



Potential Cross-Transmission of *Mycobacterium abscessus* among Non-Cystic Fibrosis Patients at a Tertiary Hospital in Japan

 Keiji Fujiwara,^{a,b,c}  Mitsunori Yoshida,^d  Yoshiro Murase,^b  Akio Aono,^b  Koji Furuuchi,^{a,c}  Yoshiaki Tanaka,^a  Ken Ohta,^a
 Manabu Ato,^d  Satoshi Mitarai,^{b,c}  Kozo Morimoto^{a,e}

^aRespiratory Disease Center, Fukujiji Hospital, Japan Anti-Tuberculosis Association, Tokyo, Japan

^bDepartment of Mycobacterium Reference and Research, The Research Institute of Tuberculosis, Japan Anti-Tuberculosis Association, Tokyo, Japan

^cDepartment of Basic Mycobacteriosis, Nagasaki University Graduate School of Biomedical Sciences, Nagasaki, Japan

^dDepartment of Mycobacteriology, Leprosy Research Center, National Institute of Infectious Diseases, Tokyo, Japan

^eDivision of Clinical Research, Fukujiji Hospital, Japan Anti-Tuberculosis Association, Tokyo, Japan

Keiji Fujiwara and Mitsunori Yoshida contributed equally to this article. Author order was determined by alphabetically.

ABSTRACT *Mycobacterium abscessus* (*M. abscessus*) is a highly antimicrobial-resistant pathogen that causes refractory pulmonary disease. Recently, the possibility of *M. abscessus* cross-transmission among cystic fibrosis (CF) patients has been reported. CF is rare in Asia, but *M. abscessus* pulmonary disease is common. Therefore, we investigated the possibility of *M. abscessus* cross-transmission in a Japanese hospital setting. Of 104 *M. abscessus* isolates, 25 isolates from 24 patients were classified into four clusters based on their variable number of tandem repeat profiles and were subjected to whole-genome sequencing (WGS). The epidemiological linkages among our patients were investigated by integrating the WGS data of previously reported nosocomial outbreak-related *M. abscessus* clinical isolates in the United Kingdom and the United States. Eight transmissible clusters (TCs) were identified. The United Kingdom and United States isolates were assigned to four clusters (TC1, TC2, TC5, and TC8) and one cluster (TC3), respectively. A total of 12 isolates from our hospital belonged to 4 clusters (TC4, TC5, TC6, and TC7). Epidemiological linkage analysis inferred direct or indirect transmission between patients in our hospital in TC4 and TC5 but not in TC6 and TC7. In TC5, the single nucleotide polymorphism distance between isolates from Japanese and United Kingdom patients was less than 21; however, there was no contact. This study revealed that genetically closely related isolates exist, even in non-CF patients. However, the transmission route remains unclear, and further research is warranted to clarify whether cross-transmission is involved.

IMPORTANCE Although the possibility of *Mycobacterium abscessus* (*M. abscessus*) cross-transmission in cystic fibrosis (CF) patients has often been reported, it is not clear whether similar events have occurred in Asian non-CF patients. Whole-genome sequencing analysis of *M. abscessus* isolates from Fukujiji Hospital in Japan indicated that genetically closely related *M. abscessus* isolates exist. In addition, according to epidemiological linkage analysis, some clusters were suspected of direct or indirect transmission between patients within our hospital. However, the transmission route of *M. abscessus* remains unclear, because interestingly, one cluster showed a single nucleotide polymorphism distance of less than 21 from the United Kingdom isolates, but no epidemiological linkage was identified.

KEYWORDS nontuberculous mycobacteria, *Mycobacterium abscessus* subsp. *massiliense*, whole-genome sequencing, variable-number tandem repeats, transmission

The prevalence of nontuberculous mycobacterial (NTM) pulmonary disease has been increasing worldwide in recent decades (1, 2). *Mycobacterium abscessus* (*M. abscessus*)

Editor Gyanu Lamichhane, Johns Hopkins University School of Medicine

Copyright © 2022 Fujiwara et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Kozo Morimoto, morimotok@fukujiji.org.

The authors declare no conflict of interest.

Received 16 January 2022

Accepted 16 April 2022

Published 10 May 2022

is a highly antimicrobial-resistant NTM that causes refractory disease (3). Two systematic reviews reported that the treatment success for *M. abscessus* pulmonary disease was 45.6% to 53.2%, which was completely unsatisfactory (4, 5). In addition to the treatment difficulty, the possibility of cross-transmission of *M. abscessus* among cystic fibrosis (CF) patients has recently been reported (6–8), indicating the importance of infection control. Thus, it is crucial to identify the transmission routes of *M. abscessus* to prevent recurrent infections.

In the context of NTM isolate genotyping, various techniques, such as pulsed-field gel electrophoresis, variable-number tandem repeats (VNTR), and multilocus sequence typing, have been utilized (7, 9–11). Recent advances in whole-genome sequencing (WGS) have enabled the detailed bacteriological and epidemiological analysis of NTM species. Bryant et al. reported the potential for cross-transmission of *M. abscessus* in a CF center in the United Kingdom by analyzing pairwise single nucleotide polymorphism (SNP) distances (6). They defined less than or equal to 25 SNP distances within the conserved genomic region as the threshold for cross-transmission in their sample set. Subsequent studies have generally used similar thresholds (12, 13). However, this SNP distance threshold was determined by a single study, and it is not clear if this threshold can be directly adapted universally.

Global genetic research on 1,080 *M. abscessus* clinical isolates from 517 CF patients in Europe, the United States, and Australia revealed that approximately 74% of CF patients worldwide were infected with two *M. abscessus* subsp. *abscessus* clones or one *M. abscessus* subsp. *massiliense* clone (14). Although the findings suggest the possibility of dominant circulating clones (DCCs) among CF patients, few studies have been conducted using WGS in combination with clinicoepidemiological profiles to investigate whether similar events occur among non-CF patients (15, 16).

A previous multicenter study involving 121 *M. abscessus* pulmonary disease patients analyzed the relationship between the VNTR profiles and clinical characteristics; however, no apparent relationship was identified (3). On the other hand, a few clusters with identical VNTR profiles were detected, suggesting the cross-transmission of a few clones (12, 17). Accordingly, in the present study, we aimed to clarify the possibility of cross-transmission among non-CF patients by not only performing WGS analysis of *M. abscessus* forming VNTR clusters but also by investigating epidemiological linkage data.

RESULTS

Characteristics of the study patients. A total of 25 isolates were obtained from the 24 patients enrolled in the present study. Of these, 23 patients met the diagnostic criteria of NTM pulmonary disease established by the American Thoracic Society/European Respiratory Society/European Society of Clinical Microbiology and Infectious Diseases/Infectious Diseases Society of America (ATS/ERS/ESCMID/IDSA), but 1 did not (18). The characteristics of the patients are shown in Table 1. A total of 15 patients were female (62.5%), the median age was 67 years (range, 54 to 71), and 6 (25.0%) had a previous NTM pulmonary disease history (4 patients had *Mycobacterium avium* complex [MAC] infection, 1 had MAC and *Mycobacterium lentiflavum* infection, and 1 had *Mycobacterium gordonae* infection). The most common radiological finding was noncavitary nodular bronchiectatic (NC-NB; 12 patients, 50.0%), followed by cavitary nodular bronchiectatic (C-NB; 6, 25.0%), fibrocavitary (FC; 5, 20.8%), and unclassified type (1, 4.2%). Colony morphotypes were classified as follows: rough, 40% (10 isolates); smooth, 44% (11); mixed, 8% (2); and intermediate, 8% (2). Regarding susceptibility to clarithromycin, all *M. abscessus* subsp. *abscessus* isolates had inducible resistance, while all but one *M. abscessus* subsp. *massiliense* isolate was susceptible. All isolates were amikacin-susceptible or intermediate.

Phylogenetic analysis results. The phylogenetic tree of the 25 isolates generated using the WGS data is shown in Fig. 1. Several phylogenetic clades (PCs) (PCs I to IV) of *M. abscessus* subsp. *abscessus* and *M. abscessus* subsp. *massiliense* with low genetic diversity were identified, all of which were consistent with each of the four VNTR clusters classified in a previous study (3). One *M. abscessus* subsp. *massiliense* isolate (RGM-96_post) obtained serially from the patient from whom RGM-96_pre was isolated at a different time point was assigned to the same PC IV as RGM-96_pre.

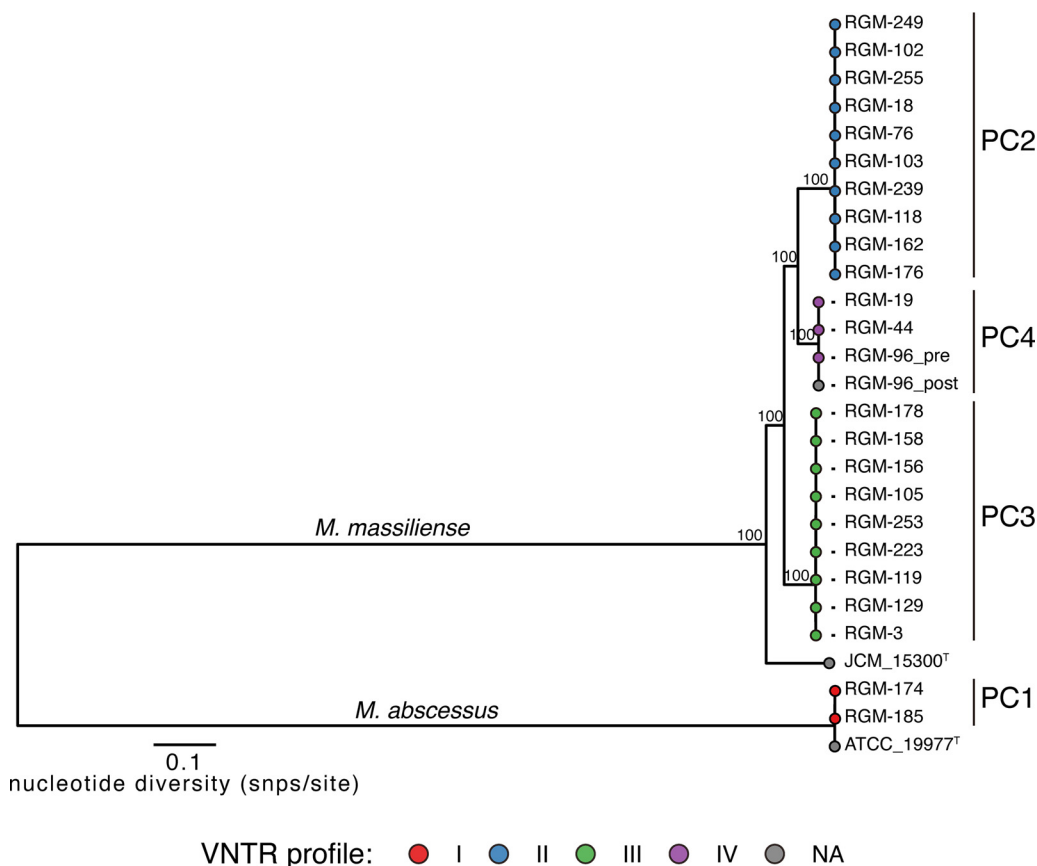


FIG 1 Phylogenetic tree of 25 *M. abscessus* isolates. The 25 isolates included 24 isolates that composed 4 clusters according to their VNTR profiles and 1 serial isolate obtained from the same patient at a different time point (RGM-96_post). The complete genome sequence of *M. abscessus* subsp. *massiliense* JCM 15300 was used as the reference. Multiple whole-genome alignments containing 113,083 recombination-free variable positions located in the core genome (4,212,890 bp of the *M. abscessus* subsp. *massiliense* JCM 15300 genome) were performed to estimate a maximum likelihood tree with 1,000 bootstrap replicates. The scale bar represents the number of substitutions per site (SNPs/site) on the respective branch. VNTR, variable-number tandem repeats; SNP, single nucleotide polymorphism; *M. abscessus*, *Mycobacterium abscessus* subsp. *abscessus*; *M. massiliense*, *Mycobacterium abscessus* subsp. *massiliense*; NA, not applicable; PC, phylogenetic clade.

SNP analysis and clustering analysis. The maximum SNP distance among *M. abscessus* subsp. *massiliense* isolates collected from the same individuals in three hospitals (Fukujuji Hospital in Japan, Papworth Hospital in the United Kingdom, and Seattle Hospital in the United States) was 25 in our analysis (Fig. 2A) (6, 11). First, we investigated the SNP distances among isolates in each PC and identified 15 combinations (17.2%) with less than 25 SNP distances (2 combinations in PC II, 9 in PC III, and 4 in PC IV) (see Fig. S1 in the supplemental material). There were 5 SNP distances between RGM-96_pre and RGM-96_post, which were isolated from the same patient. Next, we identified eight transmissible clusters (TCs) using the SNP distance as a threshold for clonal expansions (Fig. 2B). The United Kingdom isolates were assigned to four TCs (TC1, TC2, TC5, and TC8); the isolates in TC1 and TC2 were identical to those from previously reported nosocomial outbreaks (*M. abscessus* subsp. *massiliense* clusters 1 and 2), and the remaining nonclustered isolates were assigned to TC5 and TC8 (6). All isolates from a nosocomial outbreak at a CF center in the United States were assigned to TC3 (11), and 12 isolates from Fukujuji Hospital were assigned to four clusters (TC4, TC5, TC6, and TC7). TC5 consisted of four isolates from Fukujuji Hospital and two isolates from the United Kingdom, and the SNP distances between RGM-19 (Fukujuji Hospital) and the 13a and 13b isolates (from the United Kingdom) were 21 and 19, respectively (Fig. S2) (6).

Epidemiological linkages among and clinical characteristics of the clustered patients. The total numbers of overlapping days when the patients were in the outpatient department, hospital ward, and outpatient department-hospital ward on the same date

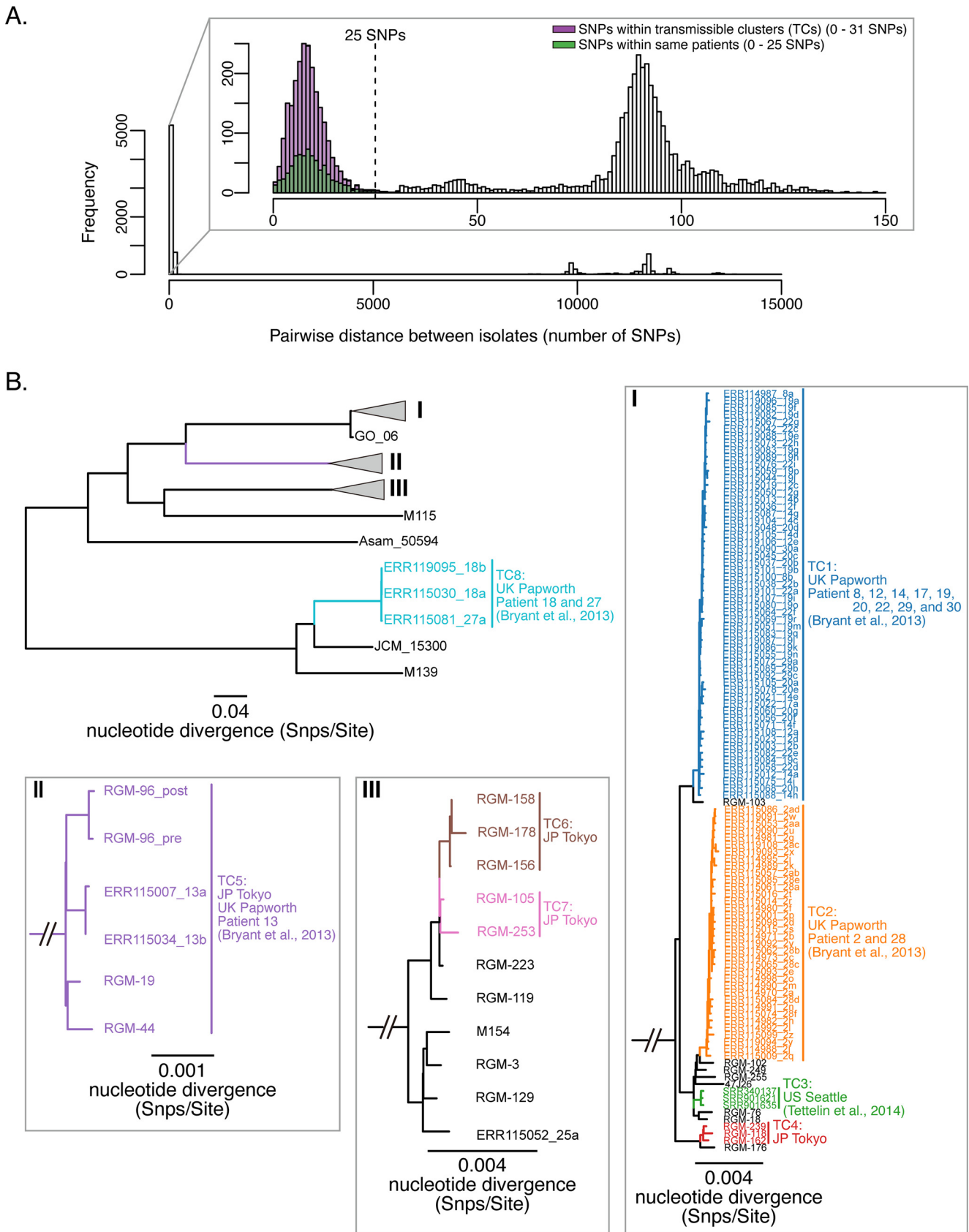


FIG 2 (A) Histogram of pairwise SNP distances among isolates collected at Fukujuji Hospital in Japan, Papworth Hospital in the United Kingdom, and Seattle Hospital in the United States. Whole-genome alignment of a total of 133 *M. abscessus* subsp. *massiliense* isolates containing 39,364 recombination-free (Continued on next page)

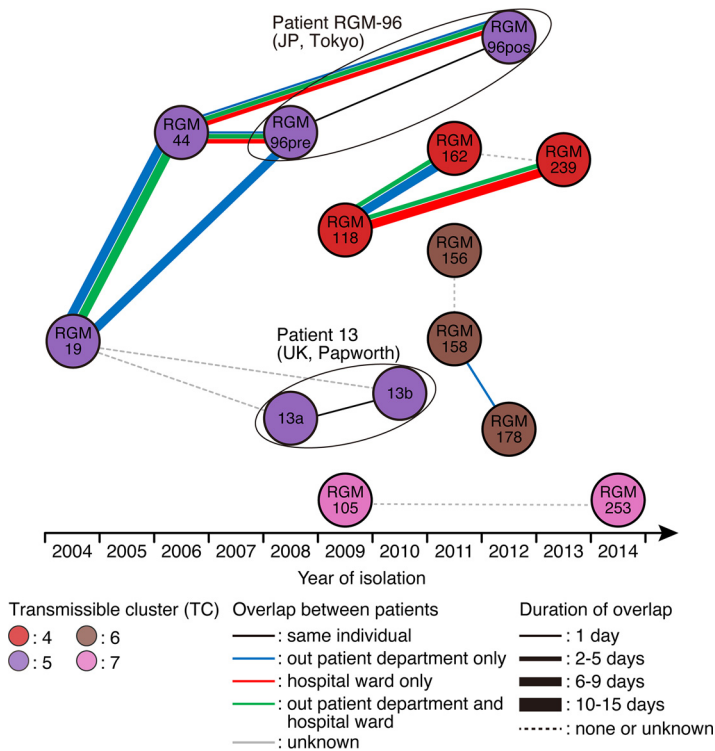


FIG 3 Contact situations among patients in each cluster. The position of each circle indicates the year that the corresponding *M. abscessus* isolate was obtained from each patient. The circled isolate pairs RGM96pre/RGM96post and 13a/13b were isolated from the same patient. Circles colored with different colors represent each transmissible cluster (TC). TC4 is colored red, TC5 is purple, TC6 is brown, and TC7 is pink. We considered opportunities for cross-transmission within the hospital to have occurred when a patient who once had a positive sputum culture was in the outpatient department or hospital ward or the outpatient department and hospital ward on the same date as another patient, and we counted these instances as overlapping days. The line thickness indicates the number of days the patients were in the hospital on the same date. TC, transmissible cluster; UK, United Kingdom.

were 6, 8, and 4 days for TC4 and 21, 3, and 13 days for TC5, respectively (Fig. 3). When hospitalized on the same dates, TC4 patients (RGM-118 and RGM-239) were hospitalized on the same floor, and TC5 patients (RGM-44 and RGM-96) were on different floors. Accordingly, patients in TC4 and TC5 shared a total of 18 and 37 days in a close setting, respectively, while TC6 patients had only one outpatient visit on the same date and TC7 patients were not in the hospital at all on the same date. In TC4, after testing positive for acid-fast bacilli (AFB) by sputum culture, patient RGM-118 was in our hospital for the first time on the same date as RGM-162, who was culture-negative. In TC5, a similar situation was observed between RGM-44 (positive) and RGM-96 (negative) (Fig. S2). In contrast, RGM-118 and RGM-239 in TC4, RGM-19, RGM-44, and RGM-96 in TC5, and RGM-158 and RGM-178 in TC6 were culture-positive before their first overlap in our hospital. Although TC5 consisted of both Japanese and United Kingdom-based patients, there was no evidence of contact among them (Fig. 3). Regarding the residences of the Japanese patients assigned to each lineage clade, patients in each cluster resided within a minimum of approximately 13 km (TC6) and a maximum of approximately 20 km (TC7) radius (Fig. S3). There was no specific cluster

FIG 2 Legend (Continued)

variable positions located in the core genome (4,124,515 bp of the *M. abscessus* subsp. *massiliense* JCM 15300 genome) was performed to calculate SNP distances among isolates. The maximum SNP distance among isolates consecutively obtained from the same individual was 25, and that between isolates assigned to transmissible clusters (TCs) in the present study was less than 31. (B) Clustering analysis for the identification of possible cross-transmission events between patients. Whole-genome alignment, as described for panel A, was used to estimate a maximum likelihood tree with 1,000 bootstrap replicates. The resulting phylogenetic tree and SNP thresholds (25 SNPs) were used to identify eight TCs using rPinecone. The United Kingdom isolates were assigned to four TCs (TC1, TC2, TC5, and TC8); TC1 and TC2 were consistent with previously reported clusters, and the remaining nonclustered isolates were assigned to TC5 and TC8. All United States isolates were assigned to TC3, and 12 isolates from Fukujiji Hospital were assigned to four clusters (TC4, TC5, TC6, and TC7). TC5 consisted of four isolates from Fukujiji Hospital and two isolates from the United Kingdom. The scale bar represents the number of substitutions per site. SNP, single nucleotide polymorphism; UK, United Kingdom; US, United States; TC, transmissible cluster; *M. massiliense*, *Mycobacterium abscessus* subsp. *massiliense*; JP, Japan.

related to the residences or water supply area. Only two patients were working at the time of sample collection, and workplace-related clusters were not identified. In addition, there were no specific clinical characteristics in each cluster, except for patients in TC4, who showed the same NC-NB radiological patterns and good treatment responses (Table 1).

DISCUSSION

In the present study, we aimed to clarify the possibility of cross-transmission of *M. abscessus* among non-CF patients. Using our analysis pipeline, WGS analysis of publicly available isolates from the United Kingdom and the United States showed that the maximum SNP distance within the same individuals was 25, consistent with a previous report (6). Using this SNP distance as a threshold for clonal expansions, we identified four clusters suspected to contain cross-transmissible isolates in our hospital. When investigating epidemiological linkages among our patients, we found that isolates in two clusters were suspected to have undergone direct or indirect transmission. Interestingly, TC5, which contained isolates from Fukujuji Hospital and the United Kingdom, had less than 21 SNP distances; the reason for this is unknown. Although we could not access the hospital or home/work environments, the present study revealed that clonal infections existed even in the non-CF population, and some cases were suspected to be cross-transmissible.

To investigate the possibility of *M. abscessus* cross-transmission events, Bryant et al. first estimated the SNP distance threshold for isolates using multiple isolates from individual patients (6). Subsequent studies applied the same threshold to their data sets to identify *M. abscessus* cross-transmission events (12, 13). However, the calculation of SNP distance based on the alignment of clinical isolates to a reference genome is highly dependent on the quality of the data set and the analysis pipeline. Therefore, we evaluated whether our analysis pipeline could investigate the possibility of cross-transmission using our data set containing both isolates obtained from two previous CF center outbreaks in the United Kingdom (6) and the United States (11) and isolates from non-CF patients in this study. Using rPinecone software, in which the SNP threshold was used to define sublineages (19), we thoroughly investigated clusters associated with possible cross-transmission events. Our analysis confirmed that the *M. abscessus* isolates from the United Kingdom and the United States had classifications similar to those of previously reported clusters (6, 11), and the 12 *M. abscessus* isolates from our hospital constituted clusters. Based on these findings, we clustered cross-transmissible isolates in CF patients using the present method and found that some isolates clustered similarly in non-CF patients using integrated SNP breakpoints.

We further investigated the epidemiological linkages of each cluster to determine whether *M. abscessus* non-CF patients who were clustered in the present study had direct or indirect contact with an infectious source. Previous reports suggested that hospitals might be transmission sites based on overlapping outpatient and inpatient dates among patients and network analysis results (6, 20). Considering that our hospital is a four-story medium-size hospital with 205 respiratory beds, direct or indirect contact between outpatients and inpatients or between inpatients on different floors could occur, and opportunities for cross-transmission between patients in clusters TC4 and TC5 within Fukujuji Hospital might have occurred, but these opportunities were limited between patients in clusters TC6 and TC7. In some cases, both patients already had positive sputum cultures when they were admitted to our hospital for the first time on the same date. Therefore, clarifying the transmission route based on only the contact conditions within the hospital was difficult. It is also difficult to identify a potential environmental source of transmission (12). *M. abscessus* exists in domestic water after sterilization and can be transmitted through aerosols in the bathroom (21, 22). However, relationships between specific clusters and patients' residences, water supply areas, or workplaces were not observed.

Interestingly, TC5, which contained the WGS data of isolates from Fukujuji Hospital in Japan and Papworth Hospital in the United Kingdom, harbored less than 21 SNP distances. However, these United Kingdom isolates were not *M. abscessus* subsp. *massiliense* DCCs (14), which have been suspected to circulate among CF patients, and there was no opportunity for close contact between Japanese and United Kingdom patients. This might suggest

the existence of successful *M. abscessus* clones worldwide; however, the global transmission of *M. abscessus* among non-CF patients has seldom been reported (16). To validate the present data, WGS studies analyzing larger samples in larger non-CF populations are needed, and prospective studies investigating detailed contact opportunities are warranted to further clarify the possibility of cross-transmission.

Our study has several limitations. First, because the present study had a retrospective design and was conducted in a single facility, the number of investigated patients was small, and detailed contact conditions could not be confirmed. Second, although Bryant et al. determined the SNP distance threshold using multiple isolates from individual CF patients (6), SNP distances between multiple isolates from an individual patient were investigated in only one patient in Fukujiji Hospital. Therefore, it was not sufficiently clear whether the threshold used in our pipeline is an appropriate threshold for non-CF patients. Considering that the genomic diversity of *M. abscessus* isolates from non-CF patients was greater than that of isolates from CF patients (23), the threshold used in our pipeline may be restricted to CF patients. To determine the appropriate threshold for non-CF patients, the SNP distances between multiple isolates from individual non-CF patients should be investigated. Third, WGS was performed mainly on isolates that formed VNTR clusters. In a previous study conducted in 2014 (3), in which we did not implement WGS, we clarified the relationships between VNTR and clinical characteristics by referring to published data (9). Unexpectedly, we found several identical VNTR genotypes in the study. We presumed that isolates without matching VNTR profiles would also not be matched in WGS (24). Thus, this study focused on isolates that formed VNTR clusters; however, less than half of the examined isolates formed transmissible clusters according to SNP analysis, suggesting that the association between VNTR and WGS clusters was not strong. If WGS was performed on all isolates, more transmissible clusters might be identified, and accordingly, direct WGS analysis should be performed in similar studies. Fourth, analysis of core genomic regions alone might not be sufficient. Recently, possible contributions of the accessory genome, regions acquired by horizontal gene transfer, have been indicated (25, 26). We therefore consider that future research, including analyses of accessory genome regions, is warranted. Fifth, we did not obtain and examine isolates from nosocomial and environmental sources that could be potential sources of infection.

In conclusion, closely related isolates were detected in different patients, even in areas with few CF patients, suggesting the possibility of *M. abscessus* cross-transmission among non-CF patients. However, the transmission route remains unclear, and further studies are needed.

MATERIALS AND METHODS

Study patients and microbiological data collection. This study analyzed 104 *M. abscessus* isolates stored at Fukujiji Hospital, Japan Anti-Tuberculosis Association, which is a specialized facility for respiratory diseases (205 exclusive beds) located in northwestern Tokyo, Japan, between September 2004 and December 2014. Basically, the first available positive culture from each patient was stored. Of the 104 *M. abscessus* isolates, 57 isolates were identified as subsp. *abscessus*, 45 as subsp. *massiliense*, and 2 as subsp. *bollettii* using multiplex PCR and *rpob* gene sequencing as previously described (27, 28). The methods for the MIC, colony morphology evaluation, and VNTR genotyping are described in the supplemental material (3, 9, 29). We performed WGS on isolates that formed VNTR clusters.

Whole-genome sequencing. According to a previous report, *M. abscessus* DNA was extracted (30). A WGS library was constructed using a QIAseq FX DNA library kit (Qiagen, Hilden, Germany). Sequencing was performed on an Illumina NextSeq 550 platform using the NextSeq reagent kit (300 cycles) with 150-mer paired-end short reads. Raw read data were deposited in the DNA Data Bank of Japan (DDBJ) under BioProject accession number [DRA012694](https://www.ncbi.nlm.nih.gov/bioproject/12694).

Phylogenetic analysis and SNP analysis. Phylogenetic analysis was performed using the core genome, and the detailed methods are described in the supplemental material (31–33). To systematically explore cross-transmission events of *M. abscessus* between patients, we made the following assumption: the SNP distance between clinical isolates from different patients is smaller than the maximum SNP distance between clinical isolates that are consecutively isolated from the same individual. In these cases, we suspected the possibility of cross-transmission via people and the environment and performed epidemiological linkage investigations. To test this assumption, we combined our data set with publicly available WGS data on 109 *M. abscessus* subsp. *massiliense* clinical isolates that were consecutively isolated from patients involved in previous nosocomial outbreaks in the United Kingdom (6) and the United States (11). Accordingly, *M. abscessus* subsp. *massiliense* clinical isolates were especially investigated in the present study. Additional raw read data were obtained from the

Sequence Read Archive (SRA) and *de novo* assembled using the Shovill pipeline. To assess the assembly quality of the 109 isolates, we calculated the genome fraction (%) of each isolate, which is the total number of bases aligned with the reference genome (*M. abscessus* subsp. *massiliense* JCM 15300), using QUAST software (34). We included only isolates with genome fractions greater than 85% in the downstream SNP analyses. The whole-genome alignment of 132 *M. abscessus* subsp. *massiliense* clinical isolates was performed as noted in the supplemental material, and these data were used to calculate the SNP distances among clinical isolates using snp-dists (<https://github.com/tseemann/snp-dists>). To identify possible cross-transmission events, we used rPinecone software to define sublineages from clonal expansions (19), in which the maximum SNP distance among isolates from the same individuals (25 SNPs; Fig. 2A) and a phylogenetic tree estimated by RAXML software were used as input parameters (33).

Clinical and epidemiological analyses. To analyze the clinical characteristics and epidemiologic linkages among patients, we collected data on sex, age, past medical histories, radiological findings, patient residences, work histories, dates of outpatient visits (including visits for outpatient examinations such as blood tests, radiographic examinations, and physiological function tests), and admission periods and wards from the first visit to Fukujiji Hospital through the end of the study. We considered that opportunities for nosocomial cross-transmission could occur if a patient who once had a positive sputum acid-fast bacilli (AFB) culture shared the same space at the same time with another patient in the ward or at the outpatient clinic. We counted those days and investigated the possibility of contact. Radiological findings based on computed tomography were classified into the following disease patterns: fibrocavitary type (FC type), cavitary nodular bronchiectatic type (C-NB type), noncavitary nodular bronchiectatic type (NC-NB type), and unclassified type (35). Regarding treatment outcomes, we referred to the Nontuberculous Mycobacteria Network European Trial Group (NTM-NET) consensus statement (36). We defined “treatment success” as more than three consecutive negative cultures collected at least 4 weeks apart and maintenance of negative culture conversion until the end of the study, which was similar to the microbiological cure criteria (36).

The study protocol was approved by the Fukujiji Hospital Institutional Review Board (protocol number 21024), and informed consent was waived because of the retrospective nature of the analysis.

Data availability. The raw read data of whole-genome sequencing were deposited in the DNA Data Bank of Japan (DDBJ) under BioProject accession number [DRA012694](https://www.ncbi.nlm.nih.gov/bioproject/5412694).

SUPPLEMENTAL MATERIAL

Supplemental material is available online only.

SUPPLEMENTAL FILE 1, PDF file, 0.9 MB.

ACKNOWLEDGMENTS

We are grateful to Tetsuya Hayashi for allowing us to use their original program for the present analysis and all the hospital staff who supported this research.

This work was supported by the Japan Agency for Medical Research and Development (19fk0108043h0403 to S.M., JP22fk0108129 to K.M.).

Author contributions: K.F. and M.Y. were responsible for data collection, statistical analysis, data analysis, and writing of the manuscript. Y.M., A.A., and K.F. contributed to data collection. S.M. contributed to the conceptualization, interpretation of the results, manuscript writing, and editing. Y.T., M.A., and K.O. contributed to the interpretation of the results. K.M. designed and conducted the study and assisted in writing the manuscript. All other authors critically read and commented on the manuscript.

We declare that we have no conflicts of interest.

REFERENCES

- Prevots DR, Marras TK. 2015. Epidemiology of human pulmonary infection with nontuberculous mycobacteria: a review. *Clin Chest Med* 36:13–34. <https://doi.org/10.1016/j.ccm.2014.10.002>.
- Koh WJ, Chang B, Jeong BH, Jeon K, Kim SY, Lee NY, Ki CS, Kwon OJ. 2013. Increasing recovery of nontuberculous mycobacteria from respiratory specimens over a 10-year period in a tertiary referral hospital in South Korea. *Tuberc Respir Dis* 75:199–204. <https://doi.org/10.4046/trd.2013.75.5.199>.
- Morimoto K, Nakagawa T, Asami T, Morino E, Fujiwara H, Hase I, Tsujimoto Y, Izumi K, Hayashi Y, Matsuda S, Murase Y, Yano R, Takasaki J, Betsuyaku T, Aono A, Goto H, Nishimura T, Sasaki Y, Hoshino Y, Kurashima A, Ato M, Ogawa K, Hasegawa N, Mitarai S. 2018. Clinico-microbiological analysis of 121 patients with pulmonary Mycobacteroides abscessus complex disease in Japan - An NTM-JRC study with RIT. *Respir Med* 145:14–20. <https://doi.org/10.1016/j.rmed.2018.10.012>.
- Pasipanodya JG, Ogbonna D, Ferro BE, Magombedze G, Srivastava S, Deshpande D, Gumbo T. 2017. Systematic review and meta-analyses of the effect of chemotherapy on pulmonary Mycobacterium abscessus outcomes and disease recurrence. *Antimicrob Agents Chemother* 61:e01206-17. <https://doi.org/10.1128/AAC.01206-17>.
- Kwak N, Dalcolmo MP, Daley CL, Eather G, Gayoso R, Hasegawa N, Jhun BW, Koh WJ, Namkoong H, Park J, Thomson R, van Ingen J, Zweijpenning SMH, Yim JJ. 2019. Mycobacterium abscessus pulmonary disease: individual patient data meta-analysis. *Eur Respir J* 54:1801991. <https://doi.org/10.1183/13993003.01991-2018>.
- Bryant JM, Grogono DM, Greaves D, Foweraker J, Roddick I, Inns T, Reacher M, Haworth CS, Curran MD, Harris SR, Peacock SJ, Parkhill J, Floto RA. 2013. Whole-genome sequencing to identify transmission of Mycobacterium abscessus between patients with cystic fibrosis: a retrospective cohort study. *Lancet* 381: 1551–1560. [https://doi.org/10.1016/S0140-6736\(13\)60632-7](https://doi.org/10.1016/S0140-6736(13)60632-7).
- Aitken ML, Limaye A, Pottinger P, Whimbey E, Goss CH, Tonelli MR, Cangelosi GA, Dirac MA, Olivier KN, Brown-Elliott BA, McNulty S, Wallace RJ Jr. 2012. Respiratory outbreak of Mycobacterium abscessus subspecies massiliense in a lung transplant and cystic fibrosis center. *Am J Respir Crit Care Med* 185: 231–232. <https://doi.org/10.1164/ajrccm.185.2.231>.

8. Davidson RM, Hasan NA, Epperson LE, Benoit JB, Kammlade SM, Levin AR, Calado de Moura V, Hunkins J, Weakly N, Beagle S, Sagel SD, Martiniano SL, Salfinger M, Daley CL, Nick JA, Strong M. 2021. Population genomics of *Mycobacterium abscessus* from U.S. cystic fibrosis care centers. *Ann Am Thorac Soc* 18:1960–1969. <https://doi.org/10.1513/AnnalsATS.202009-1214OC>.
9. Shin SJ, Choi GE, Cho SN, Woo SY, Jeong BH, Jeon K, Koh WJ. 2013. Mycobacterial genotypes are associated with clinical manifestation and progression of lung disease caused by *Mycobacterium abscessus* and *Mycobacterium massiliense*. *Clin Infect Dis* 57:32–39. <https://doi.org/10.1093/cid/cit172>.
10. Johnston DI, Chisty Z, Gross JE, Park SY. 2016. Investigation of *Mycobacterium abscessus* outbreak among cystic fibrosis patients, Hawaii 2012. *J Hosp Infect* 94:198–200. <https://doi.org/10.1016/j.jhin.2016.04.015>.
11. Tettelin H, Davidson RM, Agrawal S, Aitken ML, Shallom S, Hasan NA, Strong M, de Moura VC, De Groot MA, Duarte RS, Hine E, Parankush S, Su Q, Daugherty SC, Fraser CM, Brown-Elliott BA, Wallace RJ Jr, Holland SM, Sampaio EP, Olivier KN, Jackson M, Zelazny AM. 2014. High-level relatedness among *Mycobacterium abscessus* subsp. *massiliense* strains from widely separated outbreaks. *Emerg Infect Dis* 20:364–371. <https://doi.org/10.3201/eid2003.131106>.
12. Harris KA, Underwood A, Kenna DT, Brooks A, Kavaliunaitė E, Kapatai G, Tewolde R, Aurora P, Dixon G. 2015. Whole-genome sequencing and epidemiological analysis do not provide evidence for cross-transmission of *Mycobacterium abscessus* in a cohort of pediatric cystic fibrosis patients. *Clin Infect Dis* 60:1007–1016. <https://doi.org/10.1093/cid/ciu967>.
13. Lipworth S, Hough N, Weston N, Muller-Pebody B, Phin N, Myers R, Chapman S, Flight W, Alexander E, Smith EG, Robinson E, Peto TEA, Crook DW, Walker AS, Hopkins S, Eyre DW, Walker TM. 2021. Epidemiology of *Mycobacterium abscessus* in England: an observational study. *Lancet Microbe* 2:e498–e507. [https://doi.org/10.1016/S2666-5247\(21\)00128-2](https://doi.org/10.1016/S2666-5247(21)00128-2).
14. Bryant JM, Grogono DM, Rodriguez-Rincon D, Everall I, Brown KP, Moreno P, Verma D, Hill E, Drijkoningen J, Gilligan P, Esther CR, Noone PG, Giddings O, Bell SC, Thomson R, Wainwright CE, Coulter C, Pandey S, Wood ME, Stockwell RE, Ramsay KA, Sherrard LJ, Kidd TJ, Jabbar N, Johnson GR, Knibbs LD, Morawska L, Sly PD, Jones A, Bilton D, Laurensen I, Ruddy M, Bourke S, Bowler IC, Chapman SJ, Clayton A, Cullen M, Daniels T, Dempsey O, Denton M, Desai M, Drew RJ, Edenborough F, Evans J, Folb J, Humphrey H, Isalska B, Jensen-Fangel S, Jönsson B, Jones AM, et al. 2016. Emergence and spread of a human-transmissible multidrug-resistant nontuberculous mycobacterium. *Science* 354:751–757. <https://doi.org/10.1126/science.aaf8156>.
15. Chew KL, Octavia S, Jureen R, Ng OT, Marimuthu K, Lin RTP, Teo JWP. 2021. Molecular epidemiology and phylogenomic analysis of *Mycobacterium abscessus* clinical isolates in an Asian population. *Microb Genom* 7:000708. <https://doi.org/10.1099/mgen.0.000708>.
16. Davidson RM, Benoit JB, Kammlade SM, Hasan NA, Epperson LE, Smith T, Vasireddy S, Brown-Elliott BA, Nick JA, Olivier KN, Zelazny AM, Daley CL, Strong M, Wallace RJ Jr. 2021. Genomic characterization of sporadic isolates of the dominant clone of *Mycobacterium abscessus* subspecies *massiliense*. *Sci Rep* 11:15336. <https://doi.org/10.1038/s41598-021-94789-y>.
17. Harris KA, Kenna DT, Blauwendraat C, Hartley JC, Turton JF, Aurora P, Dixon GL. 2012. Molecular fingerprinting of *Mycobacterium abscessus* strains in a cohort of pediatric cystic fibrosis patients. *J Clin Microbiol* 50:1758–1761. <https://doi.org/10.1128/JCM.00155-12>.
18. Daley CL, Iaccarino JM, Lange C, Cambau E, Wallace RJ, Andrejak C, Böttger EC, Brozek J, Griffith DE, Guglielmetti L, Huitt GA, Knight SL, Leitman P, Marras TK, Olivier KN, Santin M, Stout JE, Tortoli E, van Ingen J, Wagner D, Winthrop KL. 2020. Treatment of nontuberculous mycobacterial pulmonary disease: an official ATS/ERS/ESCMID/IDSA clinical practice guideline. *Clin Infect Dis* 71:905–913. <https://doi.org/10.1093/cid/ciaa1125>.
19. Wailan AM, Coll F, Heinz E, Tonkin-Hill G, Corander J, Feasey NA, Thomson NR. 2019. rPinecone: define sub-lineages of a clonal expansion via a phylogenetic tree. *Microb Genom* 5:e000264. <https://doi.org/10.1099/mgen.0.000264>.
20. Gross JE, Caceres S, Poch K, Hasan NA, Jia F, Epperson LE, Lipner E, Vang C, Honda JR, Strand M, Calado Nogueira de Moura V, Daley CL, Strong M, Davidson RM, Nick JA. 2022. Investigating Nontuberculous mycobacteria transmission at the Colorado Adult Cystic Fibrosis Program. *Am J Respir Crit Care Med* <https://doi.org/10.1164/rccm.202108-1911OC>.
21. Thomson R, Tolson C, Carter R, Coulter C, Huygens F, Hargreaves M. 2013. Isolation of nontuberculous mycobacteria (NTM) from household water and shower aerosols in patients with pulmonary disease caused by NTM. *J Clin Microbiol* 51:3006–3011. <https://doi.org/10.1128/JCM.00899-13>.
22. Thomson R, Tolson C, Sidjabat H, Huygens F, Hargreaves M. 2013. *Mycobacterium abscessus* isolated from municipal water: a potential source of human infection. *BMC Infect Dis* 13:241. <https://doi.org/10.1186/1471-2334-13-241>.
23. Redondo N, Mok S, Montgomery L, Flanagan PR, McNamara E, Smyth EG, O'Sullivan N, Schaffer K, Rogers TR, Fitzgibbon MM. 2020. Genomic analysis of *Mycobacterium abscessus* complex isolates collected in Ireland between 2006 and 2017. *J Clin Microbiol* 58:e00295-20. <https://doi.org/10.1128/JCM.00295-20>.
24. Trovato A, Baldan R, Costa D, Simonetti TM, Cirillo DM, Tortoli E. 2017. Molecular typing of *Mycobacterium abscessus* isolated from cystic fibrosis patients. *Int J Mycobacteriol* 6:138–141. https://doi.org/10.4103/ijmy.ijmy_33_17.
25. Davidson RM, Hasan NA, Reynolds PR, Totten S, Garcia B, Levin A, Ramamoorthy P, Heifets L, Daley CL, Strong M. 2014. Genome sequencing of *Mycobacterium abscessus* isolates from patients in the United States and comparisons to globally diverse clinical strains. *J Clin Microbiol* 52:3573–3582. <https://doi.org/10.1128/JCM.01144-14>.
26. Davidson RM. 2018. A closer look at the genomic variation of geographically diverse *Mycobacterium abscessus* clones that cause human infection and disease. *Front Microbiol* 9:2988. <https://doi.org/10.3389/fmicb.2018.02988>.
27. Nakanaga K, Sekizuka T, Fukano H, Sakakibara Y, Takeuchi F, Wada S, Ishii N, Makino M, Kuroda M, Hoshino Y. 2014. Discrimination of *Mycobacterium abscessus* subsp. *massiliense* from *Mycobacterium abscessus* subsp. *abscessus* in clinical isolates by multiplex PCR. *J Clin Microbiol* 52:251–259. <https://doi.org/10.1128/JCM.01327-13>.
28. Adékambi T, Berger P, Raoult D, Drancourt M. 2006. rpoB gene sequence-based characterization of emerging non-tuberculous mycobacteria with descriptions of *Mycobacterium bolletii* sp. nov., *Mycobacterium phocaicum* sp. nov. and *Mycobacterium aubagnense* sp. nov. *Int J Syst Evol Microbiol* 56:133–143. <https://doi.org/10.1099/ijso.0.63969-0>.
29. Brown-Elliott BA, Woods GL. 2019. Antimycobacterial susceptibility testing of nontuberculous mycobacteria. *J Clin Microbiol* 57:e00834-19. <https://doi.org/10.1128/JCM.00834-19>.
30. Takeda K, Chikamatsu K, Igarashi Y, Morishige Y, Murase Y, Aono A, Yamada H, Takaki A, Mitarai S. 2018. Six species of nontuberculous mycobacteria carry non-identical 16S rRNA gene copies. *J Microbiol Methods* 155:34–36. <https://doi.org/10.1016/j.mimet.2018.08.012>.
31. Kurtz S, Phillippy A, Delcher AL, Smoot M, Shumway M, Antonescu C, Salzberg SL. 2004. Versatile and open software for comparing large genomes. *Genome Biol* 5:R12. <https://doi.org/10.1186/gb-2004-5-2-r12>.
32. Croucher NJ, Page AJ, Connor TR, Delaney AJ, Keane JA, Bentley SD, Parkhill J, Harris SR. 2015. Rapid phylogenetic analysis of large samples of recombinant bacterial whole genome sequences using Gubbins. *Nucleic Acids Res* 43:e15. <https://doi.org/10.1093/nar/gku1196>.
33. Stamatakis A. 2006. RAxML-VI-HPC: maximum likelihood-based phylogenetic analyses with thousands of taxa and mixed models. *Bioinformatics* 22:2688–2690. <https://doi.org/10.1093/bioinformatics/btl446>.
34. Gurevich A, Saveliev V, Vyahhi N, Tesler G. 2013. QUAST: quality assessment tool for genome assemblies. *Bioinformatics* 29:1072–1075. <https://doi.org/10.1093/bioinformatics/btt086>.
35. Furuuchi K, Fujiwara K, Uesgi F, Shimoda M, Seto S, Tanaka Y, Yoshiyama T, Yoshimori K, Kurashima A, Ohta K, Morimoto K. 2020. Posttreatment lymphopenia is associated with an increased risk of redeveloping nontuberculous lung disease in patients with *Mycobacterium avium* complex lung disease. *Clin Infect Dis* 5:e152–e157. <https://doi.org/10.1093/cid/ciaa729>.
36. van Ingen J, Aksamit T, Andrejak C, Böttger EC, Cambau E, Daley CL, Griffith DE, Guglielmetti L, Holland SM, Huitt GA, Koh WJ, Lange C, Leitman P, Marras TK, Morimoto K, Olivier KN, Santin M, Stout JE, Thomson R, Tortoli E, Wallace RJ Jr, Winthrop KL, Wagner D. 2018. Treatment outcome definitions in nontuberculous mycobacterial pulmonary disease: an NTM-NET consensus statement. *Eur Respir J* 51:1800170. <https://doi.org/10.1183/13993003.00170-2018>.