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# Seasonality and zoonotic foodborne pathogens in Canada: relationships between climate and *Campylobacter*, *E. coli* and *Salmonella* in meat products

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## Abstract

Infections due to *Campylobacter*, *Escherichia coli* and *Salmonella* pose a significant health burden in Canada, resulting in major costs to the health care system and economic impacts due to lost productivity resulting from illness. Recent literature suggests that climate may play a role in the prevalence of these pathogens along the food chain. This study used integrated surveillance data to examine associations between weather variables, serving as a proxy for climate, in agricultural areas and *Campylobacter*, generic *E. coli* and *Salmonella* contamination on samples of beef, poultry and swine meat products in Canada. Various temperature metrics (average, maximum and variability) were correlated with *Campylobacter* prevalence along the food chain. The prevalence of *E. coli* and *Salmonella* was correlated with both precipitation and temperatures metrics; however, analysis for *E. coli* was limited to beef and swine meats at retail settings, because prevalence in other combinations approached 100%, which obviated further analysis. *Campylobacter* contamination in poultry and swine at abattoir and retail settings demonstrated a seasonal trend, with increased prevalence generally from June or July through November, compared to the baseline month of December. Based on these analyses, *Campylobacter* is the most likely foodborne bacteria studied whose occurrence in meat products is affected by climatic changes in Canada. An exploratory analysis of data at the provincial scale, using Ontario as an example, revealed similar directional relationships between climate and bacterial prevalence.

## Introduction

Foodborne illness remains a global public health concern [1, 2]. In Canada, foodborne pathogens cause an estimated 4 million cases of gastrointestinal illness annually from known and unspecified agents and serotypes of *Campylobacter*, *Escherichia coli* and *Salmonella* are commonly implicated [3]. Annual cases in Canada were estimated as 447, 122 and 270 per 100 000 people for *Campylobacter*, *E. coli* and *Salmonella*, respectively [3]. These cases represent a significant cost to the Canadian economy in terms of lost production and medical treatment estimated at approximately \$325 million per annum [4–6].

The incidence of enteric infections in the Canadian population increases with air temperature and may change in response to global climate change [7]. Evidence for the link between climate and microbial foodborne disease also comes from the observed latitudinal gradients in the amount of foodborne and diarrhoeal illnesses, changing disease patterns during El Niño warming and links between foodborne illnesses and severe weather events [8, 9]. A statistical association between foodborne contamination or disease and short-term temperature changes suggests that foodborne disease will be affected by long-term climate changes such as increased average air temperatures and precipitation frequency or intensity [10, 11]. This relationship is complex, as there are many direct and indirect mechanisms whereby climate can influence foodborne illness.

Directly, the survival rates of most enteric pathogens in the environment are, within limits, positively correlated with ambient temperature [11]. Laboratory studies have shown the survival capacity of bacteria such as *E. coli* can be related to temperature, moisture and other abiotic and biotic variables such as pH and interactions with the microbial community [12]. Seasonal trends have also been identified for illnesses attributed to *Campylobacter* and *Salmonella* which have been shown to peak during summer months in temperate countries around the world [11, 13].

Indirectly, climate can influence transmission of foodborne illness by altering food consumption behaviours, susceptibility of livestock to pathogens and transmission from vectors due to range expansion, increased activity and reproduction rates [14]. Livestock stressed by

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temperature may be more susceptible to colonisation by enteric pathogens such as *E. coli* or *Salmonella*, or more likely to shed greater amounts of these bacteria [15, 16]. This can then lead to a greater risk of meat contamination from hygiene failures during processing [14]. Higher temperatures might also affect animal feeding strategies that can have a profound effect on the prevalence and shedding rate of pathogens infectious to humans by altering ecological conditions in the animal gut [17]. At higher temperatures, cattle may graze outside more frequently; consumption of grass during grazing also contributes to pathogen survival and higher shedding rates [17]. Direct or indirect effects of climate might also affect the super-shedding phenomenon (i.e. some cattle may harbour and shed bacteria at higher levels than others) [18], which strongly influences dissemination of *E. coli* O157 into the environment and ultimately to humans. Heat stress can alter the intestinal microbiota and has been associated with *Salmonella* in poultry [19] and antibiotic resistant *E. coli* in the faeces of swine [20]. The need to understand climatic drivers for current foodborne illness patterns is a key public health challenge [14]. Short-term weather variables (data collected over months or years) are considered in this paper as suitable indicators for long-term climate variables (data collected over decades) because they describe the same abiotic factors. To better understand the potential impact of climate change on enteric infections in Canada, we investigated the relationship between weather variables and sampling data of three different bacteria in Canada: *Campylobacter*, generic *E. coli* and *Salmonella*.

## Methods

Monthly bacterial presence/absence data were acquired from the Canadian Integrated Program for Antimicrobial Resistance Surveillance (CIPARS). CIPARS provides a unified approach to monitor trends in antimicrobial resistance through sampling of animals and meats at farms, abattoirs and retail locations across Canada. For our analysis, the historical database of presence/absence of bacteria was used irrespective of antimicrobial resistance profiles (in other words, antimicrobial resistance was not considered in this study). The data consisted of beef, chicken and swine samples from farm, abattoir and retail settings (Table 1). Further details about the sampling process and program design can be found in Table 1 and the CIPARS annual report [21]. All available data from 2002 to 2012 were analysed to allow for sufficient sample size for analysis, particularly for datasets with low contamination prevalence.

Data from all weather stations in Canada were downloaded from the Environment Canada website [22]. These data included average monthly temperature, maximum monthly temperature, precipitation and the station name and location coordinates for approximately 1700 weather stations across Canada. Stations for which data were available varied by month; data were available for an average of approximately 1400 stations each month (range 1300–1700). These data were placed into a geographical information system (GIS), ESRI ArcGIS 9.3 and plotted using the provided coordinates for each station, with the observed values for each month relationally joined using the station identification field. Data were then interpolated using an inverse distance weighting in ArcGIS using the Spatial Analyst function. This function transformed the point data into a continuous raster surface that used the eight nearest stations to interpolate values for the surface [23]. The transformation of point data to raster data is illustrated in Figure 1. This process was conducted for a

**Table 1.** Sample counts tested for *Campylobacter*, generic *E. coli* and *Salmonella* in retail, abattoir and on-farm settings from 2002 through 2012

	Retail			Abattoir			On-farm					
	Sample description	<i>Campylobacter</i>	<i>Salmonella</i>	<i>E. coli</i>	Sample description	<i>Campylobacter</i>	<i>Salmonella</i>	<i>E. coli</i>	Sample description	<i>Campylobacter</i>	<i>Salmonella</i>	<i>E. coli</i>
Beef	Raw ground beef from retail stores	118 <sup>a</sup>	131 <sup>a</sup>	6825	Cecal content of slaughtered animals	1005 <sup>b</sup>	207 <sup>a</sup>	1567 <sup>c</sup>	-	-	-	-
Chicken	Raw legs or wings with skin on from retail stores	8338	8361	4728 <sup>c</sup>	Cecal content of slaughtered animals	2099	8416	2543 <sup>c</sup>	-	-	-	-
Swine	Raw chops from retail stores	398	6862	8590	Cecal content of slaughtered animals	382	5432	1603 <sup>c</sup>	Pooled fecal samples from pens	-	3028	3028 <sup>c</sup>

<sup>a</sup>Excluded from analysis due to low prevalence rate (<3%).

<sup>b</sup>Excluded from analysis due to sporadic nature of sampling (8 year period across multiple provinces).

<sup>c</sup>Excluded from further analysis due to high prevalence rate (>98%).

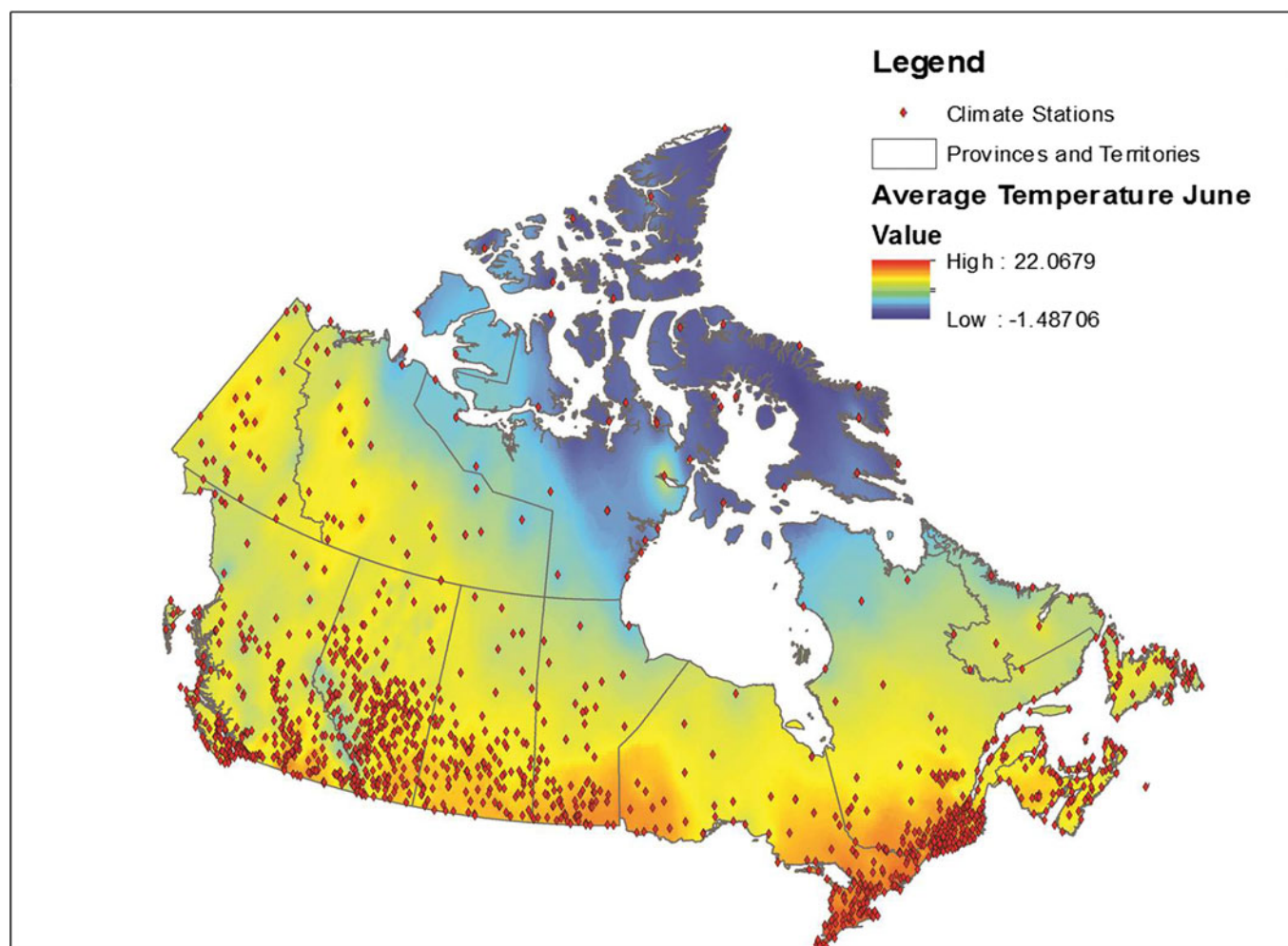


Fig. 1. Point weather station data conversion to a raster surface.

Table 2. Area (km<sup>2</sup>) of livestock zones considered in the weather analysis as compared to total provincial area

Province	Total area	Beef cattle area (%)	Chicken area (%)	Swine area (%)
Quebec	1 542 056	59 000 (3.8)	1700 (0.1)	11 700 (0.8)
Ontario	1 076 395	84 300 (7.8)	26 500 (2.5)	28 000 (2.6)
British Columbia	944 735	213 155 (22.6)	2200 (0.2)	700 (0.1)
Alberta	661 848	270 000 (40.8)	65 000 (9.8)	109 000 (16.5)
Saskatchewan	651 036	143 500 (22)	3700 (0.6)	3300 (0.5)
Manitoba	647 797	67 000 (10.3)	3800 (0.6)	17 000 (2.6)
Nova Scotia	55 284	11 000 (19.9)	650 (1.2)	(0)

Only provinces which were included in the analysis (see text) are shown.

10 year period for each month and variable, for a total 360 surfaces.

Since CIPARS data were aggregated at the provincial level and climate varies substantially within Canadian provinces, weather data were analysed only for areas where livestock operations are located in each province. Some provinces were excluded from the analysis altogether due to very low and highly dispersed livestock production. Otherwise, this would have resulted in a wide range of climate regions to consider and contribute significant

uncertainty to the model with little benefit to the overall analysis due to the relatively low contribution of these provinces to the livestock production. The delineation of livestock operation areas was determined using the Census of Agriculture data at the Census Consolidated Subdivision level [24]. Separate boundaries were created in ArcGIS for chickens, beef cows and swine, and weather data were extracted for those areas using the summarise zone feature in ArcGIS. Table 2 describes the total area of each livestock zone by province, visualised in Figure 2. This

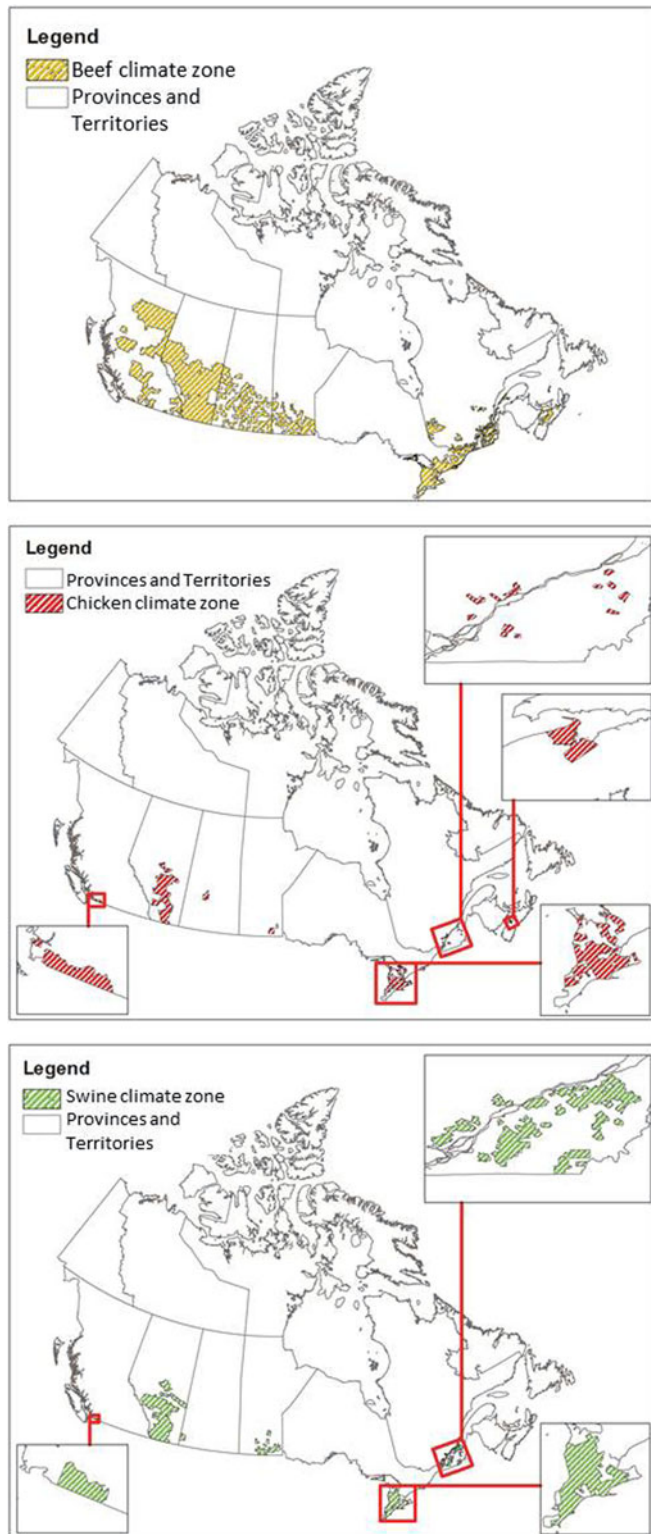


Fig. 2. Agricultural zones considered for the weather data analysis per livestock type.

effectively removed weather data from the analysis associated with areas with no livestock production in Canada. The weather and pathogen data were then combined in a single dataset. This approach reduced the variability of weather data but still provided coverage of 95.6%, 94.5% and 97.4% of cattle, chicken and swine production in Canada (calculated as the total number of livestock

in the climate zones considered divided by the total number of livestock across all provinces included in the analysis).

Regression analyses using SAS V9.2 were performed to investigate relationships between bacteria prevalence and weather variables. Over the study period, CIPARS tested for *Campylobacter*, generic *E. coli* and *Salmonella*, but not all samples were tested for all pathogens and testing varied by location along the agri-food chain and by year. Prior to running any analyses, CIPARS data were assessed for suitability. Several categories of CIPARS data (9 out of 20) were eliminated as candidates for analysis (Table 1). Generic *E. coli*, for example, were eliminated from all abattoir analyses due to the fact that nearly all samples were positive. This makes the identification of influential weather variables impossible. In some other cases, insufficient test results were available due to limited sample size or low recovery rates, resulting in too few positive results for which to make statistical associations with weather variables. Even fewer categories were included in the Ontario analysis due to more limited data.

Year and province were forced into all regression models as fixed effects to control for overall yearly and provincial effects. Weather variables included the following: average monthly temperature, monthly temperature variation from the 10 year average, average maximum temperature, total monthly precipitation and monthly precipitation variation from the 10 year average. Two separate analyses were conducted: (1) overall estimates of the impact of weather variables on bacteria prevalence were produced by grouping all data on a particular variable and bacteria/commodity/agri-food chain location and (2) a seasonal analysis was conducted to explore the impact of month on the prevalence of bacteria. Weather variables were selected using both backwards and stepwise variable selection approaches. Among weather variables, only those significant at the 5% level were retained in the final model. To explore provincial variation, a separate sub-group analysis involving Ontario chicken data only was conducted.

## Results

The significant findings from the model are shown in Table 3 for national level results. All of the significant variables associated with *Campylobacter* were related to temperature. Increased average temperatures increased odds of retail contamination with *Campylobacter* for both chicken and swine commodities; an average temperature increase of 9 °C for chicken and 10 °C for swine was associated with 10% higher odds of a positive *Campylobacter* sample. In abattoir settings, increases in the average maximum monthly temperatures of 3.6 °C and 3.5 °C were related to a 10% increase in odds of a positive *Campylobacter* sample for chicken and swine, respectively. There was also a negative association between *Campylobacter* prevalence in abattoir settings and variation from the 10 year average temperature for that month.

The prevalence of *Salmonella* was associated with precipitation and average maximum temperature in poultry products collected from the abattoir and maximum temperature and variable precipitation from on-farm swine samples in Canada. An increase of 23 and 40 mm above average precipitation during a 1 month period corresponded with a 10% increase in the odds of *Salmonella* in farm swine and abattoir swine samples, respectively. There was also a positive relationship between *Salmonella* prevalence in chicken sampled at the abattoir and total monthly precipitation, with an increase of 55 mm of the latter required to increase odds of *Salmonella* detection by 10%. A change in

**Table 3.** Statistical model coefficients for the relationship between weather variables and prevalence of bacteria along the agri-food chain in Canada from 2002 through 2012

Outcome	Independent variables				
	Temperature (°C) <sup>a</sup>	Variable temperature (°C) <sup>a</sup>	Maximum temperature (°C) <sup>a</sup>	Precipitation (mm) <sup>b</sup>	Variable precipitation (mm) <sup>b</sup>
<b>Poultry</b>					
<i>Salmonella</i> Retail	NS	NS	NS	NS	NS
<i>Salmonella</i> Abattoir	NS	NS	−0.00884 (11)	0.00174 (55)	NS
<i>Campylobacter</i> Retail	0.0105 (9)	NS	NS	NS	NS
<i>Campylobacter</i> Abattoir	NS	−0.0817 (1.4)	0.0268 (3.6)	NS	NS
<b>Beef</b>					
<i>E. coli</i> Retail	NS	NS	NS	0.00312 (31)	NS
<b>Swine</b>					
<i>Salmonella</i> Retail	NS	NS	NS	NS	NS
<i>Salmonella</i> Abattoir	NS	NS	NS	NS	0.00243 (40)
<i>Salmonella</i> On-farm	NS	NS	−0.0146 (7)	NS	0.00424 (23)
<i>Campylobacter</i> Retail	0.00817 (10)	NS	NS	NS	NS
<i>Campylobacter</i> Abattoir	NS	−0.1365 (0.7)	0.0299 (3.5)	NS	NS
<i>E. coli</i> Retail	0.0173 (6)	0.026 (3.7)	NS	NS	NS

NS, No significant relationship at significance level of 5%.

Coefficients are shown, followed by the required absolute change in the weather variable in parentheses required to increase (if positive coefficient) or decrease (if negative coefficient) odds of a positive sample by 10% (keeping all other factors constant). Only categories with sufficient data for analysis are shown (see text).

<sup>a</sup>Increase in C temperature required to increase (if + coefficient) or decrease (if − coefficient) odds of a positive sample by 10% (keeping all other factors constant).

<sup>b</sup>Increase in mm of precipitation required to increase (if + coefficient) or decrease (if − coefficient) odds of a positive sample by 10% (keeping all other factors constant).

**Table 4.** Statistical model coefficients for the relationship between weather variables and prevalence of bacteria in chicken along the agri-food chain in Ontario from 2002 through 2012

Outcome	Independent variables				
	Temperature (°C) <sup>a</sup>	Variable temperature (°C) <sup>a</sup>	Maximum temperature (°C) <sup>a</sup>	Precipitation (mm) <sup>b</sup>	Variable precipitation (mm) <sup>b</sup>
<i>Salmonella</i> Retail	NS	NS	NS	−0.0182 (6)	NS
<i>Salmonella</i> Abattoir	NS	NS	−0.00958 (11)	NS	NS
<i>Campylobacter</i> Retail	0.0544 (1.8)	NS	−0.0397 (2.5)	NS	NS
<i>Campylobacter</i> Abattoir	0.0406 (2.4)	−0.1795 (0.6)	NS	NS	NS

NS, No significant relationship at significance level of 5%.

Coefficients are shown, followed by the required absolute change in the weather variable in parentheses required to increase (if positive coefficient) or decrease (if negative coefficient) odds of a positive sample by 10% (keeping all other factors constant). Only categories with sufficient data for analysis are shown (see text).

<sup>a</sup>Increase in C temperature required to increase (if + coefficient) or decrease (if − coefficient) odds of a positive sample by 10% (keeping all other factors constant).

<sup>b</sup>Increase in mm of precipitation required to increase (if + coefficient) or decrease (if − coefficient) odds of a positive sample by 10% (keeping all other factors constant).

maximum temperature of 11 °C was required to result in a 10% increase in odds of *Salmonella* in chicken.

In retail settings, the odds of generic *E. coli* detection in Canada increased by 10% with an increase of 31 mm total precipitation for beef and a 6 °C increase in average temperature or 3.7 °C increase in monthly temperature variation for swine.

Table 4 provides the results for a sub-group analysis of Ontario chicken. The analysis of *Campylobacter* in Ontario chicken revealed similar directional relationships to those observed at the national level; however, an increase in average monthly temperature of only 2.4 °C or 1.8 °C was required to produce the same 10% increase in the odds of a positive *Campylobacter*

sample at the abattoir and retail stages, respectively. Increases in the variation from the 10 year average temperature and the average maximum temperature decreased the odds of *Campylobacter* detection at the abattoir and retail in Ontario, respectively.

In Ontario, the odds of *Salmonella* detection in retail samples decreased by 10% as precipitation increased by 6 mm (Table 4), which is the opposite relationship observed in the national-level analysis. *Salmonella* prevalence in chickens at the abattoir and pigs on farm was also negatively related to average maximum temperature. The change in maximum temperature to result in a 10% increase in odds of *Salmonella* in chicken (11 °C) in Ontario was identical to that observed at the national scale.

**Table 5.** Statistical model coefficients by month for the relationship between weather variables and prevalence of bacteria along the agri-food chain in Canada from 2002 through 2012

Outcome	Independent variables											
	Jan	Feb	Mar	Apr	May	Jun	Jul	Aug	Sep	Oct	Nov	Dec <sup>a</sup>
<b>Poultry</b>												
<i>Salmonella</i> Retail	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	0
<i>Salmonella</i> Abattoir	0.0745	-0.051	-0.166	-0.091	-0.206	-0.2645	-0.1347	-0.0311	0.0415	0.2461	0.3397	0
<i>Campylobacter</i> Retail	-0.0141	-0.2493	-0.1155	-0.2572	-0.2687	-0.0576	0.2249	0.2399	0.3471	0.1269	0.0607	0
<i>Campylobacter</i> Abattoir	-0.4373	-0.4755	-0.8203	-0.1125	-0.5445	0.1925	0.3985	0.1974	0.7626	0.6572	0.286	0
<b>Beef</b>												
<i>E. coli</i> Retail	-0.0413	-0.0924	-0.0485	-0.2403	0.1632	0.1637	-0.0761	0.0206	0.1917	-0.0313	0.1383	0
<b>Swine</b>												
<i>Salmonella</i> Retail	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	0
<i>Salmonella</i> Abattoir	0.1772	-0.0531	-0.1394	-0.0628	-0.0548	-0.2934	0.1179	-0.0474	0.0298	0.193	0.1206	0
<i>Salmonella</i> On-farm	-0.0224	0.5105	0.367	-0.518	0.328	-0.7546	0.1873	-0.0111	-0.3833	0.2313	0.2235	0
<i>Campylobacter</i> Retail	0.0866	-0.1979	-0.2447	-0.3805	-0.3513	-0.3255	0.1936	0.3511	0.428	0.2471	0.2072	0
<i>Campylobacter</i> Abattoir	-0.2907	-0.1992	-0.2033	0.2792	-0.0683	0.1115	0.2839	0.1426	0.5785	0.296	-0.176	0
<i>E. coli</i> Retail	-0.1576	-0.2137	-0.2989	0.00037	-0.0788	0.0523	0.2182	0.4276	0.1482	0.00296	-0.1045	0

NS, No significant relationship at significance level of 5%.

Coefficients indicate a positive or negative associated between month and outcome variable.

<sup>a</sup>December was selected as the baseline month to which all other months were compared.

Additional analyses were conducted to examine the seasonality of pathogen prevalence for the national and Ontario chicken subgroup datasets (Tables 5 and 6, respectively). Coefficients indicate a positive or negative association between month and outcome variable, with greater absolute values indicating the proportion of the variance in the prevalence that is predictable from the month. The effect of month on bacteria prevalence was statistically significant in all cases ( $P \leq 0.05$ ), except for *Salmonella* at retail. The strongest seasonal relationship between the pathogens investigated and climate was observed for *Campylobacter*. This finding was observed among chicken and swine samples from retail and abattoir settings both nationally and provincially. Figure 3 shows the coefficients for all the *Campylobacter* variables in chicken and swine by month and illustrates this seasonal trend, with negative coefficients generally observed in the winter months and positive coefficients from spring through the fall, indicating a greater prevalence in the latter compared to the reference month of December. There was a strong positive association between temperature and the odds of positive *Campylobacter* samples in chicken and swine in abattoir and retail environments, which has also been identified in other countries [25]. To demonstrate, results for Ontario were transformed to odds ratios and plotted with average temperatures in Figure 4. The odds of detecting *Campylobacter* in chicken at the abattoir and retail environments were greater from June through November when compared to the baseline month of December. This corresponds approximately to the period of peak average air temperatures and an approximate 2–3 month period afterwards. No clear seasonal patterns emerged for *Salmonella* or generic *E. coli*.

## Discussion

This study was designed to use large-scale national data to examine the relationship between weather and contamination of meat products with *Campylobacter*, generic *E. coli* and *Salmonella*. No single weather variable was associated with the prevalence of all bacteria across all meat products; relationships between variables and prevalence were dependent on the bacteria, location along the agri-food chain and meat product. Efforts were taken to restrict the variability of weather data for each province to only the locations where beef, chicken and swine were produced in each province. However, the use of the Census of Agriculture data reduced the area in which weather was considered with varying degrees of success. Beef production was distributed throughout much larger areas of Canada than the other livestock species (Table 2 and Fig. 2) and therefore associated with the largest variability among weather data. In Alberta, the distribution of livestock was more widespread than in other provinces, which reduced the precision of weather data. For this study, we performed a single subgroup analysis for *Campylobacter* and *Salmonella* in Ontario chicken to demonstrate how relationships change when analysing on a provincial vs. national scale, and it is expected that results would differ interprovincially as well, based on differences in animal production/processing, climate/weather and data quality.

There were some apparent differences when examining Ontario vs. national data. These differences were primarily related to the precision of relationships where provincial level data can be related to smaller weather increments. If we observed only the direct effects of temperature or precipitation on bacterial contamination, we would expect to identify consistent associations at various scales and locations. Since there was variability in the

**Table 6.** Statistical model coefficients by month for the relationship between weather variables and prevalence of bacteria in Ontario chicken from 2002 through 2012

Outcome	Independent variable											
	Jan	Feb	Mar	Apr	May	Jun	Jul	Aug	Sep	Oct	Nov	Dec <sup>a</sup>
<i>Salmonella</i> Retail	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	NS	0
<i>Salmonella</i> Abattoir	-0.0669	-0.2262	-0.2275	-0.0046	-0.3333	-0.3476	-0.096	-0.3249	0.0169	0.6976	0.4458	0
<i>Campylobacter</i> Retail	-0.1223	-0.2676	-0.3446	-0.1837	-0.3943	-0.0931	0.3138	0.4797	0.3643	0.0294	0.0495	0
<i>Campylobacter</i> Abattoir	-1.5727	-0.2529	-0.4617	-0.0433	-0.5846	0.4031	0.4018	0.435	0.7717	0.8185	0.1711	0

NS, No significant relationship at significance level of 5%.

Coefficients indicate a positive or negative associated between month and outcome variable.

<sup>a</sup>December was selected as the baseline month to which all other months were compared.

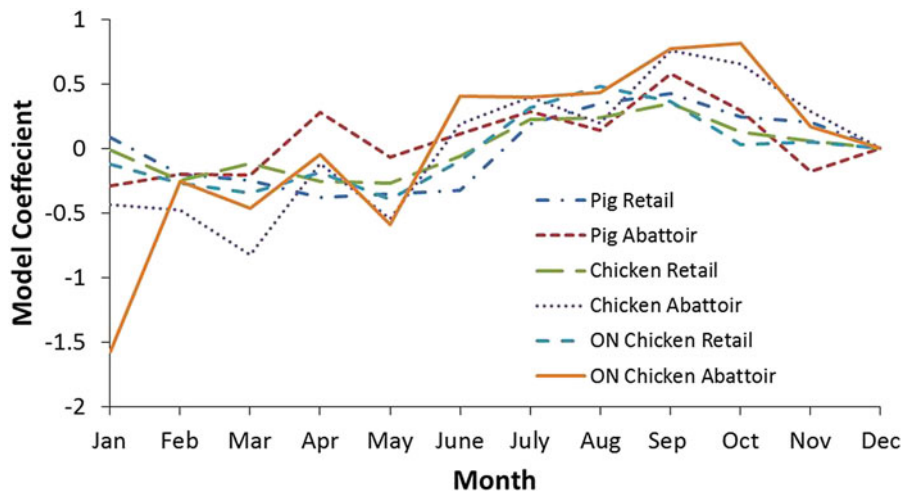


Fig. 3. Model coefficients for *Campylobacter* by month for select analyses. December was selected as the baseline month to which other months were compared.

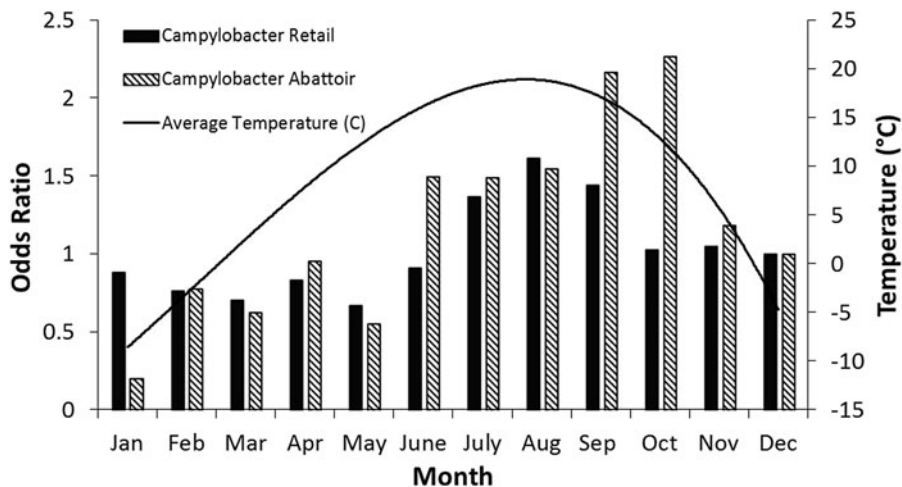


Fig. 4. Odds ratios for *Campylobacter* occurrence in chicken at abattoir and retail by month in Ontario. Average temperatures in production areas are shown for comparison. December was selected as the baseline month to which all other months were compared.

findings at the national and Ontario level and among location types (farm, abattoir, retail), we can infer that indirect effects of weather on bacteria contamination were apparent in this dataset and that other factors aside from climate impact prevalence of bacteria inter-provincially.

The mechanisms underlying the seasonality in foodborne disease are not fully understood, but they are likely a complex interplay of different factors. These include pathogen prevalence in the animal reservoir, pathogen environmental survival patterns and human behaviour and consumption patterns [26]. Much of the previous evidence of the seasonality of foodborne pathogens comes from studies which used cases of human illness as the primary outcome of interest [7, 26]. Since the outcome variable for this research was contamination along the agri-food chain instead of human cases of foodborne illness, an explanation of the relationship excludes many of the seasonal human behaviour and consumption patterns associated with foodborne illness, such as attending a barbecue or picnic and camping [14]. Since no seasonal patterns were evident in the case of *Salmonella* and generic *E. coli*, this suggests the relative importance of human behaviour and consumption patterns in their apparent seasonality of human cases. A study of one Canadian community came to a similar conclusion and the researchers suggested that contamination of retail chicken may not be driving the seasonality of salmonellosis in Canada [26].


We found that odds of contamination of chicken or swine with *Salmonella* in Canada were correlated with either precipitation or variability in precipitation, respectively. The absolute increase in these weather variables required for a 10% increase in odds were relatively high, suggesting that although significant, *Salmonella* prevalence is not sensitive to small changes to precipitation metrics. International studies have indicated that rainfall has a minor impact on the incidence of salmonellosis [27] and can even be inversely related to cases [28].

Seasonality appears to play a significant role in *Campylobacter* contamination and there is a statistical association with temperature at a range of scales amongst settings and commodities. This phenomenon makes *Campylobacter* the most likely foodborne zoonotic enteric pathogen studied whose occurrence in meat products is affected by air temperature changes in Canada. The odds of detecting *Campylobacter* in Ontario chicken, for example, are greater from June through November and July through November in the abattoir and retail environments, respectively (Fig. 4). This suggests a lag of approximately 1 month between abattoir and retail contamination in the summer. However, this pattern is not reproduced throughout the remainder of the year – abattoir contamination remained elevated in the fall whereas retail contamination decreased. This phenomenon warrants further investigation through targeted sampling.



Further research into clarifying the routes of *Campylobacter* movement pathways in the natural environment would be of great benefit to understanding the relationship between climate/weather and *Campylobacter*. The bacteria replicate primarily within the intestinal tract of warm-blooded animals [2]. Therefore, it is the post-excretion period that defines the ecology of campylobacteriosis [29]. Environmental contamination occurs primarily through feces dispersal, including by animals slaughtered at food processing, whereby the carcass can be contaminated [29]. Laboratory studies have characterised the survival of *Campylobacter* in swine manure and found a decline in the viable cells observed at higher temperatures (15, 22, 42 and 52 °C) compared to milder temperatures (4 °C) [30]. The seasonal nature of *Campylobacter* could be driven by some external and localised phenomena such as vectors (e.g. flies, rodents, wildlife, humans) [31]. Several studies have placed screened barriers on chicken houses and observed significant reductions in the prevalence of *Campylobacter* compared with control chicken houses [32, 33]. The seasonality of flies in Canada would certainly be affected by climate change leading to increased transmission due to a longer warm season, as their lifecycles are strongly regulated by temperature.

Strategic and effective food safety management requires understanding microbiological hazards and how their presence in foods can be prevented or maintained within tolerable levels when subjected to changing external factors such as climate [34]. Understanding pathogen-specific seasonal patterns is important for improving existing disease surveillance methods, generating appropriate prevention strategies, developing valid prediction models and enhancing cross-border cooperation [26]. Generally, the increases in temperature variables required to increase the odds of contamination fall within the range of projected climate change in some regions of Canada over the 21st century using medium-high future emission scenarios (and even low-emission scenarios for Ontario) [35]. The statistical associations between weather and foodborne bacteria, as well as variations in seasonal patterns, provide some insight into the complex and nonlinear nature of interactions between environmental, pathogen and host-specific factors and transmission opportunities.

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## References

- Havelaar AH *et al.* (2015) World Health Organization global estimates and regional comparisons of the burden of foodborne disease in 2010. *PLoS Medicine* **12**, e1001923.
- Newell DG *et al.* (2010) Food-borne diseases – the challenges of 20 years ago still persist while new ones continue to emerge. *International Journal of Food Microbiology* **139**, S3–15.
- Thomas MK *et al.* (2013) Estimates of foodborne illness-related hospitalizations and deaths in Canada for 30 specified pathogens and unspecified agents. *Foodborne Pathogens and Disease* **10**, 639–648.
- Copeland L and Wilcott L (2006) Evidence Review: Food Safety. British Columbia, Canada: BC Ministry of Health, CORE public health functions for BC. Available at: <https://www.health.gov.bc.ca/library/publications/year/2006/food-safety-evidence-review.pdf> (Accessed 1 August 2015).
- Thomas MK *et al.* (2006) Estimated numbers of community cases of illness due to *Salmonella*, *Campylobacter* and verotoxigenic *Escherichia coli*: Pathogen-specific community rates. *Canadian Journal of Infectious Diseases and Medical Microbiology* **17**, 229–234.
- Bank of Canada Inflation Calculator. Available at <https://www.bankofcanada.ca/rates/related/inflation-calculator/> (Accessed 15 December 2017).
- Fleury M *et al.* (2006) A time series analysis of the relationship of ambient temperature and common bacterial enteric infections in two Canadian provinces. *International Journal of Biometeorology* **50**, 385–391.
- Rose JB *et al.* (2001) Climate variability and change in the United States: potential impacts on water- and foodborne diseases caused by microbiologic agents. *Environmental Health Perspectives* **109**, 211–221.
- Park MS, Park KH and Bahk GJ (2018) Combined influence of multiple climatic factors on the incidence of bacterial foodborne diseases. *Science of the Total Environment* **610–611**, 10–16.
- Ebi K (2011) Climate change and health risks: Assessing and responding to them through adaptive management. *Health Affairs* **30**, 924–930.
- Semenza JC *et al.* (2012) Climate change impact assessment of food- and waterborne diseases. *Critical Reviews in Environmental Science and Technology* **42**, 857–890.
- Dirk Van Elsas J *et al.* (2011) Survival of *Escherichia coli* in the environment: fundamental and public health aspects. *Multidisciplinary Journal of Microbial Ecology* **580**, 173–183.
- Kovats RS *et al.* (2004) The effect of temperature on food poisoning: a time-series analysis of salmonellosis in ten European countries. *Epidemiology and Infection* **132**, 443–453.
- Health Canada (2008) Human health in a changing climate: a Canadian assessment of vulnerabilities and adaptive capacity. Ottawa, ON, Canada: Health Canada 2008. Available at <http://publications.gc.ca/site/eng/9.691579/publication.html> (Accessed August 1 2015).
- Keen J *et al.* (2003) Effect of exogenous glucocorticoids and dietary change on winter and summer STEC O157 fecal shedding in naturally-infected beef cattle. *Research Workers in Animal Diseases Conference Proceedings*. Chicago: Research Workers in Animal Diseases, Abstract No. 83.
- Pangloli P *et al.* (2008) Seasonal incidence and molecular characterization of *Salmonella* from dairy cows, calves, and farm environment. *Foodborne Pathogens and Disease* **5**, 87–96.
- Jacob ME, Callaway TR and Nagaraja TG (2009) Dietary interactions and interventions affecting *Escherichia coli* O157 colonization and shedding in cattle. *Foodborne Pathogens and Disease* **6**, 785–792.
- Williams KJ *et al.* (2015) Risk factors for *Escherichia coli* O157 shedding and super-shedding by dairy heifers at pasture. *Epidemiology and Infection* **143**, 1004–1015.
- Burkholder KM *et al.* (2008) Influence of stressors on normal intestinal microbiota, intestinal morphology, and susceptibility to *Salmonella enteritidis* colonization in broilers. *Poultry Science* **87**, 1734–1741.
- Moro MH *et al.* (2000) Effects of heat stress on the antimicrobial drug resistance of *Escherichia coli* of the intestinal flora of swine. *Journal of Applied Microbiology* **88**, 836–844.
- CIPARS. The Canadian Integrated Program for Antimicrobial Resistance Surveillance 2012 Annual Report. Available at <https://www.canada.ca/en/public-health/services/surveillance/canadian-integrated-program-antimicrobial-resistance-surveillance-cipars/canadian-integrated-program-antimicrobial-resistance-surveillance-cipars-annual-report/cipars-2012-annual-report.html> (Accessed 10 August 2018).
- Environment Canada. Canadian Climate Summaries. Available at [http://climate.weather.gc.ca/prods\\_servs/cdn\\_climate\\_summary\\_e.html](http://climate.weather.gc.ca/prods_servs/cdn_climate_summary_e.html) (Accessed 6 June 2015).
- Daly C (2006) Guidelines for assessing the suitability of spatial climate data sets. *International Journal of Climatology* **26**, 707–721.
- Statistics Canada. Census of Agriculture. Available at <http://www.statcan.gc.ca/eng/ca2011/index> (Accessed 1 August 2015).
- Nylen G *et al.* (2002) The seasonal distribution of *Campylobacter* infection in nine European countries and New Zealand. *Epidemiology and Infection* **128**, 383–390.
- Ravel A *et al.* (2010) Seasonality in human salmonellosis: assessment of human activities and chicken contamination as driving factors. *Foodborne Pathogens and Disease* **7**, 785–794.
- Aik J *et al.* (2018) Climate variability and salmonellosis in Singapore – A time series analysis. *Science of The Total Environment* **639**, 1261–1267.

28. **Zhang Y, Bi P and Hiller J** (2008) Climate variations and salmonellosis transmission in Adelaide, South Australia: a comparison between regression models. *International Journal of Biometeorology* **52**, 179–187.
29. **Skelly C and Weinstein P** (2003) Pathogen survival trajectories: an eco-environmental approach to the modeling of human campylobacteriosis ecology. *Environmental Health Perspectives* **111**, 19–28.
30. **Bui XT *et al.*** (2011) Fate and survival of *Campylobacter coli* in swine manure at various temperatures. *Frontiers in Microbiology* **2**, 262.
31. **Agunos A *et al.*** (2014) A systematic review characterizing on-farm sources of *Campylobacter* spp. for broiler chickens. *PLoS ONE* **9**, e104905.
32. **Hald B, Sommer HM and Skovgård H** (2007) Use of fly screens to reduce *Campylobacter* spp. introduction in broiler houses. *Emerging Infectious Diseases* **13**, 1951–1953.
33. **Bahrndorff S *et al.*** (2013) Foodborne disease prevention and broiler chickens with reduced *Campylobacter* infection. *Emerging Infectious Diseases* **19**, 19–21.
34. **Tirado MC *et al.*** (2010) Climate change and food safety: a review. *Food Research International* **43**, 1745–1765.
35. **Warren FJ and Lemmen DS** (eds) (2014) *Canada in a Changing Climate: Sector Perspectives on Impacts and Adaptation*. Ottawa: Government of Canada, p. 286.