

# Preliminary study of urinary arsenic concentration and arsenic methylation capacity effects on neurodevelopment in very low birth weight preterm children under 24 months of corrected age

Chuen-Bin Jiang, MD<sup>a,b</sup>, Yu-Mei Hsueh, PhD<sup>c,d</sup>, Guang-Lin Kuo, MS<sup>e</sup>, Chyong-Hsin Hsu, MD<sup>f</sup>, Jui-Hsing Chang, MD<sup>b,f,g</sup>, Ling-Chu Chien, PhD<sup>e,h,\*</sup>

## Abstract

The neurological prognoses of very low birth weight preterm (VLBWP) children during the first 2 years of life will influence their neurodevelopment during subsequent childhood years and adolescence. The objective of this study was to systematically investigate relationships of urinary arsenic (As) concentrations, the As methylation capability, and toenail As concentrations on cognitive, language, and motor development in VLBWP children under 24 months of corrected age.

Participants (n=60) in our study were recruited from October 2010 to April 2013. Urine and toenail samples were collected for evaluation to assess As exposure. The Bayley scales of infant development III were used to evaluate neurodevelopment at 2 years of corrected age. Concentrations of As species in urine and the As concentration in toenails were, respectively, analyzed using HPLC-HG-AAS and ICP-MS.

The mean concentration of total As was 28.6 µg/g creatinine, and inorganic As was 1.01 µg/L in urine. The urine contained an average of 3% inorganic As, 2% monomethylarsonic acid, and 95% dimethylarsinic acid (DMA). The mean concentration of As in toenails was 225 ng/g. Children with a longer gestational age (≥28 weeks) and higher DMA % levels appeared to have the highest unadjusted cognitive and fine motor scores.

Our study results suggest that gestational age is associated with neurodevelopment in VLBWP children. We recommend that further study simultaneously analyze multiple environmental contaminants that may have adverse effects on neurodevelopment, use biomarkers for the mother-child pair, and determine whether prenatal or postnatal As exposure has a greater influence on the neurological development of VLBWP children.

**Abbreviations:** As = arsenic, As<sup>III</sup> = Arsenite<sup>III</sup>, As<sup>V</sup> = Arsenite<sup>V</sup>, Bayley-III = the Bayley Scales of Infant and Toddler Development, DMA<sup>V</sup> = Dimethylarsonic acid, HPLC-HG-AAS = high-performance liquid chromatography with a hydride generator and atomic absorption spectrometer, iAs = inorganic As, ICP-MS = inductively coupled plasma mass spectrometry, MMA<sup>V</sup> = monomethylarsonic acid, nAs = Toenail total As level, NIST = National Institute of Standard Technology, Pb = lead, PMI = primary methylation index, SD = standard deviation, SE = standard error, SMI = secondary methylation index, TUAs = total urinary as level, VLBWP = very low birth weight preterm.

**Keywords:** arsenic methylation capability, gestational age, neurodevelopment, very low birth weight preterm children

Editor: Babak Abdinia.

This work was supported by a grant (TMU102-F-002) from Taipei Medical University.

Conflict of Interest Statement: The authors have disclosed all financial and interpersonal relationships that could present a potential conflict of interest.

<sup>a</sup> Division of Pediatric Gastroenterology, Hepatology and Nutrition, Department of Pediatrics, MacKay Children's Hospital, <sup>b</sup> Mackay Junior College of Medicine, Nursing and Management, Taipei, <sup>c</sup> Department of Family Medicine, Shuang Ho Hospital, <sup>d</sup> Department of Public Health, School of Medicine, College of Medicine, <sup>e</sup> School of Public Health, College of Public Health, Taipei Medical University, <sup>f</sup> Division of Neonatology, Department of Pediatrics, MacKay Children's Hospital, Taipei, <sup>g</sup> MacKay Medical College, New Taipei City, <sup>h</sup> Nutrition Research Center, Taipei Medical University Hospital, Taipei, Taiwan.

\* Correspondence: Ling-Chu Chien, School of Public Health, Taipei Medical University, Taipei, Taiwan, Nutrition Research Center, Taipei Medical University Hospital, Taipei, Taiwan, 250 Wuxing Street, Xinyi District, Taipei 11031, Taiwan (e-mail: lcchien@tmu.edu.tw).

Copyright © 2018 the Author(s). Published by Wolters Kluwer Health, Inc.

This is an open access article distributed under the terms of the Creative Commons Attribution-Non Commercial License 4.0 (CCBY-NC), where it is permissible to download, share, remix, transform, and build upon the work provided it is properly cited. The work cannot be used commercially without permission from the journal.

Medicine (2018) 97:43(e12800)

Received: 7 December 2017 / Accepted: 19 September 2018

<http://dx.doi.org/10.1097/MD.00000000000012800>

## 1. Introduction

Children, compared to adults, may experience higher exposure to environmental contaminants because of early life stage behaviors and physiology.<sup>[1]</sup> Arsenic (As) is a neurotoxicant that can alter cognitive function, particularly in children. One study indicated that children exposed to As and lead (Pb) have impaired central nervous system functions, including poorer performance on long-term memory and attention assessments.<sup>[2]</sup> A cross-sectional study found that long-term As exposure in drinking water led to cognitive deficits among adolescents in Taiwan.<sup>[3]</sup> Hafeman et al<sup>[4]</sup> reported an association between As concentrations and subclinical sensory neuropathy in Bangladesh. Water, urinary, and toenail As concentrations were shown to be inversely related to motor function scores among 304 children aged 8 to 11 years.<sup>[5]</sup> Hsieh et al<sup>[6]</sup> found that the As methylation capacity is associated with developmental delay among children in Taiwan. Those findings suggest that As exposure may cause neurodevelopmental damage.

A body of evidence suggests that methylation of inorganic (i) As is a detoxification mechanism.<sup>[7,8]</sup> As<sup>V</sup> is readily reduced to As<sup>III</sup><sup>[9]</sup> and is subsequently methylated to monomethylarsenic acid (MMA<sup>V</sup>) and dimethylarsinic acid (DMA<sup>V</sup>).<sup>[10,11]</sup> Approximately 55% to 80% of environmentally exposed As is excreted in urine within 3 to 4 days after being metabolized.<sup>[12,13]</sup> The As methylation capacity may particularly confer a higher risk of developmental disorders in children, as they demonstrate a poorer methylation capacity compared to adults.<sup>[14,15]</sup> Toenail clippings have been used to assess biomarkers to evaluate the long-term As exposure<sup>[16]</sup> because of the nails' growth rate of 0.03 to 0.05 mm/day and outgrowth in 12 to 18 months.<sup>[17]</sup>

Very low birth weight (birth weight <1500g) preterm (gestational age <37 weeks) (VLBWP) children are at risk of cognitive deficits, growth delays, and academic underachievement compared to those born at term and with a normal birth weight.<sup>[18–20]</sup> There are more than 200,000 newborn babies in Taiwan each year, and the incidence of prematurity is about 8% to 10%.<sup>[21]</sup> Consequences of growth lags persist not only in the neonatal period but also throughout childhood<sup>[22–24]</sup> and adulthood. The neurological prognoses of VLBWP children during the first 2 years will influence their neurodevelopment during subsequent childhood years and adolescence.

VLBWP children are at increased risk for developmental impairments.<sup>[25–27]</sup> However, studies on both preterm effects and postnatal As exposure of children are limited. To the best of the authors' knowledge, consistent evidence of relationships between As exposure and neurodevelopment in the early life of VLBWP children has not yet been well established.

Consequently, one of the aims of this study was to investigate relationships of cognitive, language, and motor development at 24 months of corrected age with As concentrations, the arsenic methylation capability, and As concentrations in various biomarkers, including the urine and toenails. The Bayley Scales of Infant and Toddler Development (Bayley-III) were calculated and validated at 6, 12, and 24 months of corrected age. The effects of variables such as gestational age and urinary As species on children's cognitive, language, and motor development at 24 months of corrected age were then better elucidated. Therefore, it is crucial to investigate the VLBWP As exposure in children, particularly in the critical early life stage, and develop optimal strategies to reduce later health effects in school-aged and adolescent children.

## 2. Materials and methods

### 2.1. Study subjects

Mackay Memorial Hospital and other nonprofit organizations jointly founded the premature baby foundation of Taiwan and provide follow-up programs for those born very prematurely or at very low birth weight of <1500g after discharge from newborn intensive care units. Children were enrolled in this study who attended the program if they met the following criteria: a birth weight of  $\leq 1500$ g and a gestational age of <37 weeks; neurodevelopment completely evaluated at 6, 12, and 24 months of corrected age; and both urine and toenail samples collected at 24 months of corrected age. In total, 60 children were recruited from Taipei MacKay Memorial Hospital between October 2010 and April 2013. Urine and toenail samples of children were collected and served as biomarkers of internal dose indices of As exposure. A structured questionnaire was administered by a trained interviewer to obtain demographic information of participants and their parents. Sociodemographic characteristics of both the mother and father were collected. Information pertaining to the children's sociodemographic characteristics (e.g., sex, gestational age, weight, head circumference, height, and Apgar scores at birth), dietary intake (e.g., rice, meat, fish, shellfish, eggs, dairy, vegetables, and fruit), parity, and family income data were also collected. This work was approved by Taipei Mackay Memorial Hospital (13MMHIS088). Written informed consent was prepared for both parents of participating infants. All of parents provided written informed consent before beginning the study.

### 2.2. Urinary As profiles and nail As concentration assessments

Urine samples were collected from each participant to analyze As<sup>III</sup>, DMA<sup>V</sup>, MMA<sup>V</sup>, and As<sup>V</sup> profiles as measured by high-performance liquid chromatography with a hydride generator and atomic absorption spectrometer (HPLC-HG-AAS) according to previously published methods.<sup>[28]</sup> Standard reference material (SRM 2670a) from the National Institute of Standard Technology (NIST) was used to assess the precision and accuracy of the urinary As analyses. Recovery rates of As<sup>III</sup>, DMA<sup>V</sup>, MMA<sup>V</sup>, and As<sup>V</sup> ranged 93.8% to 102.2% using the following equation:  $([\text{sample spiked standard solution concentration} - \text{sample concentration}] / \text{standard solution concentration}) \times 100$ . Detection limits of the 4 arsenic species were 0.02, 0.08, 0.05, and 0.07  $\mu\text{g/L}$ , respectively. All urinary As species were adjusted to urinary creatinine. The sum of As<sup>III</sup> and As<sup>V</sup> was referred to as iAs. The sum of As<sup>III</sup>, As<sup>V</sup>, MMA<sup>V</sup>, and DMA<sup>V</sup> was referred to as the total urinary As concentration. Percentages of each As species (iAs %, MMA<sup>V</sup> %, and DMA<sup>V</sup> %) were calculated by dividing the concentration of each species by the total As concentration. Primary methylation and secondary methylation indices were calculated as  $(\text{PMI} = \text{MMA}^{\text{V}}/\text{iAs})$  and  $(\text{SMI} = \text{DMA}^{\text{V}}/\text{MMA}^{\text{V}})$ , respectively. Percentages of each As species, the PMI, and SMI were internal dose indices of As exposure of the children.

Total As concentrations in toenail samples were determined by inductively coupled plasma mass spectrometry (ICP-MS; Thermo X-series II). Toenail samples were rinsed 3 times with deionized water after sonication with a neutral detergent for 30 min and dried in an oven at 37°C for 24 hours. Approximately 0.02g of samples was microwave-digested (Multiwave 3000, Anton-Paar)

with 3 mL of 65% nitric acid (Suprapur, Merck, Kenilworth, NJ). We washed residuals in a Teflon tube with 2% nitric acid and then filtered the digested fluids. Total filtered solutions were stored in 50-mL centrifuge tubes. Certified reference material (CRM GBW 09101b) was used to validate the measurements. The recovery rate ranged 92.1% to 106.6%, with a detection limit of 3.08 ng/g.

**2.3. Determination of neurodevelopment**

The Bayley Scales of Infant and Toddler Development (Bayley-III) were used to calculate and validate neurodevelopment scores to evaluate the cognitive, language, and motor development at 6, 12, and 24 months of corrected age. All participants were seen by their pediatricians, who assessed their development prior to testing.

**2.4. Statistical analysis**

Continuous variable distributions are expressed as the mean ± standard deviation (SD). Right-skewed data were normalized by logarithmic transformation for statistical analyses. The Wilcoxon signed-rank test was used to test significant differences in urinary As profiles and toenail As concentrations between extremely preterm children and preterm children. The Kruskal–Wallis test was used to assess significant differences in developmental scores in 4 groups (gestational age [ $<28$  vs  $\geq 28$  weeks]) vs DMA % (high vs low). The Wilcoxon rank sum test was conducted to test significant differences in binominal variables. All analyses were

performed using the Statistical Analysis Software (SAS) statistical package (SAS, version 9.3, Cary, NC). A  $P$  value  $< .05$  was considered significant.

**3. Results**

Table 1 shows demographic characteristics of all enrolled VLBWP children. The mean gestational age of all participants was  $28.9 \pm 3.3$  weeks, and the mean birth weight was  $1077 \pm 271$  g. In total, 68.6% of children were first parity, and the mean maternal age was  $32.55 \pm 4.34$  years. The mean rice and fish intake levels were  $19.5 \pm 14.5$  and  $6.1 \pm 3.6$  servings/week, respectively. The mean length of breast feeding was  $4.9 \pm 2.8$  months. Approximately 87% of families had a monthly family income of  $\geq$ US\$2300. Urinary As profiles and toenail As concentrations are shown in Table 2. The mean concentration of total As was  $28.5 \pm 2.72$   $\mu$ g/g creatinine, and iAs was  $0.89 \pm 0.22$   $\mu$ g/g creatinine in urine. The urine contained mean 2.42% iAs, 1.6% MMA<sup>v</sup>, and 95.9% DMA<sup>v</sup>. The primary and secondary As methylation indices were 1.85 and 188, respectively. The mean cumulative As concentration in toenails was  $225 \pm 15.8$  ng/g. Toenail As concentrations were positively correlated with urinary As<sup>III</sup> ( $r=0.35$ ,  $P<.01$ ) and urinary total As concentrations ( $r=0.33$ ,  $P<.05$ ; data not shown).

Comparisons of urinary As profiles and nail As concentrations between extremely preterm and preterm children are shown in Table 3. We divided participants into 2 groups: extremely preterm (gestational age  $<28$  weeks) and preterm (gestational age  $\geq 28$  weeks). We found no significant differences in urinary As profiles or toenail As concentrations between the 2 groups (Table 3). VLBWP children of 6 months of corrected age had lower mean Bayley-III scores in cognitive and fine motor development than those at 12 or 24 months of

**Table 1**  
**Sociodemographic and dietary characteristics of very low birth weight preterm children.**

Characteristic	All subjects (N=60) Mean ± SD or %
Children	
Age, months	28.4 ± 6.0
Male	51.7
Female	48.3
Gestational age, weeks	28.9 ± 3.3
Birth length, cm	36.1 ± 4.7
Birth weight, g	1077 ± 271
Head circumference, cm	26.1 ± 2.4
1-min Apgar score	6.6 ± 1.5
5-min Apgar score	8.3 ± 1.0
Parity	
1	68.6
2	27.5
≥3	3.9
Rice intake, servings/week	19.5 ± 14.5
Meat and poultry intake, servings/week	9.3 ± 6.7
Fish intake, servings/week	6.1 ± 3.6
Shellfish intake, servings/week	2.5 ± 2.2
Egg intake, servings/week	6.2 ± 7.4
Dairy intake, servings/week	11.9 ± 17.8
Vegetable intake, servings/week	7.5 ± 7.6
Fruit intake, servings/week	5.9 ± 6.1
Father's age, years	35.3 ± 6.2
Mother's age, years	32.6 ± 4.3
Breastfeeding, months	4.9 ± 2.8
Family income (US\$/month)	
<2300	13.3
≥2300	86.7

SD=standard deviation.

**Table 2**  
**Urinary As capacity and nail As concentration of very low birth weight preterm children.**

Variable	Mean ± SE	Medium	Minimum	Maximum.
Urinary As indices, $\mu$ g/L				
As <sup>III</sup>	0.38 ± 0.12	0.01	0.01	3.52
As <sup>V</sup>	0.05 ± 0.01	0.04	0.04	0.24
MMA <sup>V</sup>	0.34 ± 0.11	0.03	0.03	4.29
DMA <sup>V</sup>	12.37 ± 1.56	8.75	1.21	55.84
TUAs	13.14 ± 1.74	9.09	1.28	62.13
Urine creatinine, mg/dL	47.97 ± 4.36	42.85	7.70	167.00
Adjusted urinary As indices, $\mu$ g/g creatinine				
As <sup>III</sup> /creatinine	0.76 ± 0.22	0.06	0.01	6.13
As <sup>V</sup> /creatinine	0.13 ± 0.02	0.09	0.02	0.70
MMA <sup>V</sup> /creatinine	0.51 ± 0.13	0.17	0.04	5.32
DMA <sup>V</sup> /creatinine	27.05 ± 2.51	21.44	5.15	87.4
iAs /creatinine	0.89 ± 0.22	0.23	0.05	6.25
TUAs/creatinine	28.46 ± 2.72	22.30	5.26	97.23
As species percentage				
iAs %	2.42 ± 0.43	1.33	0.14	14.03
MMA <sup>V</sup> %	1.60 ± 0.28	0.78	0.12	8.63
DMA <sup>V</sup> %	95.98 ± 0.59	97.58	83.34	99.62
As methylation capacity indexes				
PMI	1.85 ± 0.56	0.56	0.02	24.08
SMI	188 ± 28.11	125	9.71	806
nAs, ng/g	225 ± 15.8	208	72.5	560

As=Arsenic, As<sup>III</sup>=arsenite<sup>III</sup>, As<sup>V</sup>=arsenite<sup>V</sup>, DMA<sup>V</sup>=dimethylarsonic acid, iAs=inorganic As, Max=maximum, Min=minimum, MMA<sup>V</sup>=monomethylarsonic acid, nAs=toenail total As level, PMI=primary methylation index, SE=standard error, SMI=secondary methylation index, total As level=As<sup>III</sup>+As<sup>V</sup>+MMA<sup>V</sup>+DMA<sup>V</sup>, TUAs=total urinary As level.

**Table 3**

Comparisons of urinary As profiles and nail As concentrations between extremely preterm and preterm children.

Variable	Extremely preterm GA <28 weeks (n=29)	Preterm GA ≥28 weeks (n=31)	P value
	Mean ± SE	Mean ± SE	
Urinary As indices			
As <sup>III</sup> , μg/L	0.48 ± 0.25	0.34 ± 0.13	.77
As <sup>V</sup> , μg/L	0.05 ± 0.01	0.04 ± 0.01	.20
MMA <sup>V</sup> , μg/L	0.43 ± 0.22	0.30 ± 0.13	.86
DMA <sup>V</sup> , μg/L	14.8 ± 3.64	11.2 ± 1.54	.61
TUAs, μg/L	15.8 ± 4.04	11.9 ± 1.73	.67
Urine creatinine, mg/dL	48.5 ± 6.77	47.7 ± 5.63	.71
TUAs, μg/g creatinine	31.5 ± 6.13	27.0 ± 2.81	.78
iAs %	2.16 ± 0.66	2.54 ± 0.55	.63
MMA <sup>V</sup> %	1.38 ± 0.39	1.71 ± 0.37	.86
DMA <sup>V</sup> %	96.5 ± 0.92	95.8 ± 0.74	.48
nAs, ng/g	203 ± 24.8	237 ± 20.2	.16

As = arsenic, As<sup>III</sup> = arsenite<sup>III</sup>, As<sup>V</sup> = arsenite<sup>V</sup>, DMA<sup>V</sup> = Dimethylarsonic acid, GA = gestational Age, iAs = inorganic As, MMA<sup>V</sup> = monomethylarsonic acid, nAs = toenail total As level, SE = standard error, TUAs = total urinary As level.

**Table 4**

Results of neurological effects evaluated by Bayley scales of infant and toddler development (Bayley-III) for very low birth weight preterm children at 6, 12, and 24 months of corrected age.

Score*		6 months	12 months	24 months
		Mean ± SD	Mean ± SD	Mean ± SD
Cognitive		7.2 ± 2.7	9.5 ± 3.1	9.1 ± 2.3
Language	Receptive	8.4 ± 2.2	8.0 ± 2.1	9.3 ± 2.1
	Expressive	7.1 ± 2.3	6.9 ± 2.3	8.2 ± 2.7
Motor	Fine motor	7.7 ± 3.5	8.9 ± 2.2	8.8 ± 2.3
	Gross motor	7.8 ± 2.4	7.1 ± 2.8	7.5 ± 2.4

SD = standard deviation.

\* Normative score: average, 10.

corrected age (Table 4). The mean gross motor scores for children at 12 and 24 months of corrected age (7.1 and 7.5, respectively) were lower than the cognitive, receptive language, and fine motor scores. We thus attempted to further examine whether an interaction existed between gestational age and As species. Due to the small numbers of subjects, we could not perform a multivariate analysis that included both dichotomized variables. However, children with a longer gestational age (≥28 weeks) and higher DMA % levels appeared to have the highest unadjusted cognitive and fine motor scores (Table 5).

**Table 5**

Mean Bayley scales of infant and toddler development (Bayley-III) for very low birth weight preterm children among children with longer or shorter gestational age and higher or lower As species levels (dimethylarsonic acid %) at 24 months of corrected age.

Score (mean ± SD)	Extremely preterm infants GA <28weeks		Preterm infants GA ≥28 weeks		P value*
	Low DMA%	High DMA%	Low DMA%	high DMA%	
Cognitive	6.5 ± 2.1	8.1 ± 2.2 <sup>†</sup>	9.5 ± 1.9	9.6 ± 2.2 <sup>†</sup>	.02
Receptive language	6.5 ± 0.7	9.0 ± 1.9	10.0 ± 1.6	9.5 ± 2.2	.09
Expressive language	6.0 ± 0.0	7.6 ± 2.5	8.8 ± 2.9	8.6 ± 2.8	.20
Fine motor	6.0 ± 1.4	7.5 ± 2.0 <sup>†</sup>	9.5 ± 1.2	9.4 ± 2.2 <sup>†</sup>	.006
Gross motor	5.0 ± 2.8	6.8 ± 2.4	9.6 ± 1.8	7.7 ± 2.3	.05

\* Kruskal–Wallis test.

<sup>†</sup> P < .05, by the Wilcoxon rank sum test.

As = arsenic, DMA = Dimethylarsonic acid, GA = gestational age, SD = standard deviation.

#### 4. Discussion

This is the first study in Taiwan to investigate the impacts of As exposure and relevant risk factors on neurodevelopment at 24 months of corrected age in VLBWP children. We followed the neurodevelopment trends of VLBWP children at 6, 12, and 24 months of corrected age. We found that they had lower mean gross motor development scores before 24 months of corrected age compared to other neurodevelopment scores. Children with a longer gestational age and higher DMA% levels had better cognitive and fine motor scores. However, a risk of As exposure related to neurodevelopment was not observed in this study. But, additional monitoring is still required in the future, especially as the growth lag persists throughout childhood and adulthood.

In Taiwan, Hsieh et al demonstrated that a high urinary total As level (19.69 ± 2.50 μg/L) was significantly associated with the risk of developmental delays in children.<sup>[6]</sup> A poor As methylation capacity (high MMA<sup>V</sup> and low DMA<sup>V</sup>) was significantly negatively associated with the health-related quality of life and functional performance in children with developmental delays. Children exposed to iAs over a long period of time may have decreased intelligence quotient scores.<sup>[29]</sup> Animal studies have shown that chronic As exposure can induce brain migration and delayed maturation, and is related to motor learning and memory impairment.<sup>[30–32]</sup> However, the maternal body burden and current exposure are major sources of As for the fetus. In a Mexican Arsenic (BEAR) Pregnancy Cohort study, the authors suggested that maternal urinary iAs was negatively

associated with the gestational age and newborn length.<sup>[33]</sup> Those authors also found that iAs exposure during the prenatal period may be a source of health effects later in life. Thus, the body of evidence indicates that children exposed to As during prenatal or postnatal periods present with signs of neurobehavioral dysfunction and growth delays.

The placenta is a selective fetal-maternal barrier that is thought to play a role in preventing environmental contaminants from transferring from the mother to the fetus.<sup>[34]</sup> However, iAs may be able to pass through the placenta and result in preterm births, low birth weights, and stillbirths.<sup>[29,35,36]</sup> Low-level As exposure during the prenatal period impairs the fetal immune regulation system, potentially causing disease later in life.<sup>[37]</sup> In a previous Taiwanese study, respective mean Bayley-III scores were 11.4, 12.5, 9.88, 12.9, and 10.4 for cognitive, receptive language, expressive language, and fine motor and gross motor development in full-term children.<sup>[38]</sup> We found that VLBWP children had lower neurodevelopment scores compared to those of full-term children. Gestational age, weight, head circumference, height, and Apgar 1- and 5-min scores at birth in VLBWP children were positively correlated with Bayley-III scores at 12 and 24 months of corrected age (data not shown). Notably, we could not rule out the possibility that preterm birth was caused by As exposure or co-exposure in mothers during pregnancy.

Arsenic concentrations in the urine and toenails in the present study were comparable to those found in other studies. The mean total As concentration was 13.14 (range: 1.28–62.1)  $\mu\text{g/L}$  in urine and  $225 \pm 15.8$  (range: 72.5–560)  $\text{ng/g}$  in toenails in the present study. In Bangladesh, urinary total As concentrations were 66 and 34  $\mu\text{g/L}$  in 1.5-year-old children who were exposed to As-contaminated drinking water.<sup>[39,40]</sup> Urinary total As concentrations in Taiwan were 19.69 and 10.22  $\mu\text{g/L}$  in children with and without developmental delays, respectively.<sup>[6]</sup> Davis et al showed that the median As concentration in infant toenails was 60 (range: 1–1210)  $\text{ng/g}$  and asserted that As concentrations in infant toenails are a reliable indicator and can serve as a reflection of As exposure from maternal water and dietary sources during the critical window of gestation.<sup>[41]</sup>

The As methylation capacity decreases with a high MMA% in urine and is related to increased cancer risk, systemic disease, and developmental delays.<sup>[6,42]</sup> In the present study, we observed low urinary (13.14  $\mu\text{g/L}$ ) and toenail As concentrations (225  $\text{ng/g}$ ) and high DMA% (96%) in VLBWP children. Toenail As concentrations were significantly positively correlated with urinary total As concentrations in the present study. Arsenic methylation is affected by age, sex, race, socioeconomic status, smoking, drinking, exposure route, As species, lifestyle, and dietary habits.<sup>[43–46]</sup> Children who consume folate, meat, eggs, red-orange vegetables, and green leafy vegetables may have higher As methylation capacities.<sup>45</sup> Measuring toenail As concentrations represents an integrated, reliable, and long-term biomarker for exposure sources. We found that toenail As concentrations were negatively associated with egg ( $r = -0.34$ ,  $P < .05$ ) and dairy intake ( $r = -0.40$ ,  $P < .05$ ; data not shown). Eggs had the highest total choline concentration,<sup>[47]</sup> which may increase the As methylation capacity.

In the present study, we demonstrated that gestational age was positively associated with receptive communication as well as fine and gross motor development scores at 24 months of corrected age. There is evidence that preterm or extremely preterm children are at risk for cognitive deficits, growth delays, academic

underachievement, and behavioral problems.<sup>[18–20]</sup> With regard to Bayley-III scores, gestational age was the most important risk factor for health effects in life in VLBWP children with low-level As exposure.

This study was a longitudinal study: urinary and toenail samples, and neurodevelopment scores were collected from VLBWP children at 6, 12, and 24 months of corrected age. However, we did not investigate other potential environmental contaminants, such as Pb, cadmium, or polychlorinated biphenyls, that also may have adverse effects on neurodevelopment.<sup>[48–50]</sup> Additionally, the number of participants was small, and a lack of recruitment of full-term children is a weakness of this study. We did not collect infant toenails, which are highly indicative of maternal As exposure during gestation.<sup>[41]</sup> Thus, additional research is necessary to determine whether prenatal or postnatal As exposure has a greater influence on neurological development in VLBWP children.

## 5. Conclusion

In conclusion, to the best of our knowledge, this is the first study analyzing low-level urinary total As, high MMA<sup>V</sup>%, and low DMA<sup>V</sup>% in VLBWP children in Taiwan. We find that they had lower mean gross motor development scores before 24 months of corrected age compared to other neurodevelopment scores. Our results suggest that gestational age is associated with neurodevelopment in VLBWP children, even without obvious As exposure. Children with a longer gestational age ( $\geq 28$  weeks) and higher DMA % levels appear to have the highest unadjusted cognitive and fine motor scores. However, this study is cross-sectional in design without a full-term control group and includes only a small number of participants. Larger sample sizes are needed to investigate neurodevelopment in future studies.

## Acknowledgments

The authors express their gratitude to staff at the Department of Pediatrics, MacKay Children's Hospital, who assisted with data collection, and all participants in this study for their time and effort.

## Author contributions

**Conceptualization:** Chuen-Bin Jiang, Ling-Chu Chien.  
**Data curation:** Chyong-Hsin Hsu, Jui-Hsing Chang.  
**Formal analysis:** Guang-Lin Kuo, Chyong-Hsin Hsu.  
**Investigation:** Yu-Mei, Hsueh, Guang-Lin Kuo, Chyong-Hsin Hsu, Jui-Hsing Chang.  
**Methodology:** Yu-Mei, Hsueh.  
**Project administration:** Ling-Chu Chien.  
**Resources:** Jui-Hsing Chang.  
**Software:** Guang-Lin Kuo.  
**Supervision:** Yu-Mei, Hsueh, Ling-Chu Chien.  
**Validation:** Ling-Chu Chien.  
**Visualization:** Guang-Lin Kuo.  
**Writing – original draft:** Chuen-Bin Jiang.  
**Writing – review & editing:** Ling-Chu Chien.

## References

- [1] USEPA Guidance on Selecting Age Groups for Monitoring and Assessing Child-Hood Exposures to Environmental Contaminants (Final). 2005; U.S. Environmental Protection Agency, Washington, DC: EPA/630/P-03/003F.

- [2] Calderon J, Navarro ME, Jimenez-Capdeville ME, et al. Exposure to arsenic and lead and neuropsychological development in Mexican children. *Environ Res* 2001;85:69–76.
- [3] Tsai SY, Chou HY, The HW, et al. The effects of chronic arsenic exposure from drinking water on the neurobehavioral development in adolescence. *Neurotoxicology* 2003;24:747–53.
- [4] Hafeman DM, Ahsan H, Louis ED, et al. Association between arsenic exposure and a measure of subclinical sensory neuropathy in Bangladesh. *J Occup Environ Med* 2005;47:778–84.
- [5] Parvez F, Wasserman GA, Factor-Litvak P, et al. Arsenic exposure and motor function among children in Bangladesh. *Environ Health Perspect* 2011;119:1665–70.
- [6] Hsieh RL, Huang YL, Shieh HS, et al. Arsenic methylation capacity and developmental delay in preschool children in Taiwan. *Int J Hyg Environ Health* 2014;217:678–86.
- [7] Vahter M. Mechanisms of arsenic biotransformation. *Toxicology* 2002;181-182:211–7.
- [8] Styblo M, Vega L, Germolec DR, et al. Metabolism and toxicity of arsenicals in cultured cells. Arsenic exposure and health effects 1st edn1999;United Kingdom Elsevier Science Ltd, Kidlington, Oxford:311-323.
- [9] Vahter M. Biotransformation of trivalent and pentavalent inorganic arsenic in mice and rats. *Environ Res* 1981;25:286–93.
- [10] Buchet JP, Lauwerys R, Roels H. Urinary excretion of inorganic arsenic and its metabolites after repeated ingestion of sodium metaarsenite by volunteers. *Int Arch Occup Environ Health* 1981;48:111–8.
- [11] Buchet JP, Lauwerys R, Roels H. Comparison of the urinary excretion of arsenic metabolites after a single oral dose of sodium arsenite, monomethylarsenate, or dimethylarsinate in man. *Int Arch Occup Environ Health* 1981;48:71–9.
- [12] Huang C, Ke Q, Costa M, et al. Molecular mechanisms of arsenic carcinogenesis. *Mol Cell Biochem* 2004;255:57–66.
- [13] Huang YK, Tseng CH, Huang YL, et al. Arsenic methylation capability and hypertension risk in subjects living in arseniasis-hyperendemic areas in southwestern Taiwan. *Toxicol Appl Pharmacol* 2007;218:135–42.
- [14] Chowdhury UK, Rahman MM, Sengupta MK, et al. Pattern of excretion of arsenic compounds [arsenite, arsenate, MMA(V), DMA(V)] in urine of children compared to adults from an arsenic exposed area in Bangladesh. *J Environ Sci Health A Tox Hazard Subst Environ Eng* 2003;38:87–113.
- [15] Concha G, Vogler G, Lezcano D, et al. Exposure to inorganic arsenic metabolites during early human development. *Toxicol Sci* 1998;44:185–90.
- [16] Slotnick MJ, Nriagu JO. Validity of human nails as a biomarker of arsenic and selenium exposure: a review. *Environ Res* 2006;102:125–39.
- [17] Fleckman P, Scher RK, Daniel CR. Basic science of the nail unit. *Nails: Therapy, Diagnosis, Surgery Saunders, Philadelphia*:1997;37–54.
- [18] Caravale B, Tozzi C, Albino G, et al. Cognitive development in low risk preterm infants at 3–4 years of life. *Arch Dis Child Fetal Neonatal Ed* 2005;90:F474–479.
- [19] McNicholas F, Healy E, White M, et al. Medical, cognitive and academic outcomes of very low birth weight infants at age 10–14 years in Ireland. *Ir J Med Sci* 2014;183:525–32.
- [20] Oliveira GE, Magalhaes LC, Salmela LF. Relationship between very low birth weight, environmental factors, and motor and cognitive development of children of 5 and 6 years old. *Rev Bras Fisioter* 2011;15:138–45.
- [21] Health Promotion Administration, Ministry of Health and Welfare. Birth Reporting Database. Taipei, Taiwan. 2015; Available at: <https://olap.hpa.gov.tw/Search.aspx?menu=100000000006&Keyword=%u65b0%u751f%u5152>. Accessed March 23, 2017.
- [22] Cooke R, Foulder-Hughes L. Growth impairment in the very preterm and cognitive and motor performance at 7 years. *Arch Dis Child* 2003;88:482–7.
- [23] Ramel SE, Demerath EW, Gray HL, et al. The relationship of poor linear growth velocity with neonatal illness and two-year neurodevelopment in preterm infants. *Neonatology* 2012;102:19–24.
- [24] Frisone MF, Mercuri E, Laroche S, et al. Prognostic value of the neurologic optimality score at 9 and 18 months in preterm infants born before 31 weeks' gestation. *J Pediatr* 2002;140:57–60.
- [25] Romeo DM, Cioni M, Scoto M, et al. Prognostic value of a scorable neurological examination from 3 to 12 months post-term age in very preterm infants: a longitudinal study. *Early Hum Dev* 2009;85:405–8.
- [26] Oros D, Altermir I, Elia N, et al. Pathways of neuronal and cognitive development in children born small-for-gestational age or late preterm. *Ultrasound Obstet Gynecol* 2014;43:41–7.
- [27] Hsueh YM, Huang YL, Huang CC, et al. Urinary levels of inorganic and organic arsenic metabolites among residents in an arseniasis-hyperendemic area in Taiwan. *J Toxicol Environ Health A* 1998;54:431–44.
- [28] Hsueh YM, Chen WJ, Lee CY, et al. Association of arsenic methylation capacity with developmental delays and health status in children: a prospective case-control trial. *Sci Rep* 2016;6:37287.
- [29] Dhar P, Mohari N, Mehra RD, et al. Preliminary morphological and morphometric study of rat cerebellum following sodium arsenite exposure during rapid brain growth (RBG) period. *Toxicology* 2007;234:10–20.
- [30] Mishra D, Flora SJ. Differential oxidative stress and DNA damage in rat brain regions and blood following chronic arsenic exposure. *Toxicol Ind Health* 2008;24:247–56.
- [31] Wang Y, Li S, Piao F, et al. Arsenic down-regulates the expression of Camk4, an important gene related to cerebellar LTD in mice. *Neurotoxicol Teratol* 2009;31:318–22.
- [32] Laine JE, Bailey KA, Rubio-Andrade M, et al. Maternal arsenic exposure, arsenic methylation efficiency, and birth outcomes in the Biomarkers of Exposure to Arsenic (BEAR) pregnancy cohort in Mexico. *Environ Health Perspect* 2015;123:186–92.
- [33] Iyengar GV, Rapp A. Human placenta as a 'dual' biomarker for monitoring fetal and maternal environment with special reference to potentially toxic trace elements. Part 1: physiology, function and sampling of placenta for elemental characterisation. *Sci Total Environ* 2001;280:195–206.
- [34] Lindgren A, Danielsson BR, Dencker L, et al. Embryotoxicity of arsenite and arsenate: distribution in pregnant mice and monkeys and effects on embryonic cells in vitro. *Acta Pharmacol Toxicol (Copenh)* 1984;54:311–20.
- [35] Vahter M. Effects of arsenic on maternal and fetal health. *Annu Rev Nutr* 2009;29:381–99.
- [36] Nadeau KC, Li Z, Farzan S, et al. In utero arsenic exposure and fetal immune repertoire in a US pregnancy cohort. *Clin Immunol* 2014;155:188–97.
- [37] Hsi HC, Jiang CB, Yang TH, et al. The neurological effects of prenatal and postnatal mercury/methylmercury exposure on three-year-old children in Taiwan. *Chemosphere* 2014;100:71–6.
- [38] Fangstrom B, Hamadani J, Nermell B, et al. Impaired arsenic metabolism in children during weaning. *Toxicol Appl Pharmacol* 2009;239:208–14.
- [39] Hamadani JD, Tofail F, Nermell B, et al. Critical windows of exposure for arsenic-associated impairment of cognitive function in pre-school girls and boys: a population-based cohort study. *Int J Epidemiol* 2011;40:1593–604.
- [40] Davis MA, Li Z, Gilbert-Diamond D, et al. Infant toenails as a biomarker of in utero arsenic exposure. *J Expo Sci Environ Epidemiol* 2014;24:467–73.
- [41] Chen Y, Wu F, Graziano JH, et al. Arsenic exposure from drinking water, arsenic methylation capacity, and carotid intima-media thickness in Bangladesh. *Am J Epidemiol* 2013;178:372–81.
- [42] Yu H, Liu S, Li M, et al. Influence of diet, vitamin, tea, trace elements and exogenous antioxidants on arsenic metabolism and toxicity. *Environ Geochem Health* 2016;38:339–51.
- [43] Kordas K, Queirolo EL, Manay N, et al. Low-level arsenic exposure: nutritional and dietary predictors in first-grade Uruguayan children. *Environ Res* 2016;147:16–23.
- [44] Shen H, Niu Q, Xu M, et al. Factors affecting arsenic methylation in arsenic-exposed humans: a systematic review and meta-analysis. *Int J Environ Res Public Health* 2016;13:205.
- [45] Jain RB. Association of arsenic exposure with smoking, alcohol, and caffeine consumption: data from NHANES 2005–2010. *Environ Toxicol Pharmacol* 2015;39:651–8.
- [46] Zeisel SH, Mar MH, Howe JC, et al. Concentrations of choline-containing compounds and betaine in common foods. *J Nutr* 2003;133:1302–7.
- [47] Wasserman GA, Liu X, Parvez F, et al. Water arsenic exposure and children's intellectual function in Araihaazar, Bangladesh. *Environ Health Perspect* 2004;112:1329–33.
- [48] Tang D, Li TY, Liu JJ, et al. Effects of prenatal exposure to coal-burning pollutants on children's development in China. *Environ Health Perspect* 2008;116:674–9.
- [49] Plusquellec P, Muckle G, Dewailly E, et al. The relation of environmental contaminants exposure to behavioral indicators in Inuit preschoolers in Arctic Quebec. *Neurotoxicology* 2010;31:17–25.
- [50] Kippler M, Tofail F, Hamadani JD, et al. Early-life cadmium exposure and child development in 5-year-old girls and boys: a cohort study in rural Bangladesh. *Environ Health Perspect* 2012;120:1462–8.