosterone and dehydroepiandrosterone (DHEA)] concentrations higher than the combined C11-oxy C₁₉ androgens. The combined C19 steroid levels were

higher in PCOS group with the combined C11-oxy androgens being similar in patients with PCOS and healthy people. Therefore, more studies on C11-oxy C₁₉ and their downstream metabolites will be needed before a conclusion can be drawn about the roles of C11-oxy C₁₉ androgens compared with the classic androgens.

Conditions characterised by androgen excess are PCOS, premature adrenarche, congenital adrenal hyperplasia, ovarian hyperthecosis, androgen secreting tumours and, to a degree, also Cushing's syndrome.^{13,14} Many of these conditions lack the necessary prevalence and frequency of life events and metabolic dysfunction to understand the overall impact of androgen excess.¹⁵ PCOS is a lifelong metabolic condition likely to affect female health from childhood through to post-menopause. Recent work by Risal *et al.* showed that

daughters of mothers with PCOS have a fivefold increased risk for PCOS.¹⁶ More importantly, they showed that prenatal androgen exposure and not obesity lead to transgenerational reproductive and metabolic dysfunction in rodent models.¹⁷ Another study by Gunning *et al.* showed subtle cardiometabolic dysfunction in early childhood in otherwise healthy weight children of women with PCOS.^{18,19} Based on its high prevalence in the general background population, and the correlation between PCOS and other features of metabolic syndrome, PCOS is a good model for studying the impact of androgen excess on metabolic risk. The origins of androgens and their mechanistic role inducing metabolic dysfunction in PCOS are discussed in detail in the following.

Review methodology

We included both *in vitro* and *in vivo* human and animal studies published in English up until March 2020 providing evidence on PCOS and metabolic dysfunctions. Articles were searched from PubMed using terms “(PCOS) OR (PCOS MeSH terms)” AND (Androgens) OR “(Androgens MeSH terms)” AND terms on metabolic dysfunctions as mentioned previously such as “Obesity”, “Diabetes” and with their associated MeSH terms. We also added specific organs such as “Adrenal”, “Ovary”, “Muscle”, “Pancreas”, “Liver” in separate searches to review their impact on PCOS and metabolic dysfunctions. Additional articles were also obtained from previously published reviews for respective subjects. Articles were then screened based on titles and abstracts and full texts were subsequently obtained.

Origins of androgen excess in PCOS and pre-receptor androgen metabolism

Androgens are responsible for the development of male characteristics such as male pattern body hair, muscle bulk, and deep voice; androgens also impact on sexual function and reproductive capacity in both sexes. In women, androgens also play an important role in general wellbeing, libido, energy levels, muscle mass and strength and bone mass.^{20–24} Apart from their role in reproduction and sexual health, androgens also play an important role in human metabolic health in both men and women.²⁵ The origins of androgen excess in PCOS have not been comprehensively delineated, but adrenals, ovaries and adipose tissue collectively contribute to the bulk of circulating

androgen burden in the disorder.⁸ Androgens are synthesized in the ovaries and adrenal glands catalysed by a series of steroidogenic enzymes, especially the rate limiting cytochrome P450 (CYP) 17A1 enzyme, responsible for the synthesis of the classic androgen pathway precursor DHEA. DHEA and its downstream product androstenedione are released into circulation, with androgenic signals further amplified after uptake into target tissues, where androgen precursor steroids are converted into the active androgens testosterone and 5 α -dihydrotestosterone (DHT), the most potent natural androgen.²⁶ Several studies have reported a systemic upregulation of 5 α -reductase activity as a significant feature in women with PCOS,^{27–31} with consequently increased activation of testosterone to 5 α -DHT. More recently, another pathway of DHT synthesis, referred to as ‘backdoor pathway’ has been discussed in the context of PCOS and congenital adrenal hyperplasia.^{32–34} This pathway involves the synthesis of DHT bypassing testosterone in the classic pathway. There are also speculations of androgens transferring *via* mother’s milk into children as an early source of androgens in neonatal and infant life. However, currently there is a lack of scientific evidence to support this theory. As PCOS is a heterogeneous condition associated with features of the metabolic syndrome, *in vivo* studies involving women with PCOS are often confounded by coexisting obesity and insulin resistance, which are also contributors to hyperandrogenism in women with and without PCOS.³⁵

Ovaries and androgen excess

The origin of androgen excess from the ovaries is complex and mainly attributed to tonic hypersecretion of Luteinizing Hormone (LH), likely due to a dysregulated hypothalamic–pituitary–gonadal axis.³⁶ A theory of insulin-potentiated accelerated gonadotropin-releasing hormone (GnRH) pulse frequency, resulting in increased LH pulse is supported by the works of Roland and Moenter.^{37,38} Insulin has also been shown to stimulate androgen productions in the ovarian theca cells of women with PCOS *via* insulin receptor rather than insulin growth factor 1 (IGF-1) receptor.^{39,40} However, the IGF-1 receptor may also play a role in conditions with extreme levels of insulin resistance such as hereditary insulin receptor mutations or lipodystrophy contributing to their hyperandrogenism and PCOS.^{41,42} High levels of IGF-1 in acromegaly have also been previously associated with

PCOS⁴³ with the observation of improvement in IGF-1 and PCOS phenotype following reduction in growth hormone levels. This was followed by the normalisation of menstrual cycle and numbers of polycystic ovary morphology suggesting that IGF-1 could play a role in hyperandrogenism and PCOS.⁴⁴ Low concentrations of progesterone have been proposed to increase LH secretion by disrupting the negative feedback of GnRH.⁴⁵ However, the presence of increased LH and androgen excess in prepubertal girls fails to support low progesterone as a sole source of LH increase in PCOS.⁴⁶ The increase in ovarian androgens in PCOS is not only LH- and insulin-dependent as some *in vitro* studies have shown that it could be due to an abnormality in primary theca cell steroidogenesis.^{47,48} Subsequent clinical studies in women with PCOS have shown that ovarian androgen production is hyperresponsive to both administration of GnRH agonists and Human Chorionic Gonadotropin (hCG) challenges.^{48–51} The role of upregulation of backdoor androgen synthesis pathway contributing to androgen excess in women with PCOS has been discussed by Marti *et al.*⁵² In their study, they have shown that the ovaries express all the enzymes required for DHT synthesis through the backdoor pathway. Comparing ovarian tissues of women with PCOS to women without PCOS, immunohistochemistry revealed higher reactivity for 3 β -hydroxysteroid dehydrogenase type 2 (HSD3B2), retinol dehydrogenase (RoDH), 5 α -reductase type 1 (SRD5A1) and aldo-keto reductase family 1 member C2 (AKR1C2) genes indicating a possible higher activities of their respective enzymes involved in the backdoor pathway. SRD5A1, the gatekeeper of the backdoor pathways converting A4 into androstenedione, showed the highest difference in PCOS compared with non-PCOS highlighting the relevance of this pathway in women with PCOS. Derangement in the Kiss-1 system resulting in increased LH release has also been postulated; however, evidence supporting this theory is lacking to date.³⁶ Recently, the role of Gamma Aminobutyric Acid (GABA) activity in the arcuate nucleus stimulating LH secretion resulting in PCOS-like reproductive dysfunction has been explored in mice by Silva *et al.*⁵³

Adrenal glands and androgen excess

Nearly 50% of women with PCOS have a predominantly adrenal hyperandrogenism phenotype as shown by elevated concentrations of

Dehydroepiandrosterone Sulphate (DHEAS) and 11 β -hydroxyandrostenedione, two androgens that are exclusively produced by the adrenal glands.^{54–56} This is particularly prominent in young women with PCOS.¹³ This may be due to upregulation of CYP17A1 activity through tonic hyperstimulation by Adrenocorticotrophic hormone (ACTH) of the adrenal gland. Lack of a significant increase in 17-hydroxyprogesterone (17OHP) after exogenous stimulation with ACTH supports this theory.^{54,57} One of the other proposed mechanisms of adrenal hyperandrogenism is a relative deficiency of HSD3B2. In women with PCOS and high DHEAS, a normal DHEA response and slightly elevated 17OHP response to ACTH suggested that elevated DHEAS was due to tonic hyperstimulation of the adrenal gland rather than deficiency of the enzymes.⁵⁴ Downregulation of 11 β -hydroxysteroid dehydrogenase type 1 (HSD11B1) activity will result in a decreased rate of cortisol activation and hence could lead to adrenal androgen stimulation *via* hypothalamic–pituitary–adrenal axis feedback; however, no convincing evidence for decreased HSD11B1 activity was found in two independent studies in women with PCOS.^{58,59} The relevance of the adrenal backdoor pathway in women with PCOS was also discussed by Saito *et al.* showing correlations between the levels of DHEAS to androstenedione and androsterone indicating a potential adrenal gland contribution in these two androgens involved in this pathway.⁶⁰

Importantly, the C11-oxy C₁₉ androgens are primarily of adrenal origin as this pathway starts with the conversion of A4 to 11-hydroxyandrostenedione (11OHA4), which can only be catalysed by 11 β -hydroxylase activity of the exclusively adrenally expressed enzyme CYP11B1.^{55,61} As C11-oxy C₁₉ androgens are particularly prominent in the serum women with PCOS¹⁰ as well as in girls with premature adrenarche,⁶² it is safe to assume that the adrenal glands provide a major contribution to circulating androgen excess in PCOS (Figure 2).

Peripheral tissues and androgen excess

After the secretion by the ovaries and/or adrenals into the circulation, androgenic precursors are further activated in peripheral target tissues into more potent androgens that bind to the cytosolic androgen receptor (AR), which subsequently translocates to the nucleus where it functions as a

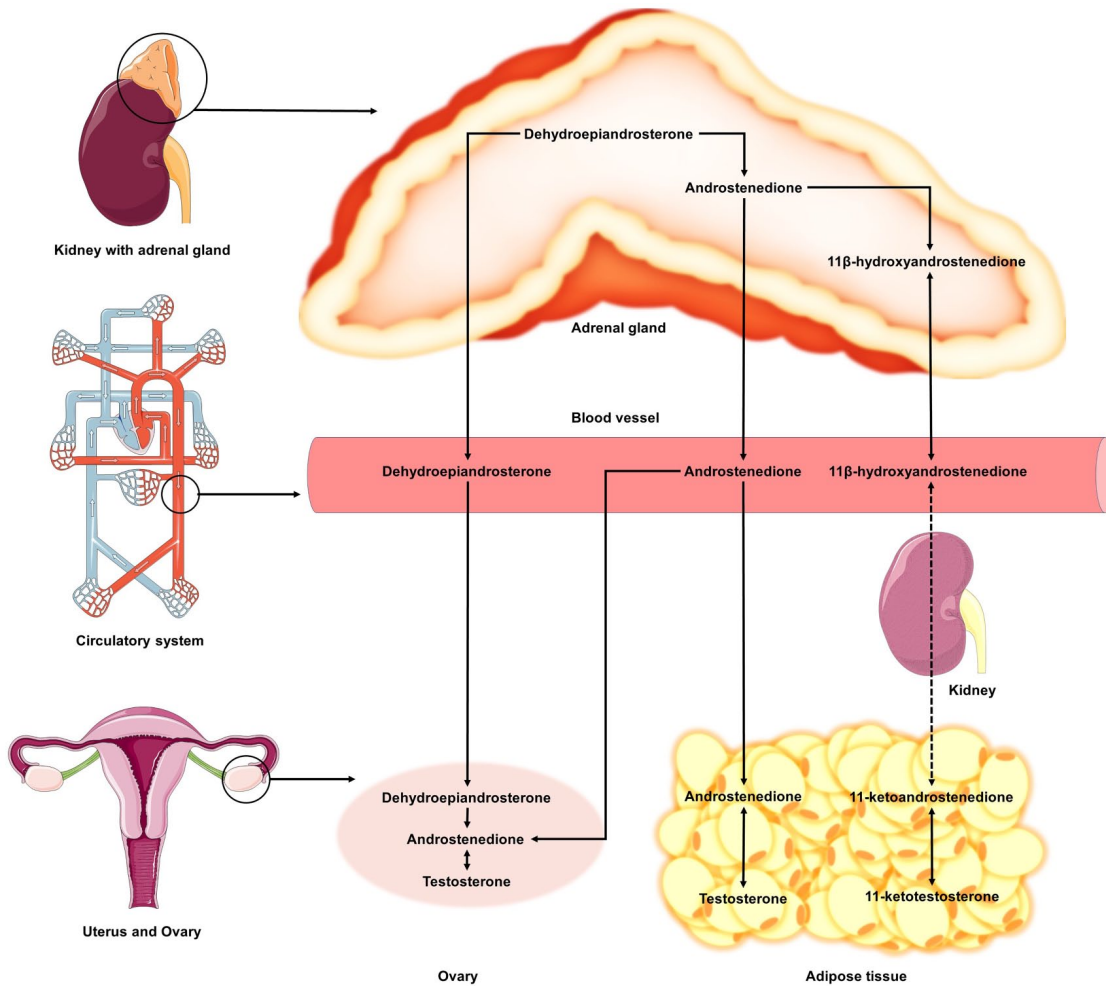


Figure 2. Adrenal, ovarian and peripheral androgen metabolism in PCOS. Androgenic precursors are secreted predominantly by the adrenal glands, and activated to potent androgens in the ovaries and peripheral tissues. Expression of key androgen-activating enzymes in peripheral tissues such as adipose tissue highlight an important role for extra-adrenal and -ovarian androgen generation.

transcription factor after binding to DNA, resulting in increased transcription of genes encoding for factors important for androgenic biologic activity. The concentration of androgens in peripheral target tissues of sex steroid action are determined by (1) the total concentration of the androgenic steroids in the circulation (2) whether they are bound to sex hormones binding globulin (SHBG) or albumin (3) important regulators of bioavailability, and the availability of mechanisms for cellular influx and efflux. Circulating androgens must cross the plasma membrane of the target cells to be metabolized by intracellular enzymes and/or to activate AR. While unconjugated steroids can freely diffuse across the plasma membrane, conjugated steroids are hydrophilic

and hence need to be actively transported inside the cell. Once in the cell, these steroids will need to be deconjugated before they can be metabolized and/or interact with the androgen receptors.^{25,63}

Combining human-based *in vivo* and *ex vivo* study approaches, our group has shown that the androgen-activating enzyme aldo-ketoreductase type 1 C3 (AKR1C3), which converts androstenedione to testosterone as well as 11OHA4 to 11KT, is a major driver of adipocyte-specific androgen excess and that its activity in adipose tissue is a major source of PCOS-related androgen excess.⁶⁴ This will be discussed in detail in the adipose tissue section in the following.

MicroRNA and androgen excess

MicroRNAs (miRNAs) are present widely in the body playing a role in the regulation of gene expression by inhibiting post-transcriptional or post-translational expression of messenger RNA (mRNA).⁶⁵ Alteration in levels of miRNA have been implicated in conditions such as cervical cancer, ovarian cancer, endometriosis and CVDs.^{66–68} Studies have shown differential expression of miRNA in women with PCOS compared with controls, suggesting a potential role of miRNA in PCOS.^{69,70} Chen *et al.* have extensively discussed the role of miRNA in PCOS.⁷¹ Muri *et al.* have previously shown that levels of miR-21, miR-103 and miR-155 were positively associated with levels of free testosterone in women with PCOS⁷² and several miRNAs have been shown to be negatively correlated with levels of testosterone and A4 in these women.^{73,74} Moreover, comparing miR-130b-3p in normal and theca cells of women with PCOS have shown a decreased expression in PCOS which was correlated with increased expression of CYP17A1 and DHEA synthesis, which may contribute to the androgen excess in PCOS.⁷⁵

In isolated porcine ovaries, overexpression of miR-378 was shown to inhibit the expression of aromatase enzymes in the ovaries, which contributed to hyperandrogenaemia.⁷⁶ Furthermore, miRNA181a also downregulates oestrogen synthesis *via* CYP19A1 expression in mouse granulosa cells.⁷⁷ However, there is no evidence of miRNA overexpression in women with PCOS, hence its role in increasing levels of androgens in this population. miR-193a-5p and miR-199a-3p have been positively associated with oestrogen and SHBG and negatively associated with free testosterone in women with PCOS with predictions of target genes, indicating the role of these miRNA in regulating some enzymes in the steroidogenesis pathways.⁷⁸ These evidences taken together suggested that more detailed studies will be needed to investigate the role of miRNA in contributing to androgen excess in women with PCOS. The role of miRNA in propagating metabolic diseases will be discussed in the relevant section in the following.

Androgens in metabolic target tissues

Critical metabolic target organs such as adipose tissue, muscle and pancreatic beta cells are also targets of androgen action (Figure 3). Here, we summarise the potential mechanisms and

metabolic dysregulation due to androgen action in each of these organs.

Adipose tissue, obesity, fat mass distribution and androgen excess

Adipose tissue forms the largest store of conserved energy, in the form of triglycerides, in the human body. The process of adipocyte differentiation from stem cells is complex and involves a variety of signals for gene regulation, histone modification, and protein modification by ubiquitin.⁷⁹ There are two distinct types of adipose tissue, brown adipose tissue and white adipose tissue. Several studies have established the role of adipose tissue beyond an energy storehouse to regulating several critical physiological processes for example, through adipokine signals such as leptin and adiponectin. The various roles of adipokines is discussed in detail by Ouchi *et al.* and Dimitriadis *et al.*^{80,81}

Women with PCOS have been shown to have more central obesity phenotypes compared with weight and body mass index (BMI)-matched controls,^{82–85} which is associated with various metabolic conditions in PCOS.^{86–88} However, these data have been inconsistent.^{89,90} Some studies have also shown that although women with PCOS have android fat mass distribution, these might not be accompanied by increased visceral fat.^{91,92} On the other hand, gluteo-femoral fat or gynecoid adiposity, low in PCOS, has been shown to be independently associated with protective lipid and glucose profile along with decreased cardiovascular and metabolic risk.⁹³ Visceral adiposity index (VAI) has similar utility to the gold-standard computed tomography scan in the evaluation of visceral adiposity.⁹⁴ VAI is a sex-specific empirical mathematical model based on BMI, waist circumference, triglycerides and high-density cholesterol in healthy normal and overweight populations.⁹⁵ Lim *et al.* reported a similar VAI in women with PCOS and healthy controls in few studies. However, subgroup analysis comparing three groups (PCOS and obesity, obesity alone, and PCOS alone) revealed that women with PCOS and obesity have higher VAI compared with the other two groups.⁹⁶ In a study on women with PCOS by Amato *et al.*, VAI was only associated with compensatory hyperinsulinemia and there was no independent association between VAI and insulin sensitivity index (by HOMA-IR and ISI Matsuda), androgen profiles and PCOS.⁹⁷ Visceral

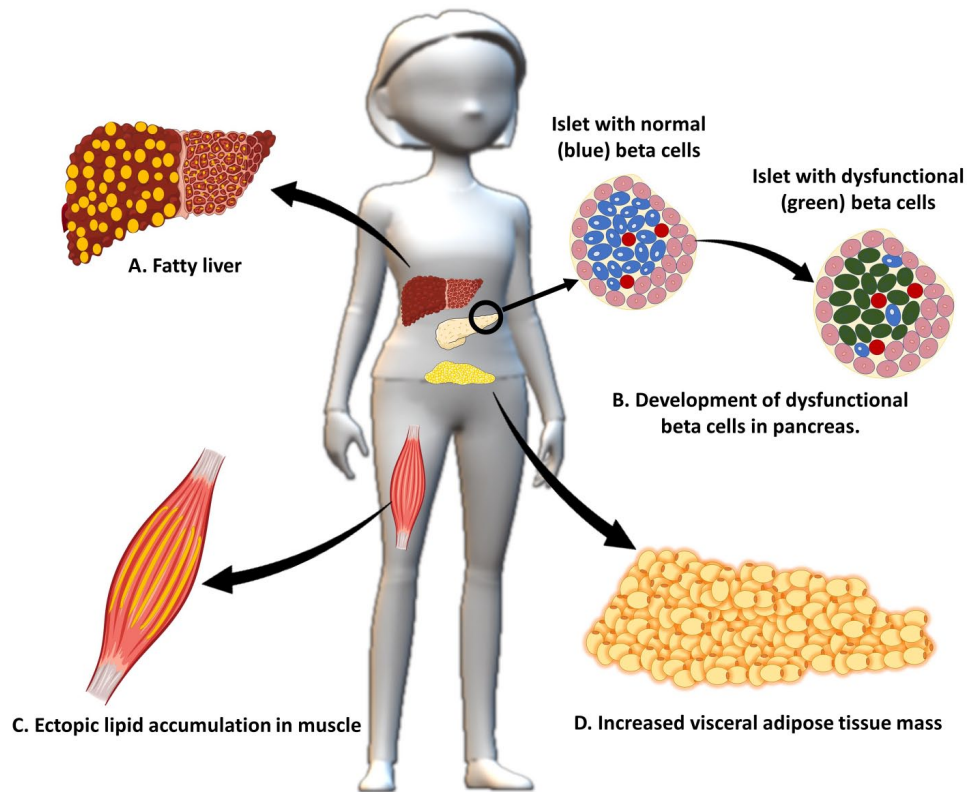


Figure 3. Impact of androgen excess on metabolic target tissues- (A) Increased fat accumulation in the hepatocytes in response to androgen excess, eventually resulting in NAFLD, (B) Androgen excess is proportionally related to beta-cell dysfunction, (C) Ectopic lipid accumulation in skeletal muscle with androgen excess influences glucose-regulating pathways resulting in insulin resistance, (D) under the influence of androgens, women with PCOS have been shown to have more central obesity phenotypes.

adiposity has also been previously, but not consistently, associated with elevated FAI⁸² and total testosterone.⁸⁵ However, the number of studies available is too low for a systematic review to examine whether there is an association between PCOS, central obesity and androgens.

Adipose tissue is a key target of androgen action. Androgens are known to inhibit adipocyte differentiation in both animal and human cell lines.^{98–101} The effects of androgen on adipose tissue can be noted from birth. Roland *et al.* noted higher fasting glucose and impaired glucose tolerance in prenatally androgenised female mice which closely mimics PCOS model.³⁷ Prior to this, Nilsson *et al.* showed that exposure to testosterone in the neonatal period resulted in insulin resistance, change in adipose tissue distribution and lean mass without significant changes in circulating androgen concentrations.¹⁰² Further, their groups showed this early exposure

to androgen resulted in long-lasting effects on insulin sensitivity, adipose tissue and lipid profile.¹⁰³ We have previously published a detailed review of the abnormalities in adipose tissue function, fat distribution and lipid metabolism in the context of androgen excess.¹⁰⁴

We have demonstrated that expression of the androgen-activating enzyme, AKR1C3, which converts the weak androgen precursor androstenedione into potent testosterone, is increased in subcutaneous (SC) adipose tissue in women with PCOS. Locally generated androgens enhance *de novo* lipogenesis within SC female adipocytes, potentially predisposing to fatty acid overspill into the systemic circulation, with consequent lipotoxicity driving an adverse metabolic phenotype.⁶⁴ We demonstrated that associated insulin resistance and hyperinsulinemia then drive AKR1C3 expression and activity within the adipocyte, raising the possibility of a vicious circle of

intra-adipose androgen activation, lipid accumulation and hyperinsulinaemia.

AKR1C3 also converts the C11-oxy C₁₉ androgen precursor 11-ketoandrostenedione (11KA4) into 11-ketotestosterone (11KT), which binds and activates the AR with similar potency to testosterone.¹⁰⁵ Recent data from Storbeck's group also suggest that 11KA4 is the preferred substrate for AKR1C3, which more efficiently converts it to active 11KT than its counterpart reaction in the classic pathway with K_M values of 0.11 μ M and 0.06 μ M for 11KA4 and A4 respectively.¹⁰⁶ Our findings of increased AKR1C3 expression in SAT of women with PCOS were corroborated by a recent study describing similar results.¹⁰⁷ Amer *et al.* have shown an upregulated AKR1C3 and CYP17A1 mRNA expression in SAT of women with PCOS as well as a higher levels of T. But, it is important to note that the authors did not assess if the higher levels of testosterone in the adipose tissue was a result of increased AKR1C3 or CYP17A1 expression. This was previously shown by our group showing a reduction of testosterone levels following inhibition of AKR1C3 by 3-4 trifluoromethyl-phenylamino-benzoic acid in healthy female adipose tissue. Studies on women with PCOS will be needed to confirm if the increased levels of testosterone in SAT are in fact related to overexpression of AKR1C3 *per se* or related to both CYP17A1 and AKR1C3 activities. These findings define a novel intra-adipose mechanism that could contribute to androgen excess and metabolic dysfunction in PCOS.

These data strongly implicate adipose tissue as a major peripheral site of activation and metabolism of C11-oxy C₁₉ androgens in PCOS and suggest that locally generated C11-oxy C₁₉ androgens play a significant role in adipose tissue lipotoxicity. To date, however, this hypothesis has not been tested in adipose tissue of human participants using *in vivo* physiology approaches.

The association between adiposity, androgen excess and PCOS has been further complicated by hyperinsulinaemia and insulin resistance, which exacerbate the metabolic, reproductive and steroidogenic abnormalities observed in the disorder.⁸ A systematic review by Lim *et al.*⁹⁶ found women with obesity and PCOS have a higher total and free testosterone and lower SHBG compared with those with normal BMI. Furthermore, women with both PCOS and obesity also have a

more severe clinical evidence of hyperandrogenism (such as hirsutism, menstrual abnormalities and anovulation) compared with women with PCOS and normal BMI. This effect tends to be more pronounced in the abdominal/visceral obesity phenotypes^{96,108-110} and weight loss in women with PCOS has been shown to improve clinical, metabolic and reproductive outcomes.¹¹⁰⁻¹¹²

Insulin resistance and compensatory hyperinsulinemia in obesity inhibits hepatic production of SHBG, with a consequent increased bioavailability of androgens. Further, insulin modulates hypothalamic-gonadal and steroid enzymatic machinery at various levels as described previously.

Pancreas, type 2 diabetes and androgen excess

Studies have shown that there is a clear sex difference in the way the foetal pancreas reacts to androgen excess. Insulin secreted from the pancreatic beta cells is critical for glucose utilisation in the peripheral tissues. Defects in insulin secretion or action predispose to insulin resistance and T2DM. Beta cells expand rapidly during late gestation and any alteration to its expansion during this period results in long-lasting deleterious effects. Navarro *et al.* demonstrated potentiating effects of DHT on male mice beta cell function *via* the glucagon-like peptide-1 (GLP-1) receptor.¹¹³ Further, they showed male mice lacking AR had decreased glucose-stimulated insulin secretion, leading to glucose intolerance. Castration of male rats resulted in approximately 30% reduction in beta cell mass.¹¹⁴ Xu *et al.* studied the transcriptome of AR-deficient islet beta cells of male mice and found altered expression of genes involved in inflammation and insulin secretion.¹¹⁵

Studies in women with PCOS have also shown that androgen excess is proportionally related to beta cell dysfunction. This has been discussed in detailed by Mauvaus-Jarvis in a review of sex steroid effects on pancreatic beta cell function.¹¹⁶ This hypothesis was further tested in female mice by exposing them to chronic androgen excess, which resulted hyperinsulinemia and insulin resistance. However, this was not observed in female β ARKO mice lacking AR expression in pancreatic beta cells.¹¹⁷ Testosterone prevents beta cell apoptosis and increases insulin mRNA levels both *in vitro* and *in vivo* in a series of studies conducted by Morimoto *et al.*^{118,119}

Harada *et al.* demonstrated an *in vivo* effect of androgens on beta cell mass and expansion in a sex-specific manner. They found reduced beta cell mass and proliferation in male rats following administration of flutamide to pregnant dams. Although beta cell mass was restored after feeding the mice a high fat diet, they had persisting glucose intolerance suggesting decreased insulin secretion.¹²⁰

Androgen exposure during intrauterine life results in several phenotypic characteristics of PCOS in nonhuman primates and sheep. When rats were exposed to testosterone during late gestation, their female offspring exhibited significant weight gain, increased adipose tissue and other traits of metabolic dysfunction.¹²¹ Exposing rhesus monkey dams to testosterone pre-conception resulted in hyperinsulinaemia and a reduction in glucose clearance following accelerated weight gain during testosterone treatment. This transient hyperglycaemic and relative hyperinsulinaemic episodes were sufficient to induce differential programming of insulin action and secretion in their female offspring.¹²² Intrauterine exposure to testosterone resulted in increased beta cell numbers in prenatally androgenised female fetuses with no effect on alpha cells.¹²³ The same group reported upregulation of genes involved in beta cell development and function.¹²⁴ Neither studies found any such changes in male offspring, even when they were exposed to androgens before the male programming window. Roland *et al.* reported fasting hyperglycaemia and impaired glucose tolerance independently of age and changes in body composition or peripheral insulin sensitivity in prenatally androgenised female mice. Once again, no such changes were noted in male counterparts.³⁷

Liu *et al.*¹²⁵ demonstrated improved differentiation of pluripotent stem cells into insulin-producing cells when testosterone was added to routine differentiation medium, resulting in an upregulation of NGN3, NEUROD1 and INS genes. Zhang *et al.* investigated the association between free-androgen index, a marker of androgen excess, with glucose tolerance in PCOS. They found a positive correlation suggestive of beta cell dysfunction in women with PCOS.¹²⁶

Glucose-stimulated insulin secretion significantly decreases in islets treated with DHT in mice studies. This is associated with significant reductions in expression of several key genes involved in islet cell

mitochondrial biogenesis and mitochondrial ATP production.¹²⁷ Chronic exposure to androgens resulted in increased oxidative stress in islet cells resulting in secondary beta cell failure in female mice fed on a western diet.^{117,128} The study also reported direct androgen receptor-dependant impairment of beta cell function by inducing mitochondrial dysfunction *in vitro*.¹²⁸ This is important in connection to skeletal muscle insulin resistance and failing insulin secretion would predispose women with PCOS to develop T2DM.

There are limitations that prevent direct translation findings from *in vitro* and animal studies into human physiology. Some of these limitations may be due to variability in animals for study, methods of randomization, choice of comparison therapy, blinding investigators to interventions and analysis, and variable duration of follow up.^{129,130} Nevertheless, these studies provide plausible pathophysiology and mechanisms linking pancreas and androgen excess, which have been explored in a few human studies described in the following.

PCOS is strongly linked with insulin resistance and T2DM in population and other large-scale studies. A meta-analysis of 35 studies of women with PCOS by Moran *et al.* reported increased odds for insulin resistance (OR 2.54, CI 1.44–4.47) and T2DM (OR 4.00, CI 1.97–8.10) compared with BMI-matched studies.¹³¹ Legro *et al.* conducted one of the earliest prospective studies to assess the prevalence of T2DM in PCOS. They found the prevalence of T2DM in women with PCOS to be 7.5%; it was 1.5% in lean women with PCOS, further supporting the synergistic harmful effect of obesity on insulin sensitivity in PCOS. Interestingly, 31.1% of their cohort had impaired glucose tolerance speculating whether standards of diagnosing T2DM may not identify all index cases in PCOS cohort.¹³² Another study by Glintborg *et al.* later found no differences in the prevalence of Impaired Glucose Tolerance (IGT) and T2DM in healthy weight women with PCOS compared with healthy controls.¹³³ Boudreaux *et al.* reported an incidence of 13.4% and 5.8% in women with PCOS and healthy controls respectively, with a relative risk of 2.3-times after prospective follow up over 8 years. Further, obesity increased the risk by fivefold in their cohort.¹³⁴ One of the largest population-based survey of T2DM in PCOS was reported from Australia by Joham *et al.* They surveyed 9145 women who self-reported the diagnosis of PCOS,

T2DM and gestational diabetes mellitus. The prevalence of T2DM was 5.1%. After adjusting to sociodemographic and other known risk factors, the odds for T2DM were significantly increased in PCOS [odds ratio 8.8, 95% confidence interval (CI) 3.9–20.1, $p=0.001$]. BMI was independently associated with T2DM in PCOS with every BMI increment increasing the risk of T2DM by 10%.¹³⁵ A Danish national register study by Rubin *et al.* studying about 18,000 women with PCOS and 54,000 controls has also shown a higher event rate of T2DM in PCOS compared with well-matched controls, and diabetes was diagnosed on average 4 years earlier than in the background population.¹³⁶ The prevalence of glucose intolerance and T2DM in a cohort of 122 women with PCOS studied by Ehrmann and colleagues was 35% and 10% respectively.¹³⁷ They further followed up a subset of the glucose-intolerant women with PCOS and found them to have higher post-prandial glucose compared with their baseline, suggesting a progressively worsening insulin resistance in the absence of any intervention. The prevalence of diabetes in the Dutch population was much less (2.3%) compared with the American cohorts described previously. However, the risk was 2.3-times higher compared with the generally healthy cohorts. And this risk increased from 1.3-times in 25–34 years to 9.4-times in 45–54 years.¹³⁸ The study by Kauffman and colleagues suggests that ethnicity plays an additive effect on insulin resistance in PCOS. Mexican American women had significantly higher insulin resistance compared with white American women, thus challenging a single population-wide screening tool.¹³⁹

First reported in 1980, the association of hyperandrogenism and hyperinsulinemia is firmly established.^{64,140–142} Our group have also shown in a retrospective cohort study in a UK primary care database that women with increased serum testosterone levels have an increased risk of incident T2DM.¹⁴³ This was recently highlighted further by Ruth *et al.* in their study of 455,097 UK Biobank samples, which showed that the risk of T2DM in women was increased by 37% for every increased standard deviation of free testosterone from baseline. A higher fasting insulin was also reported in these women.¹⁴⁴ However it is important to note that these are associations rather than causation and hence more prospective studies will be needed to investigate the impact of androgen excess on the development of T2DM.

Dunaif *et al.* were amongst the first to prove that women with PCOS have significant insulin resistance independent of obesity. However, obesity seemed to have a synergistic deleterious effect on glucose tolerance.¹⁴² Further, they proved in their study that insulin resistance was not due to decreased insulin clearance but of a different pathology.¹⁴⁵ Since then, several researchers have studied the exact mechanisms of insulin resistance and its consequent deleterious effect on metabolism. Hypotheses to explain intrinsic insulin resistance in PCOS include mitochondrial dysfunction and lipid accumulation affecting the insulin-signalling pathway. Hansen *et al.*¹⁴⁶ found that lean women with PCOS have 25% lower insulin sensitivity and 40% lower plasma adiponectin levels compared with age- and BMI-matched controls. The finding of low levels of adiponectin has been shown to predict women with PCOS who are at high risk for developing T2DM.¹⁴⁷ They also reported an increased accumulation of triacylglycerol, diacylglycerol and ceramide in skeletal muscles of PCOS women supporting the lipid accumulation theory.

Muscle

Androgen excess may play a role impacting skeletal muscles in women with PCOS by altering their insulin sensitivity. Muscle is one of the key organs responsible for disposing 70–80% of glucose load. Insulin stimulates a canonical signalling cascade composed of insulin receptor substrate (IRS)-1, phosphoinositide 3-kinase (PI3K), protein kinases PDK1, Akt and Rab. The overall cascade ends with translocation of glucose transporter (GLUT) 4 to the cell membrane which permits the intake of glucose from blood into the cell.¹⁴⁸ Several groups have tried to tease out the role of androgen excess in metabolic dysfunction using hyperglycaemic and euglycaemic-hyperinsulinaemic clamps. Most of these studies concluded that high levels of testosterone resulted in reduction in whole-body glucose uptake in healthy women.^{149–152} Furthermore, they reported that these testosterone-induced insulin resistance were not attributed to hepatic insulin resistance supporting the role of muscles in androgen-mediated insulin resistance.¹⁵³ Insulin resistance is a common finding in PCOS. Stepto *et al.* found that 75% of lean women with PCOS and 95% of overweight women with PCOS are intrinsically insulin resistant.¹⁵⁴ Various scientists have

attempted to explain insulin resistance in PCOS by one of three mechanisms: (1) dysregulation and/or dysfunction at several steps in the insulin-signalling cascade, (2) lipid accumulation and (3) mitochondrial dysfunction.^{155,156}

Most of the theories about a dysfunctional insulin-signalling pathway come from studies on diabetes. The earliest proposed mechanism of insulin resistance in skeletal muscle of women with PCOS was increased phosphorylation of serine residue on IRS-1 limiting the signal cascade.¹⁵⁷ This was also shown in a later study by Corbould *et al.*¹⁵⁸ However, these findings could not be replicated by other groups. Instead, defects distal to IRS-1/IRS-2 involving Akt substrate 160 kDa^{159,160} and other pathways were found. This theory is supported by a study on the impact of exercise on hyperandrogenized mice, showing improvement in insulin sensitivity *via* PI3K-Akt pathway with associated reduction in 5 α R1 expression in skeletal muscle of the exercise group *versus* stationary group.¹⁶¹ A cross-sectional study looking at impact of habitual physical activity on women with PCOS showed an association of having more than 7500 steps per day (active group) with reduction of BMI, waist circumference, lipid accumulation product, androgen levels and fasting and 120-min insulin levels. In their study, HOMA-IR and 2000 daily steps increment were also found to be an independent predictor of free-androgen index.¹⁶² A randomized trial on women with PCOS have also shown improvement in some androgen levels (total testosterone and SHBG) as well as insulin resistance shown by the decrease in HOMA-IR following 12 weeks of high-intensity interval training, further strengthening the link between androgens excess and insulin resistance in skeletal muscle.¹⁶³

Nilsson *et al.* found aberrant gene expression and DNA methylation in skeletal muscles of women with PCOS, mainly DYRK1A, which encodes an inhibitor of glycogen synthase kinase-3, SCP2, which is involved in lipid metabolism, SYNPO2, which encodes synaptopodin protein involved in oxidation in the muscles, KLF10, which is a transcriptional repressor regulating circadian expression of various genes involved in lipid and glucose metabolism, and NAMPT, which encodes visfatin and promotes glucose uptake into skeletal muscle.¹⁶⁴ However, many of these changes had contrasting results in the presence of androgens *in vitro* and hence remain inconclusive.

The other commonly proposed mechanisms explaining insulin resistance in PCOS include lipid accumulation and mitochondrial dysfunction. Lipid accumulation in skeletal muscle influencing glucose-regulating pathways is hypothesised to cause insulin resistance in diabetes.^{165,166} Intramuscular lipid levels (triacylglycerol, sn-1.3 diacylglycerol and ceramide) were higher in women with PCOS compared with healthy controls in a study done by Hanssen *et al.* In this study, they found women with PCOS had 25% higher whole-body insulin resistance compared with healthy controls. They also found lower AMP-activated protein kinase and Thr172 phosphorylation in association with lower plasma adiponectin levels suggesting a role of the latter in insulin resistance.¹⁴⁶ Studies on the theory of mitochondrial dysfunction causing insulin resistance has resulted in contrasting evidence over the last decades.^{167,168} A nontargeted metabolomics analysis of skeletal muscle of mice which was treated with DHEA revealed 32 metabolites and five metabolic pathways that are significantly different compared with controls.¹⁶⁹ Among these, the reduced NAD⁺/NADH ratio affecting ATP generation was a key finding, which may influence downstream activation of the insulin-signalling pathway, supporting the mitochondrial dysfunction theory. Skov *et al.* showed that impaired insulin-stimulated glucose disposal in women with PCOS was associated with downregulation of mitochondrial oxidative phosphorylation genes using global genetic pathway analysis.¹⁷⁰ Interestingly, Hutchinson *et al.* could not explain the difference in insulin resistance between women with and without PCOS with either lipid accumulation or mitochondrial dysfunction.¹⁷¹ The ability of high-intensity interval training to improve insulin sensitivity and androgen excess may also be related to increase in mitochondrial density and capacity during the training as well as increased in fat oxidation reducing lipid accumulation.^{163,172}

Taken together, the essence of these studies suggests there is perhaps a distinct mechanism of insulin resistance in PCOS. Future studies on the specific influence of androgens on insulin resistance in skeletal muscle may answer this question.

Liver

Several cross-sectional studies have shown an increased prevalence of NAFLD in women with PCOS; NAFLD is now on its way to become the most frequent cause of liver transplantation,

driven by the global obesity pandemic, and is a forerunner of CVD.¹⁷³ A study by Petta *et al.* concluded that PCOS is an independent risk factor for hepatic steatosis and possibly progression to fibrosis and cirrhosis, with hyperandrogenism and insulin resistance as main determinants.¹⁷⁴ These associations have been observed in other studies.^{175–177} Our group has carried out a cohort study utilising a large primary care database in the United Kingdom, including 63,000 women with PCOS and 121,000 matched controls, revealing that the risk of NAFLD was significantly increased in women with PCOS, even in women with a normal BMI (hazard ratio = 2.23, 95% CI 1.86–2.66, $p < 0.001$).¹⁷⁸ Androgen excess (high testosterone, low SHBG) was found to be a contributing risk factor for the development of NAFLD in PCOS in this study.¹⁷⁸ These associations have been recently supported by a meta-analysis showing a 2.3-fold increased rate of NAFLD in women with PCOS, especially those who have hyperandrogenism.¹⁷⁹

There are numerous proposed mechanisms linking this hepatic manifestation of metabolic syndrome with PCOS.^{179,180} Androgen excess in PCOS suppressed low-density lipoprotein receptor RNA expression both in the adipocytes and the liver. They speculate the suppressed receptor expression might prolong plasma half-life of Very low density lipoprotein (VLDL) and low density lipoprotein (LDL), potentially leading to lipid accumulation both in the adipocytes and liver.¹⁸¹ Tumour necrosis factor (TNF)- α levels related to hyperandrogenism have also been implicated in the development of NAFLD.¹⁸² DHEA-induced hyperandrogenism in healthy women of reproductive age resulted in increased fasting AR mRNA content and TNF- α levels, which were both potentiated by glucose ingestion.¹⁸³ TNF- α is one of the proinflammatory cytokines involved in many inflammatory disorders, including metabolic syndrome and has also been shown to induce insulin resistance *via* promotion of IRS-1.¹⁸⁴ TNF- α is also involved in inducing enzymes involved in lipid metabolism, proinflammatory cytokines and fibrosis-associated protein in the liver, thus playing a pivotal role in development of NAFLD. A study by Shimomura *et al.* showed that the adipocyte-specific nuclear form of sterol regulatory element-binding protein 1c (nSREBP-1c) transgenic mice developed hepatic changes similar to the ones seen in NAFLD, with increased TNF- α .¹⁸⁵ Follow-up studies by Kakino *et al.*

using the same mouse model showed that TNF- α is responsible for the development of NAFLD.¹⁸² In this study, TNF knockout nSREBP-1c mice showed an improved glucose tolerance and there was a significantly reduced prevalence of hepatic steatosis compared with the original nSREBP-1c model. This finding was also supported by culturing primary hepatocytes in the presence of TNF- α .¹⁸² These observations support that increased TNF- α , possibly mediated by hyperandrogenism, is an important mechanism in the development and progression of NAFLD in women with PCOS.

Our group has demonstrated androgen-mediated suppression of lipolysis and increased de novo lipogenesis *in vivo* and *in vitro*.¹⁸⁶ This would result in net positive fat accumulation beyond the adipocyte storing capacities causing fatty acid overspill, systemic lipotoxicity, insulin resistance and fat accumulation in the liver, thus leading to the development of NAFLD in women with PCOS. In the same study, serum metabolomics showed increased concentration of glycerophospholipids and lysoglycerophospholipids in women with PCOS with androgen excess, but not in controls with normal androgen concentrations, at baseline. Acute androgen exposure yielded a further increase in these metabolites whereas a decrease was observed in the BMI-matched healthy controls.⁶⁴ Both glycerophospholipids and lysoglycerophospholipids were previously observed to be increased in people with NAFLD and have been identified as potential markers of risk and progression of the condition.¹⁸⁷ These data shows that women with PCOS have a distinct metabolic response to androgens which might contribute to the development of NAFLD and other related conditions.

The role of microRNA

Interestingly, miRNA differential expression was previously associated with the increased risk in the development of T2DM.^{188,189} These expressions have been investigated in women with PCOS in the context of insulin resistance (IR).⁷¹ miR-222 has been shown to be positively associated with hyperinsulinaemia in women with PCOS, suggesting its role in IR in PCOS.⁶⁹ Treating an IR adipose tissue cell line with high levels of glucose and insulin increased levels of miR-320 significantly which in turn reversed the IR *via* increasing the expression of GLUT4. Moreover, miR-320

have been found in the follicular fluid of women with PCOS which may present a future therapeutic target for women with PCOS and IR.¹⁹⁰ Jiang *et al.* added to the literature of miRNA and IR in PCOS later showing the upregulation of miR-122, miR-193b and miR-194 in women with IGT and PCOS compared with those with normal glucose tolerance. miR-33b-5p expression in ovarian tissues has also been found to play a role in propagating IR in PCOS.¹⁹¹ This miRNA has been found to be increased in rats with PCOS and IR with levels negatively correlated with the expression GLUT4, high mobility group A2 (HMGA2) and sterol regulatory element-binding protein 1 (SREBF1). The overexpression was also shown in IR adipose tissue *in vivo* with a similar reduced expression of GLUT4, HMGA2 and SREBF1. Furthermore, expression of GLUT4, HMGA2 and SREBF1 was increased following the inhibition of miR-33b-5p, further strengthening the role of this miRNA in promoting IR in PCOS *via* this pathway.

The role of miRNA, obesity and dyslipidaemia have also been discussed by Chen *et al.* in their review.⁷¹ Several studies have also shown differential expression of miRNA with their expressions correlating to markers of adiposity such as BMI and waist-to-hip ratio.^{72,78,192} Arancio *et al.* have also shown association of miRNA levels with LDL cholesterol levels in women with PCOS and hyperandrogenism, adding into the evidence on androgen excess and miRNA in propagating metabolic diseases in women with PCOS.⁷³

In summary, miRNAs have been shown to play a role in the metabolic derangements in women with PCOS with evidences linking them to androgen excess. More studies will be needed to thoroughly investigate these associations to enable for successful therapeutic targets specifically for reducing the metabolic risk in women with PCOS.

Androgen excess and cardiovascular disease

Earlier studies investigating the association between PCOS and CVD reported no increased prevalence. Pierpoint *et al.* reviewed the case notes of 786 women who were diagnosed with PCOS between 1930 and 1979 and were followed up for an average 30 years. They did not find any increased deaths due to CVDs in this cohort compared with national rates.¹⁹³ Interestingly, the same group reported higher prevalence of cardiovascular

risk factors and nonfatal cerebrovascular disease in this cohort.¹⁹⁴ While they concluded that PCOS may have protective effects on CVD, most of these patients did not have a hormonal profile, challenging the diagnosis and the conclusion of the study. Dahlgren *et al.* reported a 7.4-times higher risk for myocardial infarction in women with PCOS compared with those without, using an early risk factor model.¹⁹⁵ However, the number of participants was relatively small. Birdsall *et al.* reported a more extensive coronary artery disease in postmenopausal women with polycystic ovaries on ultrasound.¹⁹⁶ Although the the diagnosis of PCOS in this cohort could not be ascertained from their report, the study reported associations between the polycystic ovaries with hirsutism and high levels of free testosterone. Christian *et al.* found that women with PCOS had higher coronary artery calcification compared with their age-matched controls, independently of obesity.¹⁹⁷ The incidence of coronary artery disease was four-times higher in Czech women with a history of PCOS compared with the general population.¹⁹⁸ Although these women also had higher prevalence of T2DM, the remaining cardiovascular risk factors were comparable with the rest of the population, questioning whether PCOS is an independent risk factor for coronary artery disease.¹⁹⁸ Based on these findings, the Androgen Excess and PCOS Society acknowledged that women with PCOS have a moderate-to-high risk for CVD, depending on the presence of other risk factors.¹⁹⁹

A meta-analysis evaluating the effect of androgen excess in PCOS on metabolic parameters has shown that androgen excess is associated with higher levels of total cholesterol and lower high-density cholesterol levels.²⁰⁰ A study by Luque-Ramírez *et al.* comparing hyperandrogenic with nonhyperandrogenic PCOS showed an increased mean carotid intima-media thickness independently of BMI, with the main determinant being the concentration of serum total testosterone and A4.²⁰¹ Furthermore, testosterone has also been shown to inhibit bradykinin-induced intracellular calcium kinetics resulting in endothelial dysfunction.²⁰² Androgens are also known to impact the renal system through the upregulation of sodium channels in the proximal tubules increasing the rate of fluid reabsorption, hence increasing extracellular volume and blood pressure.^{203,204} Orio *et al.*²⁰⁵ clinically confirmed endothelial dysfunction in brachial arteries and increased cardiac intima-media thickness directly proportional to androgen excess and

independently of obesity in PCOS compared with age- and BMI-matched controls. Kravariti *et al.* and Luque-Ramírez *et al.* independently confirmed both these findings in PCOS women of differing ethnicities.^{201,206} Levels of serum highly sensitive C-reactive protein (hsCRP) has been previously shown to be higher in women with PCOS, which may play a role in increased risk of CVD in this population.^{207,208} Möhlig *et al.* later found that increased levels of hsCRP were due to obesity rather than PCOS alone²⁰⁹, which was confirmed by another study showing increased hsCRP in women with PCOS that diminishes after adjustment for BMI. The study also reported no association between levels of androgens and that of hsCRP.²¹⁰ Hyperhomocysteinaemia and oxidative stress are widely known as risk factors for the development of CVD.^{211,212} Several studies have previously shown elevated levels of homocysteine and oxidative stress in women with PCOS which may contribute to their risk of developing CVD.^{213–216} A study by Yilmaz *et al.*²¹⁷ also showed a positive correlation between levels of oxidative stress and free testosterone levels in women with PCOS, suggesting the potential role of androgen excess in increasing risk of CVD *via* this mechanism. Although there is no evidence on the relationship between androgen excess and homocysteine levels, many studies using anti-androgenic oral contraceptives containing 35 µg ethynylestrodiol and 2 mg cyproterone acetate have demonstrated rapid reductions of homocysteine levels in women with PCOS.^{218–220} However, this was not the case for anti-androgen-containing drospirinone.²²¹ Recently, the role of androgens in attenuating endothelin-1-induced vasodilation and endothelin B receptor-mediated nitric oxide production resulting in endothelial dysfunction was described by Usselman *et al.* in women with PCOS.²²² These and other studies support that androgen excess is implicated either directly or indirectly in almost all proposed mechanisms to explain the increased risk for CVD in PCOS.

Conclusion

Androgen excess plays a pivotal role in increasing risk of metabolic dysfunction in women with PCOS. Currently, there are no disease-specific therapeutic options available to modify metabolic risk in women with PCOS, and treatments are limited to the use of insulin-sensitising agents, anti-hypertensives and lipid lowering agents. Consensus guidelines² have consistently acknowledged the need to identify novel biomarkers and therapies for

metabolic risk in women with PCOS, and targeting androgen excess is likely to represent the most promising future therapeutic avenue. With the emerging role of the less-well-characterised C11-oxy C₁₉ androgen subclass, it is important to establish the origins and roles of specific androgens in metabolic pathophysiology in order to identify potential therapeutic targets. There is now a strong rationale for therapies targeting androgen synthesis or action for the amelioration of metabolic risk in women with PCOS.

Author contribution(s)

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Conflict of interest statement

The authors declare that there is no conflict of interest.

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References

1. Skiba MA, Islam RM, Bell RJ, *et al.* Understanding variation in prevalence estimates of polycystic ovary syndrome: a systematic review and meta-analysis. *Hum Reprod Update* 2018; 24: 694–709.
2. Teede HJ, Misso ML, Costello MF, *et al.* Recommendations from the international evidence-based guideline for the assessment and management of polycystic ovary syndrome. *Hum Reprod* 2018; 33: 1602–1618.
3. Rotterdam ESHRE/ASRM-Sponsored PCOS Consensus Workshop Group. Revised 2003 consensus on diagnostic criteria and long-term health risks related to polycystic ovary syndrome. *Fertil Steril* 2004; 81: 19–25.
4. Azziz R, Carmina E, Dewailly D, *et al.* The androgen excess and PCOS society criteria for the polycystic ovary syndrome: the complete task force report. *Fertil Steril* 2009; 91: 456–488.
5. Azziz R, Carmina E, Dewailly D, *et al.* Criteria for defining polycystic ovary syndrome as a predominantly hyperandrogenic syndrome: an androgen excess society guideline. *J Clin Endocrinol Metab* 2006; 91: 4237–4245.
6. Azziz R, Carmina E, Dewailly D, *et al.* Positions statement: criteria for defining polycystic ovary syndrome as a predominantly hyperandrogenic syndrome: an androgen excess society guideline. *J Clin Endocrinol Metab* 2006; 91: 4237–4245.
7. O'Reilly MW, Taylor AE, Crabtree NJ, *et al.* Hyperandrogenemia predicts metabolic phenotype in polycystic ovary syndrome: the utility of serum androstenedione. *J Clin Endocrinol Metab* 2014; 99: 1027–1036.
8. Bozdag G, Mumusoglu S, Zengin D, *et al.* The prevalence and phenotypic features of polycystic ovary syndrome: a systematic review and meta-analysis. *Hum Reprod* 2016; 31: 2841–2855.
9. Pretorius E, Arlt W and Storbeck KH. A new dawn for androgens: novel lessons from 11-oxygenated C19 steroids. *Mol Cell Endocrinol* 2017; 441: 76–85.
10. O'Reilly MW, Kempegowda P, Jenkinson C, *et al.* 11-Oxygenated C19 steroids are the predominant androgens in polycystic ovary syndrome. *J Clin Endocrinol Metab* 2017; 102: 840–848.
11. Nanba AT, Rege J, Ren J, *et al.* 11-Oxygenated C19 steroids do not decline with age in women. *J Clin Endocrinol Metab* 2019; 104: 2615–2622.
12. Swart AC, du Toit T, Gourgari E, *et al.* Steroid hormone analysis of adolescents and young women with polycystic ovarian syndrome and adrenocortical dysfunction using UPC2-MS/MS. *Pediatr Res* 2020: 1–9.
13. Elhassan YS, Idkowiak J, Smith K, *et al.* Causes, patterns, and severity of androgen excess in 1205 consecutively recruited women. *J Clin Endocrinol Metab* 2018; 103: 1214–1223.
14. Idkowiak J, Elhassan YS, Mannion P, *et al.* Causes, patterns and severity of androgen excess in 487 consecutively recruited pre- and post-pubertal children. *Eur J Endocrinol* 2019; 180: 213–221.
15. Schiffer L, Kempegowda P, Arlt W, *et al.* Mechanisms in endocrinology: the sexually dimorphic role of androgens in human metabolic disease. *Eur J Endocrinol* 2017; 177: R125–R143.
16. Risal S, Pei Y, Lu H, *et al.* Prenatal androgen exposure and transgenerational susceptibility to polycystic ovary syndrome. *Nat Med* 2019; 25: 1894–1904.
17. Yan X, Dai X, Wang J, *et al.* Prenatal androgen excess programs metabolic derangements in pubertal female rats. *J Endocrinol* 2013; 217: 119–129.
18. Wilde MAD, Eising JB, Gunning MN, *et al.* Cardiovascular and metabolic health of 74 children from women previously diagnosed with polycystic ovary syndrome in comparison with a population-based reference cohort. *Reprod Sci* 2018; 25: 1492–1500.
19. Gunning MN, Sir Petermann T, Crisosto N, *et al.* Cardiometabolic health in offspring of women with PCOS compared to healthy controls: a systematic review and individual participant data meta-analysis. *Hum Reprod Update* 2020; 26: 103–117.
20. Arlt W, Callies F, Van Vlijmen JC, *et al.* Dehydroepiandrosterone replacement in women with adrenal insufficiency. *N Engl J Med* 1999; 341: 1013–1020.
21. Raisz LG, Wiita B, Artis A, *et al.* Comparison of the effects of estrogen alone and estrogen plus androgen on biochemical markers of bone formation and resorption in postmenopausal women. *J Clin Endocrinol Metab* 1996; 81: 37–43.
22. Shifren JL, Braunstein GD, Simon JA, *et al.* Transdermal testosterone treatment in women with impaired sexual function after oophorectomy. *N Engl J Med* 2000; 343: 682–688.
23. Miller KK, Sesmilo G, Schiller A, *et al.* Androgen deficiency in women with hypopituitarism. *J Clin Endocrinol Metab* 2001; 86: 561–567.

24. Snyder PJ. Editorial: the role of androgens in women. *J Clin Endocrinol Metab* 2001; 86: 1006–1007.
25. Schiffer L, Arlt W and Storbeck K-H. Intracrine androgen biosynthesis, metabolism and action revisited. *Mol Cell Endocrinol* 2018; 465: 4–26.
26. Pretorius E, Africander DJ, Vlok M, *et al.* 11-Ketotestosterone and 11-ketodihydrotestosterone in castration resistant prostate cancer: potent androgens which can no longer be ignored. *PLoS One* 2016; 11: e0159867.
27. Stewart PM, Edwards CRW, Shackleton CHL, *et al.* 5 α -Reductase activity in polycystic ovary syndrome. *Lancet* 1990; 335: 431–433.
28. Chin D, Shackleton C, Prasad VK, *et al.* Increased 5 α -reductase and normal 11 β -hydroxysteroid dehydrogenase metabolism of C19 and C21 steroids in a young population with polycystic ovarian syndrome. *J Pediatr Endocrinol Metab* 2000; 13: 253–259.
29. Fassnacht M, Schlenz N, Schneider SB, *et al.* Beyond adrenal and ovarian androgen generation: increased peripheral 5 α -reductase activity in women with polycystic ovary syndrome. *J Clin Endocrinol Metab* 2003; 88: 2760–2766.
30. Vassiliadi DA, Barber TM, Hughes BA, *et al.* Increased 5 α -reductase activity and adrenocortical drive in women with polycystic ovary syndrome. *J Clin Endocrinol Metab* 2009; 94: 3558–3566.
31. Torchen LC, Idkowiak J, Fogel NR, *et al.* Evidence for increased 5 α -reductase activity during early childhood in daughters of women with polycystic ovary syndrome. *J Clin Endocrinol Metab* 2016; 101: 2069–2075.
32. Auchus RJ. The backdoor pathway to dihydrotestosterone. *Trends Endocrinol Metab* 2004; 15: 432–438.
33. Miller WL and Auchus RJ. The molecular biology, biochemistry, and physiology of human steroidogenesis and its disorders. *Endocr Rev* 2011; 32: 81–151.
34. Kamrath C, Hochberg Z, Hartmann MF, *et al.* Increased activation of the alternative “backdoor” pathway in patients with 21-hydroxylase deficiency: evidence from urinary steroid hormone analysis. *J Clin Endocrinol Metab* 2012; 97: E367–E375.
35. Barber TM, McCarthy MI, Wass JAH, *et al.* Obesity and polycystic ovary syndrome. *Clin Endocrinol* 2006; 65: 137–145.
36. Baskind NE and Balen AH. Hypothalamic–pituitary, ovarian and adrenal contributions to polycystic ovary syndrome. *Best Pract Res Clin Obstet Gynaecol* 2016; 37: 80–97.
37. Roland AV, Nunemaker CS, Keller SR, *et al.* Prenatal androgen exposure programs metabolic dysfunction in female mice. *J Endocrinol* 2010; 207: 213–223.
38. Roland AV and Moenter SM. Prenatal androgenization of female mice programs an increase in firing activity of gonadotropin-releasing hormone (GnRH) neurons that is reversed by metformin treatment in adulthood. *Endocrinology* 2011; 152: 618–628.
39. Nestler JE, Jakubowicz DJ, Falcon de, Vargas A, *et al.* Insulin stimulates testosterone biosynthesis by human thecal cells from women with polycystic ovary syndrome by activating its own receptor and using inositolglycan mediators as the signal transduction system. *J Clin Endocrinol Metab* 1998; 83: 2001–2005.
40. Wu S, Divall S, Nwaopara A, *et al.* Obesity-induced infertility and hyperandrogenism are corrected by deletion of the insulin receptor in the ovarian theca cell. *Diabetes* 2014; 63: 1270–1282.
41. Rosenfield RL and Ehrmann DA. The pathogenesis of polycystic ovary syndrome (PCOS): the hypothesis of PCOS as functional ovarian hyperandrogenism revisited. *Endocr Rev* 2016; 37: 467–520.
42. Ehrman DA, Barnes RB and Rosenfield RL. Polycystic ovary syndrome as a form of functional ovarian hyperandrogenism due to dysregulation of androgen secretion. *Endocr Rev* 1995; 16: 322–353.
43. Kaltsas GA, Androulakis II, Tziveriotis K, *et al.* Polycystic ovaries and the polycystic ovary syndrome phenotype in women with active acromegaly. *Clin Endocrinol* 2007; 67: 917–922.
44. Hashimoto S, Yatabe J, Midorikawa S, *et al.* Inhibition of growth hormone excess reduces insulin resistance and ovarian dysfunction in a lean case of polycystic ovary syndrome with a growth-hormone-producing pituitary adenoma. *Horm Res Paediatr* 2003; 59: 149–155.
45. Soules MR, Steiner RA, Clifton DK, *et al.* Progesterone modulation of pulsatile luteinizing hormone secretion in normal women. *J Clin Endocrinol Metab* 1984; 58: 378–383.
46. Apter D, Bützow T, Laughlin GA, *et al.* Accelerated 24-hour luteinizing hormone pulsatile activity in adolescent girls with ovarian hyperandrogenism: relevance to the

- developmental phase of polycystic ovarian syndrome. *J Clin Endocrinol Metab* 1994; 79: 119–125.
47. Gilling-Smith C, Story H, Rogers V, *et al.* Evidence for a primary abnormality of thecal cell steroidogenesis in the polycystic ovary syndrome. *Clin Endocrinol* 1997; 47: 93–99.
 48. Gilling-Smith C, Willis DS, Beard RW, *et al.* Hypersecretion of androstenedione by isolated thecal cells from polycystic ovaries. *J Clin Endocrinol Metab* 1994; 79: 1158–1165.
 49. Chang RJ and Cook-Andersen H. Disordered follicle development. *Mol Cell Endocrinol* 2013; 373: 51–60.
 50. Levrant SG, Barnes RB and Rosenfield RL. A pilot study of the human chorionic gonadotrophin test for ovarian hyperandrogenism. *Hum Reprod* 1997; 12: 1416–1420.
 51. Ibañez L, Hall JE, Potau N, *et al.* Ovarian 17-hydroxyprogesterone hyperresponsiveness to gonadotropin-releasing hormone (GnRH) agonist challenge in women with polycystic ovary syndrome is not mediated by luteinizing hormone hypersecretion: evidence from GnRH agonist and human chorionic gonadotropin stimulation testing. *J Clin Endocrinol Metab* 1996; 81: 4103–4107.
 52. Marti N, Galván JA, Pandey AV, *et al.* Genes and proteins of the alternative steroid backdoor pathway for dihydrotestosterone synthesis are expressed in the human ovary and seem enhanced in the polycystic ovary syndrome. *Mol Cell Endocrinol* 2017; 441: 116–123.
 53. Silva MSB, Desrozières E, Hessler S, *et al.* Activation of arcuate nucleus GABA neurons promotes luteinizing hormone secretion and reproductive dysfunction: implications for polycystic ovary syndrome. *EBioMedicine* 2019; 44: 582–596.
 54. Carmina E, Rosato F and Janni A. Increased DHEAs levels in PCO syndrome: evidence for the existence of two subgroups of patients. *J Endocrinol Invest* 1986; 9: 5–9.
 55. Swart AC, Schloms L, Storbeck KH, *et al.* 11 β -Hydroxyandrostenedione, the product of androstenedione metabolism in the adrenal, is metabolized in LNCaP cells by 5 α -reductase yielding 11 β -hydroxy-5 α -androstenedione. *J Steroid Biochem Mol Biol* 2013; 138: 132–142.
 56. Rege J, Nakamura Y, Satoh F, *et al.* Liquid chromatography-tandem mass spectrometry analysis of human adrenal vein 19-carbon steroids before and after ACTH stimulation. *J Clin Endocrinol Metab* 2013; 98: 1182–1188.
 57. Ayers JW. Differential response to adrenocorticotropin hormone stimulation in polycystic ovarian disease with high and low dehydroepiandrosterone sulfate levels. *Fertil Steril* 1982; 37: 645–649.
 58. Walker BR, Rodin A, Taylor NF, *et al.* Endogenous inhibitors of 11 β -hydroxysteroid dehydrogenase type 1 do not explain abnormal cortisol metabolism in polycystic ovary syndrome. *Clin Endocrinol* 2000; 52: 77–80.
 59. Tsilchorozidou T, Honour JW and Conway GS. Altered cortisol metabolism in polycystic ovary syndrome: insulin enhances 5 α -reduction but not the elevated adrenal steroid production rates. *J Clin Endocrinol Metab* 2003; 88: 5907–5913.
 60. Saito K, Matsuzaki T, Iwasa T, *et al.* Steroidogenic pathways involved in androgen biosynthesis in eumenorrhic women and patients with polycystic ovary syndrome. *J Steroid Biochem Mol Biol* 2016; 158: 31–37.
 61. van Rooyen D, Gent R, Barnard L, *et al.* The in vitro metabolism of 11 β -hydroxyprogesterone and 11-ketoprogesterone to 11-ketodihydrotestosterone in the backdoor pathway. *J Steroid Biochem Mol Biol* 2018; 178: 203–212.
 62. Rege J, Turcu AF, Kasa-Vubu JZ, *et al.* 11-Ketotestosterone is the dominant circulating bioactive androgen during normal and premature adrenarche. *J Clin Endocrinol Metab* 2018; 103: 4589–4598.
 63. Giorgi EP and Stein WD. The transport of steroids into animal cells in culture. *Endocrinology* 1981; 108: 688–697.
 64. O'Reilly MW, Kempegowda P, Walsh M, *et al.* AKR1C3-mediated adipose androgen generation drives lipotoxicity in women with polycystic ovary syndrome. *J Clin Endocrinol Metab* 2017; 102: 3327–3339.
 65. Yates LA, Norbury CJ and Gilbert RJC. The long and short of microRNA. *Cell* 2013; 153: 516–519.
 66. Bu X, Zhang J, Tian F, *et al.* Value of diffusion-weighted magnetic resonance imaging combined with miR-18a level in predicting radiosensitivity of cervical cancer. *Med Sci Monit* 2018; 24: 7271–7278.
 67. Mari-Alexandre J, Carcelén AP, Agababyan C, *et al.* Interplay between microRNAs and oxidative stress in ovarian conditions with a focus on ovarian cancer and endometriosis. *Int J Mol Sci* 2019; 20: 5322.

68. Romakina VV, Zhirov IV, Nasonova SN, *et al.* MicroRNAs as biomarkers of cardiovascular diseases. *Kardiologiya* 2018; 58: 66–71.
69. Long W, Zhao C, Ji C, *et al.* Characterization of serum microRNAs profile of PCOS and identification of novel non-invasive biomarkers. *Cell Physiol Biochem* 2014; 33: 1304–1315.
70. Ding CF, Chen WQ, Zhu YT, *et al.* Circulating microRNAs in patients with polycystic ovary syndrome. *Hum Fertil* 2015; 18: 22–29.
71. Chen Z, Ou H, Wu H, *et al.* Role of microRNA in the pathogenesis of polycystic ovary syndrome. *DNA Cell Biol* 2019; 38: 754–762.
72. Murri M, Insenser M, Fernández-Durán E, *et al.* Effects of polycystic ovary syndrome (PCOS), sex hormones, and obesity on circulating miRNA-21, miRNA-27b, miRNA-103, and miRNA-155 expression. *J Clin Endocrinol Metab* 2013; 98: E1835–E1844.
73. Arancio W, Calogero Amato M, Magliozzo M, *et al.* Serum miRNAs in women affected by hyperandrogenic polycystic ovary syndrome: the potential role of miR-155 as a biomarker for monitoring the estroprogestinic treatment. *Gynecol Endocrinol* 2018; 34: 704–708.
74. Sørensen AE, Wissing ML, Salö S, *et al.* MicroRNAs related to polycystic ovary syndrome (PCOS). *Genes* 2014; 5: 684–708.
75. Mcallister JM, Han AX, Modi BP, *et al.* MiRNA profiling reveals miRNA-130b-3p mediates DENND1A variant 2 expression and androgen biosynthesis. *Endocrinology* 2019; 160: 1964–1981.
76. Xu S, Linher-Melville K, Yang BB, *et al.* MicroRNA378 (miR-378) regulates ovarian estradiol production by targeting aromatase. *Endocrinology* 2011; 152: 3941–3951.
77. Zhang Q, Sun H, Jiang Y, *et al.* MicroRNA-181a suppresses mouse granulosa cell proliferation by targeting activin receptor IIA. *PLoS One* 2013; 8: e59667.
78. Murri M, Insenser M, Fernández-Durán E, *et al.* Non-targeted profiling of circulating microRNAs in women with polycystic ovary syndrome (PCOS): effects of obesity and sex hormones. *Metabolism* 2018; 86: 49–60.
79. Lowe CE, O’Rahilly S and Rochford JJ. Adipogenesis at a glance. *J Cell Sci* 2011; 124: 2681–2686.
80. Ouchi N, Parker JL, Lugus JJ, *et al.* Adipokines in inflammation and metabolic disease. *Nat Rev Immunol* 2011; 11: 85–97.
81. Dimitriadis G, Kyrou I and Randeva H. Polycystic ovary syndrome as a proinflammatory state: the role of adipokines. *Curr Pharm Des* 2016; 22: 5535–5546.
82. Cosar E, Üçok K, Akgün L, *et al.* Body fat composition and distribution in women with polycystic ovary syndrome. *Gynecol Endocrinol* 2008; 24: 428–432.
83. Carmina E, Bucchieri S, Mansueto P, *et al.* Circulating levels of adipose products and differences in fat distribution in the ovulatory and anovulatory phenotypes of polycystic ovary syndrome. *Fertil Steril* 2009; 91: 1332–1335.
84. Strowitzki T, Halser B and Demant T. Body fat distribution, insulin sensitivity, ovarian dysfunction and serum lipoproteins in patients with polycystic ovary syndrome. *Gynecol Endocrinol* 2002; 16: 45–51.
85. Svendsen PF, Nilas L, Norgaard K, *et al.* Obesity, body composition and metabolic disturbances in polycystic ovary syndrome. *Hum Reprod* 2008; 23: 2113–2121.
86. Zheng S-H and Li X-L. Visceral adiposity index as a predictor of clinical severity and therapeutic outcome of PCOS. *Gynecol Endocrinol* 2016; 32: 177–183.
87. Techatraisak K, Wongmeerit K, Dangrat C, *et al.* Measures of body adiposity and visceral adiposity index as predictors of metabolic syndrome among Thai women with PCOS. *Gynecol Endocrinol* 2016; 32: 276–280.
88. Durmus U, Duran C and Ecirli S. Visceral adiposity index levels in overweight and/or obese, and non-obese patients with polycystic ovary syndrome and its relationship with metabolic and inflammatory parameters. *J Endocrinol Invest* 2017; 40: 487–497.
89. Barber TM, Golding SJ, Alvey C, *et al.* Global adiposity rather than abnormal regional fat distribution characterizes women with polycystic ovary syndrome. *J Clin Endocrinol Metab* 2008; 93: 999–1004.
90. Faloia E, Canibus P, Gatti C, *et al.* Body composition, fat distribution and metabolic characteristics in lean and obese women with polycystic ovary syndrome. *J Endocrinol Invest* 2004; 27: 424–429.
91. Boumosleh JM, Grundy SM, Phan J, *et al.* Metabolic concomitants of obese and nonobese

- women with features of polycystic ovarian syndrome. *J Endocr Soc* 2017; 1: 1417–1427.
92. Ribeiro VB, Kogure GS, Lopes IP, *et al.* Association of measures of central fat accumulation indices with body fat distribution and metabolic, hormonal, and inflammatory parameters in women with polycystic ovary syndrome. *Arch Endocrinol Metab* 2019; 63: 417–426.
 93. Manolopoulos KN, Karpe F and Frayn KN. Gluteofemoral body fat as a determinant of metabolic health. *Int J Obes* 2010; 34: 949–959.
 94. Brończyk-Puzoń A, Jagielski P, Kulik-Kupka K, *et al.* Usefulness of a new anthropometric indicator-VAI (visceral adiposity index) in the evaluation of metabolic and hormonal disorders in women with polycystic ovary syndrome. *Adv Clin Exp Med* 2017; 26: 825–828.
 95. Amato MC, Giordano C, Galia M, *et al.* Visceral adiposity index: a reliable indicator of visceral fat function associated with cardiometabolic risk. *Diabetes Care* 2010; 33: 920–922.
 96. Lim SS, Norman RJ, Davies MJ, *et al.* The effect of obesity on polycystic ovary syndrome: a systematic review and meta-analysis. *Obes Rev* 2013; 14: 95–109.
 97. Amato MC, Verghi M, Galluzzo A, *et al.* The oligomenorrhic phenotypes of polycystic ovary syndrome are characterized by a high visceral adiposity index: a likely condition of cardiometabolic risk. *Hum Reprod* 2011; 26: 1486–1494.
 98. Dieudonne MN, Pecquery R, Leneuve MC, *et al.* Opposite effects of androgens and estrogens on adipogenesis in rat preadipocytes: evidence for sex and site-related specificities and possible involvement of insulin-like growth factor 1 receptor and peroxisome proliferator-activated receptor 2. *Endocrinology* 2000; 141: 649–656.
 99. Gupta V, Bhasin S, Guo W, *et al.* Effects of dihydrotestosterone on differentiation and proliferation of human mesenchymal stem cells and preadipocytes. *Mol Cell Endocrinol* 2008; 296: 32–40.
 100. Chazenbalk G, Singh P, Irge D, *et al.* Androgens inhibit adipogenesis during human adipose stem cell commitment to preadipocyte formation. *Steroids* 2013; 78: 920–926.
 101. Blouin K, Nadeau M, Perreault M, *et al.* Effects of androgens on adipocyte differentiation and adipose tissue explant metabolism in men and women. *Clin Endocrinol* 2010; 72: 176–188.
 102. Nilsson C, Niklasson M, Eriksson E, *et al.* Imprinting of female offspring with testosterone results in insulin resistance and changes in body fat distribution at adult age in rats. *J Clin Invest* 1998; 101: 74–78.
 103. Alexanderson C, Eriksson E, Stener-Victorin E, *et al.* Postnatal testosterone exposure results in insulin resistance, enlarged mesenteric adipocytes, and an atherogenic lipid profile in adult female rats: comparisons with estradiol and dihydrotestosterone. *Endocrinology* 2007; 148: 5369–5376.
 104. O'Reilly MW, House PJ and Tomlinson JW. Understanding androgen action in adipose tissue. *J Steroid Biochem Mol Biol* 2014; 143: 277–284.
 105. Storbeck K-H, Bloem LM, Africander D, *et al.* 11 β -Hydroxydihydrotestosterone and 11-ketodihydrotestosterone, novel C19 steroids with androgenic activity: a putative role in castration resistant prostate cancer? *Mol Cell Endocrinol* 2013; 377: 135–146.
 106. Barnard M, Quanson JL, Mostaghel E, *et al.* 11-Oxygenated androgen precursors are the preferred substrates for aldo-keto reductase 1C3 (AKR1C3): implications for castration resistant prostate cancer. *J Steroid Biochem Mol Biol* 2018; 183: 192–201.
 107. Amer SA, Alzanati NG, Warren A, *et al.* Excess androgen production in subcutaneous adipose tissue of women with polycystic ovarian syndrome is not related to insulin or LH. *J Endocrinol* 2019; 241: 99–109.
 108. Plymate SR, Matej LA, Jones RE, *et al.* Inhibition of sex hormone-binding globulin production in the human hepatoma (Hep G2) cell line by insulin and prolactin. *J Clin Endocrinol Metab* 1988; 67: 460–464.
 109. Pasquali R, Gambineri A and Pagotto U. Review article: the impact of obesity on reproduction in women with polycystic ovary syndrome. *BJOG An Int J Obstet Gynaecol* 2006; 113: 1148–1159.
 110. Gambineri A, Pelusi C, Vicennati V, *et al.* Obesity and the polycystic ovary syndrome. *Int J Obes* 2002; 26: 883–896.
 111. Panidis D, Farmakiotis D, Rousso D, *et al.* Obesity, weight loss, and the polycystic ovary syndrome: effect of treatment with diet and orlistat for 24 weeks on insulin resistance and androgen levels. *Fertil Steril* 2008; 89: 899–906.
 112. Escobar-Morreale HF, Botella-Carretero JJ, Álvarez-Blasco F, *et al.* The polycystic ovary

- syndrome associated with morbid obesity may resolve after weight loss induced by bariatric surgery. *J Clin Endocrinol Metab* 2005; 90: 6364–6369.
113. Navarro G, Xu W, Jacobson DA, *et al.* Extranuclear actions of the androgen receptor enhance glucose-stimulated insulin secretion in the male. *Cell Metab* 2016; 23: 837–851.
 114. Harada N, Yoda Y, Yotsumoto Y, *et al.* Androgen signaling expands β -cell mass in male rats and β -cell androgen receptor is degraded under high-glucose conditions. *Am J Physiol Metab* 2018; 314: E274–E286.
 115. Xu W, Niu T, Xu B, *et al.* Androgen receptor-deficient islet β -cells exhibit alteration in genetic markers of insulin secretion and inflammation. A transcriptome analysis in the male mouse. *J Diabetes Complications* 2017; 31: 787–795.
 116. Mauvais-Jarvis F. Role of sex steroids in β cell function, growth, and survival. *Trends Endocrinol Metab* 2016; 27: 844–855.
 117. Navarro G, Allard C, Morford JJ, *et al.* Androgen excess in pancreatic β cells and neurons predisposes female mice to type 2 diabetes. *JCI Insight* 2018; 3: 6364–6369.
 118. Morimoto S, Fernandez-Mejia C, Romero-Navarro G, *et al.* Testosterone effect on insulin content, messenger ribonucleic acid levels, promoter activity, and secretion in the rat. *Endocrinology* 2001; 142: 1442–1447.
 119. Morimoto S, Mendoza-Rodríguez CA, Hiriart M, *et al.* Protective effect of testosterone on early apoptotic damage induced by streptozotocin in rat pancreas. *J Endocrinol* 2005; 187: 217–224.
 120. Harada N, Yotsumoto Y, Katsuki T, *et al.* Fetal androgen signaling defects affect β -cell mass and function, leading to glucose intolerance in high-fat diet-fed male rats. *Am J Physiol Metab* 2019; 317: E731–E741.
 121. Demissie M, Lazic M, Foecking EM, *et al.* Transient prenatal androgen exposure produces metabolic syndrome in adult female rats. *Am J Physiol Metab* 2008; 295: E262–E268.
 122. Abbott DH, Bruns CR, Barnett DK, *et al.* Experimentally induced gestational androgen excess disrupts glucoregulation in rhesus monkey dams and their female offspring. *Am J Physiol Metab* 2010; 299: E741–E751.
 123. Ramaswamy S, Grace C, Mattei AA, *et al.* Developmental programming of polycystic ovary syndrome (PCOS): prenatal androgens establish pancreatic islet α/β cell ratio and subsequent insulin secretion. *Sci Rep* 2016; 6: 27408.
 124. Rae M, Grace C, Hogg K, *et al.* The pancreas is altered by in utero androgen exposure: implications for clinical conditions such as polycystic ovary syndrome (PCOS). *PLoS One* 2013; 8: e56263.
 125. Liu H, Guo D, Ruzi A, *et al.* Testosterone improves the differentiation efficiency of insulin-producing cells from human induced pluripotent stem cells. *PLoS One* 2017; 12: e0179353.
 126. Zhang J, Hu J, Zhang C, *et al.* Analyses of risk factors for polycystic ovary syndrome complicated with non-alcoholic fatty liver disease. *Exp Ther Med* 2018; 15: 4259–4264.
 127. Wang H, Wang X, Zhu Y, *et al.* Increased androgen levels in rats impair glucose-stimulated insulin secretion through disruption of pancreatic beta cell mitochondrial function. *J Steroid Biochem Mol Biol* 2015; 154: 254–266.
 128. Liu S, Navarro G and Mauvais-Jarvis F. Androgen excess produces systemic oxidative stress and predisposes to β -cell failure in female mice. *PLoS One* 2010; 5: e11302.
 129. Pound P, Ebrahim S, Sandercock P, *et al.* Where is the evidence that animal research benefits humans? *Br Med J* 2004; 328: 514–517.
 130. Bracken MB. Why animal studies are often poor predictors of human reactions to exposure. *J R Soc Med* 2009; 102: 120–122.
 131. Moran LJ, Misso ML, Wild RA, *et al.* Impaired glucose tolerance, type 2 diabetes and metabolic syndrome in polycystic ovary syndrome: a systematic review and meta-analysis. *Hum Reprod Update* 2010; 16: 347–363.
 132. Legro RS, Kunselman AR, Dodson WC, *et al.* Prevalence and predictors of risk for type 2 diabetes mellitus and impaired glucose tolerance in polycystic ovary syndrome: a prospective, controlled study in 254 affected women. *J Clin Endocrinol Metab* 1999; 84: 165–169.
 133. Glintborg D, Henriksen JE, Andersen M, *et al.* Prevalence of endocrine diseases and abnormal glucose tolerance tests in 340 Caucasian premenopausal women with hirsutism as the referral diagnosis. *Fertil Steril* 2004; 82: 1570–1579.
 134. Boudreaux MY, Talbott EO, Kip KE, *et al.* Risk of T2DM and impaired fasting glucose among PCOS subjects: results of an 8-year follow-up. *Curr Diab Rep* 2006; 6: 77–83.
 135. Joham AE, Ranasinha S, Zoungas S, *et al.* Gestational diabetes and type 2 diabetes in reproductive-aged women with polycystic ovary

- syndrome. *J Clin Endocrinol Metab* 2014; 99: E447–E452.
136. Rubin KH, Glintborg D, Nybo M, *et al.* Development and risk factors of type 2 diabetes in a nationwide population of women with polycystic ovary syndrome. *J Clin Endocrinol Metab* 2017; 102: 3848–3857.
 137. Ehrmann DA, Barnes RB, Rosenfield RL, *et al.* Prevalence of impaired glucose tolerance and diabetes in women with polycystic ovary syndrome. *Diabetes Care* 1999; 22: 141–146.
 138. Elting MW, Korsen TJM, Bezemer PD, *et al.* Prevalence of diabetes mellitus, hypertension and cardiac complaints in a follow-up study of a Dutch PCOS population. *Hum Reprod* 2001; 16: 556–560.
 139. Kauffman RP, Baker VM, DiMarino P, *et al.* Polycystic ovarian syndrome and insulin resistance in white and Mexican American women: a comparison of two distinct populations. *Am J Obstet Gynecol* 2002; 187: 1362–1369.
 140. Burghen GA, Givens JR and Kitabchi AE. Correlation of hyperandrogenism with hyperinsulinism in polycystic ovarian disease. *J Clin Endocrinol Metab* 1980; 50: 113–116.
 141. Chang RJ, Nakamura RM, Judd HL, *et al.* Insulin resistance in nonobese patients with polycystic ovarian disease. *J Clin Endocrinol Metab* 1983; 57: 356–359.
 142. Dunaif A, Segal KR, Futterweit W, *et al.* Profound peripheral insulin resistance, independent of obesity, in polycystic ovary syndrome. *Diabetes* 1989; 38: 1165–1174.
 143. O'Reilly MW, Glisic M, Kumarendran B, *et al.* Serum testosterone, sex hormone-binding globulin and sex-specific risk of incident type 2 diabetes in a retrospective primary care cohort. *Clin Endocrinol* 2019; 90: 145–154.
 144. Ruth KS, Day FR, Tyrrell J, *et al.* Using human genetics to understand the disease impacts of testosterone in men and women. *Nat Med* 2020; 26: 1–7.
 145. Dunaif A, Segal KR, Shelley DR, *et al.* Evidence for distinctive and intrinsic defects in insulin action in polycystic ovary syndrome. *Diabetes* 1992; 41: 1257–1266.
 146. Hansen SL, Svendsen PF, Jeppesen JF, *et al.* Molecular mechanisms in skeletal muscle underlying insulin resistance in women who are lean with polycystic ovary syndrome. *J Clin Endocrinol Metab* 2019; 104: 1841–1854.
 147. Sepilian V and Nagamani M. Adiponectin levels in women with polycystic ovary syndrome and severe insulin resistance. *J Soc Gynecol Invest* 2005; 12: 129–134.
 148. Satoh T. Molecular mechanisms for the regulation of insulin-stimulated glucose uptake by small guanosine triphosphatases in skeletal muscle and adipocytes. *Int J Mol Sci* 2014; 15: 18677–18692.
 149. Oh JY, Barrett-Connor E, Wedick NM, *et al.* Endogenous sex hormones and the development of type 2 diabetes in older men and women: the Rancho Bernardo study. *Diabetes Care* 2002; 25: 55–60.
 150. Diamond MP, Grainger D, Diamond MC, *et al.* Effects of methyltestosterone on insulin secretion and sensitivity in women. *J Clin Endocrinol Metab* 1998; 83: 4420–4425.
 151. Shamma FN, Rossi G, HajHassan L, *et al.* The effect of Norplant on glucose metabolism under hyperglycemic hyperinsulinemic conditions. *Fertil Steril* 1995; 63: 767–772.
 152. Polderman KH, Gooren LJ, Asscheman H, *et al.* Induction of insulin resistance by androgens and estrogens. *J Clin Endocrinol Metab* 1994; 79: 265–271.
 153. Navarro G, Allard C, Xu W, *et al.* The role of androgens in metabolism, obesity, and diabetes in males and females. *Obesity* 2015; 20: 713–719.
 154. Stepto NK, Cassar S, Joham AE, *et al.* Women with polycystic ovary syndrome have intrinsic insulin resistance on euglycaemic-hyperinsulaemic clamp. *Hum Reprod* 2013; 28: 777–784.
 155. Carnagarin R, Dharmarajan AM and Dass CR. Molecular aspects of glucose homeostasis in skeletal muscle: a focus on the molecular mechanisms of insulin resistance. *Mol Cell Endocrinol* 2015; 417: 52–62.
 156. Stepto NK, Moreno-Asso A, McIlvenna LC, *et al.* Molecular mechanisms of insulin resistance in polycystic ovary syndrome: unraveling the conundrum in skeletal muscle? *J Clin Endocrinol Metab* 2019; 104: 5372–5381.
 157. Dunaif A, Xia J, Book CB, *et al.* Excessive insulin receptor serine phosphorylation in cultured fibroblasts and in skeletal muscle: a potential mechanism for insulin resistance in the polycystic ovary syndrome. *J Clin Invest* 1995; 96: 801–810.
 158. Corbould A, Kim Y-B, Youngren JF, *et al.* Insulin resistance in the skeletal muscle of

- women with PCOS involves intrinsic and acquired defects in insulin signaling. *Am J Physiol Metab* 2005; 288: E1047–E1054.
159. Hojlund K, Glintborg D, Andersen NR, *et al.* Impaired insulin-stimulated phosphorylation of Akt and AS160 in skeletal muscle of women with polycystic ovary syndrome is reversed by pioglitazone treatment. *Diabetes* 2008; 57: 357–366.
 160. Martens JWM, Geller DH, Arlt W, *et al.* Enzymatic activities of P450c17 stably expressed in fibroblasts from patients with the polycystic ovary syndrome. *J Clin Endocrinol Metab* 2000; 85: 4338–4346.
 161. Wu C, Jiang F, Wei K, *et al.* Exercise activates the PI3K-AKT signal pathway by decreasing the expression of 5 α -reductase type 1 in PCOS rats. *Sci Rep* 2018; 8: 1–10.
 162. Mario FM, Graff SK and Spritzer PM. Habitual physical activity is associated with improved anthropometric and androgenic profile in PCOS: a cross-sectional study. *J Endocrinol Invest* 2017; 40: 377–384.
 163. Samadi Z, Bambaiechi E, Valiani M, *et al.* Evaluation of changes in levels of hyperandrogenism, hirsutism and menstrual regulation after a period of aquatic high-intensity interval training in women with polycystic ovary syndrome. *Int J Prev Med* 2019; 10: 187.
 164. Nilsson E, Benrick A, Kokosar M, *et al.* Transcriptional and epigenetic changes influencing skeletal muscle metabolism in women with polycystic ovary syndrome. *J Clin Endocrinol Metab* 2018; 103: 4465–4477.
 165. Samuel VT, Petersen KF and Shulman GI. Lipid-induced insulin resistance: unravelling the mechanism. *Lancet* 2010; 375: 2267–2277.
 166. Borén J, Taskinen M-R, Olofsson S-O, *et al.* Ectopic lipid storage and insulin resistance: a harmful relationship. *J Intern Med* 2013; 274: 25–40.
 167. Yamada T, Ida T, Yamaoka Y, *et al.* Two distinct patterns of glucose intolerance in icteric rats and rabbits. Relationship to impaired liver mitochondria function. *J Lab Clin Med* 1975; 86: 38–45.
 168. Montgomery MK and Turner N. Mitochondrial dysfunction and insulin resistance: an update. *Endocr Connect* 2015; 4: R1–R15.
 169. Shen Q, Bi H, Yu F, *et al.* Nontargeted metabolomic analysis of skeletal muscle in a dehydroepiandrosterone-induced mouse model of polycystic ovary syndrome. *Mol Reprod Dev* 2019; 86: 370–378.
 170. Skov V, Glintborg D, Knudsen S, *et al.* Reduced expression of nuclear-encoded genes involved in mitochondrial oxidative metabolism in skeletal muscle of insulin-resistant women with polycystic ovary syndrome. *Diabetes* 2007; 56: 2349–2355.
 171. Hutchison SK, Teede HJ, Rachoń D, *et al.* Effect of exercise training on insulin sensitivity, mitochondria and computed tomography muscle attenuation in overweight women with and without polycystic ovary syndrome. *Diabetologia* 2012; 55: 1424–1434.
 172. Cassidy S, Thoma C, Houghton D, *et al.* High-intensity interval training: a review of its impact on glucose control and cardiometabolic health. *Diabetologia* 2017; 60: 7–23.
 173. Pais R, Barritt AS, Calmus Y, *et al.* NAFLD and liver transplantation: current burden and expected challenges. *J Hepatol* 2016; 65: 1245–1257.
 174. Petta S, Ciresi A, Bianco J, *et al.* Insulin resistance and hyperandrogenism drive steatosis and fibrosis risk in young females with PCOS. *PLoS One* 2017; 12: e0186136.
 175. Cai J, Wu CH, Zhang Y, *et al.* High-free androgen index is associated with increased risk of non-alcoholic fatty liver disease in women with polycystic ovary syndrome, independent of obesity and insulin resistance. *Int J Obes* 2017; 41: 1341–1347.
 176. Kim JJ, Kim D, Yim JY, *et al.* Polycystic ovary syndrome with hyperandrogenism as a risk factor for non-obese non-alcoholic fatty liver disease. *Aliment Pharmacol Ther* 2017; 45: 1403–1412.
 177. Macut D, Tziomalos K, Božić-Antić I, *et al.* Non-alcoholic fatty liver disease is associated with insulin resistance and lipid accumulation product in women with polycystic ovary syndrome. *Hum Reprod* 2016; 31: 1347–1353.
 178. Kumarendran B, O'Reilly MW, Manolopoulos KN, *et al.* Polycystic ovary syndrome, androgen excess, and the risk of nonalcoholic fatty liver disease in women: a longitudinal study based on a United Kingdom primary care database. *PLoS Med* 2018; 15: e1002542.
 179. Wu J, Yao X-Y, Shi R-X, *et al.* A potential link between polycystic ovary syndrome and non-alcoholic fatty liver disease: an update meta-analysis. *Reprod Health* 2018; 15: 77.
 180. Macut D, Božić-Antić I, Bjekić-Macut J, *et al.* Management of endocrine disease: polycystic

- ovary syndrome and nonalcoholic fatty liver disease. *Eur J Endocrinol* 2017; 177: R145–R158.
181. Baranova A, Tran T, Afendy A, *et al.* Molecular signature of adipose tissue in patients with both non-alcoholic fatty liver disease (NAFLD) and polycystic ovarian syndrome (PCOS). *J Transl Med* 2013; 11: 133.
 182. Kakino S, Ohki T, Nakayama H, *et al.* Pivotal role of TNF- α in the development and progression of nonalcoholic fatty liver disease in a murine model. *Horm Metab Res* 2018; 50: 80–87.
 183. González F, Sia CL, Bearson DM, *et al.* Hyperandrogenism induces a proinflammatory TNF α response to glucose ingestion in a receptor-dependent fashion. *J Clin Endocrinol Metab* 2014; 99: E848–E854.
 184. Hotamisligil GS, Peraldi P, Budavari A, *et al.* IRS-1-mediated inhibition of insulin receptor tyrosine kinase activity in TNF- α - and obesity-induced insulin resistance. *Science* 1996; 271: 665–670.
 185. Shimomura I, Hammer RE, Richardson JA, *et al.* Insulin resistance and diabetes mellitus in transgenic mice expressing nuclear SREBP-1c in adipose tissue: model for congenital generalized lipodystrophy. *Genes Dev* 1998; 12: 3182–3194.
 186. O'Reilly M, Gathercole L, Capper F, *et al.* Effect of insulin on AKR1C3 expression in female adipose tissue: in-vivo and in-vitro study of adipose androgen generation in polycystic ovary syndrome. *Lancet* 2015; 385(Suppl.): S16.
 187. Anjani K, Lhomme M, Sokolovska N, *et al.* Circulating phospholipid profiling identifies portal contribution to NASH signature in obesity. *J Hepatol* 2015; 62: 905–912.
 188. Shi Z, Zhao C, Guo X, *et al.* Differential expression of microRNAs in omental adipose tissue from gestational diabetes mellitus subjects reveals miR-222 as a regulator of ER α expression in estrogen-induced insulin resistance. *Endocrinology* 2014; 155: 1982–1990.
 189. Ortega FJ, Mercader JM, Moreno-Navarrete JM, *et al.* Profiling of circulating microRNAs reveals common microRNAs linked to type 2 diabetes that change with insulin sensitization. *Diabetes Care* 2014; 37: 1375–1383.
 190. Sang Q, Yao Z, Wang H, *et al.* Identification of microRNAs in human follicular fluid: characterization of microRNAs that govern steroidogenesis in vitro and are associated with polycystic ovary syndrome in vivo. *J Clin Endocrinol Metab* 2013; 98: 3068–3079.
 191. Yang Y, Jiang H, Xiao L, *et al.* MicroRNA-33b-5p is overexpressed and inhibits GLUT4 by targeting HMGA2 in polycystic ovarian syndrome: an in vivo and in vitro study. *Oncol Rep* 2018; 39: 3073–3085.
 192. Xiong W, Lin Y, Xu L, *et al.* Circulatory microRNA 23a and microRNA 23b and polycystic ovary syndrome (PCOS): the effects of body mass index and sex hormones in an Eastern Han Chinese population. *J Ovarian Res* 2017; 10: 10.
 193. Pierpoint T, McKeigue PM, Isaacs AJ, *et al.* Mortality of women with polycystic ovary syndrome at long-term follow-up. *J Clin Epidemiol* 1998; 51: 581–586.
 194. Wild S, Pierpoint T, McKeigue P, *et al.* Cardiovascular disease in women with polycystic ovary syndrome at long-term follow-up: a retrospective cohort study. *Clin Endocrinol* 2000; 52: 595–600.
 195. Dahlgren E, Janson PO, Johansson S, *et al.* Polycystic ovary syndrome and risk for myocardial infarction: evaluated from a risk factor model based on a prospective population study of women. *Acta Obstet Gynecol Scand* 1992; 71: 599–604.
 196. Birdsall MA, Farquhar CM and White HD. Association between polycystic ovaries and extent of coronary artery disease in women having cardiac catheterization. *Ann Intern Med* 1997; 126: 32.
 197. Christian RC, Dumesic DA, Behrenbeck T, *et al.* Prevalence and predictors of coronary artery calcification in women with polycystic ovary syndrome. *J Clin Endocrinol Metab* 2003; 88: 2562–2568.
 198. Cibula D, Cífková R, Fanta M, *et al.* Increased risk of non-insulin-dependent diabetes mellitus, arterial hypertension and coronary artery disease in perimenopausal women with a history of the polycystic ovary syndrome. *Hum Reprod* 2000; 15: 785–789.
 199. Wild RA, Carmina E, Diamanti-Kandarakis E, *et al.* Assessment of cardiovascular risk and prevention of cardiovascular disease in women with the polycystic ovary syndrome: a consensus statement by the Androgen Excess and Polycystic Ovary Syndrome (AE-PCOS) Society. *J Clin Endocrinol Metab* 2010; 95: 2038–2049.

200. Yang R, Yang S, Li R, *et al.* Effects of hyperandrogenism on metabolic abnormalities in patients with polycystic ovary syndrome: a meta-analysis. *Reprod Biol Endocrinol* 2016; 14: 67.
201. Luque-Ramírez M, Mendieta-Azcona C, Álvarez-Blasco F, *et al.* Androgen excess is associated with the increased carotid intima-media thickness observed in young women with polycystic ovary syndrome. *Hum Reprod* 2007; 22: 3197–3203.
202. Rubio-Gayosso I, Garcia-Ramirez O, Gutierrez-Serdan R, *et al.* Testosterone inhibits bradykinin-induced intracellular calcium kinetics in rat aortic endothelial cells in culture. *Steroids* 2002; 67: 393–397.
203. Quan A, Chakravarty S, Chen J-K, *et al.* Androgens augment proximal tubule transport. *Am J Physiol Ren Physiol* 2004; 287: F452–F459.
204. Quinkler M, Bujalska IJ, Kaur K, *et al.* Androgen receptor-mediated regulation of the α -subunit of the epithelial sodium channel in human kidney. *Hypertension* 2005; 46: 787–798.
205. Orio F, Palomba S, Cascella T, *et al.* Early impairment of endothelial structure and function in young normal-weight women with polycystic ovary syndrome. *J Clin Endocrinol Metab* 2004; 89: 4588–4593.
206. Kravariti M, Naka KK, Kalantaridou SN, *et al.* Predictors of endothelial dysfunction in young women with polycystic ovary syndrome. *J Clin Endocrinol Metab* 2005; 90: 5088–5095.
207. Kelly CCJ, Lyall H, Petrie JR, *et al.* Low grade chronic inflammation in women with polycystic ovarian syndrome. *J Clin Endocrinol Metab* 2001; 86: 2453–2455.
208. Boulman N, Levy Y, Leiba R, *et al.* Increased C-reactive protein levels in the polycystic. *J Clin Endocrinol Metab* 2004; 89: 2160–2165.
209. Möhlig M, Spranger J, Osterhoff M, *et al.* The polycystic ovary syndrome per se is not associated with increased chronic inflammation. *Eur J Endocrinol* 2004; 150: 525–532.
210. Oh JY, Lee JA, Lee H, *et al.* Serum C-reactive protein levels in normal-weight polycystic ovary syndrome. *Korean J Intern Med* 2009; 24: 350–355.
211. Clarke R, Robinson K, Graham I, *et al.* Hyperhomocysteinemia: an independent risk factor for vascular disease. *N Engl J Med* 1991; 324: 1149–1155.
212. Garibaldi S, Valentini S, Aragno I, *et al.* Plasma protein oxidation and antioxidant defense during aging. *Int J Vitam Nutr Res* 2001; 71: 332–338.
213. Yarali H, Yildirim A, Aybar F, *et al.* Diastolic dysfunction and increased serum homocysteine concentrations may contribute to increased cardiovascular risk in patients with polycystic ovary syndrome. *Fertil Steril* 2001; 76: 511–516.
214. Schachter M, Raziell A, Friedler S, *et al.* Insulin resistance in patients with polycystic ovary syndrome is associated with elevated plasma homocysteine. *Hum Reprod* 2003; 18: 721–727.
215. Loverro G, Lorusso F, Mei L, *et al.* The plasma homocysteine levels are increased in polycystic ovary syndrome. *Gynecol Obstet Invest* 2002; 53: 157–162.
216. Sabuncu T, Vural H, Harma M, *et al.* Oxidative stress in polycystic ovary syndrome and its contribution to the risk of cardiovascular disease. *Clin Biochem* 2001; 34: 407–413.
217. Yilmaz M, Bukan N, Ayvaz G, *et al.* The effects of rosiglitazone and metformin on oxidative stress and homocysteine levels in lean patients with polycystic ovary syndrome. *Hum Reprod* 2005; 20: 3333–3340.
218. Cagnacci A, Tirelli A, Renzi A, *et al.* Effects of two different oral contraceptives on homocysteine metabolism in women with polycystic ovary syndrome. *Contraception* 2006; 73: 348–351.
219. Gul OB, Somunkiran A, Yucel O, *et al.* The effect of ethinyl estradiol-cyproterone acetate treatment on homocysteine levels in women with polycystic ovary syndrome. *Arch Gynecol Obstet* 2008; 277: 25–30.
220. Luque-Ramírez M, Mendieta-Azcona C, del Rey Sánchez JM, *et al.* Effects of an antiandrogenic oral contraceptive pill compared with metformin on blood coagulation tests and endothelial function in women with the polycystic ovary syndrome: influence of obesity and smoking. *Eur J Endocrinol* 2009; 160: 469–480.
221. Mancini F, Cianciosi A, Persico N, *et al.* Drospirenone and cardiovascular risk in lean and obese polycystic ovary syndrome patients: a pilot study. *Am J Obstet Gynecol* 2010; 202: 169.
222. Usselman CW, Yarovinsky TO, Steele FE, *et al.* Androgens drive microvascular endothelial dysfunction in women with polycystic ovary syndrome: role of the endothelin B receptor. *J Physiol* 2019; 597: 2853–2865.