Next-Generation Microfluidics for Biomedical Research and Healthcare Applications

Muhammedin Deliorman¹, Dima Samer Ali^{1,2} and Mohammad A Qasaimeh^{1,2}

¹Division of Engineering, New York University Abu Dhabi (NYUAD), Abu Dhabi, UAE. ²Department of Mechanical and Aerospace Engineering, New York University, New York, NY, USA. Biomedical Engineering and Computational Biology Volume 14: 1-10 © The Author(s) 2023 Article reuse guidelines: sagepub.com/journals-permissions DOI: 10.1177/11795972231214387



ABSTRACT: Microfluidic systems offer versatile biomedical tools and methods to enhance human convenience and health. Advances in these systems enables next-generation microfluidics that integrates automation, manipulation, and smart readout systems, as well as design and three-dimensional (3D) printing for precise production of microchannels and other microstructures rapidly and with great flexibility. These 3D-printed microfluidic platforms not only control the complex fluid behavior for various biomedical applications, but also serve as microconduits for building 3D tissue constructs—an integral component of advanced drug development, toxicity assessment, and accurate disease modeling. Furthermore, the integration of other emerging technologies, such as advanced microscopy and robotics, enables the spatiotemporal manipulation and high-throughput screening of cell physiology within precisely controlled microenvironments. Notably, the portability and high precision automation capabilities in these integrated systems facilitate rapid experimentation and data acquisition to help deepen our understanding of complex biological systems and their behaviors. While certain challenges, including material compatibility, scaling, and standardization still exist, the integration with artificial intelligence, the Internet of Things, smart materials, and miniaturization holds tremendous promise in reshaping traditional microfluidic approaches. This transformative potential, when integrated with advanced technologies, has the potential to revolutionize biomedical research and healthcare applications, ultimately benefiting human health. This review highlights the advances in the field and emphasizes the critical role of the next generation microfluidic systems in advancing biomedical research, point-of-care diagnostics, and healthcare systems.

KEYWORDS: Microfluidics, 3D printing, high-throughput, automation, screening, point-of-care diagnostics, precision medicine, 3D tissue engineering, human-on-a-chip, drug development, disease modeling, personalized medicine

RECEIVED: September 28, 2023. ACCEPTED: October 30, 2023.

TYPE: Review

FUNDING: The author(s) disclosed receipt of the following financial support for the research, authorship, and/or publication of this article: This work was financially supported by NYU Abu Dhabi, UAE.

Introduction

Advancing biomedical research often depends on the use of modern problem-solving technologies that offer versatile tools and methods to enhance human convenience and health. At the forefront of these technologies is microfluidics1-an extraordinary science and enabling technology that precisely controls fluid behavior at micro-, nano-, and pico-scales.

In the past 3 decades, the technological and scientific advances in microfluidics led to the development of lab-on-achip² and organ-on-a-chip³ devices. These devices comprise closed microfluidic channels, chambers, and components for various small-scale laboratory methods and analysis. For example, portable microfluidic devices can quantitatively detect nucleic acids in blood⁴ or glucose levels in plasma and tears.⁵ This holds the potential for affordable point-of-care diagnostics, which is a critical component of precision medicine. Biomimetic microfluidic devices can replicate lung functions⁶ or cardiac tissue models⁷ on a chip. This holds the promise for drug testing, understanding disease mechanisms, and identifying potential therapeutics using physiologically relevant 3D organ-on-a-chip models. Such models are increasingly important in personalized medicine.

Further advances expanded the capabilities of microfluidic systems, resulting in the development of lab-on-a-paper⁸ DECLARATION OF CONFLICTING INTERESTS: The author(s) declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

CORRESPONDING AUTHOR: Mohammad A Qasaimeh, Division of Engineering, New York University Abu Dhabi (NYUAD), P.O. Box 129188, Abu Dhabi, UAE. Email: maq4@ nyu.edu

platforms. These platforms utilize capillary action to passively control fluids within a single sheet of paper for rapid "sampleto-diagnose" approaches. For example, paper microfluidic assays are proven effective in detecting severe acute respiratory syndrome coronavirus type 2 (SARS-CoV-2) infections when integrated with recombinase polymerase amplification⁹ or with bioconjugated gold nanoparticles.¹⁰ These assays showcase their multiplex gene and antibody detection capabilities, respectively. Additionally, they also offer rapid and semiquantitative measurements of liver enzymes¹¹ or glucose detection¹² in a fingerstick blood specimen. This functionality makes them accessible for monitoring drug-induced liver injury and diabetics in resource-limited settings, respectively.

More recently, a significant advancement in microfluidics was the development of lab-on-a-tip13 devices that integrate and execute a multitude of cell-related applications directly into the tip of an open (non-contact) microfluidic probe. These "channel-less" microfluidic systems combine a bottom glass substrate with a top microfluidic delivery system. Moreover, by utilizing the capabilities of three-dimensional (3D) printing, they offer flexibility in design, efficiency, and versatility. For example, a herringbone microfluidic probe (HB-MFP)14 can efficiently capture rare circulating tumor cells (CTCs), including clusters, from prostate cancer blood samples onto bioactivated glass substrates. Through

 $(\mathbf{\hat{n}})$

Creative Commons Non Commercial CC BY-NC: This article is distributed under the terms of the Creative Commons Attribution-NonCommercial 4.0 License (https://creativecommons.org/licenses/by-nc/4.0/) which permits non-commercial use, reproduction and distribution of the work without further permission provided the original work is attributed as specified on the SAGE and Open Access pages (https://us.sagepub.com/en-us/nam/open-access-at-sage).



Pigure 1. Laboura-crip and organ-on-a-crip devices: (a) integrated microfluidic crip for low-cost, quantitative, and portable nucleic acid testing. (b) Microfluidic platform for immunodiagnostics integrated with artificial intelligence (AI) technology. Microfluidic systems for developing advanced models of (c) lung and (d) cardiac tissues. Reproduced from Yeh et al,⁴ with permission from American Association for the Advancement of Science (a), from Bhuiyan et al,⁵ with permission from American Chemical Society (b), from Li et al,⁶ with permission from American Institute of Physics Publishing (c), and from Michas et al,⁷ with permission from American Association for the Advancement of Science (d).

affinity-based multiplexing, the HB-MFP offers insights into cellular phenotypes. In a related field, a non-contact multiphysics probe (NMP)¹⁵ can operate within the physiological tissue environment to precisely manipulate single cells. Through controlled manipulation, electropermeabilization, transfection, and cytoplasm extraction from living single cells, the NMP enhances spatiotemporal single-cell analysis within tissue samples.

As a result, next generation microfluidic systems, equipped with advanced automation and screening capabilities, are demonstrating their potential in a wide range of biomedical research applications. These applications include, but are not limited to, the manipulation and patterning of single cells, multiplex characterization of cell phenotypes, precise cell sorting, diverse bioassays and biosensing, as well as contributions to tissue engineering, drug efficacy testing, and disease modeling.

Next Generation Microfluidic Systems: Trends

At the core of microfluidics lies the precise manipulation of fluids at the microscale, which is primarily facilitated by the laminar fluid flow.¹⁶ Traditionally, microfluidic devices consist of closed microconduits designed for specific applications (see Figure 1). However, the field is currently undergoing dynamic shift toward innovative technological advances. For example, nowadays digital microfluidics¹⁷ challenges standard microchannels by providing control over fluids through discrete droplets. Moreover, the integration of microfluidics with electronics,¹⁸ photonics,¹⁹ plasmonic,²⁰ magnetics,²¹ nanomaterials,^{22,23} and advanced microscopy and spectroscopy techniques²⁴ enhances the accuracy in fluid manipulation, improves the precision in sensing and characterizing biological samples, and increases the overall automation and screening efficiency.

Notably, when microfluidics is integrated with wearable devices,²⁵ it facilitates non- to minimally invasive health and illness measurements on individuals. Whereas, when it is integrated with multifilament threads,²⁶ it enables the development of intelligent fabrics that continuously monitor human physiological condition in real time. Paper microfluidics²⁷ represents another important branch of microfluidics with applications in point-of-care diagnostics for disease detection, biomarker monitoring, and blood testing (see Figure 2). In recent years, paperbased microfluidic devices advanced into a new dimension by incorporating the art of origami (ie, paper folding) to transform from 2D microfluidics to 3D microfluidics. This innovative technology utilizes capillary action to passively "guide" 3D fluid flow on porous paper, thus eliminating the need for valves or external pumps. For example, Liu and Crooks²⁸ fabricated an entire 3D device using a flat sheet of paper in a single photolithographic step through origami approach. The device compatibility for chemical analysis was demonstrated in fluorescent and colorimetric assays. Zhu et al²⁹ developed 3D origami



Figure 2. Lab-on-a-paper devices: wearable (a) electrochemical microfluidic device and (b) antibody test assay for viral infection detection and severity monitoring, respectively. Point-of-care fingerstick devices for (c) liver function and (d) glucose level testing in blood. Reproduced from Kim et al,⁹ with permission from Elsevier (a), from Boumar et al,¹⁰ with permission from Springer Nature (b), from Pollock et al,¹¹ with permission from American Association for the Advancement of Science (c), and from Luo et al,¹² with permission from MDPI (d).

paper-based ratio-metric fluorescent microfluidic device for visual point-of-care detection of alkaline phosphatase.

Microfluidic probes,¹³ on the other hand, further expand the boundaries of microfluidics through innovative "wall-free" control of fluid, which makes them suitable for multiplexing. For example, Dorrigiv et al^{30,31} presented computer-controlled drug screening platform that utilizes pixelated MFPs to enable multiplexed delivery of multiple biochemical reagents—up to 9 distinct biochemical reagents—to 3D tumor models. This innovation significantly increased the throughput of traditional MFPs, enabling multiple testing scenarios in each experiment. Conversely, Brimmo et al³² experimented with various aperture configurations to bio-fabricate selective multicellular patterns on an open substrate without the use of confined conduits.

In recent years, microfluidics influenced various aspects of life, from applications in smartphones to outer space. In space, microfluidics requires special forces beyond gravitational forces to function effectively. These forces include capillary flow,³³ thermocapillary (Marangoni) forces,³⁴ and electrolytic gas evolution.^{34,35} To date, various types of electrospray thrusters,³⁶ based on capillary and porous surface wetting emitters, successfully demonstrated functionality both on Earth and in space, among other innovative applications.³⁷⁻³⁹ These examples highlight the significant role of microfluidics in space exploration. Furthermore, microfluidic chips are developed to track the physiological indicators of astronauts,⁴⁰ identify diseases,⁴¹ and monitor living conditions including air, water, and food quality.⁴² Additionally, the Phorm iPad mini case is a real-life example of using microfluidics to create pop-up guiding bubble buttons for providing a tactile interface on touch-screen devices.⁴³ This innovation is particularly valuable for individuals who may have difficulty with operating touchscreens, such as blind, visually impaired, elderly, and individuals with diseases that affect fine motor skills, including arthritis and Parkinson's disease.

Modern digital electronics typically rely on rigid semiconductor materials, which limit their physical configurability. This constraint makes flexible and bendable electronics desirable in several applications, including sensing and actuation. In this regard, microfluidics provides the necessary flexibility needed for various applications that are not achievable with rigid semiconductors.⁴⁴ For example, in the work by Cheng and Wu,⁴⁵ a deformable radiation sensor was achieved by fabricating its radio frequency antenna through soft lithography patterning of the microfluidic channel grid onto polydimethylsiloxane (PDMS), a silicone polymer. Room-temperature conductive liquid alloy, Galinstan, is then used to fill the channels to allow for stretchability and bending.⁴¹ The same microfluidic fabrication technique was employed by Othman et al⁴⁶ to fabricate a multi-modal microfluidic bend sensor for robotic sensing applications. These microfluidic smart sensors are proven to provide valuable insights into organ stiffness during minimally invasive surgeries, as demonstrated in the preceding works by the same authors.^{46,47} In addition to mentioned applications, microfluidic devices provide high performance in various applications, including heat sinks to cool integrated circuits,⁴⁸ simplified solutions in agriculture to study enzyme reactions and plant responses,^{49,50} and in wearable devices to monitor body movements⁵¹ or mimic human skin.⁵²

Materials, Fabrication, 3D Printing, and Commercialization

Characteristics of the used materials in manufacturing microfluidic devices significantly impact the fabrication cost, performance, and versatility of their functions. Traditional microfluidic devices commonly employ the PDMS due to its ease of production, biocompatibility, and optical transparency. Their fabrication is commonly based on soft lithography,⁵³ where master molds with microfluidic channels are initially created, followed by their replication using casting. This technique offers precise control over channel geometries and other microstructures, but is labor-intensive and time-consuming.54 In contrast, paper microfluidic platforms employ hydrophilic cellulose- or nitrocellulose-based paper substrates due to their intrinsic capillary action characteristic.55 This enables the passive flow of fluid within the porous 3D microenvironment of the paper, which is guided through paths fabricated via chemical modifications, wax printing, or other methods. Microfluidic probes, on the other hand, are classically fabricated by standard photo- and stereo-lithography approaches in silicon and glass substrates, but more recently, researchers started employing ultraviolet (UV)-curable, biocompatible, and transparent resins used in stereolithographic 3D printing.56 Their fabrication based on 3D printing emerged as an innovative approach for creating customizable device designs that otherwise would be time consuming using the conventional photolithography methods.

In the last decade, the commercialization of microfluidic devices witnessed substantial growth.^{57,58} Standard microfluidic chips are now widely available in the market, where they provide individuals and healthcare personnel with point-of-care solutions. In particular, paper microfluidic platforms, similar to pregnancy tests, are becoming preferred tools for diagnosis of infectious diseases in resource-limited settings due to their simplicity in function, flexibility in design, portability, and low cost.

After over 3 decades of development, a new concept in microfluidics, the modular microfluidics,⁵⁹ has emerged. This innovative concept was made possible by the consistency and precision of the injection-molded bricks, such as those from the famous brand LEGO, which fit together perfectly regardless of where they are manufactured in the world. Consequently, modularity has infinite potential that eliminates the borders of creativity. For example, these bricks inspired MIT researchers to use them as the foundation for a new modular microfluidic design that enabled them to form modular microfluidic devices

capable of conducting biological operations such as cell sorting, fluid mixing, and molecule filtering.⁶⁰

Applications in Biomedical Research

A key feature of microfluidics is the ability to integrate various biology, engineering, computation, chemistry, and physics related functions into a single device. In single-cell analysis, microfluidic devices enable isolation and phenotyping^{14,61,62} and sorting and patterning^{15,32,63} of rare cells, such as circulating tumor or plasma cells. These capabilities are instrumental in unraveling cellular heterogeneity and identifying biomarkers (Figure 3). In bioassays, microfluidic devices achieve high sensitivity and rapid quantification through the utilization of different phenomena and methods, such as isotachophoresis,⁶⁴ a promising approach for protein extraction, purification, and quantification.

Microfluidic devices also play a pivotal role in biosensing applications by exhibiting remarkable signal amplification capabilities.⁶⁶ In point-of-care diagnostics, on the other hand, microfluidic devices enable rapid detection of diseases and infections.⁶⁷ This expedites patient care and relieves the testing burden in laboratories, especially during unprecedented pandemic times such as the Coronavirus Disease 2019 (COVID-19).^{68,69} The environmental implications of microfluidic technology are equally promising. Monitoring water and food quality, air pollutants, and soil contaminants is achievable with improved precision.⁷⁰

Paper microfluidics is a good example of recent innovations in microfluidics. Given the advantages, such as capillarydriven fluid flow and portability, paper microfluidic platforms enable point-of-care diagnostics to be efficiently conducted with minimal sample volumes and shorter analysis times. Additionally, textile microfluidics, which is often integrated into wearable devices, extends the reach of point-of-care diagnostics. These "smart" textiles incorporate microfluidic channels that collect and analyze bodily fluids such as sweat,⁷¹ urine,⁷² and blood.⁷³ In the process, the automation through microfluidics ensures seamless data collection, while the integration into clothing offers non-invasive and continuous health monitoring in real time.

Microfluidics is rapidly integrating innovative technologies, including automation and robotics, and the Internet of Things (IoT) (Figure 4). In terms of automation, labor-intensive and time-consuming experimental tasks, such as sample preparation, mixing, and detection, can now be performed in parallel, thus reducing human error and enabling high-throughput screening and analysis. For example, in drug efficacy testing, the automation in microfluidic devices allows for a large library of compounds to be screened quickly and efficiently.⁷⁴ Furthermore, the precise control of biomolecular concentration gradients in these devices enables investigation of cellular responses and behaviors including patterning, differentiation, migration, and the development of drug resistance.^{75,76} For example, microfluidics enables the production of plasmonic nanomaterials



Figure 3. Lab-on-a-tip devices: Non-contact microfluidic probe technologies for (a) affinity-based multiplex isolation of circulating tumor cells and clusters, and (b) spatiotemporal resolved single cell manipulation and analysis. Reproduced from Deliorman et al,⁶⁵ with permission from Springer Nature (a), and from Brimmo et al,¹⁵ with permission from Wiley (b).



Figure 4. Microfluidic platforms integrated with Internet of Things (IoT) for healthcare applications: (Left) Paper-based electrochemical wearable sensors integrated with microfluidics and nanotechnology for combined sampling and data analysis. Reproduced from Deroco et al,⁷⁸ with permission from Wiley. (Right) A wearable patch, incorporating fluorescent probes on paper substrates and cotton thread microfluidics, is fabricated and paired with a smartphone-based fluorescence imaging module, a self-developed smartphone app, and an IoT-based model to enable multi-sensing of sweat biomarkers, including glucose, lactate, chloride, pH, and volume. Reproduced from Ardalan et al,⁷⁹ with permission from Elsevier.

through aqueous colloidal synthesis within a structured composite foam lattice inside a straightforward microfluidic device.⁷⁷ This approach allows for precise control over reagent dispensing and mixing, while the organized foam structure facilitates compartmentalized nanoparticle growth. Importantly, this production method operates continuously, possesses inherent digital control, is scalable, and operates fully automated.

In the context of the IoT, the integration of microfluidics with data networks results in "smart" devices that can transmit real-time information to remote locations for monitoring and in-depth analysis of the measurements.^{78,79} Microfluidic devices, with their ability to precisely manipulate small volumes of fluids and perform complex chemical and biological assays, are wellsuited for creating intelligent and responsive systems. By combining microfluidics with artificial intelligence (AI), these devices can take on advanced functionalities.^{80,81}

For example, microfluidic sensors can collect substantial data from various biological and chemical processes. AI



Figure 5. Human-on-a-chip system: The Human-on-a-chip system connects different organs within a single device through a microfluidic circulatory system. By integrating organ models like the lung, heart, gut, liver, kidney, and bone, the 4 key drug processes such as absorption, distribution, metabolism, and excretion can be simulated effectively. Reproduced from Huh et al,¹⁰⁰ with permission from Elsevier.

algorithms can then be employed to analyze this data, identify patterns, and make predictions or decisions based on the collected information. This integration enables microfluidic systems to autonomously adapt, optimize processes, and respond to changing conditions using cloud-based analysis and remote monitoring.⁸² As a result, the connectivity and analytical capability hold potential in fields such as healthcare, environmental monitoring, and diagnostics, where real-time insights and rapid decision-making are crucial.⁸³ For example, microfluidic devices integrated with environmental sensors can continuously monitor water quality and transmit data for real-time analysis.⁸⁴

In the context of the dynamic platforms, the integration of microfluidics with robotics creates platforms capable of executing even more advanced research tasks.⁸⁵ For example, a robotic arm integrated with an acoustofluidic device as its end effector induces oscillations within a capillary to create controlled flow patterns or microstreaming when placed in a liquid.⁸⁶ Moreover, self-guided movement of microrobots in fluids allows their autonomous navigation capabilities in complex environments such as blood vessels.⁸⁷

Tissue Engineering and Human-on-a-Chip

Tissue engineering merges biological cells, natural biomaterials, and dynamic biochemical and biophysical factors with microfluidic technologies to form 3D tissue constructs for drug efficacy testing,⁸⁸ disease modeling,⁸⁹ and other fundamental research.^{90,91} The formation of the tissue constructs relies on the growth and differentiation of various cell types, such as parenchymal and mesenchymal fibroblasts, to mimic cell-cell and cell-environment interactions within natural tissues.^{92,93} During this process, biomaterials provide structural integrity and mechanical support for the cells. These 3D scaffolds mimic the extracellular matrix of the target tissue for facilitating the 3D cell adhesion, migration, and organization.⁹⁴ Cellular behavior and tissue development, on the other hand, are regulated by both biochemical factors, such as growth factors, cytokines, and signaling molecules,⁹⁵ and mechanical factors, such as tissue stiffness.⁹⁶ These factors are delivered spatiotemporally to induce cellular proliferation, differentiation, and morphogenesis.⁹⁷

Merging these 3 components (ie, cells, biomaterials, and biochemical/mechanical factors) is crucial for fabricating 3D tissue constructs that mimic real organs.⁹⁸ Organ-on-a-chip devices, for this matter, are becoming increasingly successful in providing the necessary in vitro microenvironment for cells to interact with the biomaterials and get exposed to the biochemical and mechanical factors during their growth and differentiation. Nowadays, the developed organ-on-a-chip models offer accurate representation of human organs, and provide spatiotemporal organ-specific responses to drugs and diseases.99 Human-on-a-chip systems, on the other hand, connects multiple organ-on-a-chip units (Figure 5). This integration aims to mimic how various organs influence each other and how drugs or diseases impact the entire body.¹⁰⁰ Importantly, this integration also has transformative implications for pre-clinical testing. Traditionally, animal testing was a necessary step prior to clinical trials. However, with the advent

of the organ-on-a-chip platforms, these pre-clinical tests are increasingly replacing animals with microfluidic chips that closely replicate the responses of human body to drugs.¹⁰¹ This transition not only saves animal lives but also yields outcomes that are more relevant to clinical results. As a result, it enhances both the efficiency and ethical considerations of the drug development processes.

Current Challenges and Future Directions

The miniaturization of laboratory procedures and essays demands interdisciplinary collaboration between engineers, biologists, chemists, and medical experts to develop the full potential of lab-on-a-chip, organ-on-a-chip, lab-on-a-paper, and lab-on-tip devices. Whereas accurate and reproducible control of fluid flow at microscales requires detailed design, computation, fabrication, optimization, and biological testing. Particularly, the integration of conventional 3D printing with microfluidics minimizes the microfabrication workflow and enables the creation of complex designs that were previously challenging to achieve.

While the technological and scientific advances in microfluidics are exciting, several challenges still persist. One of these challenges is the physical adsorption of hydrophobic and other small molecules in PDMS microchannels that leads to altered composition of solutions and inconsistent results.¹⁰² Another challenge is the micro-to-macro scalability.¹⁰³ The complexities of organs consisting of a number of cells are not accurately reflected by relatively small number of experimental cells. Importantly, the use of a common (standardized) medium in multiorgan-on-a-chip platforms poses challenges for the growth and differentiation of cells in certain organ tissues.¹⁰⁴ In addition to these challenges, shortcoming of unified standards for various aspects of microfluidics poses a significant hurdle. The International Organization for Standardization (ISO) is the global entity responsible for international standardization efforts. In this regard, microfluidics requires standards for dimensions, connections, and initial device classification due to its diverse applications, numerous technologies, and wide array of materials.¹⁰⁵ This diversity becomes problematic when attempting to integrate microfluidic components. Moreover, there is a pressing need for standardized testing, which may be application-specific. Some of these tests include leakage testing,106 pressure testing,107 and flow throughput testing.108 Developing these test protocols must take into consideration the material of the chips as well as the temperature and pressure ranges of operation. To expedite market access, researchers have identified specific application classes with defined temperature and pressure ranges, which provide boundary conditions for the development of tests.¹⁰⁹ Standardized specifications for microfluidic devices include standardizing chip thickness, outer dimensions for compatibility with microplates and microscope slides, as well as port pitch and diameters.¹¹⁰

The integration of microfluidics with innovative technologies holds substantial promise in overcoming many challenges in the field. For example, the 3D printing can facilitate rapid prototyping toward commercialization.¹¹¹ Advanced sensing technologies can enhance the capabilities of microfluidic systems.^{112,113} Whereas, advanced wearable sensors can provide real-time and precise measurements of various parameters, ranging from molecular concentrations to cellular responses.¹¹⁴ Notably, droplet microfluidics can accelerate drug discovery workflow by coupling with high-throughput paper-based 3D cell culture and spheroids.^{115,116} This integration can expedite the assessment of drug candidates through fast compound screening, and significantly reduce reliance on animal testing by utilizing patient-specific cells.

In parallel, AI algorithms can optimize experimental protocols, automate data analysis, and predict outcomes based on large datasets, thus accelerating biological testing with improved precision in data acquisition.⁸¹ Moreover, the use of smart materials with stimuli-responsive properties can improve the functionality and reliability of microfluidic devices.¹¹⁷ Noteworthy, miniaturization can offer more accurate representations of organ function at the microscale, thus accelerating drug development, toxicity assessment, and disease modeling.¹¹⁸

In the last decade, the integration of smartphones and microfluidic devices opened up new possibilities.¹¹⁹ This integration enabled the utilization of microfluidic detection modules that rely on complementary metal oxide semiconductor cameras of the smartphones.¹²⁰⁻¹²² More than ever, they found applications in medical and public health practice where they serve as point-of-care-diagnostic tools.¹²³ For example, one notable integration is the work by Schaumburg et al,¹²⁴ where a free platform called "Appuente" was developed. This platform seamlessly combines microfluidic chips, smartphones, and cloud access. It includes a mobile application for end users that provides several functions, such as chip identification and tracking, guidance and control, data processing, smart imaging, and result reporting.

In tissue engineering, the goal is to recreate the microenvironment of natural tissues to facilitate proper cell growth, differentiation, and function. This involves replicating the biochemical and biophysical factors that cells experience in their physiological environment. In this regard, organ-on-a-chip devices successfully generate human organs in microfluidic devices, which expedite drug testing and disease modeling. In the near future, multiple organ-on-a-chip models are envisioned to be interconnected within a human-on-a-chip system. When combined with automation, robotics, and disease modeling, human-on-a-chip models can provide personalized detailed view on how drugs affect the entire body of a human.¹²⁵ Excitingly, a radical step was recently taken by the U.S. Food and Drug Administration toward ending the use of animal test

subjects in pharmaceutical research.¹²⁶ This game-changing decision can open doors for microfluidic systems to provide various complex in vitro multiorgan models for "real-life" drug development, toxicity assessment, and disease modeling as alternatives to animal testing.

Conclusion

Next generation microfluidic systems stand at the forefront of transformative technologies, bridging automation, screening, and manipulation with biomedical research. Presently, challenges still exist such as material compatibility, scaling, and standardization across multiple systems and procedures. However, the integration of AI, smart materials, and miniaturization is a promising approach to redefine the conventional approaches, and paves the way for creating transformation in biomedical research and applications that will revolutionize human health. Whereas the integration with 3D printing, advanced sensing, robotics, and clinical methods holds the key to diverse applications ranging from fundamental research to healthcare solutions.

Acknowledgements

D.S.A. acknowledges the NYU Global PhD fellowship. M.A.Q. acknowledges the financial support from NYU Abu Dhabi.

Author Contributions

M.D. and M.A.Q.: Study conception and design. M.D. and D.S.A.: Manuscript preparation including literature review, analysis, and writing. M.A.Q.: Supervision of the work and revising the manuscript.

REFERENCES

- Whitesides GM. The origins and the future of microfluidics. *Nature*. 2006;442:368-373.
- Pattanayak P, Singh SK, Gulati M, et al. Microfluidic chips: recent advances, critical strategies in design, applications and future perspectives. *Microfluid Nanofluidics*. 2021;25:99.
- Zarrintaj P, Saeb MR, Stadler FJ, et al. Human organs-on-chips: a review of the state-of-the-art, current prospects, and future challenges. *Adv Biol.* 2022;6:e2000526.
- Yeh EC, Fu CC, Hu L, Thakur R, Feng J, Lee LP. Self-powered integrated microfluidic point-of-care low-cost enabling (SIMPLE) chip. Sci Adv. 2017;3:e1501645.
- Bhuiyan NH, Hong JH, Uddin MJ, Shim JS. Artificial intelligence-controlled microfluidic device for fluid automation and bubble removal of immunoassay operated by a smartphone. *Anal Chem.* 2022;94:3872-3880.
- Li K, Yang X, Xue C, Zhao L, Zhang Y, Gao X. Biomimetic human lung-on-achip for modeling disease investigation. *Biomicrofluidics*. 2019;13:031501.
- Michas C, Karakan MÇ, Nautiyal P, et al. Engineering a living cardiac pump on a chip using high-precision fabrication. *Sci Adv.* 2022;8:eabm3791.
- Noviana E, Ozer T, Carrell CS, et al. Microfluidic paper-based analytical devices: from design to applications. *Chem Rev.* 2021;121:11835-11885.
- Kim HE, Schuck A, Lee SH, Lee Y, Kang M, Kim YS. Sensitive electrochemical biosensor combined with isothermal amplification for point-of-care COVID-19 tests. *Biosens Bioelectron*. 2021;182:113168.
- Boumar I, Deliorman M, Sukumar P, Qasaimeh MA. Spike- and nucleocapsidbased gold colloid assay toward the development of an adhesive bandage for rapid SARS-CoV-2 immune response detection and screening. *Microsyst Nanoeng*. 2023;9:82.
- Pollock NR, Rolland JP, Kumar S, et al. A paper-based multiplexed transaminase test for low-cost, point-of-care liver function testing. *Sci Transl Med.* 2012;4:152ra129.
- 12. Luo JJ, Pan SW, Yang JH, et al. Detecting glucose levels in blood plasma and

artificial tear by Au(I) complex on the carbopol polymer: a microfluidic paperbased method. *Polymers*. 2018;10:1001.

- Brimmo AT, Qasaimeh MA. Microfluidic probes and quadrupoles: a new era of open microfluidics. *IEEE Nanotechnol Mag.* 2017;11:20-31.
- Glia A, Deliorman M, Sukumar P, et al. Herringbone microfluidic probe for multiplexed affinity-capture of prostate circulating tumor cells. *Adv Mater Tech*nol. 2021;6:2100053.
- Brimmo AT, Menachery A, Sukumar P, Qasaimeh MA. Noncontact multiphysics probe for spatiotemporal resolved single-cell manipulation and analyses. *Small.* 2021;17:e2100801.
- Convery N, Gadegaard N. 30 years of microfluidics. *Micro Nano Eng.* 2019;2:76-91.
- Abdelgawad M. Digital microfluidics: automating microscale liquid handling. IEEE Nanotechnol Mag. 2020;14:6-23.
- Annese VF, Hu C. Integrating microfluidics and electronics in point-of-care diagnostics: current and future challenges. *Micromachines*. 2022;13:1923.
- Rodríguez-Ruiz I, Ackermann TN, Muñoz-Berbel X, Llobera A. Photonic labon-a-chip: integration of optical spectroscopy in microfluidic systems. *Anal Chem.* 2016;88:6630-6637.
- Wang C, Cai Y, MacLachlan A, Chen PY. Novel nanoplasmonic-structurebased integrated microfluidic biosensors for label-free in situ immune functional analysis: a review of recent progress. *IEEE Nanotechnol Mag.* 2020;14:46-53.
- Khizar S, Ben Halima H, Ahmad NM, Zine N, Errachid A, Elaissari A. Magnetic nanoparticles in microfluidic and sensing: from transport to detection. *Electrophoresis.* 2020;41:1206-1224.
- 22. Menaa F, Fatemeh Y, Vashist SK, Iqbal H, Sharts ON, Menaa B. Graphene, an interesting nanocarbon allotrope for biosensing applications: advances, insights, and prospects. *Biomed Eng Comput Biol.* 2021;12:1-19.
- Sooriyaarachchi D, Maharubin S, Tan GZ. ZnO nanowire-anchored microfluidic device with herringbone structure fabricated by maskless photolithography. *Biomed Eng Comput Biol.* 2020;11:1-8.
- Xie Y, Xu X, Wang J, Lin J, Ren Y, Wu A. Latest advances and perspectives of liquid biopsy for cancer diagnostics driven by microfluidic on-chip assays. *Lab Chip.* 2023;23:2922-2941.
- Lin PH, Nien HH, Li BR. Wearable microfluidics for continuous assay. Annu Rev Anal Chem. 2023;16:181-203.
- Nilghaz A, Ballerini DR, Shen W. Exploration of microfluidic devices based on multi-filament threads and textiles: a review. *Biomicrofluidics*. 2013;7:51501.
- 27. Li X, Ballerini DR, Shen W. A perspective on paper-based microfluidics: current status and future trends. *Biomicrofluidics*. 2012;6:011301.
- Liu H, Crooks RM. Three-dimensional paper microfluidic devices assembled using the principles of origami. JAm Chem Soc. 2011;133:17564-17566.
- Zhu Y, Tong X, Wei Q, et al. 3D origami paper-based ratiometric fluorescent microfluidic device for visual point-of-care detection of alkaline phosphatase and butyrylcholinesterase. Biosens Bioelectron. 2022;196:113691.
- Dorrigiv D, Goyette PA, St-Georges-Robillard A, Mes-Masson AM, Gervais T. Pixelated microfluidics for drug screening on tumour spheroids and ex vivo microdissected tumour explants. *Cancers*. 2023;15:1060.
- Goyette PA, Boulais É, Tremblay M, Gervais T. Pixel-based open-space microfluidics for versatile surface processing. *Proc Natl Acad Sci USA*. 2021;118:e2019248118.
- Brimmo AT, Menachery A, Qasaimeh MA. Microelectrofluidic probe for sequential cell separation and patterning. *Lab Chip.* 2019;19:4052-4063.
- Olanrewaju A, Beaugrand M, Yafia M, Juncker D. Capillary microfluidics in microchannels: from microfluidic networks to capillaric circuits. *Lab Chip*. 2018;18:2323-2347.
- Nijhuis J, Schmidt S, Tran NN. Microfluidics and macrofluidics in space: ISS-proven fluidic transport and handling concepts. Front Space Technol. 2022;2:779696.
- Matsushima H, Fukunaka Y, Kuribayashi K. Water electrolysis under microgravity. Part II. Description of gas bubble evolution phenomena. *Electrochim Acta*. 2006;51:4190-4198.
- 36. Soares JR, Cerqueira NM, Pinto de Castro RP, Santos Oliveira H, Esteves Araujo R. Design and development of new controller suitable to neighbourhood electric vehicle propulsion control. In: *IECON 2006 - 32nd Annual Conference on IEEE Industrial Electronics*, Paris, France, 6-10 November 2006.
- Xiong J, Zhou Z, Sun D, Ye X. Development of a MEMS based colloid thruster with sandwich structure. *Sens Actuators A Phys.* 2005;117:168-172.
- Anderson G, Anderson J, Anderson M, et al. Experimental results from the ST7 mission on LISA pathfinder. *Phys Rev D*. 2018;98:102005.
- Kuang S, Singh NM, Wu Y, et al. Role of microfluidics in accelerating new space missions. *Biomicrofluidics*. 2022;16:021503.
- Nascetti A, Caputo D, Scipinotti R, de Cesare G. Technologies for autonomous integrated lab-on-chip systems for space missions. *Acta Astronaut*. 2016;128:401-408.
- Nawaz AA, Mao X, Stratton ZS, Huang TJ. Unconventional microfluidics: expanding the discipline. *Lab Chip.* 2013;13:1457-1463.

- Calvo-López A, Arasa-Puig E, Puyol M, Casalta JM, Alonso-Chamarro J. Biparametric potentiometric analytical microsystem for nitrate and potassium monitoring in water recycling processes for manned space missions. *Anal Chim Acta*. 2013;804:190-196.
- 43. Saal N. Microfluidic technology enables new user interface. Inf Disp. 2012;28:6-9.
- Wu A, Wang L, Jensen E, Mathies R, Boser B. Modular integration of electronics and microfluidic systems using flexible printed circuit boards. *Lab Chip.* 2010;10:519-521.
- Cheng S, Wu Z. Microfluidic stretchable RF electronics. Lab Chip. 2010;10:3227-3234.
- Othman W, Vandyck KE, Abril C, et al. Stiffness assessment and lump detection in minimally invasive surgery using in-house developed smart laparoscopic forceps. *IEEE J Transl Eng Health Med.* 2022;10:2500410.
- Othman W, Qasaimeh MA. Smart laparoscopic grasper utilizing force and angle sensors for stiffness assessment in minimally invasive surgery. *IEEE Eng Med Bio.* 2021;2021:7336-7339.
- Hidrovo CH, Goodson KE. Active microfluidic cooling of integrated circuits. In: Bakir MS, Meindl JD, eds. *Integrated Interconnect Technologies for 3D Nanoelectronic Systems*. Artech House; 2009:293-329.
- Hammer GL, Sinclair TR, Chapman SC, van Oosterom E. On systems thinking, systems biology, and the in silico plant. *Plant Physiol.* 2004;134:909-911.
- Hansen CL, Skordalakes E, Berger JM, Quake SR. A robust and scalable microfluidic metering method that allows protein crystal growth by free interface diffusion. *Proc Natl Acad Sci USA*. 2002;99:16531-16536.
- Annabestani M, Esmaeili-Dokht P, Olyanasab A, et al. A new 3D, microfluidicoriented, multi-functional, and highly stretchable soft wearable sensor. *Sci Rep.* 2022;12:20486.
- Yin J, Santos VJ, Posner JD. Bioinspired flexible microfluidic shear force sensor skin. Sens Actuators A Phys. 2017;264:289-297.
- 53. Xia Y, Whitesides GM. Soft lithography. Angew Chem Int Ed. 1998;37:550-575.
- Deliorman M, Glia A, Qasaimeh MA. Characterizing circulating tumor cells using affinity-based microfluidic capture and AFM-based biomechanics. *Star Protoc.* 2022;3:101433.
- Kar S, Das SS, Laha S, Chakraborty S. Microfluidics on porous substrates mediated by capillarity-driven transport. *Ind Eng Chem Res.* 2020;59:3644-3654.
- 56. Brimmo A, Goyette PA, Alnemari R, Gervais T, Qasaimeh MA. 3D printed microfluidic probes. *Sci Rep.* 2018;8:10995.
- Volpatti LR, Yetisen AK. Commercialization of microfluidic devices. Trends Biotechnol. 2014;32:347-350.
- Cong H, Zhang N. Perspectives in translating microfluidic devices from laboratory prototyping into scale-up production. *Biomicrofluidics*. 2022;16:021301.
- Lai X, Yang M, Wu H, Li D. Modular microfluidics: current status and future prospects. *Micromachines*. 2022;13:1363.
- Chu J. MIT engineers make microfluidics modular using the popular interlocking blocks. Published January 30, 2018. Accessed September 25, 2023. https:// news.mit.edu/2018/microfluidics-lego-bricks-0131
- Deliorman M, Janahi FK, Sukumar P, et al. AFM-compatible microfluidic platform for affinity-based capture and nanomechanical characterization of circulating tumor cells. *Microsyst Nanoeng*. 2020;6:20.
- Qasaimeh MA, Wu YC, Bose S, et al. Isolation of circulating plasma cells in multiple myeloma using CD138 antibody-based capture in a microfluidic device. *Sci Rep.* 2017;7:45681.
- Sukunar P, Deliorman M, Brimmo AT, et al. Airplug-mediated isolation and centralization of single T cells in rectangular microwells for biosensing. *Adv Ther.* 2020;3:1900085.
- 64. Khnouf R, Han C. Isotachophoresis-enhanced immunoassays: challenges and opportunities. *IEEE Nanotechnol Mag.* 2020;14:6-17.
- 65. Deliorman M, Glia A, Qasaimeh MA. Affinity-based microfluidics combined with atomic force microscopy for isolation and nanomechanical characterization of circulating tumor cells. *Methods Mol Biol.* 2023;2679:41-66.
- Song Y, Lin B, Tian T, et al. Recent progress in microfluidics-based biosensing. *Anal Chem.* 2019;91:388-404.
- 67. Naik P, Jaitpal S, Paul D. The resurgence of paperfluidics: a new technology for cell, DNA, and blood analysis. *IEEE Nanotechnol Mag.* 2020;14:35-45.
- 68. Huang C, Wang Y, Li X, et al. Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. *J Lancet.* 2020;395:497-506.
- Li Q, Guan X, Wu P, et al. Early transmission dynamics in Wuhan, China, of novel coronavirus–infected pneumonia. *N Engl J Med*. 2020;382:1199-1207.
- Wang T, Yu C, Xie X. Microfluidics for environmental applications. Adv Biochem Eng Biotechnol. 2022;179:267-290.
- Zhang S, Tan R, Xu X, Iqbal S, Hu J. Fibers/textiles-based flexible sweat sensors: a review. ACS Mater Lett. 2023;5:1420-1440.
- Marc ME, Ignacio G, Raul FG. A smart textile system to detect urine leakage. *IEEE Sens J.* 2021;21:26234-26242.
- 73. Kurasawa S, Ishizawa H, Fujimoto K, Chino S, Koyama S. Development of

smart textiles for self-monitoring blood glucose by using optical fiber sensor. J Fiber Sci Technol. 2020;76:104-112.

- Liu Y, Sun L, Zhang H, Shang L, Zhao Y. Microfluidics for drug development: from synthesis to evaluation. *Chem Rev.* 2021;121:7468-7529.
- Kim J, Sahloul S, Orozaliev A, et al. Microfluidic electrokinetic preconcentration chips: enhancing the detection of nucleic acids and exosomes. *IEEE Nanotechnol Mag.* 2020;14:18-34.
- Qasaimeh MA, Pyzik M, Astolfi M, Vidal SM, Juncker D. Neutrophil chemotaxis in moving gradients. *Adv Biosyst* 2018; 2: 1700243.
- Duraiswamy S, Khan SA. Plasmonic nanoshell synthesis in microfluidic composite foams. *Nano Lett.* 2010;10:3757-3763.
- Deroco PB, Wachholz D, Kubota LT. Paper-based wearable electrochemical sensors: a new generation of analytical devices. *Electroanalysis*. 2023; 35:202200177.
- Ardalan S, Hosseinifard M, Vosough M, Golmohammadi H. Towards smart personalized perspiration analysis: an IoT-integrated cellulose-based microfluidic wearable patch for smartphone fluorimetric multi-sensing of sweat biomarkers. *Biosens Bioelectron*. 2020;168:112450.
- Liu L, Bi M, Wang Y, et al. Artificial intelligence-powered microfluidics for nanomedicine and materials synthesis. *Nanoscale*. 2021;13:19352-19366.
- Goda K, Lu H, Fei P, Guck J. Revolutionizing microfluidics with artificial intelligence: a new dawn for lab-on-a-chip technologies. *Lab Chip*. 2023;23:3737-3740.
- Sun H, Xie W, Mo J, Huang Y, Dong H. Deep learning with microfluidics for on-chip droplet generation, control, and analysis. *Front Bioeng Biotechnol.* 2023;11:1208648.
- 83. Pouyanfar N, Harofte SZ, Soltani M, et al. Artificial intelligence-based microfluidic platforms for the sensitive detection of environmental pollutants: recent advances and prospects. *Trends Environ Anal Chem.* 2022;34:e00160.
- Saez J, Catalan-Carrio R, Owens RM, Basabe-Desmonts L, Benito-Lopez F. Microfluidics and materials for smart water monitoring: a review. *Anal Chim Acta*. 2021;1186:338392.
- Filippi M, Yasa O, Kamm RD, Raman R, Katzschmann RK. Will microfluidics enable functionally integrated biohybrid robots? *Proc Natl Acad Sci USA*. 2022;119:e2200741119.
- Durrer J, Agrawal P, Ozgul A, Neuhauss SCF, Nama N, Ahmed D. A robotassisted acoustofluidic end effector. *Nat Commun.* 2022;13:6370.
- Schrage M, Medany M, Ahmed D. Ultrasound microrobots with reinforcement learning. Adv Mater Technol. 2023;8:2201702.
- Marei I, Abu Samaan T, Al-Quradaghi MA, et al. 3D tissue-engineered vascular drug screening platforms: promise and considerations. *Front Cardiovasc Med.* 2022;9:847554.
- Ma X, Liu J, Zhu W, et al. 3D bioprinting of functional tissue models for personalized drug screening and in vitro disease modeling. *Adv Drug Deliv Rev.* 2018;132:235-251.
- Li J, Liu Y, Zhang Y, et al. Biophysical and biochemical cues of biomaterials guide mesenchymal stem cell behaviors. *Front Cell Dev Biol.* 2021;9:640388.
- Zeng X, Meng Z, He J, et al. Embedded bioprinting for designer 3D tissue constructs with complex structural organization. *Acta Biomater*. 2022;140:1-22.
- Plikus MV, Wang X, Sinha S, et al. Fibroblasts: origins, definitions, and functions in health and disease. *Cell*. 2021;184:3852-3872.
- Bitar KN, Zakhem E. Design strategies of biodegradable scaffolds for tissue regeneration. *Biomed Eng Comput Biol.* 2014;6:13-20.
- Liu S, Yu JM, Gan YC, et al. Biomimetic natural biomaterials for tissue engineering and regenerative medicine: new biosynthesis methods, recent advances, and emerging applications. *Mil Med Res.* 2023;10:16.
- Yu M, Li YJ, Shao JY, Qin KR. Transport of dynamic biochemical signals in a microfluidic single cell trapping channel with varying cross-sections. *Eur Phys J* E. 2019;42:33.
- 96. Janmey PA, Miller RT. Mechanisms of mechanical signaling in development and disease. *J Cell Sci.* 2011;124:9-18.
- Vijayavenkataraman S, Garcia-Sabate A, Teo JC. Specialized multimaterial print heads for 3D hydrogel printing: tissue-engineering applications. *IEEE Nanotechnol Mag.* 2020;14:42-52.
- Hwang DG, Choi YM, Jang J. 3D bioprinting-based vascularized tissue models mimicking tissue-specific architecture and pathophysiology for in vitro studies. *Front Bioeng Biotechnol.* 2021;9:685507.
- Ingber DE. Human organs-on-chips for disease modelling, drug development and personalized medicine. Nat Rev Genet. 2022;23:467-491.
- Huh D, Hamilton GA, Ingber DE. From 3D cell culture to organs-on-chips. Trends Cell Biol. 2011;21:745-754.
- Konar D, Devarasetty M, Yildiz DV, Atala A, Murphy SV. Lung-on-a-chip technologies for disease modeling and drug development. *Biomed Eng Comput Biol.* 2016;7:16-26.
- Toepke MW, Beebe DJ. PDMS absorption of small molecules and consequences in microfluidic applications. *Lab Chip*. 2006;6:1484-1486.

- Wood R. The challenge of manufacturing between macro and micro. Am Sci. 2014;102:124-131.
- Picollet-D'hahan N, Zuchowska A, Lemeunier I, Le Gac S. Multiorgan-on-achip: a systemic approach to model and decipher inter-organ communication. *Trends Biotechnol.* 2021;39:788-810.
- 105. European Standards. Vol. ISO 22916:2022 (European Standards).
- Silverio V, Guha S, Keiser A, et al. Overcoming technological barriers in microfluidics: leakage testing. *Front Bioeng Biotechnol.* 2022;10:958582.
- Mair DA, Rolandi M, Snauko M, Noroski R, Svec F, Fréchet JM. Room-temperature bonding for plastic high-pressure microfluidic chips. *Anal Chem.* 2007;79:5097-5102.
- Young KM, Shankles PG, Chen T, Ahkee K, Bules S, Sulchek T. Scaling microfluidic throughput with flow-balanced manifolds to simply control devices with multiple inlets and outlets. *Biomicrofluidics*. 2022;16:034104.
- 109. Reyes DR, van Heeren H, Guha S, et al. Accelerating innovation and commercialization through standardization of microfluidic-based medical devices. *Lab Chip.* 2021;21:9-21.
- van Heeren H, Atkins T, Verplanck N, et al. Design guideline for microfluidic device and component interfaces. 2017.
- 111. Attaran M. The rise of 3-D printing: the advantages of additive manufacturing over traditional manufacturing. *Bus Horiz.* 2017;60:677-688.
- 112. Li S, Ma Z, Cao Z, Pan L, Shi Y. Advanced wearable microfluidic sensors for healthcare monitoring. *Small*. 2020;16:e1903822.
- He X, Fan C, Luo Y, Xu T, Zhang X. Flexible microfluidic nanoplasmonic sensors for refreshable and portable recognition of sweat biochemical fingerprint. *npj Flex Electron*. 2022;6:60.
- Yin R, Wang D, Zhao S, Lou Z, Shen G. Wearable sensors-enabled humanmachine interaction systems: from design to application. *Adv Funct Mater*. 2021;31:2008936.

- Alnemari R, Sukumar P, Deliorman M, Qasaimeh MA. Paper-based cell cryopreservation. Adv Biosyst. 2020;4:e1900203.
- Samara B, Deliorman M, Sukumar P, Qasaimeh MA. Cryopreservable arrays of paper-based 3D tumor models for high throughput drug screening. *Lab Chip.* 2021;21:844-854.
- 117. Guo Z, Liu H, Dai W, Lei Y. Responsive principles and applications of smart materials in biosensing. *Smart Mater Med.* 2020;1:54-65.
- Håkanson M, Cukierman E, Charnley M. Miniaturized pre-clinical cancer models as research and diagnostic tools. *Adv Drug Deliv Rev.* 2014;69:52-66.
- Martinez AW, Phillips ST, Carrilho E, Thomas SW, Sindi H, Whitesides GM. Simple telemedicine for developing regions: camera phones and paper-based microfluidic devices for real-time, off-site diagnosis. *Anal Chem.* 2008;80:3699-3707.
- Kühnemund M, Wei Q, Darai E, et al. Targeted DNA sequencing and in situ mutation analysis using mobile phone microscopy. *Nat Commun.* 2017;8:13913.
- Draz MS, Kochehbyoki KM, Vasan A, et al. DNA engineered micromotors powered by metal nanoparticles for motion based cellphone diagnostics. *Nat Commun.* 2018;9:4282.
- 122. Wood CS, Thomas MR, Budd J, et al. Taking connected mobile-health diagnostics of infectious diseases to the field. *Nature*. 2019;566:467-474.
- 123. Wang B, Li Y, Zhou M, et al. Smartphone-based platforms implementing microfluidic detection with image-based artificial intelligence. *Nat Commun.* 2023;14:1341.
- 124. Schaumburg F, Vidocevich JP, Gerlero GS, et al. A free customizable tool for easy integration of microfluidics and smartphones. *Sci Rep.* 2022;12:8969.
- Sapoznik E, Niu G, Zhou Y, Murphy SV, Soker S. Fluorescent cell imaging in regenerative medicine. *Biomed Eng Comput Biol.* 2016;7:29-33.
- Stewart A, Denoyer D, Gao X, Toh YC. The FDA modernisation act 2.0: bringing non-animal technologies to the regulatory table. *Drug Discov Today*. 2023;28:103496.