



Three Immune-Related Prognostic mRNAs as Therapeutic Targets for Pancreatic Cancer

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Objective: Pancreatic cancer is a highly lethal malignancy globally. This study aimed to probe and validate immune-related prognostic mRNAs as therapeutic targets for pancreatic cancer.

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Zhang C, Zou Y, Zhu Y, Liu Y, Feng H, Niu F, He P and Liu H (2021) Three Immune-Related Prognostic mRNAs as Therapeutic Targets for Pancreatic Cancer. Front. Med. 8:649326. doi: 10.3389/fmed.2021.649326 **Methods:** Gene transcriptome data of pancreatic cancer and normal pancreas were retrieved from TCGA-GTEx projects. Two thousand four hundred and ninety-eight immune-related genes were obtained from the IMMUPORT database. Abnormally expressed immune-related genes were then identified. Under univariate and multivariate cox models, a gene signature was constructed. Its predictive efficacy was assessed *via* ROCs. The interactions between the 21 genes were analyzed by Spearson analysis and PPI network. Using the GEPIA and The Human Protein Atlas databases, their expression and prognostic value were evaluated. The TIMER database was utilized to determine the relationships between MET, OAS1, and OASL mRNAs and immune infiltrates. Finally, their mRNA expression was externally verified in the GSE15471 and GSE62452 datasets.

Results: An immune-related 21-gene signature was developed for predicting patients' prognosis. Following verification, this signature exhibited the well predictive performance. There were physical and functional interactions between them. MET, OAS1, and OASL mRNAs were all up-regulated in pancreatic cancer and associated with unfavorable prognosis. They showed strong correlations with tumor progression. Furthermore, the three mRNAs were distinctly associated with immune infiltrates. Their up-regulation was confirmed in the two external datasets.

Conclusion: These findings identified three immune-related prognostic mRNAs MET, OAS1, and OASL, which may assist clinicians to choose targets for immunotherapy and make personalized treatment strategy for pancreatic cancer patients.

Keywords: pancreatic cancer, met, OAS1, OASL, therapeutic targets, prognosis

INTRODUCTION

Pancreatic cancer is a highly lethal disease with fairly high mortality globally (1). Annual mortality is nearly equal to incidence in many countries such as China (2). The 5-year survival is \sim 7% (3). Its unfavorable prognosis is mainly attributed to local infiltration and distant metastases. Pancreatic ductal adenocarcinoma accounts for around 95% of all cases. Only 10% of cases are

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due to genetic factors (4). Smoking, drinking, and obesity are common modifiable risk factors for this disease (5). Routine therapy methods like surgery offer dissatisfactory clinical outcomes. Only \sim 20% could benefit from curative surgical resection (6). Recently, immunotherapy has become a promising adjunct treatment for pancreatic cancer (7). Nevertheless, most of pancreatic cancer patients are resistant to most therapies including immunotherapy because tumors may evade immune surveillance (8). Moreover, there is currently no targeted therapy against driver genes in pancreatic cancer. Thus, it is of necessity to develop novel strategies to build up treatment efficacy.

High-throughput sequencing may assist us to probe therapeutic targets for cancer therapies and understand the underlying mechanisms of the anti-cancer efficacy in depth (9). Pancreatic cancer is featured by distinct immune disorders. Components in immune system contribute to the initiation and development of pancreatic cancer (10). Integrated analysis of relationships between immune-related genes and clinical outcomes of pancreatic cancer is critical to explore novel prognostic markers as well as therapeutic targets. For example, Wu et al. found that three immune-related genes CKLF, ERAP2, and EREG showed distinct correlations with pancreatic cancer patients' survival (11). In this study, we identified three immune-related prognostic genes MET, OAS1, and OASL from the 21-gene signature. Following multiple dataset verification, these genes could be promising therapeutic targets as well as prognostic markers in pancreatic cancer.

MATERIALS AND METHODS

Downloaded Datasets

Gene transcriptome data of 165 normal pancreas samples from the Genotype-Tissue Expression (GTEx) project and 178 pancreatic cancer samples from The Cancer Genome Atlas (TCGA) database were downloaded based on the UCSC Xena browser (https://xenabrowser.net/datapages/). Clinical information of these pancreatic cancer patients including gender, pathologic T, N and histologic grade was retrieved from Genomic Data Commons (GDC). Those without available follow-up data were removed. **Table 1** listed the clinical features for each patient.

Two microarray datasets GSE15471 and GSE62452 were downloaded from the Gene Expression Omnibus (GEO) database (https://www.ncbi.nlm.nih.gov/gds/). Of which, the GSE15471 dataset included 39 pairs of pancreatic cancer and normal tissues. Meanwhile, the GSE62452 dataset contained 69 pancreatic cancer and adjacent normal tissue specimens (12). **TABLE 1** | Clinical features of 178 pancreatic cancer patients from TCGA database.

Clinical features	Number
Gender	
Male	102
Female	83
Pathologic T	
Τ1	7
Τ2	24
ТЗ	148
Τ4	4
Tx	1
Pathologic N	
NO	50
N1	126
N1b	4
٨x	4
listologic grade	
31	32
G2	97
G3	51
G4	2
Эх	3

Screening for Abnormally Expressed Immune-Related Genes

After merging gene matrix of GTEx and TCGA datasets based on the Ensembl IDs, differentially expressed genes (DEGs) between pancreatic cancer and normal pancreas specimens were screened *via* the Linear Models for Microarray Data (limma) package in R (13). The screening criteria were as follows: $|\log$ fold change (FC)| > 2 and false discovery rate (FDR) < 0.05. Two thousand four hundred and ninety-eight immune-related genes were obtained from the IMMUPORT database (https:// www.immport.org/home). Following integration of DEGs and immune-related genes, abnormally expressed immune-related genes were identified for pancreatic cancer.

An Immune-Related Gene Signature Construction

For differentially expressed immune-related genes, univariate Cox regression analysis was conducted using TCGA dataset. Genes with p < 0.001 were considered related to pancreatic cancer prognosis. Then, candidate genes were screened *via* multivariate Cox regression analysis. Based on them, a risk score was established according to the following formula: risk score = $\beta_1 x_1 + \beta_2 x_2 + \ldots + \beta_i x_i$ (where β_i indicates the coefficient of gene i, and x_i indicates the expression level of gene i). The risk score of each patient was calculated and all patients were separated into high- and low-risk groups in accordance with the median value. Kaplan–Meier survival analysis was conducted between high- and low-risk groups through the survival package in R. The Receiver Operating

Abbreviations: GTEx, Genotype-Tissue Expression; TCGA, The Cancer Genome Atlas; GDC, Genomic Data Commons; GEO, Gene Expression Omnibus; DEGs, differentially expressed genes; limma, Linear Models for Microarray Data; FC, fold change; FDR, false discovery rate; ROCs, Receiver Operating Characteristic curves; AUCs, Areas under the curves; GO, Gene ontology; BP, biological processes; MF, molecular functions; CC, cellular components; KEGG, Kyoto Encyclopedia of Genes and Genomes; PPI, Protein-protein interaction; STRING, Search Tool for the Retrieval of Interacting Genes Database; GEPIA, Gene Expression Profiling Interactive Analysis; OS, overall survival; DFS, disease-free survival; HR, hazard ratio; CI, confidence interval.

Characteristic curves (ROCs) for overall survival were drawn utilizing the survivalROC package in R (14). Areas under the curves (AUCs) were calculated for detection of the efficacy to predict survival for the signature and other clinical features (age, gender, grade, pathologic T, pathologic N, and stage). Univariate and multivariate cox regression analyses were utilized to assess whether risk score could be independently predictive of patients' survival. Hazard ratio (HR), 95% confidence interval (CI) and p were calculated. HR > 1 indicated risk factors and HR < 1 indicated protective factors.

Functional Annotation Analysis

Gene ontology (GO) including biological processes (BP), molecular functions (MF) and cellular components (CC) and Kyoto Encyclopedia of Genes and Genomes (KEGG) functional annotation analyses of the survival-related genes were presented for exploring underlying biological functions *via* the clusterProfiler package in R (15). Terms with adjusted p < 0.05 were significantly enriched.

Protein-Protein Interaction (PPI) Analysis

The physical and functional interactions of proteins from the immune-related gene signature were analyzed *via* the Search Tool for the Retrieval of Interacting Genes Database (STRING) database (http://string-db.org/) (16). The degree of nodes was then calculated.

Gene Expression Profiling Interactive Analysis (GEPIA)

The expression of MET, OAS1, and OASL in pancreatic cancer and normal samples was confirmed using TCGA-GTEx projects *via* the online GEPIA database (http://gepia.cancer-pku.cn/ index.html) (17). Furthermore, overall survival (OS) and diseasefree survival (DFS) analyses were generated for high and low expression of MET, OAS1, or OASL groups among pancreatic cancer patients.

Immunohistochemistry and Immunofluorescence

Immunohistochemistry of MET, OAS1, and OASL proteins in pancreatic cancer and normal pancreas specimens was obtained from The Human Protein Atlas (https://www.proteinatlas. org/). Furthermore, their immunofluorescence images were also downloaded.

Immune Infiltration Analysis

The correlations between expression or copy number (armlevel deletion, diploid / normal, arm-level gain, and high amplification) of MET, OAS1, and OASL and the abundance of immune infiltrates composed of B cell, CD4⁺ T cell, CD8⁺ T cell, neutrophil, macrophage, and dendritic cell were assessed using the TIMER database (https://cistrome.shinyapps.io/timer/ (18)).

Statistical Analysis

All analyses were presented utilizing R version 3.5.2. Spearman correlation analysis was used to assess the associations between genes from the gene signature. The strengths of correlations

were determined as follows: 0–0.39: weak; 0.40–0.59: moderate; 0.60–0.79: strong; 0.80–1.0: very strong. The differences in gene expression between two subgroups were calculated using the Wilcox test. P < 0.05 indicated statistical significance.

RESULTS

Construction of a Prognostic Immune-Related Gene Signature for Pancreatic Cancer

From TCGA-GTEx datasets, 1,737 DEGs with |log FC| > 2 and FDR < 0.05 were identified for pancreatic cancer (n = 178) than normal pancreas specimens (n = 165), which were composed of 962 down- and 775 up-regulated genes listed in Supplementary Table 1. Among all DEGs, 229 were immune-related genes (Figure 1A; Supplementary Table 2). In Figure 1B, 50 abnormally expressed immune-related genes were significantly related to prognosis of pancreatic cancer. Using multivariate Cox regression analysis, 21 candidate genes were identified for constructing a prognostic immune-related gene signature (Table 2). The risk score was calculated for each patient by combining coefficient and expression level. One hundred and seventy-eight patients were separated into high- and lowrisk groups in accordance with the median value. Those in the high-risk group exhibited an unfavorable prognosis (p = 1.499e-14; Figure 1C). To assess whether the risk score accurately and sensitively predicted patients' survival, ROCs were established. In Figure 1D, the AUC of the risk score for overall survival was 0.833, which was much higher than other clinicopathological factors, suggesting that the risk score possessed high accuracy in predicting survival.

The Immune-Related Gene Signature as an Independent Prognostic Factor for Pancreatic Cancer

One hundred and seventy-eight pancreatic cancer patients were ranked according to their risk scores (**Figure 2A**). As the risk scores elevated, the number of dead patients gradually increased (**Figure 2B**). **Figure 2C** depicted the expression patterns of these 21 genes in high and low-risk groups. As shown in univariate cox regression analysis, age [HR (95% CI): 1.027 (1.005–1.049), p = 0.018], N [HR (95% CI): 2.180 (1.283–3.706), p = 0.004] and risk score [HR (95% CI): 1.117 (1.078–1.157), p < 0.001] were risk factors for pancreatic cancer (**Figure 2D**). Multivariate cox regression analysis revealed that risk score was an independent predictive factor for prognosis of pancreatic cancer (**Figure 2E**).

Enrichment Analysis for Survival-Related Genes

We further probed biological functions of the survival-related genes. In **Figure 3A**, these genes were distinctly associated with regulation of migration of multiple cells such as epithelial and endothelial cells. Also, they could be involved in key cellular components like endoplasmic reticulum lumen and specific granule and possess different molecular functions such as receptor activity, cytokine binding and growth factor





activity. As shown in KEGG enrichment analysis, these genes could participate in ErbB and proteoglycans in cancer pathways (**Figure 3B**).

Interactions Between Genes From the Immune-Related Gene Signature

In **Figure 4A**, genes from immune-related gene signature were all abnormally expressed in pancreatic cancer than normal samples (all p < 0.001). Except for IL22RA1 and PAK3, most of them were up-regulated in pancreatic cancer. We

further calculated their correlations at the expression levels using Spearson analysis, as shown in **Figure 4B**. S100A16 was strongly correlated to PAK3 (r = -0.68), SDC4 (r = 0.64), PLAUR (r = 0.74), MET (r = 0.66), and TMSB10 (r = 0.6). PAK3 had strong correlations with PLAUR (r = -0.64) and TMSB10 (r = -0.61). SDC4 exhibited a strong association with MET (r = 0.7). OASL was very strongly associated with OAS1 (r = 0.81). PLAUR showed a strong relationship with TMSB10 (r = 0.69). The PPI network confirmed the closely interactions between them (**Figure 4C**). **Figure 4D** listed the

TABLE 2 | Multivariate cox regression analysis for 21 candidate genes in pancreatic cancer.

ID	Coefficients	Risk ratio	95% low	95% high	Р
B2M	-0.84756	0.42846	0.269333	0.681603	0.000346
PI3	0.126991	1.135407	1.003735	1.284353	0.043462
CXCL9	0.484476	1.623325	1.293371	2.037454	2.93E-05
TMSB10	0.359616	1.43278	0.933127	2.199975	0.100254
LCN2	0.178297	1.19518	1.01854	1.402454	0.028882
S100A16	0.873443	2.395142	1.571083	3.651434	4.91 <i>E</i> -05
OASL	0.511952	1.668545	1.105363	2.518668	0.014819
PTGS2	0.242464	1.274385	1.025728	1.583322	0.028572
OAS1	0.411634	1.509282	0.980257	2.323809	0.061566
LTBP1	0.566777	1.762578	1.172497	2.649628	0.006428
PLAUR	-1.28398	0.276934	0.146236	0.524442	8.11 <i>E</i> -05
AGT	-0.36056	0.697283	0.545836	0.890749	0.003902
AREG	0.340822	1.406103	1.086432	1.819832	0.0096
CMTM6	-0.64978	0.522162	0.291347	0.935836	0.029056
HBEGF	0.51271	1.669811	1.177601	2.367752	0.004009
SPP1	0.21316	1.237583	1.050946	1.457366	0.010596
IL22RA1	0.170068	1.185385	0.937948	1.498097	0.154535
MET	0.82481	2.281447	1.443826	3.605005	0.00041
NRP2	-0.59908	0.549317	0.341531	0.88352	0.013485
SDC4	-0.62306	0.536299	0.36083	0.797097	0.002059
PAK3	0.669971	1.95418	1.058236	3.608666	0.032287



FIGURE 2 | The immune-related gene signature as an independent prognostic factor for pancreatic cancer. (A) Risk score ranking. (B) Distribution of survival status including dead (red) and alive (green) according to risk scores. (C) Heat map for the expression patterns of 21 genes in high and low risk groups. (D) Univariate and (E) multivariate cox regression analyses for risk score and other clinical indicators.



degree of each node in the network. We found that SSP1 had the highest degree.

Up-Regulation of MET, OAS1, and OASL Is Associated With Poor Clinical Outcomes of Pancreatic Cancer

Among 21 genes from the immune-related signature, MET, OAS1, and OASL were significantly associated with prognosis of pancreatic cancer patients. Patients with high MET expression were indicative of shorter DFS (p = 0.00044; Figure 5A) and OS (p = 0.00023; Figure 5B) time than those with low expression. Furthermore, high OAS1 (p = 0.047; Figure 5C) and OASL (p = 0.0072; Figure 5D) expression was distinctly related to poorer OS. MET (Figure 5E), OAS1 (Figure 5F), and OASL (Figure 5G) were all up-regulated at the mRNA and protein levels. Immunofluorescence results demonstrated that MET (Figure 5H), OAS1 (Figure 5I), and OASL (Figure 5J) were mainly distributed in cytoplasm and nucleus. This study further assessed whether MET, OAS1, and OASL expression was in association with clinical features. The data showed that MET expression was significantly higher in G3-G4 (p = 0.005; Figure 6A) and T3-T4 (p = 0.012; Figure 6B). Furthermore, higher OAS1 expression was detected in N1 (p = 0.019; **Figure 6C**) and T3-T4 (p = 0.006; **Figure 6D**). There was higher OASL expression in >65 (p = 0.048; Figure 6E) or T3-T4 (p = 0.009; Figure 6F) patients. These findings indicated that MET, OAS1, and OASL might be related to pancreatic cancer progression.

MET, OAS1, and OASL Correlates With Immune Infiltrates

Utilizing the TIMER database, we analyzed the correlation of MET, OAS1, and OASL expression with immune infiltrates. Our data suggested that MET expression was significantly

correlated to B cell (r = 0.177, p = 2.08e-02), CD8 + T cell (r = 0.35, p = 2.64e-06), CD4 + T cell (r = -0.259, p = 6.68e-04), neutrophil (r = 0.21, p = 5.79e-03), and dendritic cell (r = 0.261, p = 5.69e-04) in **Figure 7A**. OAS1 expression exhibited significant associations with neutrophil (r = 0.302, p = 5.89e-05) and dendritic cell (r = 0.185, p = 1.54e-02; **Figure 7B**). OASL expression showed a significant correlation with neutrophil (r = 0.189, p = 1.32e-02; **Figure 7C**). Moreover, copy number of MET, OAS1, and OASL was also correlated to immune infiltrates. In **Figure 7D**, there were significant associations between copy number of MET and B cell and CD4 + T cell. Copy number of OAS1 (**Figure 7E**) and OASL (**Figure 7F**) was distinctly correlated to B cell, CD4 + T cell, CD4 + T cell, and neutrophil.

Validation of MET, OAS1, and OASL Expression in External Datasets

The expression of MET, OAS1, and OASL in pancreatic cancer was verified in the GSE15471 and GSE62452 datasets. Consistently, MET was up-regulated in pancreatic cancer both in the GSE15471 (p = 1.002e-09; Figure 8A) and GSE62452 datasets (p = 2.792e-11; Figure 8B). Higher OAS1 expression was detected in pancreatic cancer than paracarcinoma tissues in the GSE15471 (p = 3.519e-07; Figure 8C) and GSE62452 datasets (p = 1.215e-11; Figure 8D). Meanwhile, OASL expression was markedly elevated in tumor specimens in the GSE15471 (p = 1.328e-05; Figure 8E) and GSE62452 datasets (p = 7.153e-10; Figure 8F).

DISCUSSION

This study constructed an immune-related gene signature for predicting clinical outcomes of pancreatic cancer patients. We identified three key genes (MET, OAS1, and OASL) that were all



up-regulated in pancreatic cancer and indicated an unfavorable prognosis. Following multiple dataset verification, the three genes might be promising therapeutic targets, which were worthy of further research.

Despite the TNM stage system as an efficient tool to detect tumor stage, there is discrepancy in prognosis based on TNM stage (19). Thus, the efficacy of TNM stage is limited. Gene-based markers have been widely explored for pancreatic cancer in recent years (20). Recently, several prognosis-related gene signatures have been established for pancreatic cancer (21–23). For example, Zhuang et al. developed a prognosis-related lncRNA signature for pancreatic cancer (24). Following comparing other clinical risk factors, the signature exhibited better predictability. Wu et al. constructed 9-gene signature

for prediction of survival time of pancreatic cancer patients (25). There is still a lack of immune-related prognostic models. Herein, the immune-related 21-gene signature could accurately and sensitively predict survival time of pancreatic cancer patients. It performed better than other clinicopathological characteristics like age, gender, grade, N, T, and stage. This signature could be independently predictive of patients' prognosis.

The molecular mechanisms of highly aggressive behaviors remain unknown. We analyzed biological functions of survivalrelated genes. These genes could regulate migration of multiple cells such as epithelial and endothelial cells and were involved in key cellular components like endoplasmic reticulum lumen and specific granule and possess different molecular functions such as receptor activity, cytokine binding, and growth factor



MET, (C) OS of OAS1, and (D) OASL *in* the GEPIA. Up-regulation of (E) MET, (F) OAS1, and (G) OASL in pancreatic cancer specimens. The left picture shows the mRINA expression levels from the GEPIA and the right one shows the immunohistochemistry images from The Human Protein Atlas. Immunofluorescence images of (H) MET, (I) OAS1, and (J) OASL in cells. The nucleus was stained blue, the microtubules were stained red, and the MET OAS1 and OASL proteins were stained green. *P < 0.05.

activity. These data were indicative that these genes were involved in tumor progression. For example, CCN1/Cyr61 secreted by pancreatic cancer cells may promote migration of endothelial cells (26). Nerve growth factors regulate CD133 functions, thereby promoting migration of pancreatic cancer cells (27). Targeting IL-1 and its receptor can prolong survival time in pancreatic cancer (28). We also found that these genes could participate in ErbB and proteoglycans in cancer pathways. It has been confirmed that dysregulated ErbB signaling promotes tumorigenesis for pancreatic cancer (29). Hence, these survival-related genes might participate in pancreatic tumor progression.

Our study revealed that there were physical and functional interactions between 21 genes from the immune-related gene signature. Among them, MET, OAS1, and OASL were verified to be markedly up-regulated in pancreatic cancer and associated with poor clinical outcomes of patients. Furthermore, MET expression was significantly correlated to infiltration

of B cell, CD8 + T cell, CD4 + T cell, neutrophil, and dendritic cell. MET gene is located on chromosome 7q21-31. Changes in MET functions have been a hallmark of multiple cancers including pancreatic cancer (30). MET overexpression induces pancreatic cancer progression (31). Consistently, MET up-regulation was markedly correlated to tumor grade and T stage (32). Dysregulated MET functions correlate with aggressive phenotypes. Consistent with previous research, MET is involved in the crosstalk between tumor cells and tumor microenvironment (30). Thus, targeting MET has been considered as an adjuvant therapy in pancreatic cancer. OAS1 and OASL, 2'-5'-oligoadenylate synthetases, are interferon-induced antiviral enzymes. We found that OAS1 expression exhibited significant associations with neutrophil and dendritic cell infiltration. OASL expression showed a significant correlation with neutrophil infiltration. Consistently, Zhang et al. also found that OAS1 and OASL were correlated to neutrophil cell infiltration in breast cancer (33). Thus,



MET, OAS1, and OASL were distinctly correlated to tumor immune microenvironment.

Collectively, we identified three immune-related prognostic genes MET, OAS1, and OASL, which could be promising therapeutic targets as well as prognostic markers for pancreatic cancer. However, several limitations of this study should be pointed out. First, the biological functions of MET, OAS1, and OASL such as their interactions with immune cells in tumor microenvironment need to be explored *in vitro* experiments. Second, their prognostic values should be verified in prospective research.

CONCLUSION

In this study, we established an immune-related 21-gene signature for prediction of pancreatic cancer prognosis. This signature could be accurately and independently predictive of



FIGURE 7 | Association of MET, OAS1, and OASL with immune infiltrates *via* the TIMER database. (A–C) Correlation of (A) MET, (B) OAS1, and (C) OASL expression with immune infiltrates. (D–F) Correlation of copy number of (D) MET, (E) OAS1, and (F) OASL with immune infiltrates. *p < 0.05; **p < 0.01; ***p < 0.001.



expression in tumor (T) and normal (N) tissue specimens in GSE15471 and GSE62452 datasets.

patients' survival. Among these genes, MET, OAS1, and OASL were validated to be up-regulated in pancreatic cancer and associated with unfavorable prognosis of patients. Also, there were closely interactions between them and immune infiltrates. Thus, MET, OAS1, and OASL could be potential therapeutic markers in pancreatic cancer.

DATA AVAILABILITY STATEMENT

The	da	itasets	genera	ated	for	this	study
can	be	found	in	online		repositories.	The

names of the repository/repositories and accession number(s) can be found in the article/**Supplementary Materials**.

AUTHOR CONTRIBUTIONS

HL conceived and designed the study. CZ, FN, and PH conducted most of the experiments, data analysis, and wrote the manuscript. YZo, YZh, YL, and HF participated in collecting data and helped to draft the manuscript. All authors reviewed and approved the manuscript.

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REFERENCES

- GBD 2017 Pancreatic Cancer Collaborators. The global, regional, and national burden of pancreatic cancer and its attributable risk factors in 195 countries and territories, 1990-2017: a systematic analysis for the Global Burden of Disease Study 2017. *Lancet Gastroenterol Hepatol.* (2019) 4:934–47. doi: 10.1016/S2468-1253(19)30347-4
- Lin QJ, Yang F, Jin C, Fu DL. Current status and progress of pancreatic cancer in China. World J Gastroenterol. (2015) 21:7988–8003. doi: 10.3748/wjg.v21.i26.7988
- Karamitopoulou E. Tumour microenvironment of pancreatic cancer: immune landscape is dictated by molecular and histopathological features. *Br J Cancer*. (2019) 121:5–14. doi: 10.1038/s41416-019-0479-5
- Barone E, Corrado A, Gemignani F, Landi S. Environmental risk factors for pancreatic cancer: an update. Arch Toxicol. (2016) 90:2617–42. doi: 10.1007/s00204-016-1821-9
- Midha S, Chawla S, Garg PK. Modifiable and non-modifiable risk factors for pancreatic cancer: a review. *Cancer Lett.* (2016) 381:269–77. doi: 10.1016/j.canlet.2016.07.022
- Lai E, Puzzoni M, Ziranu P, Pretta A, Impera V, Mariani S, et al. New therapeutic targets in pancreatic cancer. *Cancer Treat Rev.* (2019) 81:101926. doi: 10.1016/j.ctrv.2019.101926
- Banerjee K, Kumar S, Ross KA, Gautam S, Poelaert B, Nasser MW, et al. Emerging trends in the immunotherapy of pancreatic cancer. *Cancer Lett.* (2018) 417:35–46. doi: 10.1016/j.canlet.2017.12.012
- Yamamoto K, Venida A, Yano J, Biancur DE, Kakiuchi M, Gupta S, et al. Autophagy promotes immune evasion of pancreatic cancer by degrading MHC-I. *Nature*. (2020) 581:100–5. doi: 10.1038/s41586-020-2229-5
- Huang X, Tang T, Zhang G, Hong Z, Xu J, Yadav DK, et al. Genomic investigation of co-targeting tumor immune microenvironment and immune checkpoints in pan-cancer immunotherapy. *NPJ Precis Oncol.* (2020) 4:29. doi: 10.1038/s41698-020-00136-1
- Sethi V, Vitiello GA, Saxena D, Miller G, Dudeja V. The role of the microbiome in immunologic development and its implication for pancreatic cancer immunotherapy. *Gastroenterology*. (2019) 156:2097–115.e2. doi: 10.1053/j.gastro.2018.12.045
- Wu G, Deng Z, Jin Z, Wang J, Xu B, Zeng J, et al. Identification of prognostic immune-related genes in pancreatic adenocarcinoma and establishment of a prognostic nomogram: a bioinformatic study. *Biomed Res Int.* (2020) 2020:1346045. doi: 10.1155/2020/1346045
- Yang S, He P, Wang J, Schetter A, Tang W, Funamizu N, et al. A novel MIF signaling pathway drives the malignant character of pancreatic cancer by targeting NR3C2. *Cancer Res.* (2016) 76:3838–50. doi: 10.1158/0008-5472.CAN-15-2841
- Ritchie ME, Phipson B, Wu D, Hu Y, Law CW, Shi W, et al. limma powers differential expression analyses for RNA-sequencing and microarray studies. *Nucleic Acids Res.* (2015) 43:e47. doi: 10.1093/nar/gkv007
- Heagerty PJ, Lumley T, Pepe MS. Time-dependent ROC curves for censored survival data and a diagnostic marker. *Biometrics*. (2000) 56:337–44. doi: 10.1111/j.0006-341X.2000.00337.x
- Yu G, Wang LG, Han Y, He QY. clusterProfiler: an R package for comparing biological themes among gene clusters. *Omics.* (2012) 16:284–7. doi: 10.1089/omi.2011.0118

SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fmed. 2021.649326/full#supplementary-material

Supplementary Table 1 | One thousand seven hundred and thirty-seven abnormally expressed genes in pancreatic cancer.

Supplementary Table 2 | Two hundred and twenty-nine differentially expressed immune-related genes in pancreatic cancer.

- Szklarczyk D, Morris JH, Cook H, Kuhn M, Wyder S, Simonovic M, et al. The STRING database in 2017: quality-controlled protein-protein association networks, made broadly accessible. *Nucleic Acids Res.* (2017) 45(D1):D362–8. doi: 10.1093/nar/gkw937
- Tang Z, Li C, Kang B, Gao G, Li C, Zhang Z. GEPIA: a web server for cancer and normal gene expression profiling and interactive analyses. *Nucleic Acids Res.* (2017) 45(W1):W98–102. doi: 10.1093/nar/gkx247
- Li T, Fu J, Zeng Z, Cohen D, Li J, Chen Q, et al. TIMER2.0 for analysis of tumor-infiltrating immune cells. *Nucleic Acids Res.* (2020) 48(W1):W509–14. doi: 10.1093/nar/gkaa407
- Huang R, Mao M, Lu Y, Yu Q, Liao L. A novel immune-related genes prognosis biomarker for melanoma: associated with tumor microenvironment. *Aging*. (2020) 12:6966–80. doi: 10.18632/aging.103054
- Tang H, Jiang L, Stolzenberg-Solomon RZ, Arslan AA, Beane Freeman LE, Bracci PM, et al. Genome-wide gene-diabetes and gene-obesity interaction scan in 8,255 Cases and 11,900 controls from PanScan and PanC4 consortia. *Cancer Epidemiol Biomark Prev.* (2020) 29:1784–91. doi: 10.1158/1055-9965.EPI-20-0275
- Qiu X, Hou QH, Shi QY, Jiang HX, Qin SY. Identification of hub prognosis-associated oxidative stress genes in pancreatic cancer using integrated bioinformatics analysis. *Front Genet.* (2020) 11:595361. doi: 10.3389/fgene.2020.595361
- Wu M, Li X, Liu R, Yuan H, Liu W, Liu Z. Development and validation of a metastasis-related gene signature for predicting the overall survival in patients with pancreatic ductal adenocarcinoma. *J Cancer.* (2020) 11:6299– 318. doi: 10.7150/jca.47629
- Xu D, Wang Y, Liu X, Zhou K, Wu J, Chen J, et al. Development and clinical validation of a novel 9-gene prognostic model based on multiomics in pancreatic adenocarcinoma. *Pharmacol Res.* (2020) 164:105370. doi: 10.1016/j.phrs.2020.105370
- Zhuang H, Huang S, Zhou Z, Ma Z, Zhang Z, Zhang C, et al. A four prognosisassociated lncRNAs (PALnc) based risk score system reflects immune cell infiltration and predicts patient survival in pancreatic cancer. *Cancer Cell Int.* (2020) 20:493. doi: 10.1186/s12935-020-01588-y
- Wu M, Li X, Zhang T, Liu Z, Zhao Y. Identification of a nine-gene signature and establishment of a prognostic nomogram predicting overall survival of pancreatic cancer. *Front Oncol.* (2019) 9:996. doi: 10.3389/fonc.2019.00996
- Maity G, Mehta S, Haque I, Dhar K, Sarkar S, Banerjee SK, et al. Pancreatic tumor cell secreted CCN1/Cyr61 promotes endothelial cell migration and aberrant neovascularization. *Sci Rep.* (2014) 4:4995. doi: 10.1038/srep04995
- 27. Xin B, He X, Wang J, Cai J, Wei W, Zhang T, et al. Nerve growth factor regulates CD133 function to promote tumor cell migration and invasion via activating ERK1/2 signaling in pancreatic cancer. *Pancreatology*. (2016) 16:1005–14. doi: 10.1016/j.pan.2016.09.005
- Brunetto E, De Monte L, Balzano G, Camisa B, Laino V, Riba M, et al. The IL-1/IL-1 receptor axis and tumor cell released inflammasome adaptor ASC are key regulators of TSLP secretion by cancer associated fibroblasts in pancreatic cancer. J Immunother Cancer. (2019) 7:45. doi: 10.1186/s40425-019-0521-4
- Lin T, Ren Q, Zuo W, Jia R, Xie L, Lin R, et al. Valproic acid exhibits anti-tumor activity selectively against EGFR/ErbB2/ErbB3-coexpressing pancreatic cancer via induction of ErbB family members-targeting microRNAs. J Exp Clin Cancer Res. (2019) 38:150. doi: 10.1186/s13046-019-1160-9
- 30. Modica C, Tortarolo D, Comoglio PM, Basilico C, Vigna E. MET/HGF co-targeting in pancreatic cancer: a tool to provide insight into the

tumor/stroma crosstalk. Int J Mol Sci. (2018) 19:3920. doi: 10.3390/ijms191 23920

- Takiguchi S, Inoue K, Matsusue K, Furukawa M, Teramoto N, Iguchi H. Crizotinib, a MET inhibitor, prevents peritoneal dissemination in pancreatic cancer. *Int J Oncol.* (2017) 51:184–92. doi: 10.3892/ijo.2017.3992
- Delitto D, Vertes-George E, Hughes SJ, Behrns KE, Trevino JG. c-Met signaling in the development of tumorigenesis and chemoresistance: potential applications in pancreatic cancer. *World J Gastroenterol.* (2014) 20:8458–70. doi: 10.3748/wjg.v20.i26.8458
- Zhang Y, Yu C. Prognostic characterization of OAS1/OAS2/OAS3/OASL in breast cancer. BMC Cancer. (2020) 20:575. doi: 10.1186/s12885-020-07034-6

 $\label{eq:conflict} \mbox{ Conflict of Interest: YZh was employed by the company Jiangsu ALF} Biotechnology Co., Ltd.$

The remaining authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

The Handling Editor declared a shared affiliation, though no other collaboration, with one of the authors CZ.

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