European perspectives

Mediators of inflammation in response to air pollution: a focus on ozone and nitrogen dioxide

A 'CONCERTED ACTION' PROJECT IN THE EU BIOMED2 PROGRAMME FOR PROMOTING RESEARCH THAT MEETS THE PUBLIC HEALTH NEEDS OF EUROPEAN CITIZENS

ABSTRACT—Recent epidemiological and environmental chamber studies have strengthened the link between air pollution and respiratory disease. Acute exposure to ozone and nitrogen dioxide (NO₂) for short periods results in both upper and lower airway inflammation and patients suffering from asthma and allergic rhinitis are particularly at risk. Neutrophils are involved in the acute inflammatory reaction following ozone exposure whereas NO₂ poses a more complex response involving neutrophils, mast cells and lymphocytes. The prostanoids play an important role in early symptomatic and functional responses, although their source is still unclear.

Air pollution has become a matter of growing public and scientific concern as increasing motor vehicle traffic has brought a new spectrum of oxidant pollutants which include ozone and oxides of nitrogen (NO_x) [1]. They have been incriminated as factors contributing to the increasing morbidity of asthma [2–6]. We review here the role of various inflammatory mediators that contribute to airway inflammation and the current understanding of the various mechanisms that underlie airway injury following exposure to ozone and nitrogen dioxide (NO_2) .

Airways inflammatory response to ozone

Cellular response

Ozone at ambient levels evokes symptomatic and functional responses [7–14] (Table 1) and inflammation both in the upper and lower airways [14–16], with marked variation between individuals; the inflammatory response is greater in the proximal portion of

M T KRISHNA, MRCP, Clinical Research Fellow and Honorary Registrar*

D R SPRINGALL, PhD, Senior Lecturer in Histochemistry†

A J FREW, MD, MRCP, Senior Lecturer in Medicine and Honorary Consultant Physician*

J M POLAK, DSC, MD, FRCPath, FRCP, Professor of Histochemistry† S T HOLGATE, MD, DSC, FRCP, MRC Clinical Professor of

Immunopharmacology*

*Air Pollution Research Group, University Medicine, Southampton General Hospital

†Department of Histochemistry, Royal Postgraduate Medical School, London the lower airways than in the distal portion. Concentrations of ozone as low as 0.08 ppm [16] induce neutrophilic bronchitis. Inflammation is seen as early as 3 hours and lasts for up to 24 hours following exposure to ozone [17]. Because there is no direct relationship between inflammatory and functional responses following exposure to ambient levels of ozone [18], the mediators which attract neutrophils may not be the same as those producing the early symptomatic and functional responses. This also implies that it may be misleading to classify the population as 'responders' and 'non-responders' purely on the basis of lung function responses.

Prostanoids

Ozone is a powerful oxidising agent which damages tissue by generating free radicals and by peroxidation of lipids in cell membranes [19–22]. As a result, prostanoids such as PGE₂, PGF₂ α , 8-epi-PGF₂ α and TxB₂ are released into the bronchoalveolar lavage (BAL) fluid [23,24]. They stimulate pulmonary neural afferents, producing some of the characteristic responses to ozone exposure. Administration of indomethacin, a cyclo-oxygenase inhibitor [25,26] (Fig 1), prior to ozone exposure abrogates the functional responses in normal subjects and asthmatics [27]. Surprisingly, administration of steroids [28] prior to ozone challenge resulted in a paradoxical response in forced expiratory variables and numbers of BAL neutrophils.

Cytokines and adhesion molecules

The concentration of cytokines, including IL (interleukin)-6, IL-8 and GM-CSF (granulocyte macrophage colony stimulating factor) rises in the bronchoalveolar lavage (BAL) fluid following ozone exposure. IL-8 is constitutively expressed in the bronchial epithelium, and exposure to ozone provokes further synthesis of this cytokine in the bronchial epithelium, although macrophages are also an imporant source of IL-8. Exposure of mouse airways to ozone increases the expression of macrophage inflammatory protein-2 (MIP-2) [29], a chemokine capable of inducing neutrophil chemotaxis. Exposure to ambient levels of ozone also results in an upregulation of intercellular adhesion molecule-1 (ICAM-1) [30] in the vascular

Symptoms	Functional responses
Shortness of breath	Decrease in:
 Inspiratory chest pain 	• FEV1
 Tiredness 	• FVC
Eye irritation	 Inspiratory capacity
• Wheeze	Total lung capacity
	Increase in:
	Airway resistance

endothelium of the proximal airways mucosa associated with an increased expression of CD11b, a ligand for ICAM-1 on BAL neutrophils.

The nuclear transcription factor NF-κB induces the expression of several endothelial cell adhesion molecules—and of endothelially derived cytokines [31,32]. It is activated by a wide variety of extracellular stimuli including pro-inflammatory cytokines, bacterial and viral products and oxidative stress, and inhibited by treating endothelial cells with antioxidants [33]. It is therefore reasonable to speculate that 'oxidant stress' such as exposure to ozone and NO_2 will activate NF- κ B and lead to upregulation of cell adhesion molecules and pro-inflammatory cytokines in the vascular endothelium.

In addition to producing airway inflammation, ozone can induce an acute-phase systemic response seen as an increase in the levels of C-reactive proteins in the plasma and of IL-6 in BAL and plasma [34]. The significance of these observations is not fully understood although animal experiments have shown that IL-6 could play an important role in inducing 'tolerance' [35] repeated exposure to ozone.

Effects of ozone in nonadrenergic noncholinergic (NANC) nerves

Increased levels of substance P (SP) have been found in BAL fluid immediately following exposure to 0.25 ppm ozone for one hour [24]. SP can induce bronchoconstriction, increase airway permeability, activate neutrophils and increase mucociliary transport [36–38]. These observations suggest that tachykinins such as SP and neurokinin A (NKA) could contribute to early functional and symptomatic responses, and with their immunomodulatory properties augment the airway inflammation. It is plausible that ozone diminishes the activity of neutral endo-





peptidase (NEP) an epithelial enzyme responsible for the metabolism of endogenous neuropeptides, with a resultant increase in the concentration of SP.

Effects of ozone in 'high risk groups'

Recent studies have focused on the 'high risk' groups such as asthmatics and patients suffering from allergic rhinitis. Ozone potentiates the response to inhaled allergens in patients with allergic airway disease [39], and epidemiological studies [40-42] have shown strong links between exacerbations of asthma symptoms and 'ozone episodes'. In patients suffering from allergic rhinitis, exposure to ozone can induce an influx of neutrophils and eosinophils in the nasal mucosa [43], and a significant correlation was detected between neutrophil numbers and levels of IL-8 in nasal lavage fluid of asthmatics [44] following exposure to 0.24 ppm ozone. IL-8 is principally secreted by mononuclear phagocytes, fibroblasts, endothelial and epithelial cells [45]. It is an important chemoattractant for neutrophils and induces release of granule products and free radicals from neutrophils

Mediators of inflammation in response to air pollution

[46,47] which cause epithelial injury, sloughing and tissue damage.

Ozone increases epithelial permeability [48] in normal human subjects and this phenomenon is likely to be exaggerated in asthmatics in whom the underlying airway inflammation and 'epithelial shedding' would result in a greater absorption of the inhaled aero-allergens. This could explain the exacerbations of asthma and hay fever symptoms immediately following 'ozone episodes'.

Figures 1 and 2 summarise the inflammatory effects of ozone in normal and asthmatic subjects. (See Table 2 for WHO guidelines for ozone.)

Table 2: WHO guidelines for ozone. These concentrations should not be exceeded during exposures over 1 or 8 hours

1 hour	8 hours
0.076-0.100 ppm	0.050-0.060 ppm

Fig 2. Effects of combination of pollutants and allergens on allergic airways disease



Journal of the Royal College of Physicians of London Vol. 30 No. 1 January/February 1996

Airway inflammatory response to NO₂

Cellular response

Exposure to NO_2 increases the number of neutrophils in the bronchial portion of the lavage fluid (proximal airway lavage) [49], and the levels of neutrophil myeloperoxidase (MPO) [50].

Using concentrations of NO₂ between 2.25–5.0 ppm for 20 minutes in normal subjects revealed a dose dependent mastocytosis and lymphocytosis in BAL fluid 4 to 24 hours after exposure, and an increase in macrophages at concentrations greater than 4 ppm of NO₂ [51]. This effect was even more marked in smokers, suggesting that they are potentially at greater risk from the adverse effects of NO₂ [51].

Prostanoids

Healthy subjects exposed to 1 ppm NO₂ for three hours showed no significant changes in prostaglandins and histamine levels in BAL fluid [52], although TxB₂ levels were significantly higher. In asthmatics there was a decrease in the levels of 6-keto-PGF1 α (a metabolite of PGI₂, a bronchodilator), and higher levels of TxB₂ (a metabolite of TxA₂, the bronchoconstrictor) and of PGD₂ (also a bronchoconstrictor), whereas levels of leukotrienes and histamine remained unchanged. These changes support the idea that asthmatics are more susceptible to the adverse effects of NO₂ than non-asthmatics. In contrast to the study described above [51] inflammatory cell numbers did not change in either group at this low (1 ppm) concentration of NO₂.

Protease and anti-protease balance

Alpha-1-protease inhibitor (α 1-PI), inhibits plasma and lung elastase, an important factor in the pathogenesis of emphysema [53,54] and so protects the lung from proteolytic damage. Exposure to NO₂ (3 or 4 ppm) for 3 hours decreased the functional capacity of α 1-PI in BAL by 45%, although this was not accompanied by neutrophil migration in the airways; furthermore, *in vitro* exposure of α 1-PI to NO₂ did not affect its capacity to inactivate neutrophil elastase. This observation suggests that NO₂ exerts its effect indirectly, probably through the products of lipid peroxidation which are highly reactive with α 1-PI.

Cytokines

It is logical to speculate that NO₂ being a powerful oxidising agent, could damage or stimulate the cell membrane of the bronchial epithelium through the process of lipid peroxidation and generation of free radicals. NO₂ at a concentration of 0.4 ppm significantly increases the release of the pro-inflammatory cytokines GM-CSF, IL-8 and TNF- α from bronchial epithelium *in vitro* and of IL-1 α , IL-1 β , IL-6, IL-8 and Gro γ in response to 1.5 ppm NO₂ [55,56].

Effects on NANC nerves

The effect of NO_2 on the nonadrenergic noncholinergic (NANC) nerves has not been evaluated in humans. In guinea pigs, NO_2 increases bronchial responsiveness to acetylcholine and neurokinin [57] and lowers the volume density of nerves (ie the number of nerves per unit volume of tissue) which are immunoreactive to the sensory neuropeptides SP and calcitonin generelated peptide (CGRP). Because the overall density of nerves remains unchanged, this finding indicates that some of the nerves lost their immunoactivity because they had released their sensory peptides into the lung by antidromic stimulations; such a local release can lead to neurogenic inflammation with bronchoconstriction, bronchial hyperresponsiveness and vasodilatation.

Although it is not possible to evaluate the clinical implications of these changes in inflammatory medicators, their presence suggests that NO_2 at levels encountered at the sites of heavy motor vehicle traffic, or in homes where gas is used as a source for cooking and heating appliances, could have adverse effects on lung function.

NO₂ and respiratory infections

Epidemiological studies [58-60] have shown an association between exposure to NO2 and respiratory tract infections. Exposing normal subjects to 1.5 and 4 ppm NO₂ for 20 minutes on alternate days for 6 days causes a small but significant decrease in CD8+ cells, NK cells (CD16+ and CD56+) and B cells (CD19+) in the BAL fluid [61] but these changes are not seen at exposure to lower concentrations of NO2. The adverse effects of NO₂ on the immune system are reflected in the impaired ability of alveolar macrophages, obtained by BAL after exposure to 0.6 ppm NO₂ to inactivate influenza virus in vitro [62]. Exposure of nine subjects to NO₂ impaired the inactivation of influenza virus in four whose cells produced more IL-1 than the alveolar macrophages from the five subjects whose cells could inactivate the virus. Damage of cell membrane by NO2 via lipid peroxidation could alter the uptake of the virus by damaging cell surface receptors or decreasing endocytosis. The inflammatory effects of NO2 in normal and asthmatic subjects are summarised in Figs 1 and 2.

Future work

This 'concerted action' (project leader: J M Polak) is our contribution to the EU's BIOMED2 programme for promoting research that meets the public health needs of European citizens. It includes participants from UK, Italy and Belgium. We will continue to study the various mechanisms that cause airway inflammation following short-term exposures to ambient levels of ozone and high indoor levels of NO₂. Using immunohistochemical techniques we are exploring the roles of inflammatory cells, cytokines, adhesion molecules and sensory neuropeptides in airway inflammation. We are also studying the role of antioxidants by measuring the levels of vitamins A, C, E, glutathione, uric acid and the products of oxidative injury including protein carbonyls and lipid hydroperoxides. Results of these studies will shed more light on the current understanding of the health effects of air pollutants.

Exposure to air pollutants potentiates responses to aero-allergens. In future, studies should address the health effects of pollutants in a wider perspective, using a range of different concentrations of pollutants, as well as studying the effects of combinations of pollutants and allergens, and carrying out interventional studies by giving supplements of anti-oxidants and nonsteroidal anti-inflammatory drugs.

References

- 1 A first report to the RAC Foundation for Motoring and the Environment. *Cars and the environment: a view to the year 2020.* London: RAC Foundation, 1992.
- 2 Parliamentary Office of Science and Technology. Breathing in our cities-urban air pollution and respiratory health. London: HMSO 1994.
- ³ Sears MR. Epidemiology of asthma. In: O'Byrne PM (ed). Asthma as an inflammatory disease. New York: Dekker, 1990:15–48.
- 4 Lippert FW, Morris SC. Air pollution: cost benefit assessment. Science 1991;253:606–9.
- ⁵ Imai M, Yoshida K, Kitabake M. Mortalilty from asthma and chronic bronchitis associated with changes in sulphur dioxide air pollution. *Arch Environ Health* 1986;**41**:29–35.
- 6 Schmitzberger R, Rhomberg K, Buchele H, et al. Effects of air pollution on the respiratory tract of children. Paediatr Pulmonol 1993;15:68–74.
- 7 Hazucha MJ. Relationship between ozone exposure and pulmonary function changes. J Appl Physiol 1987;62:1671–80.
- 8 Silverman F, Folinsbee LJ, Barnard J, Shepherd RJ. Pulmonary function changes in ozone—interaction of concentration and ventilation. J Appl Physiol 1976;41(6):859–64.
- 9 Hazucha MJ, Bates DV, Bromberg PA. Mechanism of action of ozone on human lung. J Appl Physiol 1989;67:1535–41.
- 10 Folinsbee LJ, Hazucha MH. Persistence of ozone induced changes in lung function and airway responsiveness. In: Schneider T, Lee SD, Wolters GJR, Grant LD (eds). Atmospheric ozone research and its policy implications. Amsterdam: Elsevier Science Publishers BV, 1989;523-32.
- 11 UK Advisory Group on Medical Aspects of Air Pollution Episodes. First report ozone. London: HMSO, 1991.
- 12 McDonnell WF, Chapman RS, Leigh MW, Strope GL, Collier AM. Respiratory responses of rigorously exercising children to 0.12ppm ozone exposure. Am Rev Respir Dis 1985;132:875–9.
- 13 Dimeo MJ, Glenn MG, Holtzmann MJ, et al. Threshold concentration of ozone causing an increase in bronchial reactivity in humans and adaptation with repeated exposure. Am Rev Respir Dis 1981;124:245-8.
- 14 Krishna MT, Mudway I, Kelly FJ, Frew AJ, Holgate ST. Ozone, airways and allergic airways disease. *Clin Exp Allergy* 1995 (in press).
- 15 Koren PS, Devlin RB, Graham DE, et al. Ozone induced inflammation in the lower airways of human subjects. Am Rev Respir Dis 1989;139:407–15.

- 16 Devlin RB, McDonnell WF, Mann R, et al. Exposure of humans to ambient ozone for 6.6 hours causes cellular and biochemical changes in the lung. Am J Respir Cell Mol Biol 1991;4:72–81.
- 17 Schelegle ES, Siefkin AD, McDonald R. Time course of ozone induced neutrophilia in normal subjects. Am Rev Respir Dis 1991;141:1353-8.
- 18 Aris RM, Christian D, Hearne PQ, et al. Ozone induced airway inflammation in human subjects as determined by airway lavage and biopsy. Am Rev Respir Dis 1993;148:1363–72.
- 19 Mustafa MG. Biochemical basis of ozone toxicity. Free Rad Biol Med 1990;9:245-65.
- 20 Kelly FJ, Mudway I, Krishna MT, Holgate ST. The free radical basis of air pollution: focus on ozone. *Respir Med* 1995;89: 647-56.
- 21 Konings AWT. Mechanisms of ozone toxicity in cultured cells. I. Reduced clonogenic ability of polyunsaturated fatty acid-supplemented fibroblasts. Effect of vitamin E. J Toxicol Environ Health 1986;18:491–7.
- 22 Van-Der-Zee J, Tijssen-Christianse K, Dubbleman TM, Van-steneick J. The influence of human red blood cells. Comparison with other mechanisms of oxidative stress. *Biochem Biophys Acta* 1987;**924**:111-8.
- 23 Seltzer J, Bigby G, Stulberg M, Holtzmann MJ, Nadel JA. Ozone induced change in bronchial reactivity to methacholine and airway inflammation in humans. *J Appl Physiol* 1986;60:1321–6.
- 24 Hazbun ME, Hamilton R, Holian A, Eschenbacher WL. Ozone induced increases in substance P and 8-epi-prostaglandin F_{2a} in the airways of human subjects. Am J Respir Cell Mol Biol 1993;9:568-72.
- 25 Schelegle ES, Adams WC, Siefkin AD. Indomethacin pretreatment reduces ozone induced pulmonary function decrements in human subjects. *Am Rev Respir Dis* 1987;136:1350–4.
- 26 Ying RL, Gross KB, Terzo TS, Eschenbacher WL. Indomethacin does not inhibit the ozone induced increase in bronchial responsiveness in human subjects. Am Rev Respir Dis 1990; 142:817-21.
- 27 Alexis N, Silvermann F, Tarlo S, Covey P. Effects of indomethacin on acute ozone exposure in asthmatics. Am J Respir Crit Care Med 1995;151:A28(abstract).
- 28 Bottei GM, Devlin RB, Bromberg PA, Peden PB. The effect of corticosteroids on lower airway inflammation and pulmonary function after ozone exposure. *Am J Respir Crit Care Med* 1995; 151:A28(abstract).
- 29 Driscoll KE, Hassenbein D, Carter J, et al. Macrophage inflammatory proteins 1 and 2: expression by rat alveolar macrophages, fibroblasts and epithelial cells and in rat tissue after ozone exposure. Am J Respir Cell Mol Biol 1993;8:311–8.
- 30 Aris R, Ferrando R, Chen LC, Christian D, Balmes JR. Increased expression of ICAM on bronchial endothelium after acute exposure to ozone. Am J Respir Crit Care Med 1995;151:A23(abstract).
- 31 Baeuerle PA, Henkel T. Function and activation of NFKB in the immune system. Ann Rev Immunol 1994;12:141-79.
- 32 Manning AM, Anderson DC. Transcription factor NFκB: an emerging regulator of inflammation. In: Bristol JA (ed). Annual reports in medicinal chemistry. San Diego: Academic Press, 1994;235-44.
- 33 Marui N, Offerman MK, Swerlick R, et al. Vascular cell adhesion molecule-l gene transcription and expression are regulated through an anti-oxidant sensitive mechanism in human vascular endothelial cell. J Clin Invest 1993;92:1866–74.
- 34 Malek FY, Utell MJ, Looney RJ, et al. Does exposure to ozone induce an acute systemic phase response in humans. Am J Respir Crit Care Med 1995;151:A27 (abstract).
- 35 McKinney WJ, Dreher KL, Jascot RH, Baumann H, Costa DL. The role of IL-6 in ozone induced tolerance. Am J Respir Crit Care Med 1994;149:A156(abstract).
- 36 Solway J, Leff A. Sensory neuropeptides and airway function. J Appl Physiol 1991;71:2077–87.
- 37 Foster W, Costa D, Langenback E. Ozone exposure alters tracheobronchial mucociliary function in humans. J Appl Physiol 1987;63:996-1002.

- 38 Fuller RW, Maxwell DL, Dixon CMS, *et al.* Effect of substance P on cardiovascular and respiratory function in human subjects. J *Appl Physiol* 1987;62:1473–9.
- 39 Molfino NA, Wright SC, Ketz I, et al. Effect of low concentrations of inhaled allergen responses in asthmatic patients. Lancet 1991;338:199–203.
- 40 White MC, Etzel RA, Wilcox WD, Lloyd C. Exacerbations of childhood asthma and ozone pollution in Atlanta. *Environ Res* 1994;65:56–68.
- 41 Cody RP, Weisel CP, Beinbaum G, Lioy PJ. The effect of ozone associated with summertime photochemical smog on the frequency of asthma visits to hospital emergency departments. *Environ Res* 1992;**58**:184–94.
- 42 Bates DV, Baker-Anderson M, Sitzo R. Asthma attack periodicity: a study of hospital emergency visits to Vancouver. *Environ Res* 1990;51:51–70.
- 43 Bascom R, Nacleiro RM, Fitzgerald TK, Kagey-Sabotka A, Proud D. Effect of ozone inhalation on the response to nasal challenge with antigen of allergic subjects. Am Rev Respir Dis 1990;142:594-601.
- 44 McBride DE, Koenig JQ, Luchtel DL, Williams PV, Henderson Jr WR. Inflammatory effects of ozone in the upper airways of subjects with asthma. *Am J Respir Crit Care Med* 1994;149:1192–7.
- 45 Schroeder JM, Mrowietz U, Morita E, Christophers E. Purification and partial biochemical characterisation of human monocyte derived neutrophil-activating peptide that lacks IL-l activity. *[Immunol* 1987;139:3474–83.
- 46 Thelen M, Peveri P, Kernen P, Von Tscharner V, Walz A, Baggiolini M. Mechanism of neutrophil activation by NAF, a novel monocyte-derived peptide agonist. *FASEB J* 1988;2:2702–6.
- 47 Baggiolini M, Walz A, Kunkel SL. Neutrophil-activating peptidel/interleukin-8, a novel cytokine that activates neutrophils. J Clin Invest 1989;84:1045–9.
- 48 Kehrl HR, Vincent LM, Kowalsky RJ, et al. Ozone increases respiratory permeability in humans. Am Rev Respir Dis 1987; 135:1124–8.
- 49 Helleday R, Sandstrom T, Stjernberg N. Differences in bronchoalveolar cell response to nitrogen dioxide between smokers and non-smokers. *Eur Respir J* 1994;7:1213–20.
- 50 Sandstrom T, Helleday R, Blomberg A, Stjernberg N, Henderson R. Repeated exposure to nitrogen dioxide affects hyaluran, myeloperoxidase and methyl histamine in BAL fluid. Am J Respir Crit Care Med 1995;151:A284(abstract).
- 51 Sandstrom T, Stjernberg N, Elund A, et al. Inflammatory cell response in bronchoalveolar lavage fluid after nitrogen dioxide exposure: a dose response study. Eur Respir J 1991;4:332–9.

- 52 Jorres R, Nowak D, Grimminger F, *et al.* The effect of 1ppm nitrogen dioxide on bronchoalveolar lavage cells and inflammatory mediators in normal and asthmatic subjects. *Eur Respir J* 1995;8:416–24.
- 53 Dooley MM, Pryor WA. Free-radical pathology: inactivation of alpha-l-protease inhibitor by products from the reaction of nitrogen dioxide with hydrogen peroxide and the aetiology of emphysema. *Biochem Biophys Res Commun* 1982;106:981-7.
- 54 Pryor WA, Dooley MM, Church DF. Inactivation of human alpha-l-proteinase inhibitor by gas phase cigarette smoke. Biochem Biophys Res Commun 1984;122:676-81.
- 55 Devalia JL, Campbell AM, Sapsford RS, et al. Effect of nitrogen dioxide on synthesis of inflammatory cytokines expressed by human bronchial cells in vitro. Am J Respir Cell Mol Biol 1993;9:271-8.
- 56 Kienast K, McKinnon KP, Carter JD, Reed W, Devlin RB. Nitrogen dioxide exposure of human airway epithelial cells *in vitro* increases steady state concentrations of inflammatory mediator mRNAs. *Am J Respir Crit Care Med* 1995;151:A284 (abstract).
- 57 Lucchini RE, Springall DR, Chitano P, Fabbri LM, Polak JM, Mapp CE. In vivo exposure to nitrogen dioxide (NO₂) induces a decrease in calcitonin gene-related peptide (CGRP) and tachykinin immunoreactivity in guinea pig peripheral airways. Am J Respir Cell Mol Biol (in press).
- 58 Dijkstra L, Houthuijs D, Brunekreef B, Akkerman I, Boleij JS. Respiratory health effects of indoor environment in a population of Dutch children. Am Rev Respir Dis 1990;142:1172-8.
- 59 Samet JM, Lambert WE, Skipper BJ, et al. Nitrogen dioxide and respiratory illness in infants 1993;148:1258–65.
- 60 Hasselblad V, Eddy DM, Kotchmar DJ. Synthesis of environmental evidence: nitrogen dioxide epidemiological studies. J Air Wast Manage Assoc 1992;42:662-71.
- 61 Sandstrom T, Helleday R, Bjermer L, Stjernberg N. Effects of repeated exposure to 4ppm nitrogen dioxide on bronchoalveolar lavage fluid lymphocyte subjects and macrophages in healthy men. *Eur Respir J* 1995;5:1092–6.
- 62 Frampton MW, Smeglin AM, Roberts NJ, et al. Nitrogen dioxide exposure in vivo and human alveolar macrophage interaction of influenza virus in vitro. Env Res 1989;48:179–92.

Address for correspondence: Dr M T Krishna, University Medicine, Level D Centre Block, Southampton General Hospital, Tremona Road, Southampton SO16 6YD.