



Key inflammatory mechanisms underlying heart failure

Heart failure (HF) is a clinical syndrome based primarily on systolic or diastolic left-ventricular (LV) contractile dysfunction. The prognosis of chronic HF is poor, with about 50% of patients dying within 5 years after the initial diagnosis. There are different categories of HF, which are based on measurements of LV ejection fraction (LVEF). About half of HF patients are afflicted with HF with reduced ejection fraction (HF_rEF) with an LVEF of <40%. In contrast, HF with preserved ejection fraction (HF_pEF) is observed in roughly the other half of patients (LVEF ≥50%). Patients with an LVEF in the range of 40–49% represent a “gray area” that is defined as HF with mid-range ejection fraction (HF_{mr}EF; [1]). The prevalence of HF in industrialized nations is increasing to more than 10% among people greater 70 years of age [2]. Statistically, about one in three individuals at 55 years of age will develop HF during their remaining lifespan [3]. The increase in HF can be explained by the rising prevalence of renal failure, arterial hypertension, chronic obstructive pulmonary disease (COPD), diabetes mellitus, and metabolic syndrome. These comorbidities are characterized by chronic inflammation and are of particular importance for patients with HF_pEF [2]. Furthermore, the treatment of ischemic heart disease has significantly improved over the past few decades, which has increased the number of surviving HF patients.

In addition to playing a critical role in the development and progression of HF_pEF and HF_rEF [4, 5], the inflammatory response is also important for adverse remodeling processes following myocardial infarction (MI). The development of HF can also be directly immune-modulated, for example, following

autoimmune or infectious triggers, i.e., viral infection. Following acute myocardial injury, the inflammatory response is required to induce the regenerative response, but sustained and chronic inflammation is detrimental. Based on the dichotomous role of inflammation in cardiac tissue, the modulation of inflammatory processes has been identified as a therapeutic approach. The pathomechanisms underpinning inflammation modulation for therapeutic benefit have been investigated in numerous studies and will be summarized in this review.

HF_pEF, endothelial dysfunction, and inflammation

One hallmark of HF_pEF is impaired LV relaxation as a consequence of altered composition of the extracellular matrix and decreased cyclic guanosine monophosphate (cGMP)/protein kinase G (PKG) signaling. From a mechanistic perspective, comorbidities promote systemic inflammation, which increases reactive oxygen species (ROS) production in cardiac endothelial cells and peroxynitrite (ONOO⁻) levels. The subsequent decrease in nitric oxide (NO) in endothelial cells impairs soluble guanylate cyclase (sGC) levels and PKG activity in adjacent cardiomyocytes. This promotes adverse LV remodeling and hypophosphorylation of titin, which impairs LV relaxation. Furthermore, monocytes infiltrate cardiac tissue under conditions of chronic inflammation and differentiate into macrophages, which augment myocardial inflammation. This also promotes fibrosis by differentiation of fibroblasts into myofibroblasts following transforming growth factor beta

(TGF β) secretion by monocytes ([6; **Fig. 1**).

Several studies provide mechanistic insight into the cardioprotective effects of NO/sGC/cGMP/PKG signaling. For example, pharmacological stimulation of sGC attenuates LV remodeling after MI in mice, decreases extracellular matrix protein production in human cardiac fibroblasts following TGFβ stimulation *in vivo* [7], and attenuates vascular dysfunction in diabetic rats [8]. Similarly, the endothelial NO synthase (eNOS) transcription enhancer AVE9488 improves cardiac remodeling after MI [9] and platelet NO availability and hyperactivity in HF [10]. Senescence-accelerated-prone mice (SAMP) develop manifest HF_pEF when subjected to a high-salt, high-fat diet, which is characterized by endothelial cell dysfunction and fibrosis. These studies highlight the contribution of endothelial cell dysfunction on the age-dependent increase in HF_pEF [11]. Furthermore, increased insulin-like growth factor-1 (IGF-1) activity following growth hormone stimulation attenuates age-dependent endothelial progenitor cell dysfunction [12]. Myeloperoxidase (MPO) is a bactericidal enzyme that is released from activated polymorphonuclear neutrophils and can directly modulate the vascular inflammatory response by regulating NO bioavailability [13]. Importantly, MPO also promotes HF following ischemic injury [14], atrial structural remodeling, and increases the risk of atrial fibrillation [15].

Mice with cardiomyocyte-specific deletion of iron-regulatory proteins (Irp) 1 and 2 exhibit mitochondrial dysfunction and accelerated HF after MI [16], which underscores the importance of iron availability in cardiomyocytes. This

Hier steht eine Anzeige.



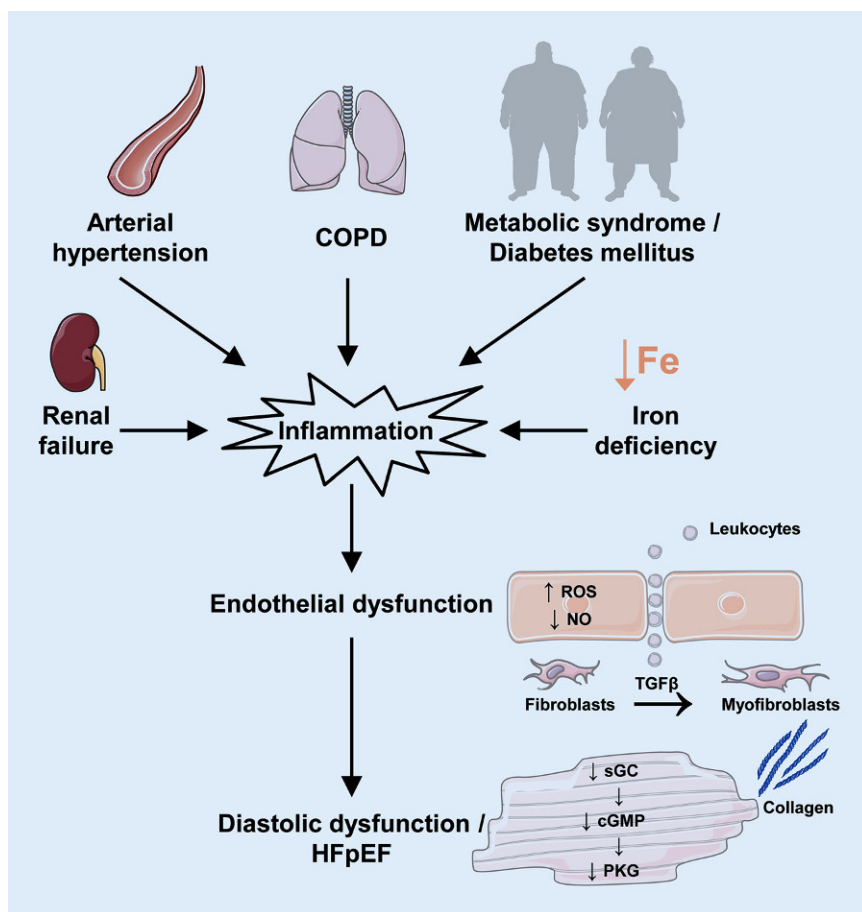


Fig. 1 ▲ Schematic depicting the impact of endothelial dysfunction and inflammation on the development of fibrosis and heart failure with preserved ejection fraction (HFpEF). Comorbidities, such as renal failure, arterial hypertension, chronic obstructive pulmonary disease (COPD), metabolic syndrome, diabetes mellitus, and iron deficiency, induce systemic inflammation. Increased mitochondrial reactive oxygen species (ROS) production, increased peroxynitrite ($ONOO^-$) levels, and decreased nitric oxide (NO) levels in endothelial cells attenuate cardiomyocyte soluble guanylate cyclase (sGC)/guanosine monophosphate (cGMP)/protein kinase G (PKG) signaling, which induces adverse left-ventricular remodeling and diastolic dysfunction. Inflammation also promotes fibrosis by differentiation of fibroblasts into myofibroblasts following transforming growth factor beta ($TGF\beta$) secretion by monocytes

is further emphasized by impaired mitochondrial capacity and contractility in human embryonic stem cell-derived cardiomyocytes following incubation with the iron chelator deferoxamine. Mitochondrial capacity and contractility were restored following enhanced intracellular iron levels, suggesting that iron levels directly mediate these effects [17]. It has been shown that LV samples from failing human hearts exhibit decreased iron content, which may impair mitochondrial capacity and ROS scavenging in these samples [18]. ROS can mediate both beneficial and deleterious effects that are based on the subcellular

localization and duration of exposure to ROS, as recently reviewed [19].

The importance of inflammation in the development of HFpEF has been demonstrated in a swine model following induction of the three most common inflammation-associated comorbidities in HFpEF patients: arterial hypertension, diabetes mellitus, and hypercholesterolemia [20]. Diabetes mellitus also increases the risk of diastolic dysfunction and HF independent of coexisting coronary artery disease and hypertension. This resulted in the term “diabetic cardiomyopathy.” Various mechanisms increase the risk of HF in diabetic patients [21, 22], including

increased inflammation. The underlying mechanisms of inflammatory-dependent HF in diabetic patients include increased expression levels of interleukins (IL-1 β , IL-6), intercellular adhesion molecule-1 (ICAM-1), and vascular cell adhesion molecule-1 (VCAM-1), and decreased activity of the collagen degrading matrix metalloproteinase (MMP).

Finally, autophagy is a highly conserved cellular process that plays important roles in the maintenance of cellular homeostasis and quality control of organelles. Depending on the extent and duration of autophagy, this cellular process can be both beneficial and detrimental [23, 24]. Perturbed cardiac autophagy has been described for several risk factors of HFpEF development, including hypertension, diabetes, and aging [25].

Inflammation following ischemic injury

Macrophages and monocytes are essential for the inflammatory response and ventricular remodeling following ischemic injury [26, 27]. The cellular response to myocardial ischemia can be categorized into different phases: the acute inflammatory phase, the healing phase, and a phase of chronic inflammation. A schematic summary of these events after MI in mice is provided in **Fig. 2**. The different phases are well characterized in murine models and require a greater time span in larger animals and humans. The cellular and inflammatory adaptations are mediated by neutrophils and monocytes, which are generated in the bone marrow and the spleen and then translocate to the injured myocardium. Three monocyte subsets have been described in humans; these are based on the expression pattern of the surface protein expression markers CD14 and CD16. On the basis of these expression patterns, monocytes can be classified as classic ($CD14^{++}CD16^{-}$; murine homolog: $Ly6C^{high}$), intermediate ($CD14^{++}CD16^{+}$), and nonclassical ($CD14^{+}CD16^{++}$; murine homolog: $Ly6C^{low}$) monocytes.

$Ly6C^{high}$ monocytes express high levels of $Ly6C$, $CCR2$, and $CD62L$ and play a predominant role in the initial inflam-

matory phase after MI. Recruitment of Ly6C^{high} monocytes is mediated by high expression levels of the cytokine CCL2. Ly6C^{high} monocytes produce high levels of pro-inflammatory cytokines, such as IL-1 β and tumor necrosis factor alpha (TNF α), which resulted in the term “inflammatory” monocytes for these cells. Ly6C^{high} monocytes recruit inflammatory macrophages, which secrete proteolytic enzymes to digest and prepare the damaged tissue for regeneration. Inflammatory macrophages are commonly termed “M1” macrophages. Decreasing the recruitment of neutrophils to the injured myocardium is essential in order to limit tissue injury and to initiate the healing process. Following ingestion of damaged and apoptotic cells, macrophages decrease their production of IL-1 β and TNF α , and increase the secretion of anti-inflammatory and pro-fibrotic cytokines, i.e., IL-10 and TGF β [28]. Following transformation and change of their activation profile, less inflammatory macrophages are termed “M2” or “reparative” macrophages. Neutrophils play a critical role in this polarization of macrophages toward a reparative M2 phenotype [29]. Although commonly used, the categorization of macrophages into “M1” and “M2” subtypes is an oversimplification to describe their heterogeneity, which is originally based on in vitro studies and is problematic for describing adaptations in vivo.

The second phase, called the “healing phase,” is dominated by Ly6C^{low} monocytes. These cells are recruited via CX₃CR₁ (fractalkine receptor)-mediated signaling to the infarcted region and are present at much lower numbers compared with Ly6C^{high} monocytes [30]. Ly6C^{high} can differentiate into Ly6C^{low} monocytes. However, the exact relationship between M1 and M2 macrophages and Ly6C^{high} and Ly6C^{low} monocytes requires further investigation [31]. A seminal study performed by the Molkentin group greatly improved our understanding of how myocardial scar tissue forms after ischemic injury [32]. Using transgenic lineage-tracing mouse lines as reporter constructs, the authors show that both cardiomyocytes and fibroblasts die in the infarcted area.

Herz 2019 · 44:96–106 <https://doi.org/10.1007/s00059-019-4785-8>
© The Author(s) 2019

C. Riehle · J. Bauersachs

Key inflammatory mechanisms underlying heart failure

Abstract

Inflammation plays a central role in the development of heart failure, especially in heart failure with preserved ejection fraction (HFpEF). Furthermore, the inflammatory response enables the induction of regenerative processes following acute myocardial injury. Recent studies in humans and animals have greatly advanced our understanding of the underlying mechanisms behind these adaptations. Importantly, inflammation can have both beneficial and detrimental effects, dependent on its extent, localization,

and duration. Therefore, modulation of cardiac inflammation has been suggested as an attractive target for the treatment of heart failure, which has been investigated in numerous clinical trials. This review discusses key inflammatory mechanisms contributing to the pathogenesis of heart failure and their potential impact as therapeutic targets.

Keywords

Cardiac failure · Inflammation · Myocardial infarction · Immune system · Cytokines

Entzündungsmechanismen bei Herzinsuffizienz

Zusammenfassung

Entzündungsprozesse spielen eine zentrale Rolle bei der Entwicklung der Herzinsuffizienz, insbesondere bei Herzinsuffizienz mit erhaltener Ejektionsfraktion (HFpEF). Darüber hinaus sind Entzündungsprozesse allerdings auch für die Reparationsvorgänge nach akutem Myokardinfarkt erforderlich. Sowohl aktuelle Studien an Tiermodellen als auch Untersuchungen an Menschen führten zu einem besseren Verständnis der zugrunde liegenden Mechanismen. Abhängig von Lokalisation, Ausmaß und der Dauer können Entzündungsprozesse sowohl vorteilhaft als auch nachteilig sein. Deshalb bietet sich deren

Beeinflussung als ein möglicher Angriffspunkt zur Behandlung der Herzinsuffizienz sowie pathologischer Umbauvorgänge an. Dies ist Gegenstand zahlreicher klinischer Studien. In der vorliegenden Übersichtsarbeit wird die Rolle wesentlicher Entzündungsprozesse in der Pathogenese der Herzinsuffizienz erörtert und deren potenzielle Bedeutung als Therapieoption diskutiert.

Schlüsselwörter

Herzinsuffizienz · Entzündung · Myokardinfarkt · Immunsystem · Zytokine

Subsequently, fibroblasts from the border zone region are activated and proliferate greatly, which results in an approximately 3.5-fold increase in the total number of fibroblasts in the infarcted area relative to uninjured conditions 3 days after MI. This elevation in count was observed for over 4 weeks. Between days 3 and 7, fibroblasts differentiate into myofibroblasts, as indicated by smooth muscle α -actin expression. Subsequently, the proliferation of myofibroblasts and smooth muscle α -actin expression decrease by days 7–10, while the scar tissue matures.

Following the healing phase, inflammation often persists or reoccurs during the development of HF. Hallmarks of chronic inflammation are the increased abundance of tissue T-lymphocytes and pro-inflammatory M1 macrophages [33,

34]. Risk factors for the development of HF include an initial inflammatory response [35] and large MI [36]. A recent study showed that the lymphatic system is required for clearing immune cells and limiting the immune response in cardiac tissue following ischemic injury. Genetic deletion of lymphatic vessel endothelial hyaluronan receptor 1 (LYVE-1) in mice decreased the clearance of leukocytes to mediastinal lymph nodes following MI [37]. This resulted in increased pathological remodeling and decreased cardiac function. These intriguing data emphasize the adverse effect of persistent inflammation following MI.

The risk of acute atherothrombotic events is increased following MI, both at culprit and nonculprit arteries [38]. Several changes in remote vessels fol-

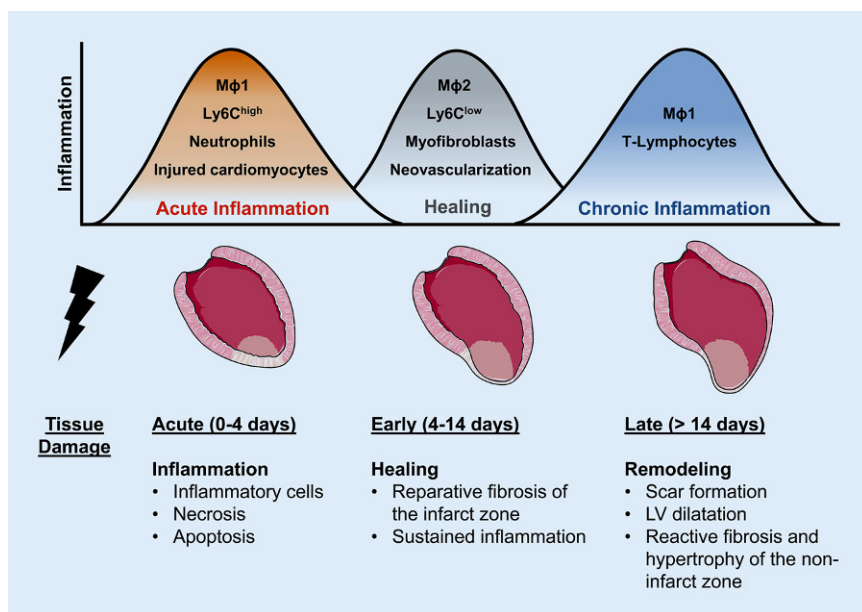


Fig. 2 ▲ Time course of inflammation and healing after myocardial infarction in mice. The acute inflammatory response is characterized by infiltration with M1 macrophages ($M\phi 1$), $Ly6C^{high}$ monocytes, and neutrophils. The main characteristics of the healing phase are infiltration with M2 macrophages ($M\phi 2$), $Ly6C^{low}$ monocytes and myofibroblasts, which contribute to wound repair, neovascularization, limitation of tissue damage, and reparative fibrosis of the infarct zone. Chronic inflammation might result from persistent inflammation following the healing phase or a second boost of inflammation. Note that the categorization of macrophages into “M1” and “M2” subtypes is an oversimplification and that the different phases require a greater time span in larger animals and humans. LV left-ventricular

lowing MI may contribute to accelerated plaque formation and inflammation: increased platelet-endothelial adhesion from endothelial-associated von Willebrand factor (VWF) multimers and increased expression of endothelial inflammatory adhesion molecules [39]. Even though the underlying molecular mechanisms require further investigation, these observations suggest increased inflammation is associated with an increased risk of future ischemic events following MI. Similarly, MI increases the risk of atherosclerosis [40]. A recently identified target for cardiac regeneration after MI is glucocorticoid receptor (GR) expression in macrophages. GR regulates factors that control inflammation and neovascularization, which are required for the preservation of contractile function and scar tissue formation [41]. Monocytes and macrophages are also important for healing and for the prevention of ventricular thrombus formation after MI [42].

Multimodality noninvasive imaging has been used to assess the inflammatory response in patients following MI and identified the spleen and bone marrow as

sources of inflammatory cells [43]. Using positron emission tomography (PET), it has also been shown that acute myocardial inflammation predicts subsequent functional outcome and neuroinflammation after MI [44]. The mechanisms contributing to chronic inflammation are incompletely understood and include persistent inflammation following the healing phase and resurgence in inflammation [45]. Stem cell transplantation following MI has been suggested as a promising therapeutic approach to limit tissue damage and preserve contractile function. While most of the transplanted cells die from apoptosis and contribute little to neovascularization, these cells may also mediate the immune response resulting in reduced scar tissue formation and improved cardiac outcome (“dying stem cell hypothesis”; [46]).

Mineralocorticoid receptor-mediated signaling

The mineralocorticoid aldosterone mediates inflammatory pathways and is

critical for adverse cardiac remodeling. Aldosterone is produced in the glomerular zone of the adrenal cortex and increases sodium reabsorption, potassium secretion, and blood pressure. This is facilitated following its binding to the mineralocorticoid receptor (MR), which is a member of the nuclear receptor transcription factor family. Following translocation to the nucleus and homodimerization, MRs promote the expression of target genes. MRs are also expressed in various cell types of the heart and the vasculature, including cardiomyocytes, fibroblasts, coronary endothelial cells, vascular smooth muscle cells, and inflammatory cells [47]. Myocardial MR expression is increased in patients with HF [48]. MR antagonists reduced mortality and morbidity rates in patients with HF in large clinical studies (RALES, EPHESUS and EMPHASIS-HF trials [49–51]) and are commonly used as a standard treatment for patients with HF. The EPHESUS trial also showed that early initiation of MR blockage after MI and concomitant HF is beneficial relative to later initiation of the treatment [52]. A prespecified meta-analysis of the ST-segment elevation myocardial infarction (STEMI) subgroup of the ALBATROSS and the REMINDER trials shows that early initiation of MR antagonist treatment in patients with STEMI reduces mortality and the composite of death or resuscitated sudden cardiac death [53]. These data highlight the benefits of MR antagonist treatment following MI.

Aldosterone and MR signaling promote inflammation, myocardial hypertrophy, adverse LV remodeling, and ischemic injury. These effects are, at least in part, independent of systemic blood pressure and transduced by increased pro-inflammatory and pro-fibrotic signaling, i.e., $TNF\alpha$, $TGF\beta$, connective tissue growth factor (CTGF), and increased oxidative stress induced by NADPH oxidases [54–56]. Several studies using mouse models have advanced our understanding of cardiac MR-mediated signaling. Mice with genetic deletion of the MR in myeloid cells are protected against cardiac hypertrophy, fibrosis, and vascular damage induced by L-NAME/angiotensin II treatment.

Table 1 Summary of major clinical trials targeting inflammatory pathways and immune-modulatory therapies in heart failure

Study	Treatment	Target	Duration (months)	Clinical setting	NYHA class	n	Primary outcome
ATTACH [66]	Infliximab	TNF α	7	DCM, ICM	III, IV	150	↑ Death and hospitalization for HF at high doses
RENEWAL (RECOVER and RENAISSANCE) [67]	Etanercept	TNF α	5.7/12.9	DCM, ICM	II–IV	2048	↔ Death and hospitalization rate for HF
Gullestad et al. [69]	Thalidomide	Multiple	3	DCM, ICM	II, III	56	↑ LVEF
Parrillo et al. [81]	Prednisone	Multiple	3	DCM	–	102	↑ LVEF
Skudicky et al. [70]	Pentoxifylline	Multiple	6	DCM	II, III	39	↑ LVEF and symptoms
Sliwa et al. [71]	Pentoxifylline	Multiple	6	DCM	II, III	28	↑ LVEF and symptoms
Sliwa et al. [72]	Pentoxifylline	Multiple	1	DCM	IV	18	↑ LVEF and ↓ TNF α
Sliwa et al. [73]	Pentoxifylline	Multiple	6	ICM	II, III	38	↑ LVEF and ↓ plasma inflammatory markers
Bahrmann et al. [74]	Pentoxifylline	Multiple	6	DCM, ICM	II, III	47	↔ LVEF
CORONA [75]	Rosuvastatin	Multiple	32.8	ICM	II–IV	5011	↔ Cardiovascular death, non-fatal MI, and nonfatal stroke
GISSI-HF [76]	Rosuvastatin	Multiple	46.9	DCM, ICM	II–IV	4574	↔ Death and cardiovascular hospitalization
Krum et al. [77]	Rosuvastatin	Multiple	6	DCM, ICM	II–IV	87	↔ LVEF
ACCLAIM [78]	Device-based immunomodulation	Nonspecific	10.2	DCM, ICM	II–IV	2426	↔ Death and cardiovascular hospitalization
Gullestad et al. [79]	Intravenous immunoglobulin	Multiple	6	DCM, ICM	II, III	40	↑ LVEF
IMAC [80]	Intravenous immunoglobulin	Multiple	12	DCM	I–IV	62	↔ LVEF
METIS [82]	Methotrexate	Multiple	3	ICM	II–IV	50	↔ 6-Minute walk test

ACCLAIM Advanced Chronic Heart Failure Clinical Assessment of Immunomodulation, **ATTACH** Anti-TNF Therapy Against Congestive Heart Failure, **CORONA** Controlled Rosuvastatin Multinational Trial in Heart Failure, **DCM** dilated cardiomyopathy, **GISSI-HF** Gruppo Italiano per lo Studio della Sopravvivenza nell'Insufficienza cardiaca-Heart Failure, **HF** heart failure, **ICM** ischemic cardiomyopathy, **IMAC** Intervention in Myocarditis and Acute Cardiomyopathy, **LVEF** left-ventricular ejection fraction, **METIS** Methotrexate Therapy on the Physical Capacity of Patients with Ischemic Heart Failure, **MI** myocardial infarction, **RECOVER** Etanercept Cytokine Antagonism in Ventricular Dysfunction, **RENAISSANCE** Randomized Etanercept North American Strategy to Study Antagonism of Cytokines, **RENEWAL** Randomized Etanercept Worldwide Evaluation

Furthermore, these mice exhibit an alternatively activated M2 macrophage phenotype. This indicates that MR expression in myeloid cells is required for efficient classic macrophage activation by pro-inflammatory cytokines [57]. Genetic deletion of the MR in cardiomyocytes, but not in fibroblasts, attenuates contractile dysfunction and HF following pressure overload induced by transverse aortic constriction. However, MR deletion in cardiomyocytes or fibroblasts has no impact on cardiac fibrosis and hypertrophy relative to wild type controls following pressure overload [58]. This suggests a potential predominant role of MR expression in myeloid cells in this context.

A series of studies performed by our laboratory has identified several MR-mediated cardioprotective mechanisms following MI. Treatment with the MR antagonist eplerenone attenuates adverse LV remodeling and contractile dysfunction in rats. The underlying mechanisms include accelerated macrophage infiltration, a transient increase in protective cytokines, and alternative M2 macrophage activation [59]. In this context, treatment with eplerenone is superior relative to spironolactone by increasing the abundance of healing Ly6C^{low} monocytes and neovessel formation [60]. Additional mechanistic insight is provided by a transgenic mouse model with cardiomyocyte-specific MR deletion, which exhibits increased healing and attenu-

ated contractile dysfunction [61]. In this model, MR deletion reduces infarct expansion and myocyte apoptosis, while infarct neovessel formation is increased in the early phase after ischemic damage. Furthermore, oxidative stress in the surviving LV myocardium is attenuated. This inflammatory cellular response is accelerated with a transient infiltration of neutrophils, which improves neovascularization and attenuates pathological remodeling. We also observed decreased expression of the MR target gene serum/glucocorticoid-regulated kinase 1 (SGK1) in MR-deficient cardiomyocytes, which mediates cardiomyocyte hypertrophy by increasing CTGF expression [62]. Notably, myeloid cell-specific MR deficiency also attenuates LV dysfunction

and LV remodeling following MI in mice by decreasing inflammation and oxidative stress [63]. Together, our studies identify a critical role of MR-transduced signaling to mediate tissue damage in ischemic heart injury. The mechanisms discovered include activation of inflammatory pathways in various cell types. Our studies also strongly support the importance of MR-antagonist treatment of patients with ischemic heart disease.

Anti-inflammatory and immune-modulatory treatment of patients with HF

Several clinical trials have tested the impact of anti-inflammatory and immune-modulatory therapies in patients with myocarditis, inflammatory cardiomyopathy, and HF [64, 65]. Despite this promising therapeutic approach, these studies have provided ambiguous results (Table 1). Based on its potential contribution to the progression of HF, the pro-inflammatory cytokine TNF α was identified as a promising pharmaceutical target. The randomized placebo-controlled ATTACH [66] and RENEWAL [67] trials tested the impact of the chimeric TNF α -antibody infliximab and the TNF α -inhibitor etanercept, respectively. The data obtained show no advantage of these treatments in patients with HF. Moreover, the ATTACH study reports adverse effects of infliximab at higher doses [66]. Potential mechanisms for these observations include binding of infliximab to TNF α -expressing cardiomyocytes, which might induce complement activation and cardiomyocyte apoptosis. Furthermore, administration of relatively high doses of infliximab might suppress TNF α below physiological concentrations, which are cardioprotective in the context of acute ischemic injury [68]. Gullestad and colleagues [69] recently described beneficial effects of the sedative and anti-nausea drug thalidomide. Despite the previously reported anti-inflammatory effects of thalidomide, the mechanisms are not completely understood and may include matrix stabilization based on decreased MMP2 expression.

Pentoxifylline is an anti-inflammatory agent that inhibits the production of TNF α and IL-6. Treatment with pentoxifylline increased LV contractile function and attenuates HF symptoms in some studies [70–73], while another report showed no difference [74]. Pentoxifylline is also a nonselective phosphodiesterase inhibitor. Therefore, pentoxifylline might mediate its cardioprotective effects by inhibiting phosphodiesterases and being, at least in part, independent of reducing inflammation. In addition to attenuating the formation of low-density lipoprotein by inhibiting the enzyme HMG-CoA reductase, statins are anti-inflammatory and improve endothelial function. However, treatment with statins is not beneficial in the context of HF unless administered in the presence of other comorbidities, such as dyslipidemia or coronary artery disease [75–77].

“Immunomodulation” therapy may provide a beneficial immune response to decrease pro-inflammatory and increase anti-inflammatory pathways. Patients with HF were subjected to “immunomodulation” by exposure of autologous blood ex vivo to controlled amounts of oxidative stress before administration by intragluteal injection. The ACCLAIM trial showed no impact of “immunomodulation” therapy on mortality and cardiovascular hospitalization [78]. Additional studies are required to understand the exact mechanisms of “immunomodulation” therapy that may contribute to potential positive effects of this treatment. In another study, HF patients were subjected to intravenous immunoglobulin (IVIg) infusions to modulate the immune response, which increased contractile function in patients with ischemic cardiomyopathy (ICM) and idiopathic dilated cardiomyopathy (DCM) [79]. By contrast, the IMAC trial [80] showed a similar increase in contractile function in patients with DCM or myocarditis who were treated with IVIg or placebo. It is important to note that the IMAC trial does not provide any data on histological sections from myocardial biopsies, inflammation, and viral persistence for a later time point. Thus, it is challenging to discern the po-

tential benefits of IVIg therapy relative to standard HF therapy, which was administered to HF patients independent of IVIg or placebo. This is of particular importance for patients with myocarditis, who might benefit from the antiviral and immune-modulatory effects of IVIg therapy the most.

Additional therapeutic approaches to modulate the immune response in patients with HF include treatment with prednisone and methotrexate as well as by reduction in the abundance of auto-antibodies by immunoabsorption. In summary, the results of most studies targeting anti-inflammatory and immune-modulatory therapy are ambiguous. Table 1 summarizes major published clinical trials with anti-inflammatory and immune-modulatory treatment in patients with HF. Future research is warranted to identify additional targets for the modulation of inflammation in HF.

Conclusion

Inflammation plays a central role in the development of the different etiologies of HF, especially in HFpEF. Importantly, the inflammatory response following ischemic damage is also required to induce the regenerative response and is transduced by MR-mediated signaling. Decreased MR signaling is beneficial following ischemic damage because of the attenuation of pathological remodeling and MR antagonists are a well-established standard treatment for HF. A variety of key inflammatory markers have been identified that have been subsequently tested as potential targets for the treatment of HF. Even though clinical trials have provided inconclusive results, modulation of inflammation remains a promising target for the treatment of HF. Additional studies are required to further delineate the mechanisms and to identify novel target molecules, which is the subject of ongoing research in this field.

Hier steht eine Anzeige.



Corresponding address

Prof. Dr. J. Bauersachs

Department of Cardiology and Angiology,
Hannover Medical School
Carl-Neuberg-Str. 1, 30625 Hannover, Germany
bauersachs.johann@mh-hannover.de

Acknowledgements. This work was supported by the German Research Foundation, Clinical Research Unit (KFO) 311. Figures were produced using templates from Servier Medical Art (www.servier.com).

Compliance with ethical guidelines

Conflict of interest. C. Riehle received travel support from Abiomed. J. Bauersachs received honoraria for lectures and advice from Novartis, Pfizer, Vifor, Bayer, BMS, Servier, AstraZeneca, Orion, CVRx, Abiomed, Abbott, and Medtronic, and research support from Zoll, CVRx, Bayer, Vifor, Abiomed, and Medtronic.

This article does not contain any studies with human participants or animals performed by any of the authors.

Open Access This article is distributed under the terms of the Creative Commons Attribution 4.0 International License (<http://creativecommons.org/licenses/by/4.0/>), which permits unrestricted use, distribution, and reproduction in any medium, provided you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made.

References

- Bauersachs J, Maisch B (2018) Heart failure 2.0 or 0.1? *Herz* 43(5):381–382. <https://doi.org/10.1007/s00059-018-4720-4>
- Ponikowski P, Voors AA, Anker SD et al (2016) 2016 ESC Guidelines for the diagnosis and treatment of acute and chronic heart failure: The Task Force for the diagnosis and treatment of acute and chronic heart failure of the European Society of Cardiology (ESC). Developed with the special contribution of the Heart Failure Association (HFA) of the ESC. *Eur J Heart Fail* 18(8):891–975. <https://doi.org/10.1002/ehf.592>
- Bleumink GS, Knetsch AM, Sturkenboom MC et al (2004) Quantifying the heart failure epidemic: Prevalence, incidence rate, lifetime risk and prognosis of heart failure. The Rotterdam Study. *Eur Heart J* 25(18):1614–1619. <https://doi.org/10.1016/j.ehj.2004.06.038>
- Frantz S, Falcao-Pires I, Balligand JL et al (2018) The innate immune system in chronic cardiomyopathy: a European Society of Cardiology (ESC) scientific statement from the Working Group on Myocardial Function of the ESC. *Eur J Heart Fail* 20(3):445–459. <https://doi.org/10.1002/ehf.1138>
- Lourenco AP, Leite-Moreira AF, Balligand JL et al (2018) An integrative translational approach to study heart failure with preserved ejection fraction: A position paper from the Working Group on Myocardial Function of the European Society of Cardiology. *Eur J Heart Fail* 20(2):216–227. <https://doi.org/10.1002/ehf.1059>
- Paulus WJ, Tschope C (2013) A novel paradigm for heart failure with preserved ejection fraction: Comorbidities drive myocardial dysfunction and remodeling through coronary microvascular endothelial inflammation. *J Am Coll Cardiol* 62(4):263–271. <https://doi.org/10.1016/j.jacc.2013.02.092>
- Fraccarollo D, Galuppo P, Motschenbacher S et al (2014) Soluble guanylyl cyclase activation improves progressive cardiac remodeling and failure after myocardial infarction. Cardioprotection over ACE inhibition. *Basic Res Cardiol* 109(4):421. <https://doi.org/10.1007/s00395-014-0421-1>
- Schafer A, Galuppo P, Fraccarollo D et al (2010) Increased cytochrome P4502E1 expression and altered hydroxyeicosatetraenoic acid formation mediate diabetic vascular dysfunction: Rescue by guanylyl-cyclase activation. *Diabetes* 59(8):2001–2009. <https://doi.org/10.2337/db09-1668>
- Fraccarollo D, Widder JD, Galuppo P et al (2008) Improvement in left ventricular remodeling by the endothelial nitric oxide synthase enhancer AVE9488 after experimental myocardial infarction. *Circulation* 118(8):818–827. <https://doi.org/10.1161/CIRCULATIONAHA.107.717702>
- Schafer A, Fraccarollo D, Widder J et al (2009) Inhibition of platelet activation in rats with severe congestive heart failure by a novel endothelial nitric oxide synthase transcription enhancer. *Eur J Heart Fail* 11(4):336–341. <https://doi.org/10.1093/eurjhf/hfp005>
- Gevaert AB, Shakeri H, Leloup AJ et al (2017) Endothelial senescence contributes to heart failure with preserved ejection fraction in an aging mouse model. *Circ Heart Fail*. <https://doi.org/10.1161/CIRCHEARTFAILURE.116.003806>
- Thum T, Hoerber S, Froese S et al (2007) Age-dependent impairment of endothelial progenitor cells is corrected by growth-hormone-mediated increase of insulin-like growth-factor-1. *Circ Res* 100(3):434–443. <https://doi.org/10.1161/01.RES.0000257912.78915.af>
- Eiserich JP, Baldus S, Brennan ML et al (2002) Myeloperoxidase, a leukocyte-derived vascular NO oxidase. *Science* 296(5577):2391–2394. <https://doi.org/10.1126/science.1106830>
- Askari AT, Brennan ML, Zhou X et al (2003) Myeloperoxidase and plasminogen activator inhibitor 1 play a central role in ventricular remodeling after myocardial infarction. *J Exp Med* 197(5):615–624
- Rudolph V, Andrie RP, Rudolph TK et al (2010) Myeloperoxidase acts as a profibrotic mediator of atrial fibrillation. *Nat Med* 16(4):470–474. <https://doi.org/10.1038/nm.2124>
- Haddad S, Wang Y, Galy B et al (2017) Iron-regulatory proteins secure iron availability in cardiomyocytes to prevent heart failure. *Eur Heart J* 38(5):362–372. <https://doi.org/10.1093/eurheartj/ehw333>
- Hoes MF, Grote Beverborg N, Kijlstra JD et al (2018) Iron deficiency impairs contractility of human cardiomyocytes through decreased mitochondrial function. *Eur J Heart Fail* 20(5):910–919. <https://doi.org/10.1002/ehf.1154>
- Melenovsky V, Petrak J, Mracek T et al (2017) Myocardial iron content and mitochondrial function in human heart failure: A direct tissue analysis. *Eur J Heart Fail* 19(4):522–530. <https://doi.org/10.1002/ehf.640>
- Aldosari S, Awad M, Harrington EO et al (2018) Subcellular reactive oxygen species (ROS) in cardiovascular pathophysiology. Antioxidants (Basel). <https://doi.org/10.3390/antiox7010014>
- Sorop O, Heinonen I, van Kranenburg M et al (2018) Multiple common comorbidities produce left ventricular diastolic dysfunction associated with coronary microvascular dysfunction, oxidative stress, and myocardial stiffening. *Cardiovasc Res* 114(7):954–964. <https://doi.org/10.1093/cvr/cvy038>
- Riehle C, Bauersachs J (2018) Of mice and men: models and mechanisms of diabetic cardiomyopathy. *Basic Res Cardiol* 114(1):2. <https://doi.org/10.1007/s00395-018-0711-0>
- Riehle C, Abel ED (2016) Insulin signaling and heart failure. *Circ Res* 118(7):1151–1169. <https://doi.org/10.1161/CIRCRESAHA.116.306206>
- Riehle C, Abel ED (2014) Insulin regulation of myocardial autophagy. *Circ J* 78(11):2569–2576
- Riehle C, Wende AR, Sena S et al (2013) Insulin receptor substrate signaling suppresses neonatal autophagy in the heart. *J Clin Invest* 123(12):5319–5333. <https://doi.org/10.1172/JCI71171>
- Dick SA, Epelman S (2016) Chronic heart failure and inflammation: What do we really know? *Circ Res* 119(1):159–176. <https://doi.org/10.1161/CIRCRESAHA.116.308030>
- Zhang Y, Bauersachs J, Langer HF (2017) Immune mechanisms in heart failure. *Eur J Heart Fail* 19(11):1379–1389. <https://doi.org/10.1002/ehf.942>
- Nahrendorf M (2018) Myeloid cell contributions to cardiovascular health and disease. *Nat Med* 24(6):711–720. <https://doi.org/10.1038/s41591-018-0064-0>
- Hulsmans M, Sam F, Nahrendorf M (2016) Monocyte and macrophage contributions to cardiac remodeling. *J Mol Cell Cardiol* 93:149–155. <https://doi.org/10.1016/j.jmcc.2015.11.015>
- Horckmans M, Ring L, Duchene J et al (2017) Neutrophils orchestrate post-myocardial infarction healing by polarizing macrophages towards a reparative phenotype. *Eur Heart J* 38(3):187–197. <https://doi.org/10.1093/eurheartj/ehw002>
- Nahrendorf M, Swirski FK, Aikawa E et al (2007) The healing myocardium sequentially mobilizes two monocyte subsets with divergent and complementary functions. *J Exp Med* 204(12):3037–3047. <https://doi.org/10.1084/jem.20070885>
- Dutta P, Nahrendorf M (2015) Monocytes in myocardial infarction. *Arterioscler Thromb Vasc Biol* 35(5):1066–1070. <https://doi.org/10.1161/ATVBAHA.114.304652>
- Fu X, Khalil H, Kanisak O et al (2018) Specialized fibroblast differentiated states underlie scar formation in the infarcted mouse heart. *J Clin Invest* 128(5):2127–2143. <https://doi.org/10.1172/JCI98215>
- Devaux B, Scholz D, Hirche A et al (1997) Upregulation of cell adhesion molecules and the presence of low grade inflammation in human chronic heart failure. *Eur Heart J* 18(3):470–479
- Ismahil MA, Hamid T, Bansal SS et al (2014) Remodeling of the mononuclear phagocyte network underlies chronic inflammation and disease progression in heart failure: Critical importance of the cardiosplenic axis. *Circ Res* 114(2):266–282. <https://doi.org/10.1161/CIRCRESAHA.113.301720>
- van Diepen S, Newby LK, Lopes RD et al (2013) Prognostic relevance of baseline pro- and anti-

- inflammatory markers in STEMI: An APEX AMI substudy. *Int J Cardiol* 168(3):2127–2133. <https://doi.org/10.1016/j.ijcard.2013.01.004>
36. Larose E, Rodes-Cabau J, Pibarot P et al (2010) Predicting late myocardial recovery and outcomes in the early hours of ST-segment elevation myocardial infarction: traditional measures compared with microvascular obstruction, salvaged myocardium, and necrosis characteristics by cardiovascular magnetic resonance. *J Am Coll Cardiol* 55(22):2459–2469. <https://doi.org/10.1016/j.jacc.2010.02.033>
 37. Vieira JM, Norman S, Villa Del Campo C et al (2018) The cardiac lymphatic system stimulates resolution of inflammation following myocardial infarction. *J Clin Invest* 128(8):3402–3412. <https://doi.org/10.1172/JCI97192>
 38. Jernberg T, Hasvold P, Henriksson M et al (2015) Cardiovascular risk in post-myocardial infarction patients: Nationwide real world data demonstrate the importance of a long-term perspective. *Eur Heart J* 36(19):1163–1170. <https://doi.org/10.1093/eurheartj/ehu505>
 39. Moccetti F, Brown E, Xie A et al (2018) Myocardial infarction produces sustained proinflammatory endothelial activation in remote arteries. *J Am Coll Cardiol* 72(9):1015–1026. <https://doi.org/10.1016/j.jacc.2018.06.044>
 40. Dutta P, Courties G, Wei Y et al (2012) Myocardial infarction accelerates atherosclerosis. *Nature* 487(7407):325–329. <https://doi.org/10.1038/nature11260>
 41. Galuppo P, Vettorazzi S, Hovelmann J et al (2017) The glucocorticoid receptor in monocyte-derived macrophages is critical for cardiac infarct repair and remodeling. *FASEB J* 31(11):5122–5132. <https://doi.org/10.1096/fj.201700317R>
 42. Frantz S, Hofmann U, Fraccarollo D et al (2013) Monocytes/macrophages prevent healing defects and left ventricular thrombus formation after myocardial infarction. *FASEB J* 27(3):871–881. <https://doi.org/10.1096/fj.12-214049>
 43. Wollenweber T, Roentgen P, Schafer A et al (2014) Characterizing the inflammatory tissue response to acute myocardial infarction by clinical multimodality noninvasive imaging. *Circ Cardiovasc Imaging* 7(5):811–818. <https://doi.org/10.1161/CIRCIMAGING.114.001689>
 44. Thackeray JT, Hupe HC, Wang Y et al (2018) Myocardial inflammation predicts remodeling and neuroinflammation after myocardial infarction. *J Am Coll Cardiol* 71(3):263–275. <https://doi.org/10.1016/j.jacc.2017.11.024>
 45. Prabhu SD, Frangogiannis NG (2016) The biological basis for cardiac repair after myocardial infarction: From inflammation to fibrosis. *Circ Res* 119(1):91–112. <https://doi.org/10.1161/CIRCRESAHA.116.303577>
 46. Thum T, Bauersachs J, Poole-Wilson PA et al (2005) The dying stem cell hypothesis: Immune modulation as a novel mechanism for progenitor cell therapy in cardiac muscle. *J Am Coll Cardiol* 46(10):1799–1802. <https://doi.org/10.1016/j.jacc.2005.07.053>
 47. Bauersachs J, Jaissner F, Toto R (2015) Mineralocorticoid receptor activation and mineralocorticoid receptor antagonist treatment in cardiac and renal diseases. *Hypertension* 65(2):257–263. <https://doi.org/10.1161/HYPERTENSIONAHA.114.04488>
 48. Yoshida M, Ma J, Tomita T et al (2005) Mineralocorticoid receptor is overexpressed in cardiomyocytes of patients with congestive heart failure. *Congest Heart Fail* 11(1):12–16
 49. Pitt B, Zannad F, Remme WJ et al (1999) The effect of spironolactone on morbidity and mortality in patients with severe heart failure. Randomized Aldactone Evaluation Study Investigators. *N Engl J Med* 341(10):709–717. <https://doi.org/10.1056/NEJM199909023411001>
 50. Pitt B, Remme W, Zannad F et al (2003) Eplerenone, a selective aldosterone blocker, in patients with left ventricular dysfunction after myocardial infarction. *N Engl J Med* 348(14):1309–1321. <https://doi.org/10.1056/NEJMoa030207>
 51. Zannad F, McMurray JJ, Krum H et al (2011) Eplerenone in patients with systolic heart failure and mild symptoms. *N Engl J Med* 364(1):11–21. <https://doi.org/10.1056/NEJMoa1009492>
 52. Adamopoulos C, Ahmed A, Fay R et al (2009) Timing of eplerenone initiation and outcomes in patients with heart failure after acute myocardial infarction complicated by left ventricular systolic dysfunction: Insights from the EPHESUS trial. *Eur J Heart Fail* 11(11):1099–1105. <https://doi.org/10.1093/eurjhf/hfp136>
 53. Beygui F, Van Belle E, Ecollan P et al (2018) Individual participant data analysis of two trials on aldosterone blockade in myocardial infarction. *Heart*. <https://doi.org/10.1136/heartjnl-2018-312950>
 54. Sun Y, Zhang J, Lu L et al (2004) Tissue angiotensin II in the regulation of inflammatory and fibrogenic components of repair in the rat heart. *J Lab Clin Med* 143(1):41–51. <https://doi.org/10.1016/S0022214303001914>
 55. Lopez-Andres N, Martin-Fernandez B, Rossignol P et al (2011) A role for cardiotrophin-1 in myocardial remodeling induced by aldosterone. *Am J Physiol Heart Circ Physiol* 301(6):H2372–2382. <https://doi.org/10.1152/ajpheart.00283.2011>
 56. Johar S, Cave AC, Narayanapanicker A et al (2006) Aldosterone mediates angiotensin II-induced interstitial cardiac fibrosis via a Nox2-containing NADPH oxidase. *FASEB J* 20(9):1546–1548. <https://doi.org/10.1096/fj.05-4642fje>
 57. Usher MG, Duan SZ, Ivaschenko CY et al (2010) Myeloid mineralocorticoid receptor controls macrophage polarization and cardiovascular hypertrophy and remodeling in mice. *J Clin Invest* 120(9):3350–3364. <https://doi.org/10.1172/JCI41080>
 58. Lother A, Berger S, Gilsbach R et al (2011) Ablation of mineralocorticoid receptors in myocytes but not in fibroblasts preserves cardiac function. *Hypertension* 57(4):746–754. <https://doi.org/10.1161/HYPERTENSIONAHA.110.163287>
 59. Fraccarollo D, Galuppo P, Schraut S et al (2008) Immediate mineralocorticoid receptor blockade improves myocardial infarct healing by modulation of the inflammatory response. *Hypertension* 51(4):905–914. <https://doi.org/10.1161/HYPERTENSIONAHA.107.100941>
 60. Fraccarollo D, Galuppo P, Sieweke JT et al (2015) Efficacy of mineralocorticoid receptor antagonism in the acute myocardial infarction phase: Eplerenone versus spironolactone. *ESC Heart Fail* 2(3):150–158. <https://doi.org/10.1002/ehf2.12053>
 61. Fraccarollo D, Berger S, Galuppo P et al (2011) Deletion of cardiomyocyte mineralocorticoid receptor ameliorates adverse remodeling after myocardial infarction. *Circulation* 123(4):400–408. <https://doi.org/10.1161/CIRCULATIONAHA.110.983023>
 62. Vallon V, Wyatt AW, Klingel K et al (2006) SGK1-dependent cardiac CTGF formation and fibrosis following DOCA treatment. *J Mol Med* 84(5):396–404. <https://doi.org/10.1007/s00109-005-0027-z>
 63. Fraccarollo D, Thomas S, Scholz CJ et al (2018) Macrophage mineralocorticoid receptor is a pleiotropic modulator of myocardial infarct healing. *Hypertension* 73(1):102–111. <https://doi.org/10.1161/HYPERTENSIONAHA.118.12162>
 64. Panahi M, Papanikolaou A, Torabi A et al (2018) Immunomodulatory interventions in myocardial infarction and heart failure: A systematic review of clinical trials and meta-analysis of IL-1 inhibition. *Cardiovasc Res* 114(11):1445–1461. <https://doi.org/10.1093/cvr/cvy145>
 65. Maisch B, Alter P (2018) Treatment options in myocarditis and inflammatory cardiomyopathy: Focus on i.v. immunoglobulins. *Herz* 43(5):423–430. <https://doi.org/10.1007/s00059-018-4719-x>
 66. Chung ES, Packer M, Lo KH et al (2003) Randomized, double-blind, placebo-controlled, pilot trial of infliximab, a chimeric monoclonal antibody to tumor necrosis factor- α , in patients with moderate-to-severe heart failure: Results of the anti-TNF Therapy Against Congestive Heart Failure (ATTACH) trial. *Circulation* 107(25):3133–3140. <https://doi.org/10.1161/01.CIR.0000077913.60364.D2>
 67. Mann DL, McMurray JJ, Packer M et al (2004) Targeted anticytokine therapy in patients with chronic heart failure: Results of the Randomized Etanercept Worldwide Evaluation (RENEWAL). *Circulation* 109(13):1594–1602. <https://doi.org/10.1161/01.CIR.0000124490.27666.B2>
 68. Heymans S, Hirsch E, Anker SD et al (2009) Inflammation as a therapeutic target in heart failure? A scientific statement from the Translational Research Committee of the Heart Failure Association of the European Society of Cardiology. *Eur J Heart Fail* 11(2):119–129. <https://doi.org/10.1093/eurjhf/hfn043>
 69. Gullestad L, Ueland T, Fjeld JG et al (2005) Effect of thalidomide on cardiac remodeling in chronic heart failure: Results of a double-blind, placebo-controlled study. *Circulation* 112(22):3408–3414. <https://doi.org/10.1161/CIRCULATIONAHA.105.564971>
 70. Skudicky D, Bergemann A, Sliwa K et al (2001) Beneficial effects of pentoxifylline in patients with idiopathic dilated cardiomyopathy treated with angiotensin-converting enzyme inhibitors and carvedilol: Results of a randomized study. *Circulation* 103(8):1083–1088
 71. Sliwa K, Skudicky D, Candy G et al (1998) Randomised investigation of effects of pentoxifylline on left-ventricular performance in idiopathic dilated cardiomyopathy. *Lancet* 351(9109):1091–1093. [https://doi.org/10.1016/S0140-6736\(97\)09338-0](https://doi.org/10.1016/S0140-6736(97)09338-0)
 72. Sliwa K, Woodiwiss A, Candy G et al (2002) Effects of pentoxifylline on cytokine profiles and left ventricular performance in patients with decompensated congestive heart failure secondary to idiopathic dilated cardiomyopathy. *Am J Cardiol* 90(10):1118–1122
 73. Sliwa K, Woodiwiss A, Kone VN et al (2004) Therapy of ischemic cardiomyopathy with the immunomodulating agent pentoxifylline: Results of a randomized study. *Circulation* 109(6):750–755. <https://doi.org/10.1161/01.CIR.0000112568.48837.60>
 74. Bahrmann P, Hengst UM, Richartz BM, Figulla HR (2004) Pentoxifylline in ischemic, hypertensive and idiopathic-dilated cardiomyopathy: Effects on left-ventricular function, inflammatory cytokines and

- symptoms. Eur J Heart Fail 6(2):195–201. <https://doi.org/10.1016/j.ejheart.2003.09.005>
75. Kjekshus J, Apetrei E, Barrios V et al (2007) Rosuvastatin in older patients with systolic heart failure. N Engl J Med 357(22):2248–2261. <https://doi.org/10.1056/NEJMoa0706201>
 76. Tavazzi L, Maggioni AP, Marchioli R et al (2008) Effect of rosuvastatin in patients with chronic heart failure (the GISSI-HF trial): A randomised, double-blind, placebo-controlled trial. Lancet 372(9645):1231–1239. [https://doi.org/10.1016/S0140-6736\(08\)61240-4](https://doi.org/10.1016/S0140-6736(08)61240-4)
 77. Krum H, Ashton E, Reid C et al (2007) Double-blind, randomized, placebo-controlled study of high-dose HMG CoA reductase inhibitor therapy on ventricular remodeling, pro-inflammatory cytokines and neurohormonal parameters in patients with chronic systolic heart failure. J Card Fail 13(1):1–7. <https://doi.org/10.1016/j.cardfail.2006.09.008>
 78. Torre-Amione G, Anker SD, Bourge RC et al (2008) Results of a non-specific immunomodulation therapy in chronic heart failure (ACCLAIM trial): A placebo-controlled randomised trial. Lancet 371(9608):228–236. [https://doi.org/10.1016/S0140-6736\(08\)60134-8](https://doi.org/10.1016/S0140-6736(08)60134-8)
 79. Gullestad L, Aass H, Fjeld JG et al (2001) Immunomodulating therapy with intravenous immunoglobulin in patients with chronic heart failure. Circulation 103(2):220–225
 80. McNamara DM, Holubkov R, Starling RC et al (2001) Controlled trial of intravenous immune globulin in recent-onset dilated cardiomyopathy. Circulation 103(18):2254–2259
 81. Parrillo JE, Cunnion RE, Epstein SE et al (1989) A prospective, randomized, controlled trial of prednisone for dilated cardiomyopathy. N Engl J Med 321(16):1061–1068. <https://doi.org/10.1056/NEJM198910193211601>
 82. Moreira DM, Vieira JL, Gottschall CA (2009) The effects of METHotrexate therapy on the physical capacity of patients with ISchemic heart failure: A randomized double-blind, placebo-controlled trial (METIS trial). J Card Fail 15(10):828–834. <https://doi.org/10.1016/j.cardfail.2009.06.439>

Deutscher Herzbericht: Verbesserte Behandlungsmöglichkeiten führen zu einem Rückgang der Sterberate

Obwohl die Häufigkeit der Herzinsuffizienz anstieg, gelang es, die Sterberate von Herzinsuffizienz-Patienten um nahezu 11 % zu senken. „Das ist natürlich ein großartiger Erfolg, der nicht ohne Grund erreicht wurde“, erklärt Katus. „Zum einen haben wir deutliche Fortschritte bei den Behandlungsmöglichkeiten sowohl im medikamentösen als auch im interventionellen und technologischen Bereich erreicht. Zum anderen konnten wir in der Ärzteschaft das Bewusstsein dafür verbessern, das für die Herzinsuffizienz und die damit einhergehenden Rhythmusstörungen bessere Behandlungsmöglichkeiten zur Verfügung stehen.“ So wurden mehr Patienten bei niedergelassenen Kardiologen und in den Kliniken vorstellig.

Zertifizierte Patientenversorgung

Die Etablierung spezialisierter und durch die DGK initiiertes und zertifizierter Versorgungseinheiten, den sogenannten „Heart Failure Units“, hat ganz wesentlich dazu beigetragen. Dennoch stellt die Herzschwäche weiterhin die häufigste Ursache für eine stationäre Behandlung in Deutschland dar. Dies wird voraussichtlich künftig weiter zunehmen. Der Qualitätsoffensive durch die Zertifizierungen der DGK kommt somit auf dem Weg zu einer verbesserten Versorgung von Patienten mit Herzinsuffizienz eine enorme Wichtigkeit zu.

Ähnlich positive Behandlungserfolge können durch die nicht-invasive Herzklappenimplantation (TAVI) beim älteren Menschen mit erhöhtem Risiko berichtet werden. Auch für die TAVIs hat die DGK gemeinsam mit der herzchirurgischen Fachgesellschaft DGTHG in Deutschland durch die Zertifizierung von spezialisierten TAVI-Zentren die Einhaltung von hohen qualitativen Standards initiiert und verbessert.

Weniger Herzinfarkte

Die DGK zeigt sich sehr erfreut, dass das Auftreten der ischämischen Herzerkrankungen, also Ereignisse, die auf Durchblutungsstörungen am Herzen zurückzuführen sind, abgenommen haben (-2,2 %). „Dies zeigt uns, dass unsere Präventionsstrategien zu greifen beginnen, und das finde ich sehr

beeindruckend“, so der DGK-Präsident. „Dies ist das beste Beispiel für den hohen Stellenwert der Vorbeugung von Herz-Kreislauf-Krankheiten.“

Nach den großen Erfolgen in den Jahren zuvor ist die Sterberate nach einem Herzinfarkt jedoch seit einiger Zeit nahezu unverändert geblieben. Sie sank 2017 um nur 0,6 %. „Wir scheinen hier ein Plateau erreicht zu haben. Dies zeigen uns auch Daten aus Schweden. Dort konnte die Sterblichkeit aufgrund eines Herzinfarktes zuletzt auch kaum noch gesenkt werden. Wir müssen daher neue alternative Therapieansätze entwickeln, um die Sterberate im Herzinfarkt noch weiter senken zu können“, fordert er.

**Deutscher Herzbericht
Presstext DGK 02/2019**