

# *CFH* 184G as a genetic risk marker for anterior uveitis in Chinese females

Ming-ming Yang, Timothy Y.Y. Lai, Pancy O.S. Tam, Sylvia W.Y. Chiang, Carmen K.M. Chan, Fiona O.J. Luk, Tsz Kin Ng, Chi-Pui Pang

Department of Ophthalmology and Visual Sciences, The Chinese University of Hong Kong, Hong Kong, China

**Objective:** To investigate the association of three single nucleotide polymorphisms (SNPs) in the complement factor H (*CFH*), *KIAA1109*, and interleukin-27 (*IL-27*) genes in patients with anterior uveitis (AU).

**Methods:** A case-control study was performed in 98 Chinese AU patients and 308 healthy controls. Three SNPs including *CFH*-rs800292, *KIAA1109*-rs4505848, and *IL27*-rs4788084 were detected using TaqMan SNP Genotyping Assays. Analyses were also stratified according to gender, clinical features and human leukocyte antigen (HLA)-B27 status of the patients.

**Results:** No significant association was found between all three SNPs and AU. However, when stratified by gender, there were significant increases in the frequency of the *CFH*-rs800292 184G allele and GG homozygosity in female patients compared with control subjects (p=0.003 and p=0.009, respectively). Similar association was not detected in males. No significant association was found between AU and *KIAA1109*-rs4505848 or *IL27*-rs4788084 even stratified by gender. There was no significant difference in genotypes of AU patients stratified by various clinical features. Subgroup analyses showed that all three SNPs (rs800292, rs4505848, and rs4788084) were not associated with AU in HLA-B27–positive patients, neither in HLA-B27–negative patients.

**Conclusions:** Our results showed an association between AU and *CFH* polymorphism in Chinese female patients but not in males, indicating gender-specific genetic differences in *CFH*. Gender should be considered in genetic studies of anterior uveitis even extending to other immunologic diseases.

Uveitis is an intraocular inflammatory disease involving the uveal tract and can be anatomically classified as anterior, intermediate, posterior, and panuveitis [1]. Anterior uveitis (AU) is the most common form, accounting for 50% to 92% of cases in most Western countries and 28% to 50% in Asian countries [2]. AU involves the anterior part of the uveal tract and can be idiopathic or associated with other immunologic disorders such as ankylosing spondylitis (AS) and rheumatoid arthritis (RA). Although the exact pathogenesis of AU is unclear, clinical and animal studies have demonstrated that the inflammation in AU is regulated by various endogenous immunological factors. In addition, an important conceptual consideration is that the disease can manifest in individuals with genetic predisposition coupled with environmental trigger [3,4]. AU is closely associated with human leukocyte antigen (HLA)-B27, the major histocompatibility complex (MHC) type I gene [5,6]. Gene polymorphisms of various non-MHC genes such as cytokines and antioxidant enzyme genes have also been implicated to play important roles in the pathogenesis of AU [7-9].

Interleukins are potent inflammatory mediators produced by white blood cells and play important roles in the development of uveitis [10-16]. Studies have shown that the levels of interleukin 2 (IL-2), interleukin 21 (IL-21) and their receptors were upregulated in both experimental autoimmune uveitis (EAU) animals and in uveitis patients [10-13]. Recently, several studies have revealed that IL-27 expression was upregulated during uveitis and retinal cells could suppress uveitis through production of IL-27 [15,16]. Several single nucleotide polymorphisms (SNPs) in interleukin genes have been found to be associated with various types of uveitis but only a few of them were found to have a strong association [17-19]. In recent years, based on genome-wide association study (GWAS), several candidate SNPs were successively found to be associated with a range of immune-mediated diseases, such as Behçet's disease, type 1 diabetes mellitus, RA, celiac disease, and Graves' disease [20-24]. Some of these genetic loci were replicated reciprocally in various diseases, implying that these loci might be the general risk factors for multiple autoimmune diseases [25-27].

The complement system is another important pathway of innate immunity. The system can be divided into the classic, lectin and alternative pathways and plays an important role in many immunological diseases including uveitis. Various studies have demonstrated that the activation of complement system is critical for the development of experimental autoimmune anterior uveitis (EAAU) and the depletion of the

Correspondence to: Timothy Y.Y. Lai, M.D., FRCS, FRCOphth, Department of Ophthalmology & Visual Sciences, The Chinese University of Hong Kong, Hong Kong Eye Hospital, 147K Argyle Street, Kowloon, Hong Kong; Phone; +852 27623134; FAX: +852 27159490; email: tyylai@cuhk.edu.hk

		CONTROL SUBJECTS.		
Polymorphism rs800292 ( <i>CFH</i> 184G/A)	Anterior uveitis (n=98)	Controls (n=308)	p-value	Odds ratio
Genotype				
AA	10 (10.2)	48 (15.6)	0.17§	
AG	42 (42.9)	145 (47.1)	0.091*	
GG	46 (46.9)	115 (37.3)	0.19‡	
Allele			•	
А	62 (31.6)	241 (39.1)	0.059§	1.39 (0.99–1.96)
G	134 (68.4)	375 (60.9)	Ŭ	
rs4505848 (KIAA1109 G/A)				
Genotype				
GG	12 (12.2)	50 (16.2)	0.42§	
AG	57 (58.2)	157 (51.0)	Ŭ	
AA	29 (29.6)	101 (32.8)		
Allele				
G	81 (41.3)	257 (41.7)	0.92§	
А	115 (58.7)	359 (58.3)	Ŭ	
rs4788084 (IL27 T/C)				
Genotype				
TT	9 (9.2)	21 (6.8)	0.74§	
СТ	39 (39.8)	126 (40.9)	0	
CC	50 (51.0)	161 (52.3)		
Allele	~ /			
Т	57 (29.1)	168 (27.3)	0.62§	
С	139 (70.9)	448 (72.7)	U	

 TABLE 1. COMPARISON OF GENOTYPE AND ALLELE FREQUENCIES OF Rs800292, rs4505848, and rs4788084 polymorphisms in patients with AU and control subjects.

Data are the number of subjects (% of the total group). §x<sup>2</sup> test. \*p-value for dominant model. ‡p-value for recessive model.

host's complement system could result in complete inhibition of EAAU [28,29]. Under normal conditions, complement system is continuously active at a low level and is tightly regulated by intraocular complement regulatory proteins (CRegs) such as complement factor H (CFH), decayaccelerating factor and S-protein. Studies have shown that suppression of CRegs can exacerbate EAAU due to unregulated complement activation [30,31]. CFH acts as a key regulator in the complement alternative pathway and is involved in the pathogenesis of many immunological diseases [32-34]. Recent studies have suggested that variants in the CFH gene, as well as in the genes of other CFH-related proteins might be involved in many immune-mediated diseases [35-37]. We hypothesized that rs4505848 within the KIAA1109/Testis nuclear RNA-binding protein (Tenr)/IL2/ IL21 gene cluster, rs4788084 in IL-27, and rs800292 in CFH might be involved in the pathogenesis of AU. The purpose of our study is to determine the association of these immune-associated SNPs in Chinese patients with AU.

### **METHODS**

*Study design and subjects:* Subjects were recruited from the Hong Kong Eye Hospital. The study protocol was approved by an institutional review board and all the procedures were conducted according to the tenets of the Declaration of Helsinki. Informed consent was obtained from all study subjects after explanation of the nature of the study.

All patients underwent detailed ocular examination including visual acuity, intraocular pressure, slit lamp, and dilated fundus examinations. Clinical details were also collected from the case notes including age, sex, medical history such as systemic illness, age at initial presentation, laterality, pattern of anterior uveitis (acute, recurrent or chronic), clinical features, and complications of AU. The definition of uveitis was based on the Standardization Uveitis Nomenclature (SUN) classification [38]. Acute AU was defined as AU resolving completely within 3 months, chronic AU as AU not fully resolved within 3 months, and recurrent AU as the development of AU more than once. Patients with any of the following situations were excluded from the study (1) AU secondary to ocular or systemic infections; (2) AU secondary to specific syndromes (e.g., Posner-Schlossman's syndrome, Fuchs' uveitis, Vogt-Koyanagi-Harada (VKH) or Behcet's disease); (3) patients who were unable to cooperate during ocular examination and with chronic uveitis at the onset of the study. Three hundred and eight subjects aged 50 years older with no ophthalmic eye disease except senile cataract were included as control subjects.

*DNA extraction and genotyping:* Venous blood was obtained from each subject and genomic DNA was extracted with a DNA extraction kit (QIAamp; Qiagen, Hilden, Germany) according to the manufacturer's instructions. *CFH*-rs800292, *KIAA1109*-rs4505848, and *IL27*-rs4788084 SNPs were genotyped by TaqMan allelic discrimination assay (TaqMan;

Polymorphism	Female AU patients (n=53)	Female controls (n=183)	p-value	Odds ratio (95% confidence
rs800292 ( <i>CFH</i> 184G/A)				
Genotype	t u			
AA	3 (5.7)	31 (16.9)	0.015§	
AG	19(35.8)	82 (44.8)	0.009*	2.28 (1.22–4.24)
GG	31 (58.5)	70 (38.3)	0.039	3.40 (1.00–11.6)
Allele				
Α	25 (23.6)	144 (39.3)	0.003§	2.10 (1.28–3.45)
IJ	81 (76.4)	222 (60.7)	2	
rs4505848 (KIAA1109 G/A)	~	~		
Genotype				
CG	7 (13.2)	27 (14.8)	0.54§	
AG	32 (60.4)	95 (51.9)	٥	
AA	14 (26.4)	61 (33.3)		
Allele				
IJ	46 (43.4)	149 (40.7)	0.62§	
Α	60 (56.6)	217 (59.3)	٥	
rs4788084 ( <b>IL27</b> T/C)	~	~		
Genotype				
LL	8 (15.1)	12 (6.6)	0.13§	
CT	19(35.8)	78 (42.6)	2	
CC	26 (49.1)	93 (50.8)		
Allele	~	~		
T	35 (33.0)	102 (27.9)	0.30§	
C	71 (67.0)	264 (72.1)		

•	
nodel	
ive n	
ecess	
for r	
value	
-d‡ :	
nodel	
iant r	
lomir	
for c	
value	
-d*	
test.	
$\$\chi^2$	
oup).	
al gre	
e tota	
the	
J	
(% of	
jects (% of	
subjects (% of	
er of subjects (% of	
umber of subjects (% of	
he number of subjects (% of	
are the number of subjects (% of	

TS.

2657

Polymorphism	Male AU patients (n=45)	Male controls (n=125)	p-value
rs800292 (CFH 184G/A)			
Genotype			
AA	7 (15.6)	17 (13.6)	0.92§
AG	23 (51.1)	63 (50.4)	0.75*
GG	15 (33.3)	45 (36.0)	0.75‡
Allele			
А	37 (41.1)	97 (38.8)	0.70§
G	53 (58.9)	153 (61.2)	-
rs4505848 (KIAA1109 G/A)			
Genotype			
GG	5 (11.1)	23 (18.4)	0.52§
AG	25 (55.6)	62 (49.6)	
AA	15 (33.3)	40 (32.0)	
Allele			
G	35 (38.9)	108 (43.2)	0.48§
А	55 (61.1)	142 (56.8)	U U
rs4788084 (IL27 T/C)			
Genotype			
TT	1 (2.2)	9 (7.2)	0.43§
СТ	20(44.4)	48 (38.4)	0
CC	24 (53 3)	68 (54 4)	
Allele	- (555)		
Т	22 (24 4)	66 (26 4)	0 728
Ċ	68 (75 6)	184 (73.6)	0.723
C	00 (75.0)	101 (75.0)	

 TABLE 3. COMPARISON OF GENOTYPE AND ALLELE FREQUENCIES OF RS800292, RS4505848, AND RS4788084 POLYMORPHISMS IN MALE PATIENTS WITH

 AU AND MALE CONTROL SUBJECTS.

Data are the number of subjects (% of the total group).  $\xi \chi^2$  test. \*p-value for dominant model.  $\pm$ p-value for recessive model.

Applied Biosystems [ABI], Foster City, CA) according to the manufacturer's instructions. All PCR amplifications were performed with the following thermal cycling conditions: 95 °C for 10 min followed by 40 cycles of 92 °C for 15 s, and 62 °C for 1.5 min (rs4505848 and rs4788084); and 60 °C for 1 min (rs800292), respectively. The HLA-B27 allele was detected by nested PCR as described by Konno et al. [39]. The B locus of HLA was first amplified, the B27 allele was then amplified from the diluted PCR product of the B locus by using sequence-specific primers. The B27 allele was further detected and confirmed as described previously [39]. All PCR reactions were performed with Taq polymerase (HotStarTaq Plus; Qiagen) in an automated thermal cycler (model 9700; ABI). Pre- and post-PCR plate readings were performed on a sequence detection system (Prism 7000; ABI), and the allele types were confirmed by the system software (Prism 7000 SDS software version 1.1; ABI).

Statistical analysis: Hardy–Weinberg equilibrium (HWE) for genotype frequencies of the SNPs in the control group were tested by  $\chi^2$  test. The genotype frequencies for each polymorphism were determined by direct counting and allele frequencies were calculated. Allelic and genotypic frequencies between patients with AU and controls were compared by  $\chi^2$  test. Analyses were also performed by stratifying AU patients based on gender and HLA-B27 status and compared with control subjects. Odds ratios (OR) and 95% confidence intervals (CI) were calculated. Bonferroni correction was applied to adjust for the number of comparisons performed based on the total number of loci and thus a p value of 0.017 was considered as statistically significant.

#### RESULTS

*Patients demographics:* Ninety-eight patients with AU were recruited, including 45 (45.9%) males and 53 (54.1%) females. The mean±SD age of the patients was 49.7±16.0 years (range, 11–87 years). Forty-seven (48.0%) patients had unilateral uveitis, and 51 (52.0%) had bilateral involvement. Systemic diseases associated with the patients included AS (n=19, 19.4%); psoriasis (n=1, 1.0%); systemic lupus erythematosus (n=1, 1.0%); ulcerative colitis (n=1, 1.0%), and interstitial nephritis (n=1, 1.0%). Ninety-two (93.9%) patients had acute AU only, of which 59 (60.2%) had recurrent episodes of AU.

Associations between SNPs and AU: All genotype frequencies of the 3 SNPs in the control subjects conformed to the Hardy– Weinberg equilibrium. There was no significant difference in allelic and genotypic frequencies for the 3 SNPs (*CFH*rs800292, *KIAA1109*-rs4505848, and *IL27*-rs4788084) in AU patients compared with controls. There was a trend toward higher 184G allele frequency for rs800292 in AU patients

Polymorphism	Recurrent (n=59)	Non-recurrent (n=39)	p-value
rs800292 (CFH 184G/A)			
Genotype			
AA	5 (8.5)	5 (12.8)	0.38§
AG	23 (39.0)	19 (48.7)	
GG	31 (52.5)	15 (38.5)	
Allele			
А	33 (28.0)	29 (37.2)	0.18§
G	85 (72.0)	49 (62.8)	-
rs4505848 (KIAA1109 G/A)			
Genotype			
GG	7 (11.9)	5 (12.8)	0.51§
AG	32 (54.2)	25 (64.1)	-
AA	20 (33.9)	9 (23.1)	
Allele			
G	46 (39.0)	35 (44.9)	0.41§
А	72 (61.0)	43 (55.1)	
rs4788084 (IL27 T/C)			
Genotype			
TT	7 (11.9)	2 (5.1)	0.50§
СТ	22 (37.3)	17 (43.6)	0
CC	30 (50.8)	20 (51.3)	
Allele			
Т	36 (30.5)	21 (26.9)	0.59§
С	82 (69.5)	57 (73.1)	U
Data are the number of su	bjects (% of the total group). $\xi\chi$	<sup>2</sup> test.	

TABLE 4. COMPARISON OF GENOTYPE AND ALLELE FREQUENCIES OF RS800292, RS4505848, AND RS4788084 POLYMORPHISMS IN AU PATIENTS STRATIFIED BY RECURRENCE STATUS.

compared with controls but the difference did not reach the

#### DISCUSSION

level of statistical significance (31.6% versus 39.1%, p=0.059; Table 1).

Associations between SNPs and AU stratified by gender and clinical features: When the analyses were stratified on the basis of gender, there was a significant increase in the frequency of 184G allele and GG homozygosity for the CFH-rs800292 SNP in female AU patients compared with control female subjects (p=0.003, OR=2.10 and p=0.009, OR=2.28, respectively). However, similar difference was not observed in male patients. For the KIAA1109-rs4505848 and IL27-rs4788084, there was no significant difference in both male and female AU patients compared with controls (Table 2 and Table 3). There was no significant difference in allelic and genotypic frequencies in AU patients stratified by recurrence and laterality (Table 4 and Table 5).

Association of SNPs genotypes and allele frequencies stratified by HLA-B27 status: Forty-two (42.9%) of the 98 patients with AU were HLA-B27-positive and fifty-six (57.1%) were HLA-B27-negative. When analyzed on the basis of HLA-B27 status, there was no significant difference in allelic and genotypic frequencies for the 3 SNPs (CFHrs800292, KIAA1109-rs4505848, and IL27-rs4788084) between either HLA-B27-positive or HLA-B27-negative patients compared with control subjects (Table 6).

In our study, we investigated the association of three immuneassociated SNPs in the CFH, KIAA1109, and IL27 in Chinese patients with AU. Our results demonstrated there was a trend toward higher 184G allele frequency in the CFH-rs800292 SNP among AU patients compared with controls. Although the difference failed to reach the level of statistical significance (p=0.059), the findings suggested that CFH might be associated with the susceptibility of developing AU. Since genetic and environmental factors might be associated with some gender-specific differences in the severity and type of immunological diseases, we therefore further evaluated the association stratified by gender. Our results showed a significant increase in the frequency of 184G allele and GG homozygousity in female AU patients compared with control subjects. To our knowledge, the associations between genetic variants in CFH and AU and the gender differences have not been described previously. Activated complement is a "double-edged sword" which might cause self-tissue damage especially in a sensitive organs like the eyes and they must be carefully regulated by CRegs such as CFH [40]. CFH is located in the long arm of chromosome 1 (1q32), which is a major soluble inhibitor of the alternative pathway in controlling complement activation [30]. Furthermore, in vivo studies have revealed that human retinal pigment epithelial (RPE) cells can synthesize and express CFH, which

	51	A105.	
Polymorphism rs800292 (CFH 184G/A)	Bilateral AU (n=51)	Unilateral AU (n=47)	p-value
Genotype			
AA	4 (7.8)	6 (12.8)	0.70§
AG	23 (45.1)	19 (40.4)	
GG	24 (47.1)	22 (46.8)	
Allele			
А	31 (30.4)	31 (33.0)	0.70§
G	71 (69.6)	63 (67.0)	Ŭ
rs4505848 (KIAA1109 G/A)	× ,		
Genotype			
GG	7 (13.7)	5 (10.6)	0.55§
AG	27 (52.9)	30 (63.8)	, i i i i i i i i i i i i i i i i i i i
AA	17 (33.3)	12 (25.5)	
Allele	× ,		
G	41 (40.2)	40 (42.6)	0.74§
А	61 (59.8)	54 (57.4)	Ŭ
rs4788084 (IL27 T/C)		· · · · · · · · · · · · · · · · · · ·	
Genotype			
TT	4 (7.8)	5 (10.6)	0.53§
СТ	23 (45.1)	16 (34.0)	Ŭ
CC	24 (47.1)	26 (55.3)	
Allele		· · · · · · · · · · · · · · · · · · ·	
Т	31 (30.4)	26 (27.7)	0.67§
С	71 (69.6)	68 (72.3)	0

 TABLE 5. COMPARISON OF GENOTYPE AND ALLELE FREQUENCIES OF RS800292, RS4505848, AND RS4788084 POLYMORPHISMS STRATIFIED BY LATERALITY

 STATUS.

Data are the number of subjects (% of the total group).  $\xi \chi^2$  test for 2×3.

upregulates various CRegs including CFH to suppress the development of EAU [31,41,42].

In our previous study, we found SNP rs800292 in CFH was associated with age-related macular degeneration (AMD) in Chinese patients [43]. In addition, CFH has also been found to be associated with other immune-mediated diseases such as multifocal choroiditis, hemolytic-uremic syndrome (HUS) and glomerulonephritis, where some risk loci overlap with those of AMD [35,36]. In this study, the SNP rs800292 was found to be associated with AU in female patients. The change of G184A nucleotide of rs800292 in the CFH gene results in the synthesis of Isoleucine instead of Valine at codon 62. This might lead to structural changes affecting the ability of complement component 3b (C3b) binding and reducing the activation of the alternative pathway C3-convertase (C3bBb). This subsequently causes excessive activation of the complement system to induce immunologic disorder and the gender bias may be due to different pathway of CFH in AU. The exact mechanism is still unclear and further studies will be needed to investigate the functional interaction of CFH and AU and the difference in gender-susceptibility in AU.

SNP rs4505848 is in the region encompassing *KIAA1109/ Tenr/IL2/IL21* which is located on the chromosome 4q27. This 480 kB block of linkage disequilibrium includes *IL2* and *IL21*, which are both functional candidate loci for autoimmune diseases. As mentioned previously, levels of IL2, IL21, and their receptors were significantly increased during uveitis in patients and animal models and they may participate in the regulation of T-cell responses [11-13]. Current evidences indicated that both T helper 1 (Th1) and T helper 17 (Th17) effector cells can independently induce uveitis in animal models [44,45]. Functional studies have revealed that Th17 cells contribute to uveitis through expanded IL-2, meanwhile IL-21 was highly expressed and promoted the differentiation of Th17 cells both in vitro and in vivo studies [13,15]. IL27 is a recently discovered cytokine belonging to the IL6/IL12 families, which consists of EBI3 and p28 subunits. Studies have shown that IL27 was constitutively expressed in retinal ganglion and photoreceptor cells in EAU and IL27 could promote Th1 but inhibit Th17 cells differentiation, which lead to a mutual antagonism between the two pathways [15]. Taken together, these studies indicate that IL2, IL21, and

IL27 play important roles in the development of uveitis, predicting an association of these two SNPs (rs4505848 and rs4788084) with AU. However in our study, no such association was found even stratified by gender. This might be due to ethnic differences in susceptibility to AU; the small number of subjects; the wide variety of AU syndromes; the complex regulation mechanism in inflammatory activity, as well as the candidate polymorphisms chosen among these plausible genes. Therefore, further investigations are needed in more subtypes of uveitis and other ethnic groups. In addition, considering HLA-B27 is the strongest association

rs800292 ( <i>CFH</i> 184G/A) Genotype AA AG GG Allele A rs4505848 ( <i>KIAA1109</i> G/A)	3 (7.1) 20 (47.6) 19 (45.2)		CONC-II) (IN 1110)	p-value1	p-value,
Genotype AG GG Allele A G rs4505848 (KIAA1109 G/A)	3 (7.1) 20 (47.6) 19 (45.2)				
AA AG GG Allele A G rs4505848 ( <i>KIAA1109</i> G/A)	3 (7.1) 20 (47.6) 19 (45.2)				
AG GG Allele A G rs4505848 ( <i>KIAA1109</i> G/A)	20 (47.6) 19 (45.2)	7 (12.5)	48(15.6)	0.30§	0.31§
GG Allele A G rs4505848 ( <i>KIAA1109</i> G/A)	19 (45.2)	22 (39.3)	145 (47.1)		
Allele A G rs4505848 ( <i>KIAA1109</i> G/A)	n.	27 (48.2)	115 (37.3)		
A G Is4505848 ( <i>KIAA110</i> 9 G/A)		~			
G 154505848 ( <b>K</b> IAA1109 G/A)	26 (31.0)	36 (32.1)	241 (39.1)	0.15§	0.16
rs4505848 (KIAA1109 G/A)	58 (69.0)	76 (67.9)	375 (60.9)	•	2
	~		× ·		
Genotype					
GG	4 (9.5)	8 (14.3)	50 (16.2)	0.24§	0.91
AG	27 (64.3)	30 (53.6)	157(51.0)		
AA	11 (26.2)	18 (32.1)	101 (32.8)		
Allele					
G	35 (41.6)	46(41.1)	257 (41.7)	§66.0	0.90
А	49 (58.4)	66 (58.9)	359 (58.3)		
rs4788084 (IL27 T/C)					
Genotype					
TT	4 (9.5)	5(8.9)	21 (6.8)	0.75§	0.80
CT	18 (42.9)	21 (37.5)	126 (40.9)		
CC	20 (47.6)	30 (53.6)	161 (52.3)		
Allele					
Т	26 (31.0)	31 (27.7)	168 (27.3)	0.48§	0.93
C	58 (69.0)	81 (72.3)	448 (72.7)		

5, ŵ 2 4 5, 2 ۶۲ ż b ン 2 Data are the numb versus controls.

 $\mathbf{ts}$ 

## Molecular Vision 2011; 17:2655-2664 < http://www.molvis.org/molvis/v17/a287>

© 2011 Molecular Vision

with AU and the interaction with these immune-associated genes, further analyses were performed in these AU patients stratified by HLA-B27 status. We found all the 3 SNPs were not associated with AU regardless of HLA-B27 positivity. The exact reason is unclear but it may account for the weaker association of these SNPs with AU, or the influence of these genes on AU independent on HLA-B27.

In conclusion, we found the association of *CFH* 184G with AU in Chinese female patients. The genetic association between the complement system and AU was identified with gender susceptibility. Further studies to replicate the candidate SNPs in others ethnic groups and determine the biologic roles of these polymorphisms in immune mechanisms involved in uveitis are worthwhile.

#### REFERENCES

- Gritz DC, Wong IG. Incidence and prevalence of uveitis in Northern California; the Northern California Epidemiology of Uveitis Study. Ophthalmology 2004; 111:491-500. [PMID: 15019324]
- Chang JH, Wakefield D. Uveitis: a global perspective. Ocul Immunol Inflamm 2002; 10:263-79. [PMID: 12854035]
- Martin TM, Kurz DE, Rosenbaum JT. Genetics of uveitis. Ophthalmol Clin North Am 2003; 16:555-65. [PMID: 14740996]
- Martin TM, Rosenbaum JT. Genetics in uveitis. Int Ophthalmol Clin 2005; 45:15-30. [PMID: 15791155]
- Brewerton DA, Caffrey M, Nicholls A, Walters D, James DC. Acute anterior uveitis and HL-A 27. Lancet 1973; 302:994-6. [PMID: 4127279]
- Chang JH, McCluskey PJ, Wakefield D. Acute anterior uveitis and HLA-B27. Surv Ophthalmol 2005; 50:364-88. [PMID: 15967191]
- Lan C, Tam PO, Chiang SW, Chan CK, Luk FO, Lee GK, Ngai JW, Law JS, Lam DS, Pang CP, Lai TY. Manganese superoxide dismutase and chemokine genes polymorphisms in chinese patients with anterior uveitis. Invest Ophthalmol Vis Sci 2009; 50:5596-600. [PMID: 19628738]
- Kuo NW, Lympany PA, Menezo V, Lagan AL, John S, Yeo TK, Liyanage S, du Bois RM, Welsh KI, Lightman S. TNF-857T, a genetic risk marker for acute anterior uveitis. Invest Ophthalmol Vis Sci 2005; 46:1565-71. [PMID: 15851552]
- Yeo TK, Ahad MA, Kuo NW, Spagnolo P, Menezo V, Lympany P, Lightman S. Chemokine gene polymorphisms in idiopathic anterior uveitis. Cytokine 2006; 35:29-35. [PMID: 16950632]
- Yang PZ. Effects of interleukin-2 on experimental uveitis and pinealitis. Zhonghua Yan Ke Za Zhi 1993; 29:175-9. [PMID: 8223049]
- Lacomba MS, Martin CM, Chamond RR, Galera JM, Omar M, Estevez EC. Aqueous and serum interferon gamma, interleukin (IL) 2, IL-4, and IL-10 in patients with uveitis. Arch Ophthalmol 2000; 118:768-72. [PMID: 10865312]
- Arocker-Mettinger E, Asenbauer T, Ulbrich S, Grabner G. Serum interleukin 2-receptor levels in uveitis. Curr Eye Res 1990; 9:25-9. [PMID: 2384010]

- Liu L, Xu Y, Wang J, Li H. Upregulated IL-21 and IL-21 receptor expression is involved in experimental autoimmune uveitis (EAU). Mol Vis 2009; 15:2938-44. [PMID: 20057909]
- Lee YS, Amadi-Obi A, Yu CR, Egwuagu CE. Retinal cells suppress intraocular inflammation (uveitis) through production of interleukin-27 and interleukin-10. Immunology 2011; 132:492-502. [PMID: 21294722]
- Amadi-Obi A, Yu CR, Liu X, Mahdi RM, Clarke GL, Nussenblatt RB, Gery I, Lee YS, Egwuagu CE. TH17 cells contribute to uveitis and scleritis and are expanded by IL-2 and inhibited by IL-27/STAT1. Nat Med 2007; 13:711-8. [PMID: 17496900]
- Lee YS, Amadi-Obi A, Yu CR, Egwuagu CE. Retinal cells suppress intraocular inflammation (uveitis) through production of interleukin-27 and interleukin-10. Immunology 2011; 132:492-502. [PMID: 21294722]
- Zhou H, Jiang Z, Yang P, Hou S, Li F, Shu Q, Chen Y, Chen F. Polymorphisms of IL23R and Fuchs' syndrome in a Chinese Han population. Mol Vis 2010; 16:2585-9. [PMID: 21151597]
- Cordeiro CA, Moreira PR, Costa GC, Dutra WO, Campos WR, Orefice F, Teixeira AL. Interleukin-1 gene polymorphisms and toxoplasmic retinochoroiditis. Mol Vis 2008; 14:1845-9. [PMID: 18941541]
- Wallace GR, Kondeatis E, Vaughan RW, Verity DH, Chen Y, Fortune F. IL-10 genotype analysis in patients with Behcet's disease. Hum Immunol 2007; 68:122-7. [PMID: 17321902]
- Garner CP, Murray JA, Ding YC, Tien Z, van Heel DA, Neuhausen SL. Replication of celiac disease UK genomewide association study results in a US population. Hum Mol Genet 2009; 18:4219-25. [PMID: 19648293]
- 21. Teixeira VH, Pierlot C, Migliorini P, Balsa A, Westhovens R, Barrera P, Alves H, Vaz C, Fernandes M, Pascual-Salcedo D, Bombardieri S, Dequeker J, Radstake TR, Van Riel P, van de Putte L, Lopes-Vaz A, Bardin T, Prum B, Cornélis F, Petit-Teixeira E, European Consortium on Rheumatoid Arthritis Families. Testing for the association of the KIAA1109/Tenr/ IL2/IL21 gene region with rheumatoid arthritis in a European family-based study. Arthritis Res Ther 2009; 11:R45. [PMID: 19302705]
- 22. Todd JA, Walker NM, Cooper JD, Smyth DJ, Downes K, Plagnol V, Bailey R, Nejentsev S, Field SF, Payne F, Lowe CE, Szeszko JS, Hafler JP, Zeitels L, Yang JH, Vella A, Nutland S, Stevens HE, Schuilenburg H, Coleman G, Maisuria M, Meadows W, Smink LJ, Healy B, Burren OS, Lam AA, Ovington NR, Allen J, Adlem E, Leung HT, Wallace C, Howson JM, Guja C, Ionescu-Tîrgovişte C. Genetics of Type 1 Diabetes in Finland, Simmonds MJ, Heward JM, Gough SC, Wellcome Trust Case Control Consortium, Dunger DB, Wicker LS, Clayton DG. Robust associations of four new chromosome regions from genomewide analyses of type 1 diabetes. Nat Genet 2007; 39:857-64. [PMID: 17554260]
- 23. Remmers EF, Cosan F, Kirino Y, Ombrello MJ, Abaci N, Satorius C, Le JM, Yang B, Korman BD, Cakiris A, Aglar O, Emrence Z, Azakli H, Ustek D, Tugal-Tutkun I, Akman-Demir G, Chen W, Amos CI, Dizon MB, Kose AA, Azizlerli G, Erer B, Brand OJ, Kaklamani VG, Kaklamanis P, Ben-Chetrit E, Stanford M, Fortune F, Ghabra M, Ollier WE, Cho

YH, Bang D, O'Shea J, Wallace GR, Gadina M, Kastner DL, Gül A. Genome-wide association study identifies variants in the MHC class I, IL10, and IL23R–IL12RB2 regions associated with Behçet's disease. Nat Genet 2010; 42:698-702. [PMID: 20622878]

- Mizuki N, Meguro A, Ota M, Ohno S, Shiota T, Kawagoe T, Ito N, Kera J, Okada E, Yatsu K, Song YW, Lee EB, Kitaichi N, Namba K, Horie Y, Takeno M, Sugita S, Mochizuki M, Bahram S, Ishigatsubo Y, Inoko H. Genome-wide association studies identify IL23R–IL12RB2 and IL10 as Behçet's disease susceptibility loci. Nat Genet 2010; 42:703-6. [PMID: 20622879]
- 25. Zhernakova A, Alizadeh BZ, Bevova M, van Leeuwen MA, Coenen MJ, Franke B, Franke L, Posthumus MD, van Heel DA, van der Steege G, Radstake TR, Barrera P, Roep BO, Koeleman BP, Wijmenga C. Novel association in chromosome 4q27 region with rheumatoid arthritis and confirmation of type 1 diabetes point to a general risk locus for autoimmune diseases. Am J Hum Genet 2007; 81:1284-8. [PMID: 17999365]
- Cooper JD, Smyth DJ, Smiles AM, Plagnol V, Walker NM, Allen JE, Downes K, Barrett JC, Healy BC, Mychaleckyj JC, Warram JH, Todd JA. Meta-analysis of genome-wide association study data identifies additional type 1 diabetes risk loci. Nat Genet 2008; 40:1399-401. [PMID: 18978792]
- Barrett JC, Clayton DG, Concannon P, Akolkar B, Cooper JD, Erlich HA, Julier C, Morahan G, Nerup J, Nierras C, Plagnol V, Pociot F, Schuilenburg H, Smyth DJ, Stevens H, Todd JA, Walker NM, Rich SS. Type 1 Diabetes Genetics Consortium. Genome-wide association study and meta-analysis find that over 40 loci affect risk of type 1 diabetes. Nat Genet 2009; 41:703-7. [PMID: 19430480]
- Jha P, Sohn JH, Xu Q, Nishihori H, Wang Y, Nishihori S, Manickam B, Kaplan HJ, Bora PS, Bora NS. The complement system plays a critical role in the development of experimental autoimmune anterior uveitis. Invest Ophthalmol Vis Sci 2006; 47:1030-8. [PMID: 16505038]
- Read RW, Szalai AJ, Vogt SD, McGwin G, Barnum SR. Genetic deficiency of C3 as well as CNS-targeted expression of the complement inhibitor sCrry ameliorates experimental autoimmune uveoretinitis. Exp Eye Res 2006; 82:389-94. [PMID: 16143328]
- Liszewski MK, Farries TC, Lublin DM, Rooney IA, Atkinson JP. Control of the complement system. Adv Immunol 1996; 61:201-83. [PMID: 8834497]
- Jha P, Sohn JH, Xu Q, Wang Y, Kaplan HJ, Bora PS, Bora NS. Suppression of complement regulatory proteins (CRPs) exacerbates experimental autoimmune anterior uveitis (EAAU). J Immunol 2006; 176:7221-31. [PMID: 16751365]
- Fakhouri F, de Jorge EG, Brune F, Azam P, Cook HT, Pickering MC. Treatment with human complement factor H rapidly reverses renal complement deposition in factor H-deficient mice. Kidney Int 2010; 78:279-86. [PMID: 20445496]
- Bao L, Haas M, Quigg RJ. Complement factor H deficiency accelerates development of lupus nephritis. J Am Soc Nephrol 2011; 22:285-95. [PMID: 21148254]
- Skerka C, Zipfel PF. Complement factor H related proteins in immune diseases. Vaccine 2008; 26:I9-14. [PMID: 19388158]

- 35. Caprioli J, Castelletti F, Bucchioni S, Bettinaglio P, Bresin E, Pianetti G, Gamba S, Brioschi S, Daina E, Remuzzi G, Noris M, International Registry of Recurrent and Familial HUS/ TTP. Complement factor H mutations and gene polymorphisms in haemolytic uraemic syndrome: the C-257T, the A2089G and the G2881T polymorphisms are strongly associated with the disease. Hum Mol Genet 2003; 12:3385-95. [PMID: 14583443]
- 36. Neary JJ, Conlon PJ, Croke D, Dorman A, Keogan M, Zhang FY, Vance JM, Pericak-Vance MA, Scott WK, Winn MP. Linkage of a gene causing familial membranoproliferative glomerulonephritis type III to chromosome 1. J Am Soc Nephrol 2002; 13:2052-7. [PMID: 12138136]
- 37. Dieguez-Gonzalez R, Akar S, Calaza M, Gonzalez-Alvaro I, Fernandez-Gutierrez B, Lamas JR, de la Serna AR, Caliz R, Blanco FJ, Pascual-Salcedo D, Velloso ML, Perez-Pampin E, Pablos JL, Navarro F, Narvaez J, Lopez-Longo FJ, Herrero-Beaumont G, Gomez-Reino JJ, Gonzalez A. Lack of association with rheumatoid arthritis of selected polymorphisms in 4 candidate genes: CFH, CD209, eotaxin-3, and MHC2TA. J Rheumatol 2009; 36:1590-5. [PMID: 19567623]
- Jabs DA, Nussenblatt RB, Rosenbaum JT. Standardization of uveitis nomenclature for reporting clinical data. Results of the First International Workshop. Am J Ophthalmol 2005; 140:509-16. [PMID: 16196117]
- Konno Y, Numaga J, Tsuchiya N, Ogawa A, Islam SM, Mochizuki M, Mitsui H, Oda H, Maeda H. HLA-B27 subtypes and HLA class II alleles in Japanese patients with anterior uveitis. Invest Ophthalmol Vis Sci 1999; 40:1838-44. [PMID: 10393058]
- Atkinson JP, Oglesby TJ, White D, Adams EA, Liszewski MK. Separation of self from non-self in the complement system: a role for membrane cofactor protein and decay accelerating factor. Clin Exp Immunol 1991; 86:27-30. [PMID: 1718640]
- Bando Y, Tanouchi Y, Fukuyado K, Matsuda S, Mimura Y. The dynamics of leucocytes and complements in endotoxin induced uveitis. Nippon Ganka Gakkai Zasshi 1989; 93:369-74. [PMID: 2788980]
- Kim YH, He S, Kase S, Kitamura M, Ryan SJ, Hinton DR. Regulated secretion of complement factor H by RPE and its role in RPE migration. Graefes Arch Clin Exp Ophthalmol 2009; 247:651-9. [PMID: 19214553]
- 43. Ng TK, Chen LJ, Liu DT, Tam PO, Chan WM, Liu K, Hu YJ, Chong KK, Lau CS, Chiang SW, Lam DS, Pang CP. Multiple gene polymorphisms in the complement factor h gene are associated with exudative age-related macular degeneration in Chinese. Invest Ophthalmol Vis Sci 2008; 49:3312-7. [PMID: 18421087]
- 44. Luger D, Silver PB, Tang J, Cua D, Chen Z, Iwakura Y, Bowman EP, Sgambellone NM, Chan CC, Caspi RR. Either a Th17 or a Th1 effector response can drive autoimmunity: conditions of disease induction affect dominant effector category. J Exp Med 2008; 205:799-810. [PMID: 18391061]
- 45. Mizuki N, Meguro A, Ota M, Ohno S, Shiota T, Kawagoe T, Ito N, Kera J, Okada E, Yatsu K, Song YW, Lee EB, Kitaichi N, Namba K, Horie Y, Takeno M, Sugita S, Mochizuki M, Bahram S, Ishigatsubo Y, Inoko H. Genome-wide association studies identify IL23R–IL12RB2 and IL10 as Behçet's

Molecular Vision 2011; 17:2655-2664 < http://www.molvis.org/molvis/v17/a287>

© 2011 Molecular Vision

disease susceptibility loci. Nat Genet 2010; 42:703-6. [PMID: 20622879]

Articles are provided courtesy of Emory University and the Zhongshan Ophthalmic Center, Sun Yat-sen University, P.R. China. The print version of this article was created on 10 October 2011. This reflects all typographical corrections and errata to the article through that date. Details of any changes may be found in the online version of the article.